



# Frontotemporal Dementia: Implications for Understanding Alzheimer Disease

Michel Goedert<sup>1</sup>, Bernardino Ghetti<sup>2</sup>, and Maria Grazia Spillantini<sup>3</sup>

<sup>1</sup>MRC Laboratory of Molecular Biology, Cambridge CB2 0QH, United Kingdom

<sup>2</sup>Indiana University School of Medicine, Department of Pathology and Laboratory Medicine, Indianapolis, Indiana 46202

<sup>3</sup>Centre for Brain Repair, Department of Clinical Neurosciences, University of Cambridge, Cambridge CB2 0PY, United Kingdom

Correspondence: mg@mrc-lmb.cam.ac.uk

Frontotemporal dementia (FTD) comprises a group of behavioral, language, and movement disorders. On the basis of the nature of the characteristic protein inclusions, frontotemporal lobar degeneration (FTLD) can be subdivided into the common FTLD-tau and FTLD-TDP as well as the less common FTLD-FUS and FTLD-UPS. Approximately 10% of cases of FTD are inherited in an autosomal-dominant manner. Mutations in seven genes cause FTD, with those in tau (*MAPT*), chromosome 9 open reading frame 72 (*C9ORF72*), and progranulin (*GRN*) being the most common. Mutations in *MAPT* give rise to FTLD-tau and mutations in *C9ORF72* and *GRN* to FTLD-TDP. The other four genes are transactive response–DNA binding protein-43 (*TARDBP*), fused in sarcoma (*FUS*), valosin-containing protein (*VCP*), and charged multivesicular body protein 2B (*CHMP2B*). Mutations in *TARDBP* and *VCP* give rise to FTLD-TDP, mutations in *FUS* to FTLD-FUS, and mutations in *CHMP2B* to FTLD-UPS. The discovery that mutations in *MAPT* cause neurodegeneration and dementia has important implications for understanding Alzheimer disease.

## THE CONCEPT OF FRONTOTEMPORAL DEMENTIA: HISTORICAL OVERVIEW

Frontotemporal dementia (FTD) results from degeneration of the cortex of the frontal and temporal lobes, often in conjunction with the degeneration of subcortical brain regions. This gives rise to a spectrum of behavioral, language, and movement disorders. A link exists between FTD and forms of motor neuron disease (MND). Work on FTD stretches back to the end of the 19th century.

Arnold Pick was Professor of Neuropsychiatry at the German University in Prague from 1886 to 1921. In 1892, he described a 71-year-old man with behavioral disturbances, aphasia, and dementia (Pick 1892). At autopsy, marked atrophy of the left temporal lobe rather than the diffuse atrophy characteristic of senile dementia was present. Although Pick was doubtful of the primacy of these observations, his paper is considered to be the first description of lobar cortical atrophy. At the time, there was much interest in language abnormalities,

---

Editors: Dennis J. Selkoe, Eckhard Mandelkow, and David M. Holtzman  
Additional Perspectives on The Biology of Alzheimer Disease available at [www.perspectivesinmedicine.org](http://www.perspectivesinmedicine.org)

Copyright © 2012 Cold Spring Harbor Laboratory Press; all rights reserved; doi: 10.1101/cshperspect.a006254  
Cite this article as *Cold Spring Harb Perspect Med* 2012;4:a006254

following the description of motor and sensory aphasias (Broca 1861; Wernicke 1874; see also Freud 1891). A few years later, Déjerine and Sériex (1897) described a case of sensory aphasia with bilateral temporal atrophy. Pick went on to report four additional cases with temporal lobe atrophy and language disturbances (Pick 1901, 1904). In 1906, he described a patient with disinhibition and mixed apraxia who had severe bilateral frontal and left-sided parietal atrophy, with a more moderate atrophy of the left temporal lobe (Pick 1906). Pick was mainly interested in comparing the clinical picture with the macroscopic appearance of the brain. He made no systematic attempt at identifying histopathological abnormalities. Alzheimer discovered the association of argyrophilic intracytoplasmic inclusions and ballooned neurons with lobar cortical atrophy, in the absence of the plaques and tangles he had described four years earlier (Alzheimer 1907, 1911). This revealed the existence of a second type of intraneuronal inclusion and established that different inclusions can characterize distinct clinical entities.

Richter proposed that lobar cortical atrophies are hereditary diseases (Richter 1918) and Gans, a pupil of Pick, linked his mentor's name to cases of lobar cortical atrophy (Gans 1923). Additional examples of frontal and/or temporal cortical atrophy with or without argyrophilic inclusions were subsequently reported and the clinical condition was called "Pick's disease" (Onari and Spatz 1926; Stertz 1926). Unlike Pick, who believed to have described atypical forms of senile dementia, Onari and Spatz considered Pick's disease to be a distinct entity. One of their patients (Therese Mühlich) had already been described by Alzheimer. Carl Schneider proposed a three-stage model for the clinical course of Pick's disease (Schneider 1927, 1929). In most individuals, the first stage is characterized by disinhibition and impaired judgement, although Schneider recognized that amnesic aphasia is the presenting symptom of temporal lobe atrophy. The second stage is dominated by progressive dementia and focal symptoms, such as apathy in frontal lobe atrophy and sensory aphasia in temporal lobe atrophy. Stereotyped

perseverations of speech, movement, and facial expression also appear. The third stage is characterized by dementia and severe language problems, resulting in a vegetative state with flexion contractures. Schneider concluded that the argyrophilic inclusions and ballooned cells described by Alzheimer were diagnostic of Pick's disease. Similar cases were described in the 1930s, when it became clear that lobar cortical atrophy has a high degree of heritability, irrespective of the presence of argyrophilic inclusions (Grünthal 1930; Verhaart 1930; Von Braunmühl and Leonhard 1934). The link between frontal lobe dementia and MND was also recognized (Meyer 1929; Von Braunmühl 1932). The early work was summarized and discussed by Van Mansvelt (1954) and Lüers and Spatz (1957).

Interest in the focal dementias waned after World War II, before it was rekindled in the 1970s and 1980s. Cases of Pick's disease with argyrophilic inclusions and ballooned neurons (type A) were now distinguished from those with ballooned neurons lacking argyrophilic inclusions (type B) and those lacking both ballooned neurons and argyrophilic inclusions (type C) (Constantinidis 1974). Work by Brun, Gustafson, and Neary showed that some individuals with frontal lobe atrophy lacked a distinctive histopathology (Brun 1987; Gustafson 1987; Neary et al. 1988). Clinically, these patients suffered from a severe personality disorder, which is now known as behavioral-variant FTD (bvFTD). Mesulam described primary progressive aphasia (PPA), with an isolated language deficit as the most prominent presenting feature, in the absence of strokes or tumors (Mesulam 1982, 1987, 2001). PPA has been divided into three syndromes (Gorno-Tempini et al. 2011): (1) Semantic dementia (SD), also known as semantic variant PPA, a fluent aphasia with loss of word meaning (Snowden et al. 1989); (2) progressive nonfluent aphasia (PNFA), also known as nonfluent/agrammatic variant PPA, a disorder characterized by effortful, nonfluent speech (Grossman et al. 1996); and (3) logopenic progressive aphasia (LPA), also known as logopenic variant PPA, a nonfluent aphasia with deficits in word

retrieval and sentence repetition (Gorno-Tempini et al. 2004b).

Symptoms correlate better with specific patterns of brain atrophy than with the underlying neuropathology. Prediction of the neuropathology based on clinical picture remains challenging. FTD is the third most common cause of early-onset dementia (disease onset < 65 years), after Alzheimer disease and vascular dementia (Rossor et al. 2010). Approximately 40% of patients with FTD have a family history, but only 10% of cases are inherited in a dominant manner. Links exist with the corticobasal syndrome (CBS), progressive supranuclear palsy (PSP), parkinsonism, and MND.

## CLINICAL PRESENTATIONS OF FRONTOTEMPORAL DEMENTIA

### Behavioral-Variant Frontotemporal Dementia (bvFTD)

bvFTD comprises more than half of the cases of FTD and is the most heritable form. Presenting features are insidious and include progressive changes in the patient's personality, interpersonal conduct, and emotional modulation (Gustafson 1987; Neary et al. 1988; Piguet et al. 2011a). A variable degree of language impairment is also present. Apathy manifesting as passivity, inertia, reduced motivation, and social withdrawal, associated with a lack of insight, is common. Disinhibition often coexists alongside apathy and manifests itself by impulsivity. Emotional blunting characterized by a lack of empathy is common. Abnormal eating behavior can be extensive, resulting in marked weight gain. Stereotypic and ritualistic behavior is common, as expressed by motor stereotyping, including humming, lip smacking, hand ruffling, and foot tapping. Neglect of self-care and impairment of other activities of daily living are common. Most patients are unable to manage their financial affairs. Memory is relatively spared in the early stages of bvFTD. By neuroimaging, four subtypes have been identified based on relative grey matter loss: frontal-dominant, frontotemporal, temporofrontoparietal, and temporal-dominant (Whitwell et al. 2009a). Combined

frontotemporal and basal ganglion atrophy can also be present, as can atrophy of a number of other subcortical regions. bvFTD and Alzheimer disease lead to divergent network activity patterns, with atrophy in an anterior salience network in bvFTD and a posterior default mode network in Alzheimer disease (Zhou et al. 2010).

### Semantic Dementia (SD)

SD is a progressive fluent aphasia, which is characterized by the loss of word meaning (Snowden et al. 1989; Hodges and Patterson 2007). Patients have difficulty in finding words, with anomia being a defining feature. They also complain of memory loss, but this does not in general reflect true amnesia. Although language deficits predominate, behavioral alterations also occur. SD is the least heritable FTD syndrome. A deficit in naming and word comprehension predominates, with the patient's vocabulary being depleted of all but the most common words. However, speech is fluent and the syntax correct. This is often coupled with deficits in person recognition, especially when the right temporal lobe is affected. Although patients are insightful and can be distressed by their condition, lack of empathy and mental inflexibility are common. Restriction of food preferences is present without the overeating characteristic of bvFTD. Compulsive behavior is prominent and centers on visual objects (left temporal lobe predominance) or on letters, words, and symbols (right temporal lobe predominance). By neuroimaging of grey matter, bilateral, often asymmetric, anterior temporal lobe atrophy is most prominent. The hippocampus can also be affected (Mummery et al. 1999). White matter changes are found in the ventral language pathways and the temporal components of the dorsal language pathways (Galantucci et al. 2011).

### Progressive Nonfluent Aphasia (PNFA)

PNFA is a disorder of expressive language. Patients lose the ability to speak fluently, with relative preservation of word comprehension and nonlinguistic cognition (Grossman et al.

