



Published in final edited form as:

*Am J Physiol Regul Integr Comp Physiol.* 2003 September ; 285(3): R701–R708. doi:10.1152/ajpregu.00009.2003.

## Hypertrophy changes the muscarinic receptor subtype mediating bladder contraction from M<sub>3</sub> toward M<sub>2</sub>

Alan S. Braverman<sup>1</sup> and Michael R. Ruggieri Sr.<sup>1,2</sup>

<sup>1</sup>Temple University School of Medicine, Department of Urology

<sup>2</sup>Temple University School of Medicine, Department of Pharmacology, Philadelphia, Pennsylvania 19140

### Abstract

Major pelvic ganglion electrocautery (MPGE) and spinal cord injury in the rat induce bladder hypertrophy and a change in muscarinic receptor subtypes mediating bladder contraction from predominantly M<sub>3</sub> to a combination of M<sub>2</sub> and M<sub>3</sub>. To determine whether this is a result of bladder hypertrophy or denervation, we studied the following groups: sham-operated controls, urinary diversion (DIV), MPGE together with urinary diversion (DIV-DEN), bilateral MPGE (DEN), bladder outlet obstruction (BOO), and MPG decentralization (MPG-DEC). The degree of bladder denervation was determined by the maximal carbachol response normalized to the response to electric field stimulation. Receptor subtype density was determined by immunoprecipitation. The affinity of subtype-selective muscarinic antagonists for inhibition of carbachol-induced contractions was used to determine the subtype-mediating contraction. DEN, MPG-DEC, and BOO bladders were hypertrophic whereas DIV bladders were atrophic compared with sham operated. Bladder contraction in sham-operated, DIV, and DIV-DEN was mediated by the M<sub>3</sub> receptor subtype, whereas the M<sub>2</sub> subtype participated in contraction in the DEN, MPG-DEC, and BOO groups. The hypertrophied bladders had an increase in total and M<sub>2</sub> receptor density while all experimental groups showed a reduction in M<sub>3</sub> receptor density. Thus bladder hypertrophy, independent from bladder denervation, causes a shift in the muscarinic receptor subtype mediating bladder contraction from M<sub>3</sub> toward M<sub>2</sub>.

### Keywords

denervation; outlet obstruction; urinary diversion

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Pharmacological data, based on the actions of subtype-selective antimuscarinic agents, can distinguish four subtypes of muscarinic acetylcholine receptors (M<sub>1</sub>–M<sub>4</sub>). Molecular techniques have identified five muscarinic receptor subtypes (M<sub>1</sub>–M<sub>5</sub>) arising from five separate genes (7, 8). Both M<sub>2</sub> and M<sub>3</sub> muscarinic receptor subtypes are found in most smooth muscles. The M<sub>2</sub> receptor preferentially couples to the inhibition of adenylyl cyclase through the G<sub>i</sub> family of proteins, while the M<sub>3</sub> receptor preferentially couples to IP<sub>3</sub> generation and calcium mobilization through the G<sub>q</sub> family of proteins (7, 8). Pertussis toxin (PTX), which ADP ribosylates and therefore inactivates the G<sub>i</sub> family of proteins, has no apparent effect on contraction (23). Even though the M<sub>2</sub> muscarinic receptor density is greater than the M<sub>3</sub> receptor density in bladder and other smooth muscles, the affinity of

subtype-selective muscarinic receptor antagonist drugs indicates that contraction is mediated by the M<sub>3</sub> receptor in most smooth muscles under normal conditions (7, 9).

A number of studies have shown that under certain conditions the M<sub>2</sub> receptor subtype can contribute to the contractile response. This includes selective alkylation of M<sub>3</sub> receptors in an environment of increased intracellular levels of cAMP in the rat urinary bladder (5, 16), guinea pig ileum (10), and trachea (29) or after alkylation without increasing intracellular cAMP levels in other tissues such as the guinea pig gallbladder (2) and colon (22). Other studies of smooth muscle contraction after experimentally induced pathologies, for example in a cat model of experimentally induced esophagitis (25), in the denervated rat bladder (4), and in a model of acute cholecystitis in the guinea pig gallbladder (2), also suggest that the M<sub>2</sub> receptor participates in mediation of contraction. In addition, in otherwise normal tissues, the M<sub>2</sub> receptor appears to mediate contraction after inhibition of the sarcoplasmic reticulum calcium ATPase, G<sub>q</sub>, phosphatidylinositol-specific phospholipase C, phosphatidylcholine-specific phospholipase C, or protein kinase C (PKC; Refs. 2, 25). Additional evidence for an M<sub>2</sub> receptor-mediated contractile pathway was demonstrated by the synergistic effects of M<sub>2</sub>- and M<sub>3</sub>-selective antagonists for inhibition of bladder contraction in normal bladders treated with thapsigargin and denervated bladders (6).

Our previous studies showed that both bilateral major pelvic ganglion electrocautery (DEN) and spinal cord injury (SCI) in the rat induce bladder hypertrophy and a change in muscarinic receptor subtype mediating bladder contraction from M<sub>3</sub> toward M<sub>2</sub> (1, 4). To determine whether this change is a result of bladder hypertrophy or denervation, additional experimental pathologies were studied. These include major pelvic ganglion decentralization (MPG-DEC), bladder outlet obstruction (BOO), urinary diversion (DIV), and urinary diversion with denervation (DIV-DEN).

## METHODS

### Materials

The following drugs or chemicals were obtained from the sources indicated: carbachol, methocramine, and *para*-fluoro-hexahydro-sila-diphenidol (*p*-F-HHSiD) were from Sigma Chemical (St. Louis, MO), and darifenacin was a generous gift from Pfizer (Sandwich, UK).

### Surgery

Rats (200–250 g female Sprague-Dawley rats from Ace Animals, Boyertown, PA) were anesthetized with 25 µg/kg buprenorphine and 2% isoflurane in oxygen, and a midline incision was made in the lower abdomen. The pelvic plexus was exposed. For bilateral denervation, both the left and right major pelvic ganglion were cauterized with a hand stitching pencil attached to a model SSE 2 solid-state electrosurgery device (Valleylab, Boulder, CO). For sham-operated animals, the plexus was exposed but left intact. For urinary diversions, both ureters were dissected free, cut, and sutured into the colon. For BOO, the urethra was exposed, a 21-gauge syringe needle was placed parallel to the urethra, a 3–0 silk suture was tied around both the needle and urethra, and the needle was then removed. For MPG-DEC, the nerve fibers entering the ganglion from the spinal cord were severed. The subcutaneous tissue, muscle, and skin were sutured. After surgery, urine was expressed with manual pressure on the lower abdomen twice daily for 3 days.

