
The structure of the yeast ribosomal RNA genes. 4. Complete sequence of the 25 S rRNA gene from *Saccharomyces cerevisiae*

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ABSTRACT

The complete nucleotide sequence of the 25 S rRNA gene from one rDNA repeating unit of *Saccharomyces cerevisiae* has been determined. The corresponding 25 S rRNA molecule contains 3392 nucleotides, and has an estimated relative molecular mass (M_r , Na-salt) of 1.17×10^6 . Striking sequence homology is observed with known 5'- and 3'-end terminal segments of L-rRNA from other eukaryotes. Possible models of interaction with 5.8 S rRNA are discussed.

INTRODUCTION

Determination of the primary structure of rRNA molecules and the respective genes is important for understanding ribosome structure, function, biogenesis and evolution. It is established that in *Saccharomyces cerevisiae* the structural genes for 5 S, 18 S, 5.8 S and 25 S rRNA are organized in one rDNA repeating unit (about 9.1 kb), present in 100 to 120 copies per haploid genome (1). The structure of the yeast rDNA repeating unit is studied in details and all four rRNA genes are mapped within seven Eco RI restriction fragments designated A to G according to their length (2-5). A large part of the rDNA repeating unit has been sequenced, including the 5 S (6,7), 18 S (8) and 5.8 S (2,9) rRNA genes. Further, the 5'- and 3'-ends of the 25 S rRNA gene are now precisely mapped within the Eco RI fragments A and E, respectively (10, 24).

In the present work we report the complete sequence of the 25 S rRNA gene from one rDNA repeating unit of *Saccharomyces cerevisiae*.

MATERIALS AND METHODS

The recombinant plasmids pY1rA9 and pY1rB3, containing the rDNA Eco RI fragments A, E and F, are used (3). Restriction endonucleases are prepared in our laboratories by standard procedures or obtained as a gift from Dr. A.Yanulaitis. The [32 P]-orthophosphate is a product of The Radiochemical

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Centre, Amersham, U.K. The [γ ³²P]ATP (about 1000 Ci/mmol) is prepared by the method of Glynn and Chappel (11). Plasmid DNA is isolated according to Tanaka and Weissblum (12). Restriction rDNA fragments are purified by preparative electrophoresis in 4 or 6 % acrylamide gels. End labelling is carried out with T₄ polynucleotide kinase (Boehringer-Mannheim) as described by Maxam and Gilbert (13,14). The end-labelled fragments are subjected to strand separation (15) or to secondary restriction endonuclease cleavage. Sequencing of DNA is carried out according to Maxam and Gilbert (13,14).

RESULTS AND DISCUSSION

Sequencing strategy. The entire 25 S rRNA gene is contained in the Eco RI fragments A, F and E (Figure 1). These fragments have estimated lengths of about 2.85, 0.36 and 0.59 kb, respectively. Therefore, first of all, a strategy for the sequencing of fragment Eco RI-A had to be devised. Analysis of the cleavage patterns of fragment Eco RI-A with different restriction endonucleases revealed that digestion with Msp I and Sau 3A provides two sets of 11 and 8 subfragments, respectively, which appear to be convenient for sequencing (see Fig.1). Most of the sequences in fragments Eco RI-A, as well as Eco RI-F and Eco RI-E, were determined in both strands and independently confirmed by the use of overlapping restriction fragments. In some cases the orientation of sequenced rDNA chains was defined by hybridization with 25 S rRNA.

Sequence results. The complete sequence of 25 S rRNA (deduced from the

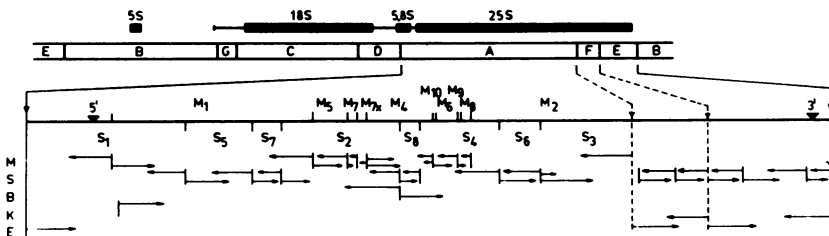


FIGURE 1. Above - Endonuclease Eco RI restriction map of the *S.cerevisiae* rDNA repeating unit. Arrangement of the Eco RI fragments A to G and the location of rRNA genes are indicated.

Below - Expanded map of the 25 S rRNA gene. Restriction sites are indicated by vertical lines: M, Msp I; S, Sau 3A; B, Bgl II; K, Kpn I; E, Eco RI. The subfragments obtained upon digestion of fragment Eco RI-A with endonucleases Msp I [M] and Sau 3A [S] are numbered by size. The sequenced strands are shown as horizontal arrows pointing from the labelled 5'-end.

corresponding rDNA) is shown in Figure 2. The identification of the 5'- and 3'-terminal nucleotides is based on our previous results (10). The total length of 25 S rRNA is 3392 nucleotides, yielding a relative molecular mass (M_r , Na-salt) of 1.17×10^6 . This figure is smaller than the ones derived from physicochemical measurements (16,17), but it is in close agreement with results obtained by R-loop and hybridization analyses (4).