M. Goedert et al.

1996). Several speech changes characterize PNFA. At an early stage, patients speak less than normal and in shorter sentences. They show speech apraxia, with effortful speech and phonological errors. Word finding difficulty is commonly observed, often resulting in muteness. Behavioral features similar to those of bvFTD may occur, but they are usually mild, with apathy being most common. As the disease progresses, extrapyramidal features become widespread and can lead to a change in diagnosis (Gorno-Tempini et al. 2004a). Heritability of PNFA is intermediate between that of bvFTD and SD. Neuroimaging shows a widening of the Sylvian fissure, with atrophy of left posterior frontal and insular regions (Neary et al. 2003; Nestor et al. 2003). In white matter, the most prominent changes are found in the dorsal language pathways (Galantucci et al. 2011).

#### Logopenic Progressive Aphasia (LPA)

LPA is a progressive nonfluent aphasia, which is characterized by a slow speech rate and word retrieval difficulties (Gorno-Tempini et al. 2004b, 2008). Repetition of phrases is also markedly impaired, in part as a result of limited auditory-verbal short-term memory. Single-word comprehension and semantic association are largely preserved. It differs from the fast output typical of patients with SD and the agrammatism and articulation deficits characteristic of PNFA. However, a language variant of Alzheimer disease overlaps with LPA (Galton et al. 2000; Alladi et al. 2007). It has been suggested that LPA and posterior cortical atrophy are clinical presentations of sporadic, early-onset Alzheimer disease (Migliaccio et al. 2009). This nonmemory phenotype characterizes about one quarter of patients (Van der Flier et al. 2011). Inheritance of the *APOE*  $\epsilon$ 4 allele appears not to be a risk factor for LPA and posterior cortical atrophy, distinguishing them from the more common amnesic forms of Alzheimer disease (Strittmatter and Roses 1995). Neuroimaging of LPA shows atrophy or hypoperfusion of the left posterior superior and middle temporal regions, and of the inferior parietal region (Gorno-Tempini et al.

2004b). Brain atrophy is located more posteriorly than in SD and PNFA. White matter changes are most marked in the temporoparietal component of the dorsal language pathway (Galantucci et al. 2011).

#### Frontotemporal Dementia and Corticobasal Syndrome (CBS)

CBS and PSP are atypical parkinsonian disorders. CBS is characterized by extrapyramidal symptoms consisting of progressive asymmetric rigidity and dystonia, and by signs of cortical dysfunction in the form of PNFA, apraxia, cortical sensory loss, alien limb syndrome, myoclonus, and hemineglect. For many years, the emphasis was on the extrapyramidal component, even though similarities with Pick's disease were noticed early on (Rebeiz et al. 1968). More recent work has shown that patients with CBS can have aphasia or a behavioral disorder characteristic of bvFTD (Lippa et al. 1991; Kertesz et al. 1994). Pathologically, CBS is heterogenous, but its most common form is corticobasal degeneration (CBD). Some cases of CBS are dominantly inherited. Neuroimaging shows variable frontoparietal and basal ganglion atrophy (Whitwell et al. 2010).

#### Frontotemporal Dementia and Progressive Supranuclear Palsy (PSP)

The clinical presentation of PSP includes vertical supranuclear ophthalmoplegia with difficulty looking up, bradykinesia, axial dystonia and rigidity, pseudobulbar palsy and postural instability with backward falls (Steele et al. 1964; Litvan et al. 1996). More than half of the patients develop cognitive impairment. Apathy and emotional blunting, accompanied by mental slowness and reduced verbal fluency, are common. A small percentage of cases of PSP is inherited. By neuroimaging, atrophy in premotor and supplemental motor areas is observed, with sparing of the inferior frontal lobe (Whitwell et al. 2010). Some patients show PNFA with early apraxia of speech (Josephs et al. 2006). Three subtypes of PSP have been described: Richardson's syndrome,

PSP-parkinsonism, and PSP-pure akinesia with gait freezing (Williams and Lees 2009). Cognitive impairment and cortical atrophy are most prominent in Richardson's syndrome, which corresponds to classical PSP.

### Frontotemporal Dementia and Parkinsonism Linked to Chromosome 17 (FTDP-17)

In the 1980s and 1990s, dominantly inherited forms of FTD were identified (Ghetti et al. 2011). Extrapyramidal signs resembling CBS and PSP also featured prominently. Amyotrophy was present in some cases. These forms of inherited FTD were given different names according to their predominant clinical and pathological features, including familial Pick's disease, disinhibition-dementia-parkinsonism-amyotrophy complex, familial progressive subcortical gliosis, familial presenile dementia with tangles, autosomal-dominant parkinsonism, dementia with pallido-ponto-nigral degeneration, and multiple system tauopathy with presenile dementia. Despite this heterogeneity, disease was linked to the long arm of chromosome 17 (Wilhelmsen et al. 1994). The syndrome received its name at a consensus conference during which 13 families were presented (Foster et al. 1997). FTDP-17 is divided into a dementia-dominant and a parkinsonism-dominant type. Neuroimaging shows variable frontotemporo-parietal and basal ganglion atrophy (Whitwell et al. 2009b).

### Frontotemporal Dementia with Motor Neuron Disease (MND)

MND can be associated with cognitive dysfunction (Morita et al. 1987). Mild frontal lobe involvement is found in 30% of cases and in ~3% of cases FTD is present (Shaw 2010). A psychotic phase consisting of vivid delusions is an early sign. Behavioral and cognitive changes tend to predate MND. Bulbar signs are common and electromyography is as in MND. Inherited cases of FTD-MND have been linked to chromosome 9p21 (Vance et al. 2006). Neuroimaging shows posterior frontal

lobe atrophy (Whitwell et al. 2006). Based on the presence of isolated upper MND in some cases, further subclassification of FTD-MND has been proposed (Josephs and Dickson 2007).

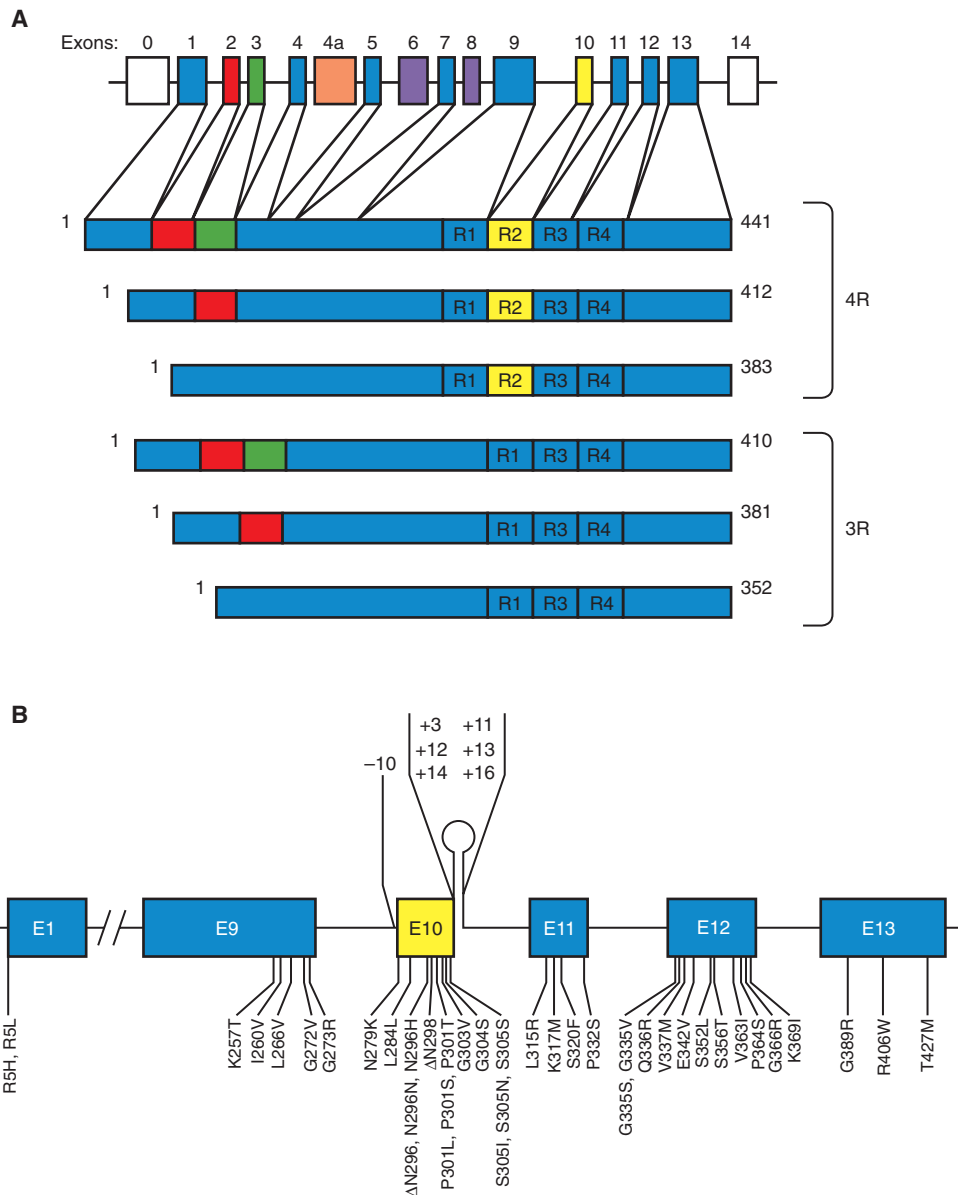
### HISTOPATHOLOGY OF FRONTOTEMPORAL LOBAR DEGENERATION (FTLD)

#### FTLD-Tau

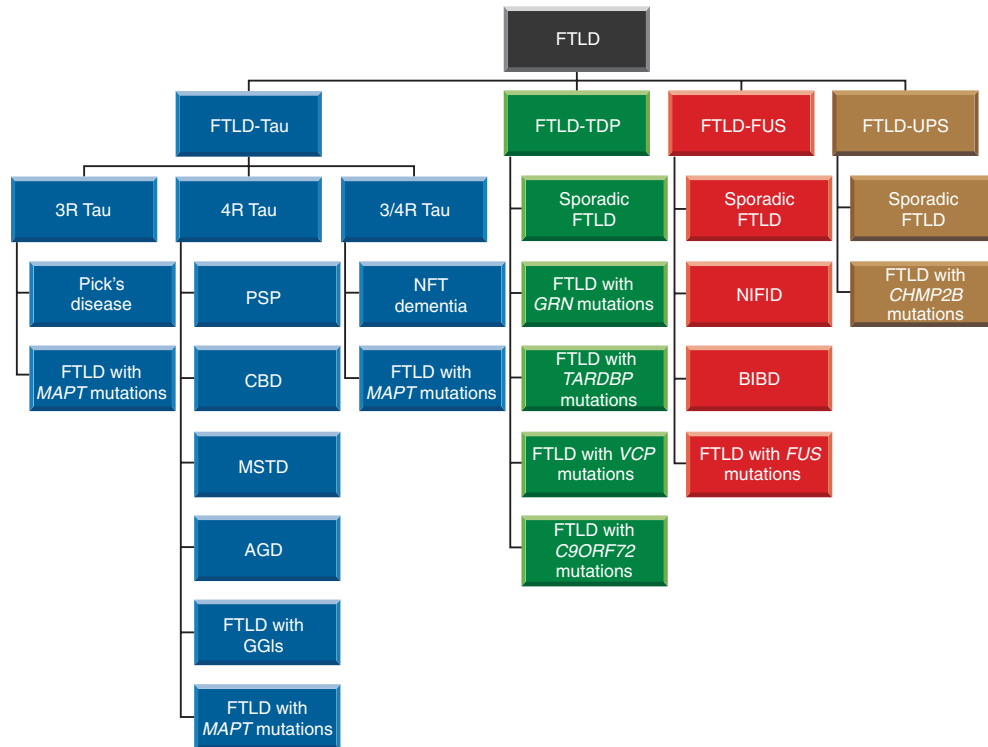
In 1986, the argyrophilic inclusions of Pick's disease were shown to be immunoreactive for hyperphosphorylated tau (Pollock et al. 1986), a normally soluble microtubule-associated protein that stabilizes microtubules and promotes microtubule assembly. It followed the finding that the intracellular inclusions of Alzheimer disease stain for hyperphosphorylated tau (Brion et al. 1985; Grundke-Iqbal et al. 1986). In adult human brain, six tau isoforms are expressed from a single *MAPT* gene through alternative mRNA splicing (Fig. 1A) (Goedert et al. 1989a,b). Three isoforms have three repeats each and three isoforms have four repeats each. By 1992, the paired helical filament of Alzheimer disease had been shown to be made of the six tau isoforms, each full-length and hyperphosphorylated (Goedert et al. 1988, 1992; Wischik et al. 1988; Lee et al. 1991). Filamentous tau inclusions were subsequently shown to be characteristic of many cases of FTDP-17 (Spillantini et al. 1996, 1998a).