Immediately before bladder harvesting of the MPG-DEC group, they were tested to ensure that spinal stimulation was ineffective in inducing a bladder contraction while MPG stimulation caused an increase in bladder pressure. For this determination, the bladder was catheterized per urethra with PE-50 tubing connected to a pressure transducer. The bladder was emptied and filled manually until an intravesical pressure of 5 cm H<sub>2</sub>O was induced. A

unipolar pith electrode grounded to the abdomen was inserted into the spinal column between the L2 and L3 vertebral bodies and stimulated with a 2- to 5-s train of square wave pulses at 2 V, 30 Hz, 1-ms duration delivered by a Grass S-88 stimulator (Astro-Med, West Warwick, RI). The major pelvic ganglion was stimulated with bipolar electrodes separated by 4 mm with the same stimulation parameters. These stimulations consistently induced a marked bladder contraction in neurally intact animals. Any MPG-DEC animals that showed a bladder contraction to the spinal stimulation or did not show a bladder contraction to the MPG stimulation were not used.

### Muscle strips

Urinary bladders were removed from rats euthanized by decapitation. The urinary bladder body (tissue above the ureteral orifices) was dissected free of the serosa and surrounding fat. The bladder was divided in the midsagittal plane, then cut into longitudinal smooth muscle strips (~4 × 10 mm). The muscle strips were then suspended with 1 g of tension in tissue baths containing 15 ml of modified Tyrode solution (125 mM NaCl, 2.7 mM KCl, 0.4 mM NaH<sub>2</sub>PO<sub>4</sub>, 1.8 mM CaCl<sub>2</sub>, 0.5 mM MgCl<sub>2</sub>, 23.8 mM NaHCO<sub>3</sub>, and 5.6 mM glucose) and equilibrated with 95 O<sub>2</sub>-5% CO<sub>2</sub> at 37°C.

### Carbachol dose response

After equilibration to the bath solution for 30 min, bladder strips were incubated for 30 min in the presence or absence of antagonist. Dose-response curves were derived from the peak tension developed after cumulative addition of carbachol (10 nM to 300 μM final bath concentration). Only one concentration of antagonist was used for each muscle strip ( $n = 6-8$  strips per antagonist concentration). Dose ratios were determined based on the average of the responses of antagonist free strips. An EC<sub>50</sub> value was determined for each strip using a sigmoidal curve fit of the data (Origin, Originlab Northampton, MA). The EC<sub>50</sub> values determined in the presence of antagonist were used to generate Schild plots to calculate pA<sub>2</sub> values for each antagonist. If the slope of the Schild plot was not significantly different from unity (95% confidence interval), the slope of the Schild plot was constrained to unity to calculate the pK<sub>b</sub> value. To construct the Schild plot for methoctramine and p-F-HHSiD, doses of 0.3 and 3.0 μM were used. The Schild plot for 4-diphenacetoxy-*N*-methylpiperidine methiodide (4-DAMP) was done using 3.0, 10.0, and 30.0 nM. Because higher doses of darifenacin appeared unsurmountable and lower doses did not produce a significant shift in the concentration- response curve, a single dose of 30 nM darifenacin was used. The estimated pK<sub>b</sub> for darifenacin was calculated using the formula  $pK_b = - [\log(\text{darifenacin concentration}) - \log(\text{dose ratio} - 1)]$ .

### Immunoprecipitations

Immunoprecipitations were performed using antibodies as previously described. (4, 30). Protein concentration in the solubilized receptor preparation was determined by a dye binding assay (Bio-Rad). Muscarinic receptor density is reported as mean ± SE femtomoles per milligram protein in this solubilized receptor preparation. Total muscarinic receptor levels were determined by desalting over Sephadex G-50 minicolumns. At least four determinations were performed on two different pools of bladders for all groups.

### Statistical and data analysis

For Fig. 1, contractile force is presented as absolute millinewtons of force generated. Tension displayed in Fig. 2, *B* and *C*, is normalized to cross-sectional area defined as weight divided by length (which assumes a tissue density of 1). Statistical analysis of multiple group comparison was performed by ANOVA with a post hoc Newman-Keuls test or a Student's *t*-test where appropriate (GB-STAT, Dynamic Microsystems, Silver Spring, MD).

Statistically significant differences in the affinity values and departure from unity in the slopes derived from the Schild plots were determined using the 95% confidence intervals.

## RESULTS

### Hypertrophy

The time course of denervation-induced changes reveals no increase in bladder weight after 6 h but a statistically significant increase after 24 h, which continues to increase at 3 days and 3 wk (data not shown). Even though there is a large increase in bladder weight, no differences in the carbachol-induced maximal contraction are seen at any time point (Fig. 1). Because the rat bladder has no intramural ganglion cells (14), after denervation and degeneration of the nerve terminals in the bladder wall, no response to electric field stimulation (EFS) was expected. At 6 h postdenervation, the nerve terminals are apparently still intact because the muscle strips are able to contract to EFS (8 V, 30 Hz, 1 ms). A significant reduction in the EFS contractile response is seen at 24 h, the next time point measured. No further reduction is seen at time points after 24 h. The change in affinity of muscarinic receptor subtype-selective antagonists is not evident at 24 h postsurgery but occurs at 3 days postsurgery (data not shown). Consequently, the 3-day postsurgery time point was chosen for all subsequent studies.

Neither DIV nor DIV-DEN induces hypertrophy as evidenced by an increase in bladder weight-to-body weight ratio compared with sham-operated controls; however, DEN, BOO, and MPG-DEC induce significant hypertrophy by 3 days postsurgery. Interestingly, DIV bladders are significantly smaller than sham-operated or DIV-DEN bladders (Fig. 2A).

### EFS responses

To determine the effect of short-term hypertrophy on EFS contractile force, the contractile force normalized to cross-sectional area was determined for each group. Figure 2B shows that no differences in the EFS contraction to a submaximal stimulation of 8 V, 30 Hz, 1 ms is seen with DIV, BOO, or MPG-DEC compared with sham-operated control bladders. DIV-DEN and DEN bladders contract less to EFS than sham-operated controls. Unexpectedly, DIV-DEN bladders contract greater than DEN bladders to EFS, suggesting that the nerve terminals in these bladders are not completely degenerated at 3 days (Fig. 2B).