The use of the chemical method of Maxam and Gilbert (14) permitted confident reading of sequences extending 200-250 nucleotides from the labelled end. However, the anomalous behaviour of nucleotide 1642 is noteworthy. This nucleotide behaves simultaneously as C and G (data not shown). The absence of a purine band and analysis of the complementary rDNA strand identify this nucleotide as a C. Most likely, this cytidine residue is modified by *E.coli* enzymes in such a way that it is split by the G-specific dimethyl sulfate reaction. Such modification of a cytidine residue in plasmid DNA has not been encountered previously (14) and could be of more general interest.

Homology with other eukaryotic L-rRNA. The 25 S rRNA of *S.cerevisiae* is the first major eukaryotic L-rRNA with a known complete sequence. Previous indirect evidence indicates that strongly conserved regions of homology exist in eukaryotic L-rRNA (18-20). Comparison with published sequence data on the 5'-end of *Xenopus laevis* (21) and the 3'-end of *Neurospora crassa* (22) L-rRNA, reveals that such highly conserved sequences may be present at both ends of eukaryotic L-rRNA molecules. The sequence encompassing nucleotides 2-113 of *S.cerevisiae* L-rRNA shows 81 % homology with the respective sequence of *X.laevis*, while the 3'-end segment (nucleotides 3290-3392) is 74 % homologous with the respective sequence in *Neurospora crassa*.

Interaction with 5.8 S rRNA. It is well known that in the eukaryote ribosome 5.8 S rRNA is hydrogen-bonded to L-rRNA (1,18). Recently, it was reported that 5.8 S rRNA interacts with the 3'-terminal fragment of *Neurospora crassa* L-rRNA and a model of possible complementary sequences was proposed (22). Our preliminary computer search, including the full length of *S.cerevisiae* 25 S rRNA, failed to reveal uninterrupted complementary sequences between L-rRNA and 5.8 S rRNA longer than eight nucleotides. Further, linear alignment of both sequences revealed numerous regions in 25 S rRNA with 35 to 38 % complementarity to 5.8 S rRNA. The best fit is observed for the interaction between nucleotides 5-20 of L-rRNA and the 3'-end half of 5.8 S rRNA. Also, another region of high complementarity involves nucleotides 3265-3333 of L-rRNA and the 5'-end half of 5.8 S rRNA. These results appear to favor a model in which 5.8 S rRNA interacts simultaneously with segments