Around 40% of patients with FTD show tau inclusions (Fig. 2). They include most cases of PNFA, ~45% of cases of bvFTD and some cases of SD (Piguet et al. 2011a). Most cases of LPA are characterized by focal Alzheimer disease pathology (A $\beta$  plaques and tau tangles), as are some cases of SD and PNFA (Mesulam et al. 2008; Rabinovici et al. 2008). Focal Alzheimer disease pathology accounts for ~25% of autopsy cases of PPA. A frontal variant of Alzheimer disease has also been described (Johnson et al. 1999). Tau inclusions are characteristic of Pick's disease, PSP, and CBD, which belongs to the CBS spectrum (Goedert and Spillantini 2006). They are not typical of FTD-MND, even though MND can occur in conjunction with FTD and tauopathy (Fu et al. 2010).





**Figure 1.** *MAPT* and the six tau isoforms expressed in adult human brain and mutations in *MAPT* in fronto-temporal dementia and parkinsonism linked to chromosome 17. (A) *MAPT* consists of 16 exons (E). Alternative mRNA splicing of E2 (red), E3 (green), and E10 (yellow) gives rise to six tau isoforms (352–441 amino acids). The constitutively spliced exons (E1, E4, E5, E7, E9, E11, E12, E13) are indicated in blue. E0, which is part of the promoter, and E14 are noncoding (white). E6 and E8 (violet) are not transcribed in human brain. E4a (orange) is only expressed in the peripheral nervous system. The repeats of tau (R1–R4) are shown, with three isoforms having four repeats each (4R) and three isoforms having three repeats each (3R). Each repeat is 31 or 32 amino acids in length. (B) Shown are 39 coding region mutations in E1, E9, E10, E11, E12, and E13 as well as seven intronic mutations flanking E10.

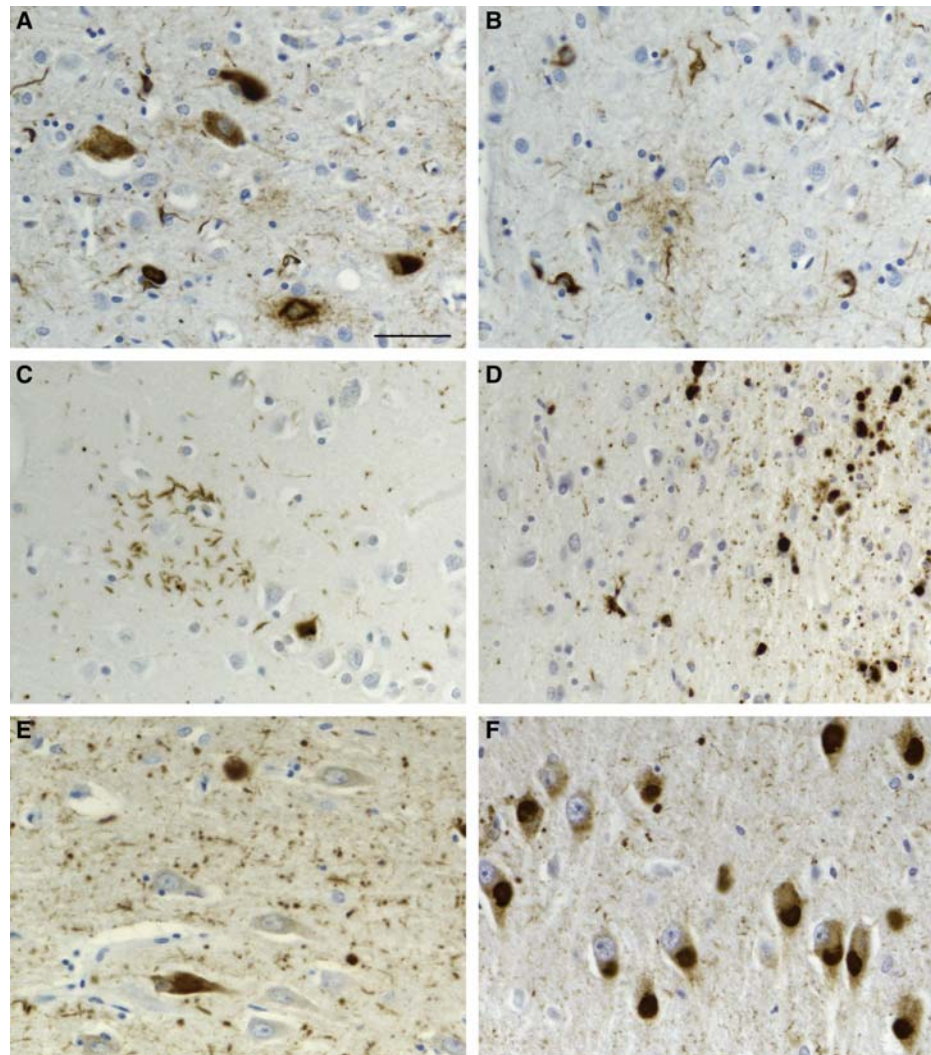


**Figure 2.** Frontotemporal lobar degeneration (FTLD) molecular classification. Four subtypes (FTLD-Tau, FTLD-TDP, FTLD-FUS, and FTLD-UPS) can be distinguished, based on what is known about the major components that make up the pathological deposits (Tau protein, TDP-43, FUS and unknown protein). FTLD-Tau and FTLD-TDP are more common than FTLD-FUS and FTLD-UPS. Tau deposits are made of either three-repeat (3R), four-repeat (4R) or all six (3/4R) brain isoforms of tau. Together, FTLD-TDP, FTLD-FUS, and FTLD-UPS make up FTLD-U, which is characterized by the presence of tau-negative, ubiquitin-positive inclusions. In some cases of FTLD-U, the ubiquitinated protein is unknown; they are classified as FTLD-UPS, to indicate that the inclusions can currently only be identified by markers of the ubiquitin-proteasome system. Abbreviations: PSP, progressive supranuclear palsy; CBD, corticobasal degeneration; MSTD, multiple system tauopathy with presenile dementia; AGD, argyrophilic grain disease; GGI, globular glial inclusion; NIFID, neuronal intermediate filament inclusion disease; BIBD, basophilic inclusion body disease; UPS, ubiquitin-proteasome system.

The anatomical distribution of pathology rather than its molecular identity determines the nature of the clinical syndromes. In most cases of Alzheimer disease, the locus coeruleus, entorhinal cortex, and hippocampus are the initial targets of neurofibrillary pathology, with the neocortex becoming affected later (Braak and Del Tredici 2011). In Pick's disease, tau inclusions predominate in the cerebral cortex, resulting in FTD (Piguet et al. 2011b). In PSP, patients with Richardson's syndrome have a higher tau burden and a different distribution

of inclusions than patients with PSP-parkinsonism (Williams et al. 2007). The subthalamic nucleus, substantia nigra, and globus pallidus are the most affected brain regions. In CBD, apraxia, rigidity, dystonia, and frontal lobe signs reflect the presence of neuronal and glial tau deposits in brainstem, basal ganglia, and cerebral cortex (Feany and Dickson 1995).

The repeats of tau form the core of the filaments whose isoform composition varies between diseases. The assembly of four-repeat tau into filaments is characteristic of PSP,



**Figure 3.** FTLD-Tau. Inclusions in progressive supranuclear palsy (A,B), corticobasal degeneration (C), white matter tauopathy with globular glial inclusions (D), argyrophilic grain disease (E), and Pick's disease (F). Progressive supranuclear palsy, corticobasal degeneration, white matter tauopathy with globular glial inclusions and argyrophilic grain disease are four-repeat tauopathies with abundant neuronal and glial tau filaments. Pick's disease is a three-repeat tauopathy with abundant neuronal tau filaments. Scale bar, 50  $\mu$ m.

CBD, and many cases of FTDP-17 (Fig. 3). It is also typical of argyrophilic grain disease and white matter tauopathy with globular glial inclusions, which belong to the FTD spectrum. A combination of neuronal and glial tau pathology is in evidence, with the glial pathology predominating in white matter tauopathy with globular glial inclusions (Kovacs et al. 2008).

In contrast, in Pick's disease and some cases of FTDP-17, three-repeat tau predominates in the neuronal inclusions (Fig. 3), whereas in Alzheimer disease, other diseases with extracellular deposits, Guam Parkinsonism-dementia complex, tangle-only dementia, and some cases of FTDP-17, both three- and four-repeat tau isoforms make up the neurofibrillary lesions.