### Carbachol responses

The carbachol-induced maximal contraction (Fig. 2C) is not different between any groups except that the BOO group contracts significantly greater than every other group. As the nerves degenerate or are unable to induce a maximal contraction, the ratio of the carbachol maximum contractile response to the EFS contractile response will increase. This can be used as a measure of functional denervation. DEN, DIV-DEN, MPG-DEC, and BOO bladders are functionally denervated compared with sham-operated bladders. DEN bladders are significantly more functionally denervated than the DIV-DEN, BOO, and MPG-DEC bladders. DIV-DEN bladders are functionally denervated similar to BOO, both of which are more functionally denervated than the MPG-DEC bladders. Comparing the carbachol potency between the groups, a lower carbachol  $EC_{50}$  is found in the DEN bladders compared with sham operated (Fig. 3).

### Muscarinic receptor-subtype protein density

Total,  $M_2$ , and  $M_3$  receptor protein density was determined by subtype-selective immunoprecipitation (Fig. 4). Sham-operated bladders have a significantly different total receptor density (~470 fmol/mg solubilized protein) than every group, with an  $M_2$ -to- $M_3$  ratio of ~6:1. Total muscarinic receptor density in the DIV and DIV-DEN bladders is

significantly less than sham operated with M<sub>2</sub>-to-M<sub>3</sub> ratios of about 9:1 and 10:1, respectively. DEN, BOO, and MPG-DEC bladders have greater total receptor densities than sham-operated with M<sub>2</sub>-to-M<sub>3</sub> ratios of about 34:1, 24:1, and 11:1, respectively.

The hypertrophied bladders (DEN, BOO) have an increase in M<sub>2</sub> receptors, a decrease in M<sub>3</sub> receptors, and an increase in total receptor density, while the hypertrophied MPG-DEC bladders have an increase in M<sub>2</sub> and total receptors, with no change in M<sub>3</sub> receptors compared with sham operated. The atrophied bladders (DIV) have a decrease in both M<sub>2</sub> and M<sub>3</sub> receptors, which is reflected in a decrease in total receptor density. The DIV-DEN bladders, which are neither hypertrophied nor atrophied, have a decrease in M<sub>2</sub>, M<sub>3</sub>, and total receptor density.

### Correlation of muscarinic receptor-subtype density with functional denervation

As can be seen in Fig. 5, the density of the M<sub>2</sub> receptor protein correlates ( $R = 0.77$ ,  $P = 0.05$ ) with the degree of functional denervation. However, there is no correlation between the density of the M<sub>3</sub> subtype and the degree of functional denervation.

### Affinity of antagonists for inhibition of carbachol-induced contractions

Based on the affinity of a series of muscarinic receptor antagonists for inhibition of carbachol-induced contractions, sham-operated bladder contractions are mediated by the M<sub>3</sub> receptor subtype (Table 1). This is based on a high affinity for the M<sub>3</sub>-selective antagonists p-F-HHSiD ( $7.7 \pm 0.2$ ), 4-DAMP ( $9.1 \pm 0.2$ ), and darifenacin ( $8.5 \pm 0.1$ ) and a low affinity for the M<sub>2</sub>-selective antagonist methoctramine ( $6.2 \pm 0.2$ ). The affinities in DIV and DIV-DEN are also consistent with M<sub>3</sub> receptors mediating contraction. The M<sub>3</sub>-selective antagonists have a lower affinity in the hypertrophied bladders (DEN, MPG-DEC, and BOO), which is consistent with participation of the M<sub>2</sub> receptor in mediation of contraction. Paradoxically, the M<sub>2</sub>-selective antagonist methoctramine has a low affinity for inhibition of contraction in all groups, which seems to preclude participation of M<sub>2</sub> receptors in contraction. This phenomenon was also seen in our earlier studies (1, 4), which prompted speculation of the existence of two contractile pathways, one mediated by the M<sub>3</sub> receptor subtype, the other mediated by the M<sub>2</sub> subtype. Further evidence for the existence of two pathways was provided by studies in which antagonists showed superadditive inhibitory effects in blocking bladder contraction (6).

## DISCUSSION

Hypertrophy occurs rapidly after either DEN, MPG-DEC, or BOO in the rat urinary bladder. The bladder-to-body weight ratios (mg/g) in these groups increased by over twofold within 3 days. These results are in general agreement with previous published increases in bladder weight after 1 wk of denervation, decentralization, or outlet obstruction (11, 13). Similarly, nerve terminals in the denervated bladders degenerate rapidly as evidenced by an almost complete absence of EFS-induced contraction by 24 h after surgery. Although there is virtually no response to EFS at 24 h postsurgery and significant hypertrophy, there is no reduction in the maximal response to carbachol in the DEN and the MPG-DEC bladders. However, the BOO bladders, while having no reduction in EFS-induced contractility, have a significantly greater response to carbachol with no change in agonist affinity. As expected, using the ratio of the maximal carbachol contraction to the EFS-induced contraction as a measure of functional denervation, the DEN and DIV-DEN bladders are functionally denervated. Unexpectedly though, the MPG-DEC and the BOO bladders are also somewhat functionally denervated. It is possible that the increased bladder pressures in these groups leads to local ischemia and thus degeneration of nerve terminals. The DIV-DEN group, while functionally denervated compared with sham-operated animals, has a greater response



to EFS than DEN alone, providing evidence that the nerve terminals do not degenerate as rapidly in this group where the bladder is not exposed to increased mechanical stress. In addition, because bladders from the DIV-DEN group do not atrophy as do the DIV bladders, an intact innervation appears to be required for atrophy to occur. This may indicate a role for bladder innervation in control of hypertrophy, possibly via the release of paracrine factors that act in opposition to bladder hypertrophy and thereby induce atrophy.

The only group with denervation-induced supersensitivity is the DEN group, not the MPG-DEC or the DIV-DEN groups (Fig. 3). While the DIV-DEN group does not have increased bladder pressure because of the diversion, the MPG-DEC does. However, the MPG-DEC group has intact innervation from the major pelvic ganglion to the bladder. This suggests that, for at least the 3-day postoperative time point, both a complete absence of nerve terminals and increased pressure with concomitant hypertrophy are required for development of carbachol supersensitivity. Muscarinic agonist supersensitivity after 1–3 wk of urinary diversion or MPG decentralization has been reported (12), but we found that this does not occur at the 3-day time point. Our results are somewhat different from previously reported effects of bladder hypertrophy for 1 wk where no increase in the methacholine-induced contraction was seen but supersensitivity to methacholine was noted. These differences may be attributed to the duration of hypertrophy or possibly the method of inducing the hypertrophy [urethral ligature used here vs. paraffin injected into the lumen of the bladder used by Ekstrom et al. (13)]. The duration of the hypertrophy is more likely the reason for the differences, because the supersensitivity was transient and not present 4 wk after paraffin injection (13).