pGUUUGACCUC¹AAUCAGGUAGGAGUACCCGCGUAAUCUUAAGCAUUAAGCGAGGAAAGAAACCC²AACCGGAUUGCCUUAUUAACGGCGAGUGAAG³ 100
 CGGCAAAAGCCUC⁴AAAUUUGAAACUCUGGUAC⁵CUUCGGUGCCAGUUGUA⁶AAUUGGAGAGGGCAACU⁷UUGGGGCGGUCCUUGUCUUAUGU⁸UCCUUGGAACA⁹ 200
 GGACGUC¹⁰AUAGAGGGUGAGCAUCCCGUGUGGGGAGG¹¹CGGUUCUUGUA¹²AAAGUGCCU¹³UCGAGAGUCGAGUUGU¹⁴UUGGGAUUGCAGCUCUUAAGUGGG¹⁵ 300
 UGGUAAAUUCC¹⁶AUCUAAAAGC¹⁷UAAAUAUUGGGCAGAGACC¹⁸CAUGCGCAAC¹⁹AAAGUACAGUG²⁰AAGUAGAAAGAAAGAAACU²¹UUGAAAAGAGAGUGAAAAAGU²² 400
 ACGUAAAUUGUUGAAGGGAAGGCAU²³UUGAUCAGAC²⁴AUGGGUJUUGUG²⁵CCUCUGCCU²⁶CGUAGGAAAGGGAUUG²⁷CGAAUUCACUGGGCCAGCA²⁸ 500
 UCAGUUUGGGCAGGAUAAAUC²⁹CAUAGGAUUGAUCU³⁰GGCCUGGUAAGUAUUAUAGCCU³¹UGGGAAUACUGCCAGC³²UGGGACUAGGACUGGCGACGU³³ 600
 AAGUCAAGG³⁴AUGCGUCAUAAUGGUUAU³⁵AUGCCCGCCGCU³⁶UUGAAACACGGACC³⁷AAGGAGUCU³⁸AACGUCU³⁹AUAGCGAGU⁴⁰UUGGGUGUA⁴¹AAACCCCAUACG⁴² 700
 CGUAUUGAAAGUGA⁴³CGUAAGGUUGGGCCUC⁴⁴CGAAGAGGUG⁴⁵CACA⁴⁶AUCCACCGAUC⁴⁷UGU⁴⁸UUCGG⁴⁹AUGGAUUGA⁵⁰UAGAAAGCAU⁵¹AGCUGUUGGG⁵²A 800
 CCCGAAAG⁵³AUGGUGAACU⁵⁴AUGCCUGAAUAGGGUGAAGCC⁵⁵AGAGGAGG⁵⁶UAGCCU⁵⁷UGGGGAAACU⁵⁸UGGUGGAGGC⁵⁹UCUGA⁶⁰GGGU⁶¹UCAGCGG⁶²U⁶³AAUCCG⁶⁴AUCCGUGAAUUGG⁶⁵ 900
 GUUAGGGGCC⁶⁶AAAGACU⁶⁷AAUCGAAACC⁶⁸AUCU⁶⁹AGUAGGUGGUUCCUGCCGAAGU⁷⁰UCCUC⁷¹AGGAUAGCAGAAAGCUC⁷²GU⁷³UUAUUG⁷⁴AGGUAAGG⁷⁵UAAAGCG⁷⁶ 1000
 AAUGAUUAG⁷⁷AGGUUCCGGGCGCAAAUGA⁷⁸CCUUGACC⁷⁹UAUUC⁸⁰AAACU⁸¹UUA⁸²AAAUUAGU⁸³A⁸⁴AGAAGUCCU⁸⁵U⁸⁶GUUACU⁸⁷UAAU⁸⁸UUGAACCGUGG⁸⁹ACAUIUUGA⁹⁰AUG⁹¹ 1100
 AAGAGCUUU⁹²UAGUGGGCCAU⁹³UUUUGGUAAGC⁹⁴AGAACUG⁹⁵CGAGUGGGGAU⁹⁶GAACCGAACCAGAGAGU⁹⁷UAAGGUGCGGGAU⁹⁸AC⁹⁹ACGCU¹⁰⁰CAUCAGACACCAC¹⁰¹ 1200
 AAAAGGUGU¹⁰²AGUUC¹⁰³AUCUAGAC¹⁰⁴AGCCGGAC¹⁰⁵GGUGGCC¹⁰⁶AUGGAAGUCGGAUCCGCU¹⁰⁷AAAGGAGUGU¹⁰⁸AAACAACU¹⁰⁹CCGGCGGAU¹¹⁰GAACUAGCCCCUGA¹¹¹ 1300
 AAUUGAUGGC¹¹²CCU¹¹³CAAGCGUGU¹¹⁴UACCU¹¹⁵UAU¹¹⁶ACU¹¹⁷ACCUC¹¹⁸AGGGUUG¹¹⁹AU¹²⁰UAGUAGCCCU¹²¹AGCAGUAGG¹²²CGGGUGG¹²³AGU¹²⁴CAAGGCCUAG¹²⁵A 1400
 CCGUAGAGU¹²⁶CGGUCGAC¹²⁷CGCCU¹²⁸CUAGU¹²⁹GCAGAU¹³⁰UUGGUGUAGUAGCA¹³¹AAUAU¹³²U¹³³CA¹³⁴AAUAGAACU¹³⁵U¹³⁶UAGAGACU¹³⁷U¹³⁸UAGGUGGGA¹³⁹AAAGGUUCCACGU¹⁴⁰ 1500
 CAACAGCAGU¹⁴¹UGACGUGG¹⁴²U¹⁴³UAGUCGAU¹⁴⁴CCUAAGAGAU¹⁴⁵GGGAAGCUC¹⁴⁶CGU¹⁴⁷UUC¹⁴⁸UUAAGG¹⁴⁹CCU¹⁵⁰GUUUA¹⁵¹UAGCAGGCCCA¹⁵²CAUCGAAAGGGAU¹⁵³CCGGU¹⁵⁴A 1600
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 UAUACCC¹⁶⁷CGGAU¹⁶⁸UUGGUUUAU¹⁶⁹CCGGAGAU¹⁷⁰UGGGU¹⁷¹CUUAU¹⁷²UGGCU¹⁷³UAGGAGAGGCCAC¹⁷⁴CCU¹⁷⁵UUGCUGGCUC¹⁷⁶CGGUGCCU¹⁷⁷UGACGGCC¹⁷⁸CGU¹⁷⁹AAAAU¹⁸⁰C 1800
 CACAGGAAGG¹⁸¹AAUAGUUUUC¹⁸²AUGCUAGGU¹⁸³CGUACUGAUAA¹⁸⁴CCCGCAGCAGGUCU¹⁸⁵CCAAGG¹⁸⁶U¹⁸⁷GAACAGCCU¹⁸⁸CU¹⁸⁹AGUUGUAGUAGAAU¹⁹⁰AUAGUAGAAUAGGGGAAG¹⁹¹ 1900

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FIGURE 2. Nucleotide sequence of 25 S rRNA of *Saccharomyces cerevisiae* derived from the sequence of the 25 S rRNA gene of one rDNA repeating unit. The restriction fragment Eco RI-A terminates at nucleotide 2533. The fragment Eco RI-F is from nucleotide 2534 to nucleotide 2897.

located at both ends of the L-rRNA molecule. Such a model is compatible with the available experimental data on 5.8 S rRNA:L-rRNA interactions (22,23), but further evidence is needed in order to clarify the interaction within the ribosome of these two rRNA molecules.

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