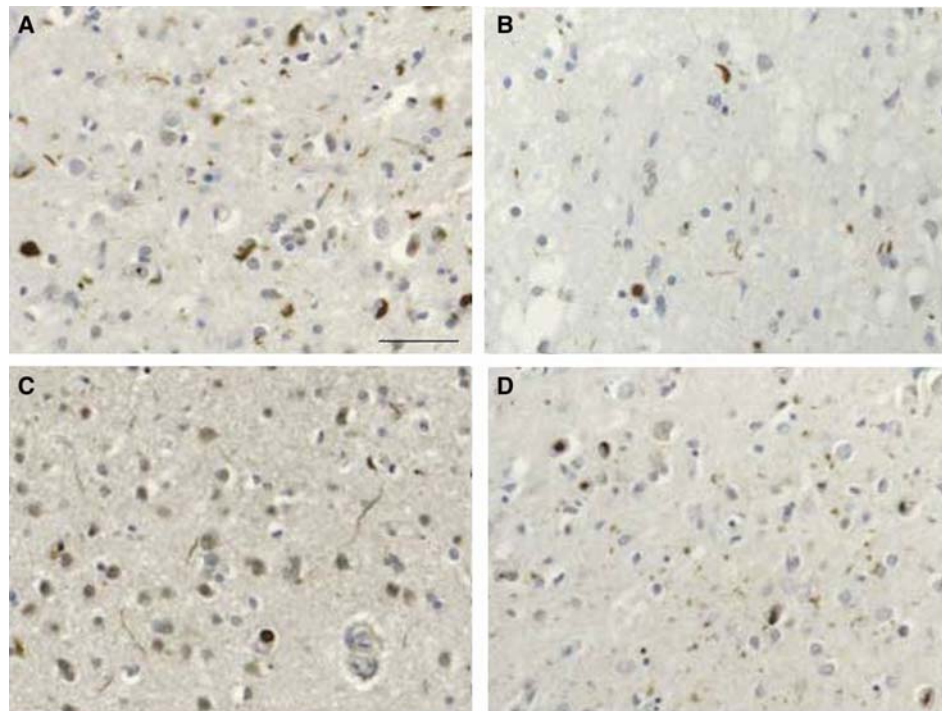
Distinct sets of tau isoforms in different neurodegenerative diseases and the presence of morphologically distinct filaments have led to the suggestion that self-propagating conformers of tau may exist (Goedert et al. 2010), akin to the prion strains accounting for the conformational variability of PrP<sup>Sc</sup> (Colby and Prusiner 2011). In support, experimental evidence for the intercellular transfer of tau aggregates has been adduced (Clavaguera et al. 2009; Frost et al. 2009).

### FTLD-TDP

By 2006, most cases of FTLD were known to exhibit either tau-positive or tau-negative inclusions (FTLD-U) (Fig. 2). The latter were first described in patients with MND (Okamoto et al. 1991). Four histological subtypes (A–D)

of FTLD-U can be distinguished (Fig. 4) (Mackenzie et al. 2006, 2011; Sampathu et al. 2006). Type A is associated with bvFTD and PFNA, type B with bvFTD and FTD-MND, type C with SD, and type D with familial inclusion body myopathy with Paget's disease of the bone and frontotemporal dementia (IBMPFD).

In 2006, transactive response-DNA binding protein-43 (TDP-43) was identified as the major component of the inclusions in most cases of FTLD-U (Figs. 2 and 4), MND, and FTLD-MND (Arai et al. 2006; Neumann et al. 2006). Around 50% of patients with FTD have TDP-43 inclusions. They include most cases of SD, ~45% of cases of bvFTD, as well as some cases of PNFA, LPA, FTDP-17, CBS, and PSP (Pigué et al. 2011a). Most cases of FTD-MND belong to the TDP-43 proteinopathy group. In Alzheimer disease, TDP-43 deposits



**Figure 4.** FTLD-TDP: Histological subtypes (A–D). Type A is characterized by abundant TDP-43-immunoreactive compact neuronal cytoplasmic inclusions and short dystrophic neurites, often with neuronal intranuclear inclusions (A); type B by abundant compact and granular cytoplasmic inclusions (B); type C by abundant long dystrophic neurites (C); and type D by abundant lentiform neuronal intranuclear inclusions and many short dystrophic neurites (D). Scale bar, 50  $\mu$ m.



M. Goedert et al.

are found in a minority of cases in conjunction with the characteristic plaques and tangles (Amador-Ortiz et al. 2007).

TDP-43 is a ubiquitously expressed 414 amino acid RNA-binding protein of the heterogeneous nuclear ribonucleoprotein (hnRNP) family with two RNA recognition motifs, nuclear localization and export signals, and a carboxy-terminal glycine-rich region. The glycine-rich region is predicted to show a prion domain, based on an algorithm that identifies yeast prion domains (Cushman et al. 2010). TDP-43 binds to noncoding RNAs, introns, and the 3'-untranslated regions of mRNAs, indicating a role in the integration of gene regulation. It functions as a transcriptional repressor and splicing modulator. UV-cross-linking and immunoprecipitation analysis has shown that TDP-43 has thousands of potential targets, with a preference for long clusters of UG-rich intronic sequences (Polymenidou et al. 2011; Tollerey et al. 2011). It binds to ~30% of the mouse transcriptome. TDP-43 negatively regulates its own mRNA and protein through binding to a long UG-rich region in its 3'-untranslated region (Ayala et al. 2011). It is predominantly nuclear, even though it normally shuttles between nucleus and cytoplasm. In FTD, TDP-43 it is found mainly in the cytoplasm in a hyperphosphorylated, ubiquitinated, and truncated form (Hasegawa et al. 2008).

### FTLD-FUS

Most cases of FTLD-U show TDP-43 inclusions. In 2009, inclusions made of fused in sarcoma (FUS) were shown to account for the bulk of TDP-43-negative FTLD-U (Fig. 2) (Neumann et al. 2009a), following the discovery that mutations in *FUS* cause familial forms of ALS (Kwiatkowski et al. 2009; Vance et al. 2009). Less than 10% of cases of FTLD have FUS inclusions. They include atypical cases of FTLD-U, basophilic inclusion body disease, and neuronal intermediate filament inclusion body disease (Munoz et al. 2009; Neumann et al. 2009a,b). Neuroimaging of FTLD-FUS shows atrophy of frontoinsular and cingulate cortex, and of the head of the caudate nucleus (Josephs et al. 2010;

Seelaar et al. 2010). FTD-FUS should be suspected when disease onset is before 40 years of age, in the absence of a family history of FTD, and the presence of caudate atrophy. This is reminiscent of a case from the early literature (Bonfiglioglio 1938). The existence of cases of PPA with FUS inclusions remains to be demonstrated.

FUS is a widely expressed 526 amino acid protein with an amino-terminal region rich in QGSY residues, a glycine-rich region, an RNA recognition motif, two RGG domains, and a zinc finger motif. Like TDP-43, it is a DNA/RNA-binding protein that is involved in transcriptional and translational regulation, as well as in mRNA splicing and transport. The predicted prion domain of FUS resides in the amino-terminal region (Cushman et al. 2010). In normal brain, FUS is concentrated in the nucleus, with smaller amounts in the cytoplasm. In FTD, the ability of FUS to shuttle to the nucleus is impaired, resulting in its cytoplasmic accumulation. FUS inclusions contain the full-length protein (Neumann et al. 2009a). Staining for TDP-43 and FUS appears to be mutually exclusive, suggestive of distinct subtypes of FTLD-U.

### FTLD-UPS

FTLD-TDP and FTLD-FUS account for the majority of cases of FTLD-U. Additional forms remain to be discovered, because inclusions that are negative for TDP-43 and FUS, but positive for components of the ubiquitin-proteasome system (UPS), have been described (Fig. 2) (Holm et al. 2009).

### GENETICS OF FRONTOTEMPORAL DEMENTIA

Dominantly inherited FTD is caused by mutations in seven genes. Mutations in *MAPT* (Hutton et al. 1998; Poorkaj et al. 1998; Spillantini et al. 1998b), chromosome 9 open reading frame 72 (*C9ORF72*) (DeJesus-Hernandez et al. 2011; Renton et al. 2011), and progranulin (*GRN*) (Baker et al. 2006; Cruets et al. 2006) are the most common. The other four genes are *TARDBP* (Benajiba et al. 2009; Kovacs et al. 2009), *FUS*

(Ticozzi et al. 2009), valosin-containing protein (VCP) (Watts et al. 2004), and charged multivesicular body protein 2B (*CHMP2B*) (Skibinski et al. 2005).

### Mutations in *MAPT*

Mutations in *MAPT* account for ~5% of cases of FTD and are believed to cause disease through a gain of toxic function mechanism. Most mutations are located in exons 9–12 (which encode the repeats) and the adjacent introns (Fig. 1B). It remains to be determined whether the amino acid changes in codon 5 of exon 1 are pathogenic. Mutations fall into two largely nonoverlapping groups: those with a primary effect at the protein level and those influencing the alternative splicing of tau pre-mRNA. Mutations acting at the protein level change or delete single amino acids in tau. This reduces the ability of tau to interact with microtubules, suggesting that this interaction is crucial for preventing the self-assembly of tau (Hasegawa et al. 1998). Some mutations also promote the assembly of tau into filaments (Goedert et al. 1999; Nacharaju et al. 1999). Mutations having their primary effect at the RNA level are intronic or exonic and increase the alternative mRNA splicing of exon 10. This changes the ratio of 3- to 4-repeat isoforms, resulting in the relative overproduction of 4-repeat tau and the formation of filamentous inclusions made of 4-repeat tau.

Cases with *MAPT* mutations show abundant filamentous inclusions made of hyperphosphorylated tau in either nerve cells or in both nerve cells and glial cells. Clinical and neuropathological phenotypes similar or identical to those of Pick's disease, PSP, CBD, and argyrophilic grain disease have been described. A given mutation can lead to different clinical syndromes in an individual family. Thus, mutation P301S in exon 10 of *MAPT* caused bvFTD in a father and CBD in his son (Bugiani et al. 1999), supporting the view that FTD and CBS are part of the same disease spectrum (Kertesz et al. 2000).

Haplotypes H1 and H2 characterize *MAPT* in populations of European descent. They result

from a 900 kb inversion/noninversion (H1/H2) polymorphism (Stefansson et al. 2005). Inheritance of the H1 haplotype is a risk factor for PSP and CBD (Williams and Lees 2009). This was confirmed in a genome-wide association study of PSP, which also implicated proteins involved in vesicle traffic, the unfolded protein response and the innate immune system (Höglinger et al. 2011). Heterozygous microdeletions in the chromosomal region, which defines the H1 and H2 haplotypes, give rise to a clinical phenotype consisting of mental retardation, hypotonia, and a characteristic face (Koolen et al. 2006; Sharp et al. 2006; Shaw-Smith et al. 2006). Besides *MAPT*, the deleted region comprises five additional genes [corticotrophin-releasing hormone receptor 1 (*CRHR1*), intramembrane protease 5 (*IMP5*), *NP 689679.1*, *NP 787078.1*, and *KIAA1267*]. Deletions occur on the H2 haplotype through low-copy repeat-mediated nonallelic homologous recombination. An association has also been described between the H1 haplotype and idiopathic Parkinson's disease (Pastor et al. 2000), a disease without tau inclusions. The elevated risk of PSP and CBD conferred by the H1 haplotype appears to promote *MAPT* transcription and incorporation of exon 10, resulting in increased levels of four-repeat tau (Caffrey et al. 2006).

### Mutations in *C9ORF72*

The cause of chromosome 9p21-linked FTD-MND has been identified as a hexanucleotide (GGGGCC) expansion in the noncoding region of *C9ORF72*, a gene that encodes a protein of unknown function (DeJesus-Hernandez et al. 2011; Renton et al. 2011). The hexanucleotide expansion leads to the loss of an alternatively spliced transcript and the formation of nuclear RNA foci. The latter may be toxic. The repeat expansion in *C9ORF72* is also a common cause of isolated FTD and MND.