Analysis of the density of muscarinic receptors reveals an increase in total receptor density in the hypertrophied bladders regardless of whether they are denervated. All of this increase is accounted for by an increase in the  $M_2$  subtype. All experimental groups except MPG-DEC have a significantly lower density of the  $M_3$  subtype. The nonhypertrophied groups (DIV and DIV-DEN) have a decrease in total,  $M_2$ , and  $M_3$  receptor density. Therefore, hypertrophy leads to an increase in density of total and  $M_2$  receptors while urinary diversion results in a decrease in density of total and  $M_2$  receptors regardless of whether the diversion is accompanied by tissue atrophy (DIV group) or not (DIV-DEN group). Our results are consistent with those reported by Nilvebrant et al. (20), who showed increases in total muscarinic receptor density after 1 wk of denervation and decreases in total receptor density after 1 wk of diversion or diversion with denervation. While a trend for an increase in total receptor density was reported in the bladders induced to hypertrophy by paraffin injection, the increase was not significant (20), whereas we found a significant increase in total receptor density as a result of hypertrophy induced by outlet obstruction. This discrepancy may be the result of either the duration of hypertrophy or the method used to induce the hypertrophy.

In the DIV and DIV-DEN groups, despite decreased  $M_3$  receptor density (Fig. 4), the contractions are mediated by the  $M_3$  subtype (Table 1) and there is no change in agonist potency (Fig. 3). In other words there is no correlation between  $M_3$  receptor density and the potency of agonist or antagonist. The potency of carbachol is increased only in the DEN group (Fig. 3), which also has the largest increase in  $M_2$  receptor density (Fig. 4). The potency of carbachol is not increased in the BOO and MPG-DEC groups despite bladder hypertrophy and increased  $M_2$  receptor density. Thus the potency of carbachol for inducing contraction is not related to total receptor density or the density of the  $M_2$  or  $M_3$  subtype. This finding is similar to that reported by Nilvebrant et al. (20), who found that total receptor density does not correlate with supersensitivity in denervated, hypertrophied, diverted, or diverted and denervated bladders.

The significance of alterations in receptor density is not completely clear. However, the density of the M<sub>2</sub> receptor subtype correlates with functional denervation. The greater the degree of functional denervation, the greater is the density of the M<sub>2</sub> receptor subtype (Fig. 5A). No such correlation exists for the M<sub>3</sub> subtype (Fig. 5B). However, all groups with bladder hypertrophy have an increase in M<sub>2</sub> receptor density and low affinities for M<sub>3</sub>-selective antagonists, which suggests an M<sub>2</sub>-mediated component of contraction. The DIV and DIV-DEN bladders, which are not hypertrophied, have higher affinities for these M<sub>3</sub>-selective antagonists, suggesting that the M<sub>3</sub> subtype mediates bladder contraction. Another report on the effect of BOO on muscarinic receptor-mediated bladder contraction found no differences in carbachol-mediated contraction (18). The differences in results reported by this group and our results may be due to the use of males as opposed to females or the duration of obstruction (4 wk as opposed to 3 days in our study).

Prejunctional autoreceptors have been identified on the nerves innervating the rat bladder (3, 26, 28). Activation of the M<sub>1</sub> subtype increases acetylcholine release and contraction while activation of the M<sub>2</sub> subtype reduces acetylcholine release and inhibits contraction. Somogyi and de Groat (27) have shown that the prejunctional facilitatory mechanism is upregulated after chronic spinal transection; furthermore, this facilitation appears to be primarily mediated by M<sub>3</sub> receptors as opposed to M<sub>1</sub> receptors in normal animals. Thus previous evidence exists for plasticity in the neural mechanism governing bladder contraction, and our results provide evidence for plasticity in the smooth muscle mechanism mediating bladder contraction.

The *in vitro* results reported in this manuscript, namely an M<sub>2</sub> receptor-mediated component of contraction, is superficially in agreement with *in vivo* experiments implicating the M<sub>2</sub> receptor subtype in bladder contraction. The amplitude of volume-induced bladder contractions in the urethane-anesthetized rat is inhibited by subtype-selective muscarinic antagonists with potencies that correlate most favorably with pK<sub>i</sub> estimates of these compounds at human recombinant M<sub>2</sub> receptors (16). In addition, the M<sub>2</sub>- and M<sub>4</sub>-selective antagonist AQ-RA 741, similar to the nonselective antagonist tolterodine, has a greater selectivity for inhibition of bladder contraction than for inhibition of salivation in the  $\alpha$ -chloralose-anesthetized cat, suggesting M<sub>2</sub> receptor involvement in bladder contraction (15). Intravenous injection of AF-DX116 (M<sub>2</sub>-selective antagonist) reduces contraction pressure but not frequency or the duration of bladder contraction (21). In these *in vivo* studies of systemic administration of antagonists, the sites of action of the antagonists are not known and inhibition of bladder contraction or salivation could be due to additional effects on the central or the peripheral nervous system and not strictly due to effects of the antagonist on the end organ itself.

Intracerebroventricular injection of the M<sub>3</sub>-selective antagonist 4-DAMP inhibits both the amplitude and the duration of volume-induced bladder contractions, whereas AF-DX116 decreases contraction frequency while prolonging the duration of contraction (21). On the other hand, intracerebroventricular injection of darifenacin, an M<sub>3</sub>-selective antagonist, has no effect on voiding parameters in normal conscious rats, while intracerebroventricular injection of tolterodine, a non-subtype-selective antagonist, decreases voiding pressure and increases bladder capacity (17). Thus evidence exists for the central nervous system control of bladder contraction by both M<sub>2</sub> and M<sub>3</sub> muscarinic receptor subtypes. Because the prejunctional facilitatory autoreceptors have been shown to change from M<sub>1</sub> to predominantly M<sub>3</sub> after spinal cord transection in the rat (27), it may also be possible that pathophysiological conditions could induce alterations in the central mechanisms governing micturition.