### Mutations in *GRN*

Mutations in *GRN* account for ~5% of cases of FTD and cause disease by a loss of function mechanism. Progranulin is a 593 amino acid

glycoprotein consisting of 7.5 tandem repeats of a 12-cysteine granulin motif. Although its function is only incompletely understood, progranulin may be a physiological antagonist of tumor necrosis  $\alpha$  signaling (Tang et al. 2011). It has been reported to act on nerve cells by binding to sortilin following release from activated microglial cells (Hu et al. 2010). Mutations in *GRN* include gene deletions, as well as nonsense, frameshift, and splice-site mutations that cause premature termination, creating null alleles with the mutant RNAs being degraded by nonsense-mediated decay (Van Swieten and Heutink 2008). Known mutations result in haploinsufficiency, implying that progranulin is critical for the survival of neurons in adult brain. Reduced levels of plasma progranulin have been used to identify mutation carriers (Ghidoni et al. 2008). Mutations in *GRN* cause diseases belonging to the whole spectrum of FTD, with a predominance of bvFTD and PNEA (Yu et al. 2010). Parietal deficits and CBS have been observed (Spina et al. 2007). This is reflected in frontotemporoparietal cortical atrophy. Cases with *GRN* mutations show type A TDP-43 inclusions (Mackenzie et al. 2006), showing that a reduction in progranulin levels causes the accumulation of TDP-43. Unlike *TARDBP* mutations, mutations in *GRN* do not appear to cause MND. In a genome-wide study of FTLD-TDP, significant association was detected with three single nucleotide polymorphisms in the transmembrane protein 106B locus (*TMEM106B*) (Van Deerlin et al. 2010). It was most significant in patients with *GRN* mutations.

#### Mutations in *TARDBP*

Mutations in *TARDBP* are mostly associated with inherited forms of MND, consistent with the presence of TDP-43 inclusions in upper and lower motor neurons in patients with the disease (Gitcho et al. 2008; Sreedharan et al. 2008; Yokoseki et al. 2008). *TARDBP* mutations have also been described in two patients with bvFTD and SD who went on to develop MND (Benajiba et al. 2009). Histopathological changes were not documented. One patient

with a K263E change in *TARDBP* developed FTD, supranuclear palsy, and chorea, in the absence of MND. Abundant neuronal and glial TDP-43 deposits were in evidence, especially in brainstem and subcortical nuclei (Kovacs et al. 2009). The mechanisms by which mutations in *TARDBP* cause neurodegeneration are unclear. Pathological assembly is associated with a marked reduction in nuclear TDP-43 staining (Neumann et al. 2006) and the cytoplasmic accumulation of TDP-43 is believed to be an early event (Giordana et al. 2010). A combination of loss of function and gain of toxic function mechanisms may be at play. Wild-type TDP-43 is prone to aggregation and disease-causing mutations increase its aggregation and toxicity (Johnson et al. 2009). Many disease-causing mutations are located in the carboxy-terminal domain of TDP-43.

#### Mutations in *FUS*

Mutations in *FUS* cause inherited forms of MND (Kwiatkowski et al. 2009; Vance et al. 2009). Patients have *FUS* inclusions in spinal cord and cerebral cortex. Cases of FTD and/or FTD-MND may also be caused by mutations in *FUS* (Ticozzi et al. 2009), but larger clinicopathological series must be awaited. Like mutant TDP-43, mutant *FUS* accumulates in the cytoplasm, where it is found in stress granules (Dormann et al. 2010; Nishimoto et al. 2010). Wild-type *FUS* is prone to aggregation, but disease-causing mutations do not increase its aggregation or toxicity (Sun et al. 2011). The mutations appear to cause cytoplasmic mislocalization instead. Although several disease-causing mutations are present in its amino-terminal region, most mutations are located in the carboxy-terminus of *FUS*.

#### Mutations in *VCP*

Mutations in *VCP* cause IBMPFD through what appears to be a gain of function (Watts et al. 2004), possibly as the result of a dominant negative effect. IBMPFD affects skeletal muscle, bone, and nervous system, with dementia developing in  $\sim 30\%$  of patients. It is characterized





by the presence of type D TDP-43 inclusions (Neumann et al. 2007). Some missense mutations in *VCP* cause inherited MND (Johnson et al. 2010) and motor neuron abnormalities are present in many patients with IBMPFD. Furthermore, a missense mutation in *vacuolar protein sorting 54 (Vps54)*, the homolog of *VCP*, causes motor neuron degeneration in the wobbler mouse, a model of MND (Schmitt-John et al. 2005). *VCP* belongs to the type II AAA<sup>+</sup> (ATPases associated with a variety of activities) family and takes part in multiple cellular processes, including protein quality control, nuclear functions, and the regulation of membrane dynamics. It extracts ubiquitinated proteins from complexes, so that they can be degraded by the proteasome. *VCP* promotes autophagic protein degradation, with disease-causing mutations giving rise to defective autophagosome maturation (Ju and Weihl 2010). Transgenic mice expressing mutant *VCP* show many characteristics of IBMPFD, including involvement of skeletal muscle, bone and brain, and show increased activation of NF- $\kappa$ B signaling (Custer et al. 2010). In the brain of these mice, TDP-43 is redistributed from the nucleus to the cytoplasm, in the absence of nuclear inclusions. In a *Drosophila* model of IBMPFD, a genetic screen has identified TBPH, the fly ortholog of TDP-43, as one of three RNA-binding proteins that dominantly suppress degeneration (Ritson et al. 2010). In this model, *VCP* mutations lead to the redistribution of TDP-43 to the cytoplasm.

### Mutations in *CHMP2B*

Mutations in *CHMP2B* appear to cause disease through a gain of toxic function mechanism (Skibinski et al. 2005). The early signs are those of bvFTD, with extrapyramidal symptoms developing later, resulting in a clinical picture of CBS (Gydesen et al. 2002). In a Danish family with a truncating *CHMP2B* mutation, the intracytoplasmic inclusions are ubiquitin-positive, but negative for TDP-43 and FUS (Holm et al. 2009). *CHMP2B* is a component of the endosomal-sorting complex required for transport-III (ESCRT-III), which is involved in the

degradation of proteins in the endocytic and autophagic pathways. A disruption of these processes results in the accumulation of autophagosomes, possibly leading to FTD (Lee et al. 2007).

### IMPLICATIONS FOR UNDERSTANDING ALZHEIMER DISEASE

For a long time, tau inclusions were believed by many to be epiphenomena of little relevance. Reasons underlying this negative stance were the absence of genetic evidence linking dysfunction of tau to neurodegeneration and the presence of tau pathology in diseases other than Alzheimer disease. Things changed with the identification of mutations in *MAPT* in cases of FTDP-17 with filamentous tau pathology, establishing that dysfunction of tau is sufficient to cause neurodegeneration and dementia (Hutton et al. 1998; Poorkaj et al. 1998; Spillantini et al. 1998b). Thus, a pathway leading from soluble to insoluble, filamentous tau is central to the neurodegenerative process in the human tauopathies. It is therefore important to understand the mechanisms underlying tau aggregation and its downstream consequences for cell function. With the benefit of hindsight, it is clear that Alzheimer's description of silver-positive inclusions in cases with either presenile dementia or lobar cortical atrophy marked the beginning of the tauopathy field.

The crucial importance of FTDP-17T is that it proves that mutations in *MAPT* can lead to neurofibrillary assembly, neurodegeneration and dementia, in the absence of A $\beta$  amyloid deposits. The morphologies of tau filaments observed in the various forms of FTDP-17T vary (Crowther and Goedert 2000). Some mutations, such as V337M and R406W, produce filaments that appear identical to the paired helical and straight filaments of Alzheimer disease (Spillantini et al. 1996; Reed et al. 1997; Hutton et al. 1998; Poorkaj et al. 1998). All six tau isoforms are affected by the mutations and are incorporated into the filaments, which give rise to a pattern of tau bands on SDS-PAGE identical to that seen in Alzheimer disease. Mutation G389R also affects all six tau isoforms



and the majority of filaments resemble the straight filaments of Alzheimer disease (Murrell et al. 1999), despite the presence of Pick-like bodies by light microscopy. In contrast, in the case of mutations that increase the splicing of exon 10, the filaments appear as irregularly twisted ribbons, which are made of tau isoforms with four repeats (Spillantini et al. 1997). In the case of mutation P301L in exon 10, which affects only four-repeat tau isoforms, the majority of filaments consists of narrow, irregularly twisted ribbons, with a smaller number of straight filaments (Spillantini et al. 1998c).

Unlike Alzheimer disease and several other neurodegenerative diseases with tau inclusions, most cases of FTD lack extracellular deposits. However, focal Alzheimer disease pathology is diagnostic of a significant proportion of cases of PPA. Crossing mice transgenic for human mutant amyloid precursor protein with mice transgenic for human mutant tau results in increased tau deposition in some brain regions (Lewis et al. 2001). Similarly, in mice transgenic for the Danish mutant form of human BRI2 and mutant tau, the extracellular deposition of Dan-amyloid promotes tau phosphorylation and aggregation (Coomaraswamy et al. 2010). Phosphorylation of tau by GSK3 $\beta$  and AMP-activated protein kinase is a potential mechanism. This is consistent with the coexistence of extracellular amyloid deposits and intraneuronal tau inclusions in Alzheimer disease, familial British and Danish dementias, and in some diseases caused by mutations in the prion protein gene (Ghetti et al. 1994; Vidal et al. 2004; Goedert and Spillantini 2006). It suggests that extracellular amyloid deposits with a certain conformation trigger the intraneuronal assembly of tau into filaments.

Tau is required for A $\beta$  toxicity in experimental models (Roberson et al. 2007). The absence of A $\beta$  toxicity in mice lacking *MAPT* may result from a reduction in excitotoxicity, because of the decreased dendritic localization of the tyrosine kinase Fyn, resulting in hypophosphorylation of the NMDA receptor and a reduced interaction with postsynaptic density protein-95 (Ittner et al. 2010). Haploinsufficiency of p73, a member of the p53 protein

family, has been found to be associated with the formation of tau aggregates in nerve cells and to potentiate A $\beta$  toxicity, possibly through the activation of stress-activated protein kinases (Wetzel et al. 2008).

The intraneuronal pathology of Alzheimer disease may originate in a single cell and become self-sustaining, irrespective of upstream factors. Thus, injection of sonicated brain extract from mice with abundant tau inclusions into the cerebral cortex and hippocampus of transgenic mice lacking inclusions induces the assembly of human wild-type tau into filaments and leads to the spreading of pathology from the injection sites to neighboring brain regions (Clavaguera et al. 2009). Injection of brain extract immunodepleted of tau or divided into soluble and insoluble fractions shows that insoluble tau induces aggregation, in the absence of obvious signs of neurodegeneration. Parallel work has shown the transfer of aggregated tau between transfected cells (Frost et al. 2009). It thus appears that the tau species responsible for transmission and toxicity are not identical. An uncoupling of prion infective titre and neurotoxicity has been described (Sandberg et al. 2011).

Although tau inclusions form in many neurodegenerative diseases, their relevance for neurotoxicity remains a subject for debate. Studies using transgenic mice overexpressing human mutant tau in a conditional manner have reported a dissociation between tangle formation and nerve cell death (Santacruz et al. 2005). It appears that soluble hyperphosphorylated tau can contribute to nerve cell dysfunction prior to assembly into filaments. This is reminiscent of *Drosophila* and *Caenorhabditis elegans* lines expressing human wild-type or mutant tau, in which nerve cell loss and a reduced lifespan are observed, in the apparent absence of tau filaments (Wittmann et al. 2001; Kraemer et al. 2003). In genetic modifier screens in *Drosophila*, an increase in kinase activity enhanced tau toxicity, whereas an increase in phosphatase activity was beneficial (Feany et al. 2010). Activation of oxidative defences was also beneficial. In *C. elegans*, loss of *Sut-2* (suppressor of tau pathology-2),



eliminated the toxic effects of human mutant tau, possibly via an increase in autophagic clearance (Guthrie et al. 2009).

The main goal behind work on tauopathies is to transform the treatment of common neurodegenerative diseases through an understanding of the underlying molecular pathways. There is an unmet need for mechanism-based therapies of Alzheimer disease. Tau binds to microtubules and boosting this interaction may be beneficial. This may be achieved through a reduction of the hyperphosphorylation of tau (Le Corre et al. 2006). In Alzheimer disease, tau assembles into paired helical and straight filaments. The assembly pathway is being defined and inhibitors of aggregation are being developed (Pickhardt et al. 2005; Taniguchi et al. 2005). Immunotherapy has been shown to clear tau aggregates from transgenic mouse brain and to reduce functional impairment (Asuni et al. 2007). Because aggregation is a concentration-dependent process, a reduction in production and/or increased clearance of tau are also potential targets (Morris et al. 2011).

## REFERENCES

- Alladi S, Xuereb J, Bak T, Nestor P, Knibb J, Patterson K, Hodges JR. 2007. Focal cortical presentations of Alzheimer's disease. *Brain* **130**: 2636–2645.
- Alzheimer A. 1907. Über eine eigenartige Erkrankung der Hirnrinde. *Allg Z Psychiat* **64**: 146–148.
- Alzheimer A. 1911. Über eigenartige Krankheitsfälle des späteren Alters. *Z ges Neurol Psychiat* **4**: 356–385.
- Amador-Ortiz C, Lin WL, Ahmed Z, Personett D, Davies P, Duara R, Graff-Radford NR, Hutton ML, Dickson DW. 2007. TDP-43 immunoreactivity in hippocampal sclerosis and Alzheimer's disease. *Ann Neurol* **61**: 435–445.
- Arai T, Hasegawa M, Akiyama H, Ikeda K, Nonaka T, Mori H, Mann D, Tsuchiya K, Yoshida M, Hashizume Y, et al. 2006. TDP-43 is a component of ubiquitin-positive tau-negative inclusions in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Biochem Biophys Res Commun* **351**: 602–611.
- Asuni AA, Boutajangout A, Quatrain D, Sigurdsson EM. 2007. Immunotherapy targeting pathological tau conformers in a transgenic mouse model reduces brain pathology associated with functional improvements. *J Neurosci* **27**: 9115–9129.
- Ayala YM, De Conti L, Avendano-Vázquez SE, Dhir A, Romano M, D'Ambrogio A, Tollervey J, Ule J, Baralle M, Buratti E, et al. 2011. TDP-43 regulates its mRNA levels through a negative feedback loop. *EMBO J* **30**: 277–288.
- Baker M, Mackenzie IR, Pickering-Brown SM, Gass J, Rademakers R, Lindholm C, Snowden J, Adamson J, Sadvnick AD, Rollinson S, et al. 2006. Mutations in progranulin cause tau-negative frontotemporal dementia linked to chromosome 17. *Nature* **442**: 916–919.
- Benajiba L, Le Ber I, Camuzat A, Lacoste M, Thomas-Anterion C, Couratier P, Legallic S, Salachas F, Hannequin D, Decousus M, et al. 2009. *TARDBP* mutations in motoneuron disease with frontotemporal lobar degeneration. *Ann Neurol* **65**: 470–474.
- Bonfiglio F. 1938. Die umschriebene Atrophie der Basalganglien. *Z Neurol* **160**: 306–333.
- Braak H, Del Tredici K. 2011. The pathological process underlying Alzheimer's disease in individuals under thirty. *Acta Neuropathol* **121**: 171–181.
- Brion JP, Passareiro H, Nunez J, Flament-Durand J. 1985. Mise en évidence immunologique de la protéine tau au niveau des lésions de dégénérescence neurofibrillaire de la maladie d'Alzheimer. *Arch Biol* **95**: 229–235.
- Broca P. 1861. Perte de la parole, ramollissement chronique et destruction partielle du lobe antérieur gauche du cerveau. *Bull Soc Anthropol* **2**: 235–238.
- Brun A. 1987. Frontal lobe degeneration of non-Alzheimer type. I. Neuropathology. *Arch Gerontol Geriatr* **6**: 193–208.
- Bugiani O, Murrell JR, Giaccone G, Hasegawa M, Ghigo G, Tabaton M, Morbin M, Primavera A, Carella F, Solaro C, et al. 1999. Frontotemporal dementia and corticobasal degeneration in a family with a P301S mutation in *Tau*. *J Neuropathol Exp Neurol* **58**: 667–677.
- Caffrey TM, Joachim C, Paracchini S, Esiri MM, Wade-Martins R. 2006. Haplotype-specific expression of exon 10 at the human *MAPT* locus. *Hum Mol Genet* **15**: 3529–3537.
- Clavaguera F, Bolmont T, Crowther RA, Abramowski D, Frank S, Probst A, Fraser G, Stalder AK, Beibel M, Staufenbiel M, et al. 2009. Transmission and spreading of tauopathy in transgenic mouse brain. *Nat Cell Biol* **11**: 909–913.
- Colby DW, Prusiner SB. 2011. Prions. *Cold Spring Harb Perspect Biol* **3**: a006833.
- Constantinidis J, Richard J, Tissot R. 1974. Pick's disease. Histological and clinical correlations. *Eur Neurol* **11**: 208–217.
- Coomaraswamy J, Kilger E, Wöfling H, Schäfer C, Kaeser SA, Wegenast-Braun BM, Hefendehl JK, Wolburg H, Mazzella M, Ghiso J, et al. 2010. Modeling familial Danish dementia in mice supports the concept of the amyloid hypothesis of Alzheimer's disease. *Proc Natl Acad Sci* **107**: 7969–7974.
- Crowther RA, Goedert M. 2000. Abnormal tau-containing filaments in neurodegenerative diseases. *J Struct Biol* **130**: 271–279.
- Cruts M, Gijselink I, van der Zee J, Engelborghs S, Wils H, Pirici D, Rademakers R, Vandenberghe R, Dermaut B, Martin JJ, et al. 2006. Null mutations in progranulin cause ubiquitin-positive frontotemporal dementia linked to chromosome 17q21. *Nature* **442**: 920–924.
- Cushman M, Johnson BS, King OD, Gitler AD, Shorter J. 2010. Prion-like disorders: Blurring the divide between transmissibility and infectivity. *J Cell Sci* **123**: 1191–1201.