The question arises as to how M<sub>2</sub> receptors directly mediate smooth muscle contraction. M<sub>2</sub> receptors are traditionally thought to preferentially couple to PTX-sensitive G proteins such as the G<sub>i</sub> subfamily, resulting in inhibition of adenylyl cyclase, while M<sub>3</sub> receptors preferentially couple to G<sub>q</sub> and stimulation of phosphoinositol hydrolysis leading to an increase in cytosolic calcium. Pharmacological studies demonstrate that most smooth muscle contraction is mediated by the M<sub>3</sub> subtype (7, 9). However, an M<sub>2</sub>-mediated contractile response in bladder muscle can be demonstrated after the majority of M<sub>3</sub> receptors are inactivated in an environment of increased intracellular cAMP such as during stimulation with a β-adrenergic agonist (5, 16). This pathway has been proposed to mediate contraction indirectly, merely by blocking β-adrenergic agonist-induced relaxation via increased cAMP (9). However, M<sub>2</sub> receptors acting through G<sub>i</sub> may also stimulate bladder contraction directly via PKC activation as previously found in the cat lower esophageal sphincter smooth muscle. A low degree of muscarinic stimulation and, consequently, a low degree of calcium mobilization result in activation of PKC, whereas PKC activation is inhibited at higher intracellular calcium concentrations (24). Thus in the face of normal calcium mobilization mediated by the M<sub>3</sub> receptor subtype, the signal transduction pathway mediated by the M<sub>2</sub> subtype may be inhibited. One hypothesis to explain the shift in muscarinic receptor subtype mediating contraction from M<sub>3</sub> to M<sub>2</sub> in the hypertrophied bladders is a deficit in calcium mobilization.

We have previously shown quantitative evidence for an interaction between the second messenger systems activated by the M<sub>2</sub> and the M<sub>3</sub> receptor subtypes in the denervated rat bladder (6). The simultaneous action of M<sub>2</sub>-selective and M<sub>3</sub>-selective antagonists induces a synergistic inhibition of contraction in denervated bladders, which indicates a facilitatory interaction of the two subtypes in inducing contraction. However, results in the normal rat bladder show no facilitatory interaction between M<sub>2</sub> and M<sub>3</sub> subtypes for inducing contraction. Only after either denervation or blocking the sarcoplasmic reticulum calcium pump with thapsigargin does this interaction become facilitatory. These two results provide further support for an interaction between subtypes mediating contraction as previously reported in the guinea pig colon where the M<sub>2</sub> and M<sub>3</sub> receptor subtypes are thought to interact in a facilitatory manner to mediate contraction (23). However, our results in the normal rat bladder do not support such a facilitatory interaction; actually, the opposite appears to occur, namely that the M<sub>3</sub> pathway seems to inhibit the M<sub>2</sub> pathway, possibly via calcium mobilization. It is possible that the interaction between subtypes is different in the guinea pig colon or that after 3 days of in vivo PTX treatment (23), the interaction between subtypes becomes altered.

Experimental pathologies that interfere with the normal functioning of the bladder induce a decrease in density of the M<sub>3</sub> receptor subtype. Conditions that lead to hypertrophy induce an increase in density of the M<sub>2</sub> receptor subtype and a shift in the mechanism of contraction such that the M<sub>2</sub> subtype can be shown to at least partially mediate contraction.

## Acknowledgments

### DISCLOSURES

This work was supported by National Institutes of Diabetes and Digestive and Kidney Diseases Grant R01-DK-4333.