M. Goedert et al.

- Custer SK, Neumann M, Lu H, Wright AC, Taylor JP. 2010. Transgenic mice expressing mutant forms of VCP/p97 recapitulate the full spectrum of IBMPFD including degeneration in muscle, brain and bone. *Hum Mol Genet* **19**: 1741–1755.
- Déjerine J, Sériex P. 1897. Un cas de surdit  verbale pure termin e par aphasie sensorielle, suivi d'autopsie. *CR Acad Sci Paris* **49**: 1074–1077.
- DeJesus-Hernandez M, Mackenzie IR, Boeve BF, Boxer AL, Baker M, Rutherford NJ, Nicholson AM, Finch NA, Flynn H, Adamson J, et al. 2011. Expanded GGGGCC hexanucleotide repeat in noncoding region of *C9ORF72* causes chromosome 9p-linked FTD and ALS. *Neuron* doi: 10.1016/j.neuron.2011.09.011.
- Dormann D, Rodde R, Edbauer D, Bentmann E, Fischer I, Hruscha A, Than ME, Mackenzie IRA, Capell A, Schmid B, et al. 2010. ALS-associated fused in sarcoma (*FUS*) mutations disrupt transportin-mediated nuclear import. *EMBO J* **29**: 21841–21857.
- Feany MB. 2010. New approaches to the pathology and genetics of neurodegeneration. *Am J Pathol* **176**: 2058–2066.
- Feany MB, Dickson DW. 1995. Widespread cytoskeletal pathology characterizes corticobasal degeneration. *Am J Pathol* **146**: 1388–1396.
- Foster NL, Wilhelmsen K, Sima AA, Jones MZ, D'Amato CJ, Gilman S. 1997. Frontotemporal dementia and parkinsonism linked to chromosome 17: A consensus conference. *Ann Neurol* **41**: 706–715.
- Freud S. 1891. *Zur Auffassung der Aphasien*. Franz Deuticke, Leipzig und Wien.
- Frost B, Jacks RL, Diamond MI. 2009. Propagation of tau misfolding from the outside to the inside of a cell. *J Biol Chem* **284**: 12845–12852.
- Fu YJ, Nishihara Y, Kuroda S, Toyoshima Y, Ishihara T, Shinozaki M, Miyashita A, Piao YS, Tan CF, Tani T, et al. 2010. Sporadic four-repeat tauopathy with frontotemporal lobar degeneration, Parkinsonism, and motor neuron disease: A distinct neuropathological and biochemical disease entity. *Acta Neuropathol* **120**: 21–32.
- Galantucci S, Tartaglia MC, Wilson SM, Henry ML, Filippi M, Agosta F, Dronkers NF, Henry RG, Ogar JM, Miller BL, et al. 2011. White matter damage in primary progressive aphasia: A diffusion tensor tractography study. *Brain* **134**: 3011–3029.
- Galton CJ, Patterson K, Xuereb JH, Hodges JR. 2000. Atypical and typical presentations of Alzheimer's disease: A clinical, neuropsychological, neuroimaging and pathological study of 13 cases. *Brain* **123**: 484–498.
- Gans A. 1923. Betrachtungen  ber Art und Ausbreitung des krankhaften Prozesses in einem Fall von Pick'scher Atrophie des Stirnhirns. *Z Neurol* **80**: 10–28.
- Ghetti B, Tagliavini F, Giaccone G, Bugiani O, Frangione B, Farlow MR, Dlouhy SR. 1994. Familial Gerstmann-Str ussler-Scheinker disease with neurofibrillary tangles. *Mol Neurobiol* **8**: 41–48.
- Ghetti B, Wszolek ZW, Boeve BF, Spina S, Goedert M. 2011. Frontotemporal dementia and parkinsonism linked to chromosome 17. In *Neurodegeneration: The molecular pathology of dementia and movement disorders*, 2nd ed. (ed. Dickson D, Weller RO), pp. 110–134. Blackwell, Oxford, UK.
- Ghidoni R, Benussi L, Glionna M, Franzoni M, Binetti G. 2008. Low plasma progranulin levels predict progranulin mutations in frontotemporal lobar degeneration. *Neurology* **71**: 1235–1239.
- Giordana MT, Piccini M, Grifoni S, De Marco G, Vercellino M, Magistrello M, Pellerino A, Buccinna B, Lupino E, Rinaudo MT. 2010. TDP-43 redistribution is an early event in sporadic amyotrophic lateral sclerosis. *Brain Pathol* **20**: 351–360.
- Gitcho MA, Baloh RH, Chakraverty S, Mayo K, Norton JB, Levitch D, Hatanpaa KJ, White CL, Bigio EH, Caselli R, et al. 2008. TDP-43 A315 mutation in familial motor neuron disease. *Ann Neurol* **63**: 535–538.
- Goedert M, Spillantini MG. 2006. A century of Alzheimer's disease. *Science* **314**: 777–781.
- Goedert M, Wischik CM, Crowther RA, Walker JE, Klug A. 1988. Cloning and sequencing of the cDNA encoding a core protein of the paired helical filament of Alzheimer disease. *Proc Natl Acad Sci* **85**: 4051–4055.
- Goedert M, Spillantini MG, Potier MC, Ulrich J, Crowther RA. 1989a. Cloning and sequencing of the cDNA encoding an isoform of microtubule-associated protein tau containing four tandem repeats: Differential expression of tau protein mRNAs in human brain. *EMBO J* **8**: 393–399.
- Goedert M, Spillantini MG, Jakes R, Rutherford D, Crowther RA. 1989b. Multiple isoforms of human microtubule-associated protein tau: Sequences and localization in neurofibrillary tangles of Alzheimer's disease. *Neuron* **3**: 519–526.
- Goedert M, Spillantini MG, Cairns NJ, Crowther RA. 1992. Tau proteins of Alzheimer paired helical filaments: Abnormal phosphorylation of all six brain isoforms. *Neuron* **8**: 159–168.
- Goedert M, Jakes R, Crowther RA. 1999. Effects of frontotemporal dementia FTDP-17 mutations on heparin-induced assembly of tau filaments. *FEBS Lett* **450**: 306–311.
- Goedert M, Clavaguera F, Tolnay M. 2010. The propagation of prion-like protein inclusions in neurodegenerative diseases. *Trends Neurosci* **33**: 317–325.
- Gorno-Tempini ML, Murray RC, Rankin KP, Weiner MW, Miller BL. 2004a. Clinical, cognitive and anatomical evolution from nonfluent progressive aphasia to corticobasal syndrome: a case report. *Neurocase* **10**: 426–436.
- Gorno-Tempini ML, Dronkers NF, Rankin KP, Ogar JM, Phengrasamy L, Rosen HJ, Johnson JK, Weiner MW, Miller BL. 2004b. Cognition and anatomy in three variants of primary progressive aphasia. *Ann Neurol* **55**: 335–346.
- Gorno-Tempini ML, Brambati SM, Ginex V, Ogar J, Dronkers NF, Marcone A, Perani D, Garibotto V, Cappa SF, Miller BL. 2008. The logopenic/phonological variant of primary progressive aphasia. *Neurology* **71**: 1227–1234.
- Gorno-Tempini ML, Hillis AE, Weintraub S, Kertesz A, Mendez M, Cappa SF, Ogar JM, Rohrer JD, Black S, Boeve BF, et al. 2011. Classification of primary progressive aphasia and its variants. *Neurology* **76**: 1006–1014.
- Grossman M, Mickanin J, Onishi K, Hughes E, D'Esposito M, Ding XS, Alavi A, Reivich M. 1996. Progressive non-fluent aphasia: Language, cognitive, and PET measures



- contrasted with probable Alzheimer disease. *J Cogn Neurosci* **8**: 135–154.
- Grundke-Iqbal I, Iqbal K, Tung YC, Quinlan M, Wisniewski HM, Binder LI. 1986. Abnormal phosphorylation of the microtubule-associated protein tau in Alzheimer cytoskeletal pathology. *Proc Natl Acad Sci* **83**: 4913–4917.
- Grünthal E. 1930. Über ein Brüderpaar mit Pickscher Krankheit. *Z Neurol* **129**: 350–375.
- Gustafson L. 1987. Frontal lobe degeneration of non-Alzheimer type. II. Clinical picture and differential diagnosis. *Arch Gerontol Geriatr* **6**: 209–223.
- Guthrie CR, Schellenberg GD, Kraemer BC. 2009. SUT-2 potentiates tau-induced neurotoxicity in *Caenorhabditis elegans*. *Hum Mol Genet* **18**: 1825–1838.
- Gydesen S, Brown JM, Brun A, Chakrabarti L, Gade A, Johannsen P, Rossor M, Thusgaard T, Grove A, Yancopoulos D, et al. 2002. Chromosome 3 linked frontotemporal dementia (FTD-3). *Neurology* **59**: 1585–1594.
- Hasegawa M, Smith MJ, Goedert M. 1998. Tau proteins with FTDP-17 mutations have a reduced ability to promote microtubule assembly. *FEBS Lett* **437**: 207–210.
- Hasegawa M, Arai T, Nonaka T, Kametani F, Yoshida M, Hashizume Y, Beach TG, Buratti E, Baralle F, Morita M, et al. 2008. Phosphorylated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Ann Neurol* **64**: 60–70.
- Hodges JR, Patterson K. 2007. Semantic dementia: A unique clinicopathological syndrome. *Lancet Neurol* **6**: 1004–1014.
- Höglinger GU, Melhem NM, Dickson D, Sleiman PMA, Wang LS, Klei L, Rademakers R, de Silva R, Litvan I, Riley DC, et al. 2011. Common variants affect risk for the tauopathy progressive supranuclear palsy. *Nat Genet* **43**: 699–705.
- Holm IE, Isaacs AM, Mackenzie IRA. 2009. Absence of FUS-immunoreactive pathology in frontotemporal dementia linked to chromosome 3 (FTD-3) caused by mutation in the *CHMP2B* gene. *Acta Neuropathol* **118**: 719–720.
- Hu F, Padukkavidana T, Vaegter CB, Brady OA, Zheng Y, Mackenzie IR, Feldman HH, Nykjaer A, Strittmatter SM. 2010. Sortilin-mediated endocytosis determines levels of the frontotemporal dementia protein, progranulin. *Neuron* **68**: 654–667.
- Hutton M, Lendon CL, Rizzu M, Baker M, Froelich S, Houlden H, Pickering-Brown S, Chakraverty S, Isaacs A, Grover A, et al. 1998. Association of missense and 5'-splice-site mutations in *tau* with the inherited dementia FTDP-17. *Nature* **393**: 702–705.
- Ittner LM, Ke YD, Delerue F, Bi M, Gladbach A, Van Eersel J, Wölfing H, Chieng BC, Christie J, Napier IA, et al. 2010. Dendritic function of tau mediates amyloid- $\beta$  toxicity in Alzheimer's disease mouse models. *Cell* **142**: 387–397.
- Johnson JK, Head E, Kim R, Starr A, Cotman CW. 1999. Clinical and pathological evidence for a frontal variant of Alzheimer disease. *Arch Neurol* **56**: 1233–1239.
- Johnson BS, Snead D, Lee JJ, McCaffery MM, Shorter J, Gitler AD. 2009. TDP-43 is intrinsically aggregation-prone, and amyotrophic lateral sclerosis-linked mutations accelerate aggregation and toxicity. *J Biol Chem* **284**: 20329–20339.
- Johnson JO, Mandrioli J, Benatar M, Abramzon Y, Van Deerlin VM, Trojanowski JQ, Gibbs JR, Brunetti M, Gronka S, Wu J, et al. 2010. Exome sequencing reveals *VCP* mutations as a cause of familial ALS. *Neuron* **68**: 857–864.
- Josephs KA, Dickson DW. 2007. Frontotemporal lobar degeneration with upper motor neuron disease/primary lateral sclerosis. *Neurology* **69**: 1800–1801.
- Josephs KA, Duffy JR, Strand EA, Whitwell JL, Layton KF, Parisi JE, Hauser ME, Witte RJ, Boeve BF, Knopman DS, et al. 2006. Clinicopathological and imaging correlates of progressive aphasia and apraxia of speech. *Brain* **129**: 1385–1398.
- Josephs KA, Whitwell JL, Parisi JE, Petersen RC, Boeve BF, Jack CR, Dickson DW. 2010. Caudate atrophy on MRI is a characteristic feature of FTL-D-FUS. *Eur J Neurol* **17**: 969–975.
- Ju JS, Wehl CC. 2010. Inclusion body myopathy, Paget's disease of the bone and frontotemporal dementia: A disorder of autophagy. *Hum Mol Genet* **19**: R38–R45.
- Kertesz A, Hudson L, Mackenzie IRA, Munoz DG. 1994. The pathology and nosology of primary progressive aphasia. *Neurology* **44**: 2065–2072.
- Kertesz A, Martinez-Lage P, Davidson W, Munoz DG. 2000. The corticobasal degeneration syndrome overlaps progressive aphasia and frontotemporal dementia. *Neurology* **55**: 1368–1375.
- Koolen DA, Vissers LELM, Pfundt R, de Leeuw N, Knight SJL, Regan R, Kooy RF, Reyniers E, Romano C, Fichera M, et al. 2006. A new chromosome 17q21.31 microdeletion syndrome associated with a common inversion polymorphism. *Nat Genet* **38**: 999–1001.
- Kovacs GG, Majtenyi K, Spina S, Murrell JR, Gelpi E, Höftberger R, Fraser G, Crowther RA, Goedert M, Budka H, et al. 2008. White matter tauopathy with globular glial inclusions: A distinct sporadic frontotemporal lobar degeneration. *J Neuropathol Exp Neurol* **67**: 963–975.
- Kovacs GG, Murrell JR, Horvath S, Haraszti L, Majtenyi K, Molnar MJ, Budka H, Ghetti B, Spina S. 2009. *TARDBP* variation associated with frontotemporal dementia, supranuclear gaze palsy, and chorea. *Mov Disord* **24**: 1843–1847.
- Kraemer BC, Zhang B, Leverenz JB, Thomas JH, Trojanowski JQ, Schellenberg GD. 2003. Neurodegeneration and defective neurotransmission in a *Caenorhabditis elegans* model of tauopathy. *Proc Natl Acad Sci* **100**: 9980–9985.
- Kwiatkowski TJ, Bosco DA, LeClerc AL, Tamrazian E, Vandenberg CR, Russ C, Davis A, Gilchrist J, Kasarskis EJ, Munsat T, et al. 2009. Mutations in the *FUS/TLN1* gene on chromosome 16 cause familial amyotrophic lateral sclerosis. *Science* **323**: 1205–1211.
- Le Corre S, Klafki HW, Plesnila N, Hübinger G, Obermeier A, Sahagún H, Monse B, Seneci P, Lewis J, Eriksen J, et al. 2006. An inhibitor of tau hyperphosphorylation prevents severe motor impairments in tau transgenic mice. *Proc Natl Acad Sci* **103**: 9673–9678.
- Lee VMY, Balin BJ, Otvos L, Trojanowski JQ. 1991. A68—a major subunit of paired helical filaments and derivatized forms of normal tau. *Science* **251**: 675–678.
- Lee JA, Beigneux A, Tariq Ahmad S, Young SG, Gao FB. 2007. ESCRT-III dysfunction causes autophagosome