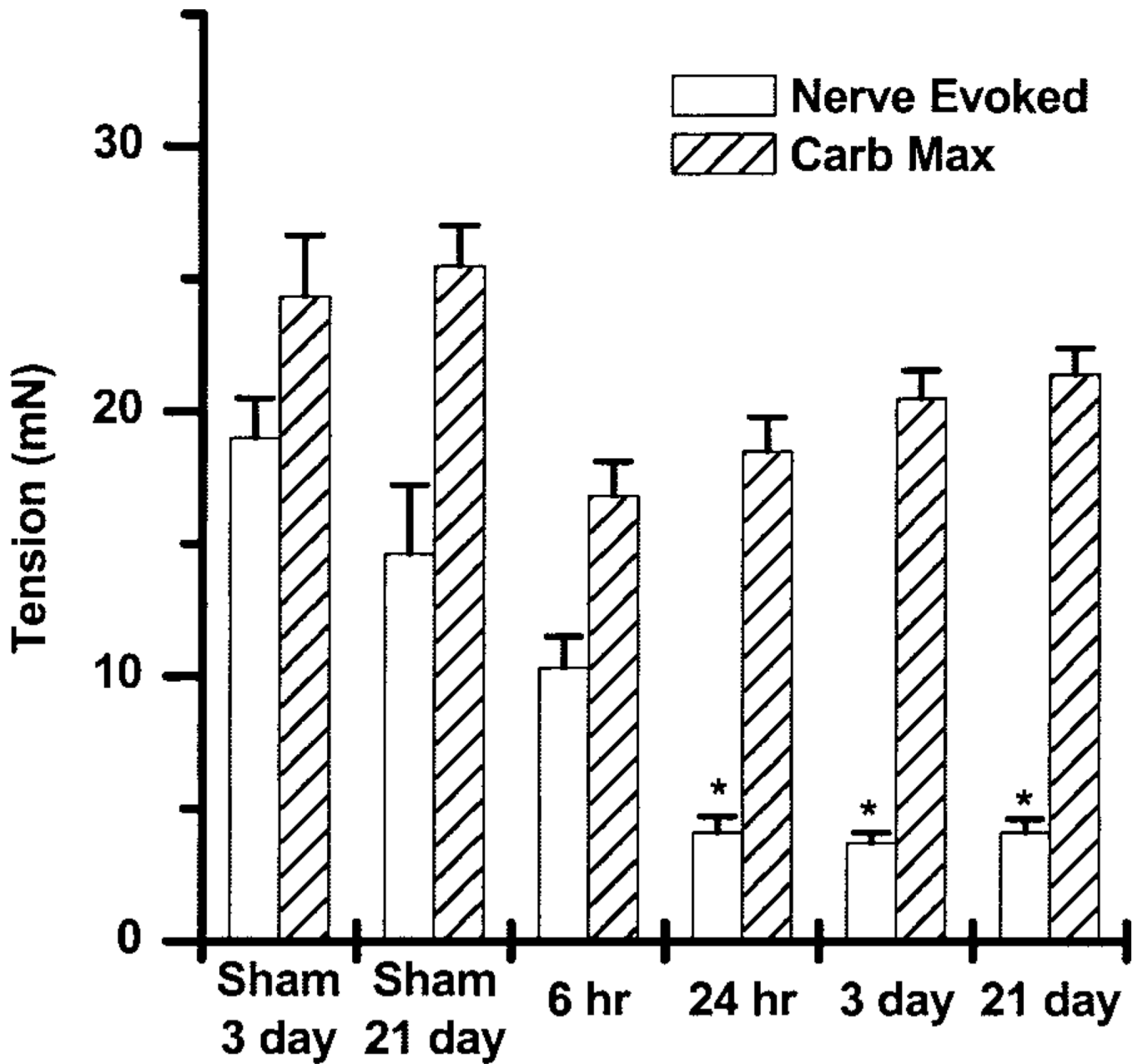
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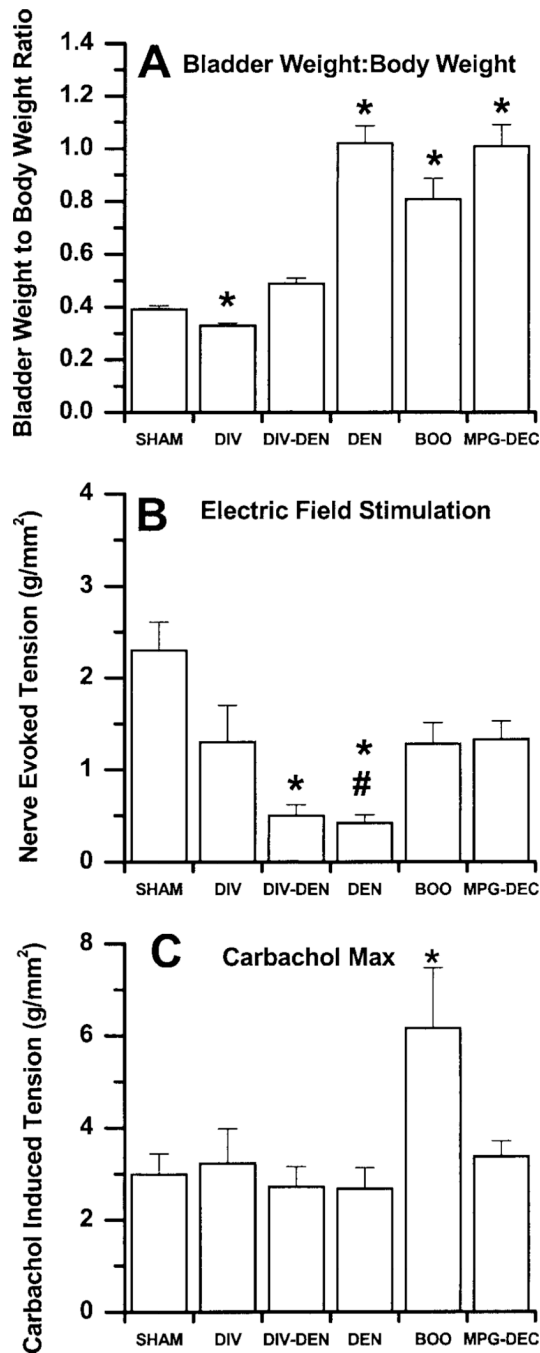


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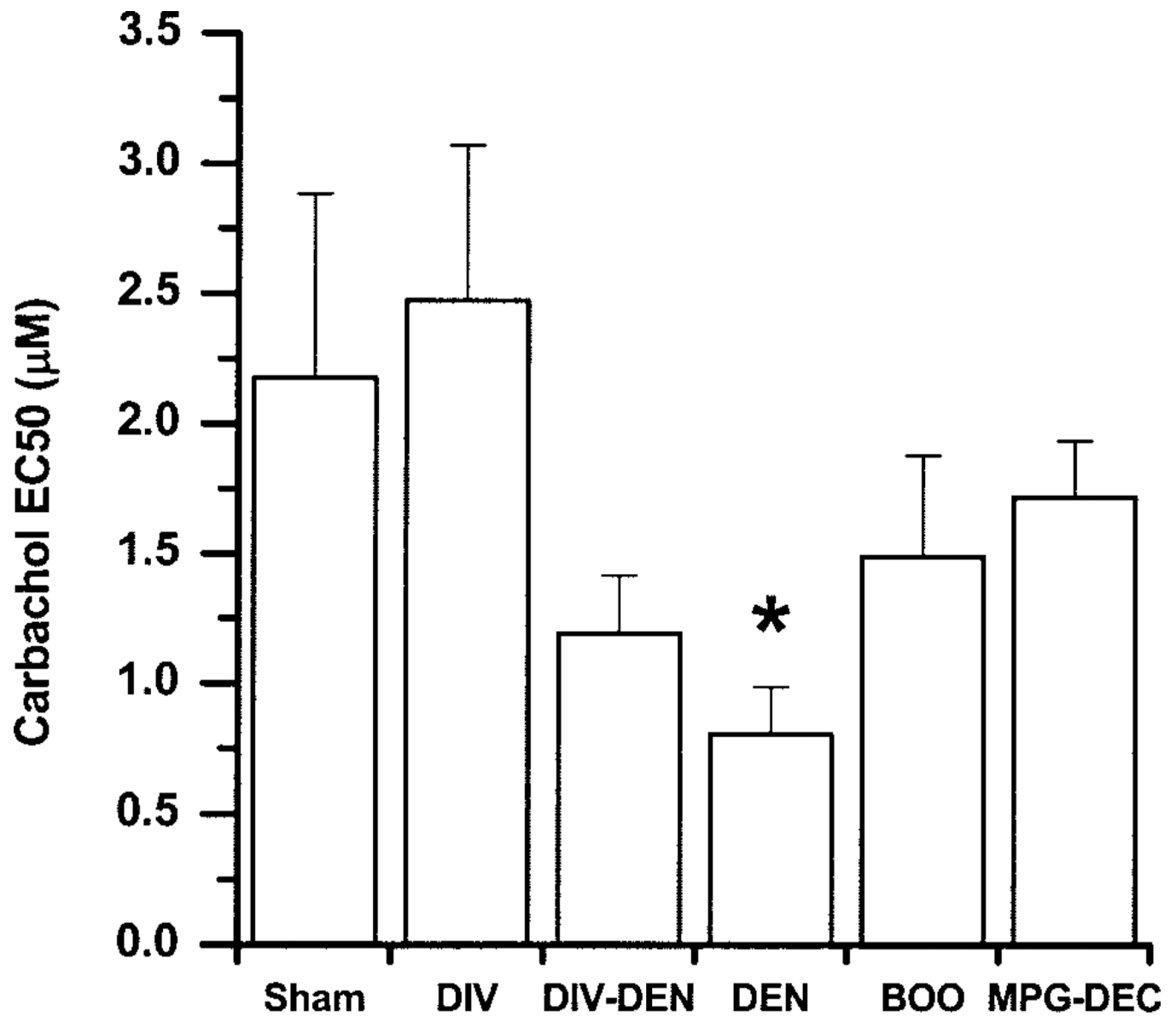
**Fig. 1.** Time course of denervation-induced changes in the maximal carbachol response and the nerve-evoked responses to electric field stimulation (EFS) of 8 V, 30 Hz, 1 ms presented as means  $\pm$  SE. There were no significant differences in the carbachol-evoked maximum (Carb Max). However, by 24 h the nerve-evoked contractions were significantly reduced.  
\*Significantly different from control,  $P < 0.05$ .

**Fig. 2.**

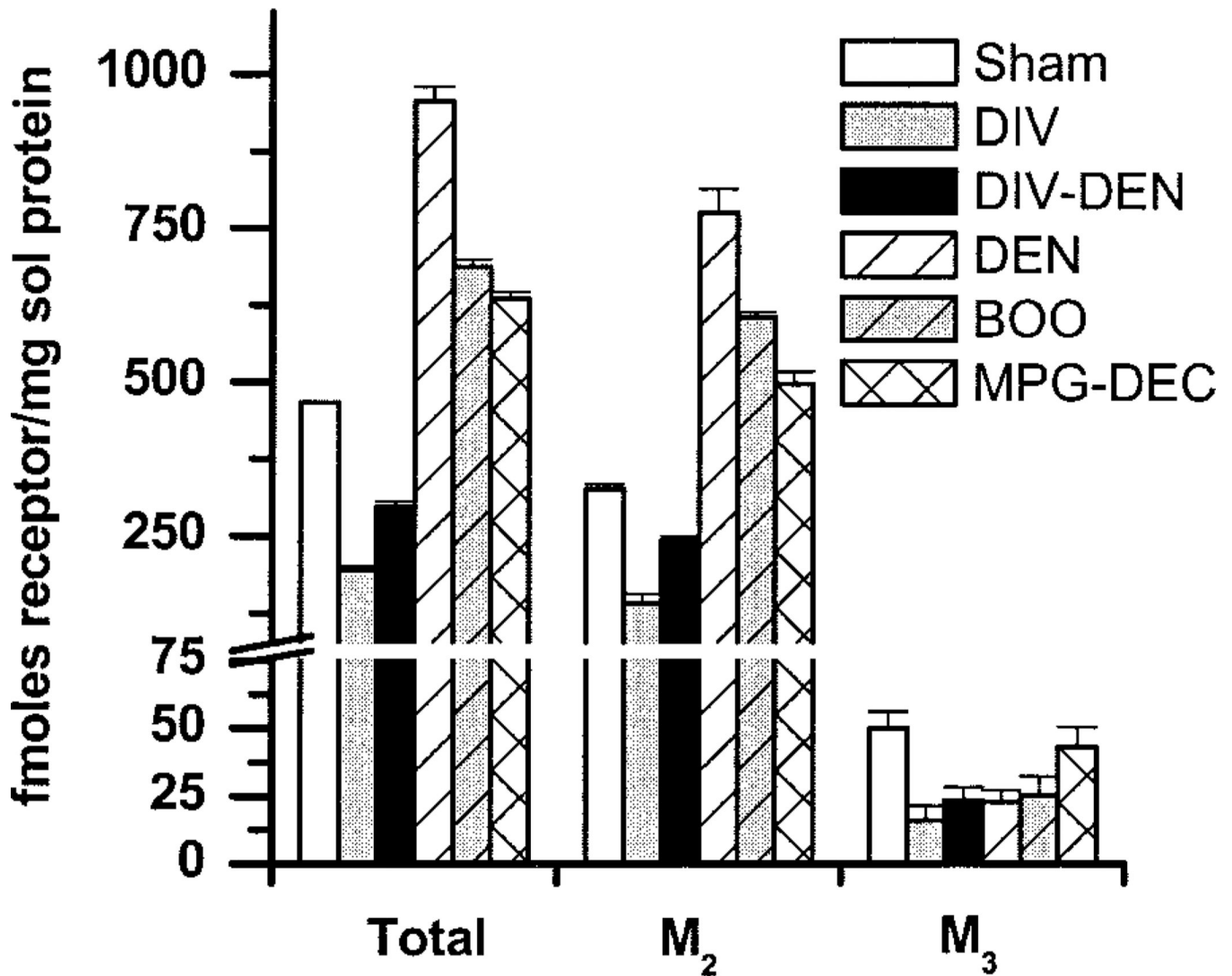
DIV, urinary diversion; DIV-DEN, major pelvic ganglion (MPG) electrocautery (MPGE) together with urinary diversion; DEN, bilateral MPGE; BOO, bladder outlet obstruction; MPG-DEC; MPG decentralization. *A*: effect of experimental pathologies on bladder hypertrophy presented as means  $\pm$  SE (mg/g). \*DEN, BOO, and MPG-DEC are significantly greater than sham operated while DIV is significantly less than sham operated ( $n = 9-24$  rats/group). *B*: nerve-evoked contractile response to EFS of 8 V, 30 Hz, 1 ms presented as means  $\pm$  SE. \*DIV-DEN and DEN are significantly less than sham operated. #DIV-DEN is significantly greater than DEN ( $n = 40-60$  muscle strips/group). *C*: maximal carbachol

contractile response presented as means  $\pm$  SE. \*BOO is significantly greater than every other group ( $n = 40\text{--}60$  muscle strips/group).



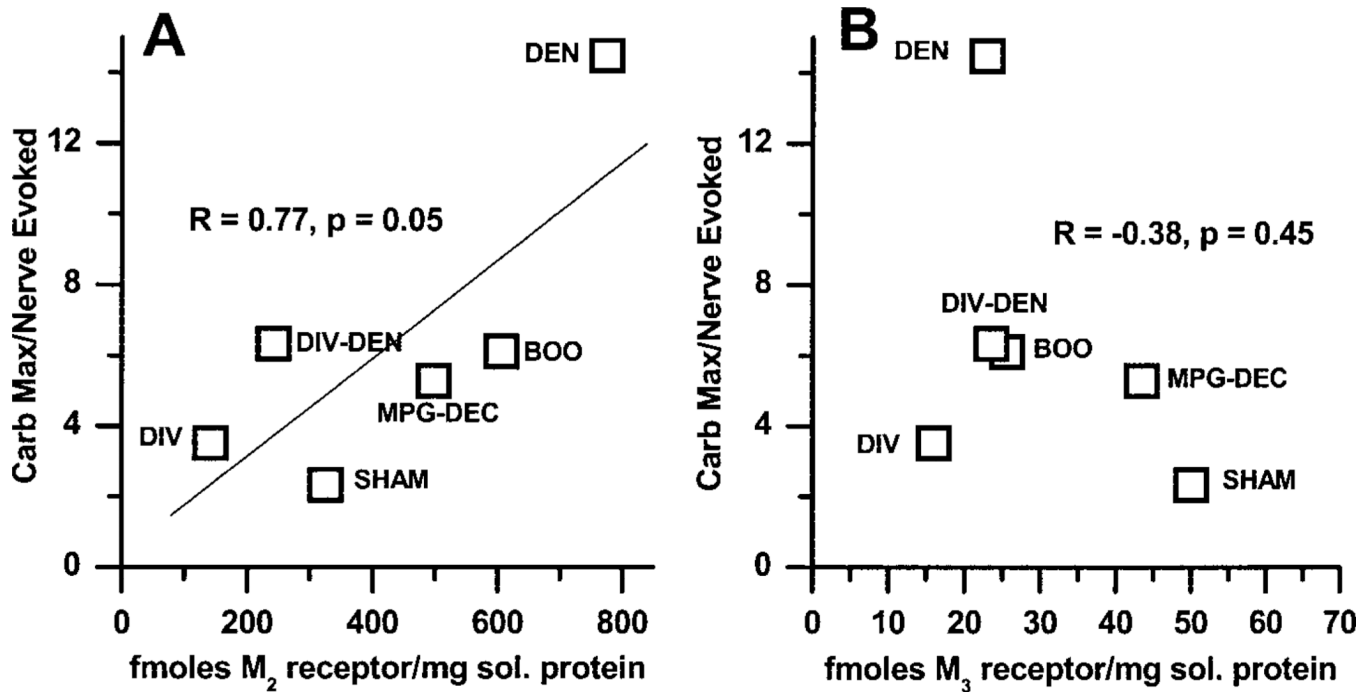


**Fig. 3.** Effect of experimental pathologies on the potency of carbachol to induce contraction presented as means  $\pm$  SE. \*DEN is significantly less than sham operated, DIV, and MPG-DEC ( $n = 7-36$  muscle strips/group).



**Fig. 4.**

Effect of experimental pathologies on the density of urinary bladder muscarinic receptor subtypes. M<sub>2</sub> and M<sub>3</sub> receptors were labeled with [<sup>3</sup>H]QNB and solubilized as described in Luthin et al. (19). Data shown are average fmol ( $\pm$ SE) of receptor/mg solubilized protein from sham operated, DIV, DIV-DEN, DEN, BOO, and MPG-DEC ( $n = 4-5$  determinations from 6-18 bladders). Protein concentration in the solubilized receptor preparation was ~8% of the protein concentration in the crude homogenate. Compared with filtration binding, ~50% of the muscarinic receptors were solubilized (data not shown). Group differences were determined using ANOVA with a post hoc Newman-Keuls test. For total receptor density all groups are different from each other. For M<sub>2</sub> receptor density, all groups are different from each other. For M<sub>3</sub> receptor density, sham operated is different from DIV, DIV-DEN, DEN, and BOO.



**Fig. 5.** Correlation of muscarinic receptor subtype density with functional denervation. For each individual muscle strip the ratio of the carbachol maximum:EFS-induced contraction was determined. The average of the individual muscle strips for each experimental group for functional denervation is plotted vs. the immunoprecipitation data for M<sub>2</sub> receptor density (A) and M<sub>3</sub> receptor density (B). The ratio of the average carbachol maximum to the average EFS contraction displayed in Fig. 2 does not equal the average of the individual ratios for the strips. The density of M<sub>2</sub> receptors correlates with functional denervation. No correlation between the M<sub>3</sub> receptor density and functional denervation is seen.

**Table 1**

Affinity (95% confidence interval) values of subtype-selective muscarinic receptor antagonists for inhibition of carbachol-induced bladder contraction and their reported affinity for the M<sub>2</sub> and M<sub>3</sub> receptor subtypes

Group	Methoctramine	4-DAMP	<i>p</i> -F-HHSiD	Darifenacin
Sham operated	6.0–6.4	8.9–9.1	7.6–7.8	8.3–8.7
DIV	6.0–6.4		7.2–7.6	8.7–9.1
DIV-DEN	6.1–6.5		7.2–7.6	8.6–9.2
DEN	6.3–6.7	8.3–8.7	6.8–7.2	7.5–8.5
BOO	5.9–6.5		6.5–7.1	7.8–8.4
MPG-DEC	4.4–5.6		6.5–7.1	7.5–7.9
M <sub>2</sub> *	7.8–8.3	8.0–8.4	6.0–6.9	7.0–7.4
M <sub>3</sub> *	6.3–6.9	8.9–9.3	7.8–7.9	8.4–8.9

Values are 95% confidence intervals; 6–8 muscle strips were used per antagonist concentration. Nonoverlapping confidence intervals denote a statistically significant difference ( $P < 0.05$ ). The affinities for 4-DAMP, methoctramine, and *p*-F-HHSiD were determined by Schild analysis. Concentrations of 0.3 and 3.0  $\mu$ M methoctramine and *p*-F-HHSiD were used, while 3.0, 10.0, and 30.0 nM 4-DAMP were used. The estimated  $pK_b$  for darifenacin was determined based on a single 30 nM darifenacin concentration.

\* Adapted from Caulfield (7) and Caulfield and Birdsall (8), which include both functional and ligand binding studies. DIV, urinary diversion; DIV-DEN, major pelvic ganglion (MPG) electrocautery (MPGE) together with urinary diversion; DEN, bilateral MPGE; BOO, bladder outlet obstruction; MPG-DEC, MPG decentralization; 4-DAMP, 4-diphenacetoxyl-*N*-methylpiperidine methiodide; *p*-F-HHSiD, *para*-fluoro-hexahydrosila-diphenidol.