M. Goedert et al.

- accumulation and neurodegeneration. *Curr Biol* **17**: 1561–1567.
- Lewis J, Dickson DW, Lin WL, Chisholm L, Corral A, Jones G, Yen SH, Sahara N, Skipper L, Yager D, et al. 2001. Enhanced neurofibrillary degeneration in transgenic mice expressing mutant tau and APP. *Science* **293**: 1487–1491.
- Lippa CE, Cohen R, Smith TW, Drachman DA. 1991. Primary progressive aphasia with focal neuronal achromasia. *Neurology* **41**: 882–886.
- Litvan I, Agid Y, Calne D, Campbell G, Dubois B, Duvoisin RC, Goetz CG, Golbe LI, Grafman J, Growdon JH, et al. 1996. Clinical research criteria for the diagnosis of progressive supranuclear palsy (Steele-Richardson-Olszewski syndrome). *Neurology* **47**: 1–9.
- Lüers T, Spatz H. 1957. Pickische Krankheit. In *Handbuch der speziellen Anatomie und Histologie* (ed. Henke F, Lubarsch O), Vol. 13, pp. 614–715. Springer, Berlin.
- Mackenzie IRA, Baborie A, Pickering-Brown S, Du Plessis D, Jaros E, Perry RH, Neary D, Snowden JS, Mann DMA. 2006. Heterogeneity of ubiquitin pathology in frontotemporal lobar degeneration. Classification and relation to clinical phenotype. *Acta Neuropathol* **112**: 539–549.
- Mackenzie IRA, Neumann M, Baborie A, Sampathu DM, Du Plessis D, Jaros E, Perry RH, Trojanowski JQ, Mann DMA, Lee VMY. 2011. A harmonized classification system for FTLT-DTP pathology. *Acta Neuropathol* **122**: 111–113.
- Mesulam MM. 1982. Slowly progressive aphasia without generalized dementia. *Ann Neurol* **11**: 592–598.
- Mesulam MM. 1987. Primary progressive aphasia—differentiation from Alzheimer's disease. *Ann Neurol* **22**: 533–534.
- Mesulam MM. 2001. Primary progressive aphasia. *Ann Neurol* **49**: 425–432.
- Mesulam MM, Wicklund A, Johnson N, Rogalski E, Léger GC, Rademaker A, Weintraub S, Bigio EH. 2008. Alzheimer and frontotemporal pathology in subsets of primary progressive aphasia. *Ann Neurol* **63**: 709–719.
- Meyer A. 1929. Über eine der amyotrophischen Lateralsklerose nahestehende Erkrankung mit psychischen Störungen. *Z Neurol* **121**: 107–128.
- Migliaccio R, Agosta F, Rascovsky K, Karydas A, Bonasera S, Rabinovici GD, Miller BL, Gorno-Tempini ML. 2009. Clinical syndromes associated with posterior atrophy. *Neurology* **73**: 1571–1578.
- Morita K, Kaiya H, Ikeda T, Namba M. 1987. Presenile dementia combined with amyotrophy: A review of 34 Japanese cases. *Arch Gerontol Geriatr* **6**: 263–277.
- Morris M, Maeda S, Vossel K, Mucke L. 2011. The many faces of tau. *Neuron* **70**: 410–426.
- Mummery CJ, Patterson K, Wise RJS, Vandenberghe R, Price CJ, Hodges JR. 1999. Disrupted temporal lobe connections in semantic dementia. *Brain* **122**: 61–73.
- Munoz DG, Neumann M, Kusaka H, Yokota O, Ishihara K, Terada S, Kuroda S, Mackenzie IR. 2009. FUS pathology in basophilic inclusion body disease. *Acta Neuropathol* **118**: 617–627.
- Murrell JR, Spillantini MG, Zolo P, Guazzelli M, Smith MJ, Hasegawa M, Redi F, Crowther RA, Pietrini P, Ghetti B, et al. 1999. Tau gene mutation G389R causes a tauopathy with abundant Pick body-like inclusions and axonal deposits. *J Neuropathol Exp Neurol* **58**: 1207–1226.
- Nacharaju P, Lewis J, Easson C, Yen S, Hackett J, Hutton M, Yen SH. 1999. Accelerated filament formation from tau protein with specific FTDP-17 mutations. *FEBS Lett* **447**: 195–199.
- Neary D, Snowden JS, Northen B, Goulding P. 1988. Dementia of frontal type. *J Neurol Neurosurg Psychiatry* **51**: 353–361.
- Neary D, Snowden JS, Gustafson L, Passant U, Stuss D, Black S, Freedman M, Kertesz A, Robert PH, Albert M, et al. 2003. Progressive non-fluent aphasia is associated with hypometabolism centred on the left anterior insula. *Brain* **126**: 2406–2418.
- Nestor PJ, Graham NL, Fryer TD, Williams GB, Patterson K, Hodges JR. 2003. Progressive non-fluent aphasia is associated with hypometabolism centred on the left anterior insula. *Brain* **126**: 2406–2418.
- Neumann M, Sampathu DM, Kwong LK, Truax AC, Micsenyi MC, Chou TT, Bruce J, Schuck T, Grossman M, Clark CM, et al. 2006. Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science* **314**: 130–133.
- Neumann M, Mackenzie IR, Cairns NJ, Boyer PJ, Markesbery WR, Smith CD, Taylor JP, Kretschmar HA, Kimonis VE, Forman MS. 2007. TDP-43 in the ubiquitin pathology of frontotemporal dementia with VCP gene mutations. *J Neuropathol Exp Neurol* **66**: 152–157.
- Neumann M, Rademakers R, Roeber S, Baker M, Kretschmar HA, Mackenzie IRA. 2009a. A new subtype of frontotemporal lobar degeneration with FUS pathology. *Brain* **132**: 2922–2931.
- Neumann A, Roeber S, Kretschmar HA, Rademakers R, Baker M, Mackenzie IRA. 2009b. Abundant FUS-immunoreactive pathology in neuronal intermediate filament inclusion disease. *Acta Neuropathol* **118**: 605–616.
- Nishimoto Y, Ito D, Yagi T, Nihei Y, Tsunoda Y, Suzuki N. 2010. Characterization of alternative isoforms and inclusion body of the TAR DNA-binding protein-43. *J Biol Chem* **285**: 608–619.
- Okamoto K, Hirai S, Yamazaki T, Sun X, Nakazato Y. 1991. New ubiquitin-positive intraneuronal inclusions in the extra-motor cortices in patients with amyotrophic lateral sclerosis. *Neurosci Lett* **129**: 233–236.
- Onari K, Spatz H. 1926. Anatomische Beiträge zur Lehre von der Pickschen umschriebenen Grosshirnrinden-Atrophie ("Pickische Krankheit"). *Z Neurol* **101**: 470–511.
- Pastor P, Ezquerro M, Munoz E, Marti MJ, Blesa R, Tolosa E, Oliva R. 2000. Significant association between the tau gene A0/A0 genotype and Parkinson's disease. *Ann Neurol* **47**: 242–245.
- Pick A. 1892. Ueber die Beziehungen der senilen Hirnatrophie zur Aphasie. *Prager med Wschr* **17**: 165–167.
- Pick A. 1901. Senile Hirnatrophie als Grundlage von Herderscheinungen. *Wiener klin Wschr* **14**: 403–404.
- Pick A. 1904. Zur Symptomatologie der linksseitigen Schläfenlappenatrophie. *Mschr Psychiat Neurol* **16**: 378–388.
- Pick A. 1906. Über einen weiteren Symptomencomplex im Rahmen der Dementia senilis, bedingt durch umschriebene stärkere Hirnatrophie (gemischte Apraxie). *Mschr Psychiat Neurol* **19**: 97–108.





- Pickhardt M, Gazova Z, von Bergen M, Khlistunova I, Wang Y, Hascher A, Mandelkow EM, Biernat J, Mandelkow E. 2005. Anthraquinones inhibit tau aggregation and dissolve Alzheimer paired helical filaments in vitro and in cells. *J Biol Chem* **280**: 3628–3635.
- Piguot O, Hornberger M, Mioshi E, Hodges JR. 2011a. Behavioural-variant frontotemporal dementia: Diagnosis, clinical staging, and management. *Lancet Neurol* **10**: 162–172.
- Piguot O, Halliday GW, Reid WGJ, Casey B, Carman R, Huang Y, Xuereb JH, Hodges JR, Kril JJ. 2011b. Clinical phenotypes in autopsy-confirmed Pick disease. *Neurology* **76**: 253–259.
- Pollock NJ, Mirra SS, Binder LI, Hansen LA, Wood JG. 1986. Filamentous aggregates in Pick's disease, progressive supranuclear palsy, and Alzheimer's disease share antigenic determinants with microtubule-associated protein, tau. *Lancet* **328**: 1211.
- Polymenidou M, Lagier-Tourenne C, Hutt KR, Huelga SC, Moran J, Liang TY, Ling SC, Sun E, Wanczewicz E, Mazur C, et al. 2011. Long pre-mRNA depletion and RNA mis-splicing contribute to neuronal vulnerability from loss of TDP-43. *Nat Neurosci* **14**: 459–468.
- Poorkaj P, Bird TD, Wijsman E, Nemens E, Garruto RM, Anderson L, Andreadis A, Wiederholt WC, Raskind M, Schellenberg GD. 1998. Tau is a candidate gene for chromosome 17 frontotemporal dementia. *Ann Neurol* **43**: 815–825.
- Rabinovici GD, Jagust WJ, Furst AJ, Ogar JM, Racine CA, Mormino EC, O'Neil JP, Lal RA, Dronkers NF, Miller BL, et al. 2008. A $\beta$  amyloid and glucose metabolism in three variants of primary progressive aphasia. *Ann Neurol* **64**: 388–401.
- Rebeiz JJ, Kolodny EM, Richardson EP. 1968. Corticodentonigral degeneration with neuronal achromasia. *Arch Neurol* **18**: 20–33.
- Reed LA, Grabowski TJ, Schmidt ML, Morris JC, Goate A, Solodkin A, Van Hoesen GW, Schelper RL, Talbot CJ, Wragg MA, et al. 1997. Autosomal dominant dementia with widespread neurofibrillary tangles. *Ann Neurol* **42**: 564–572.
- Renton AE, Majounie E, Waite A, Simón-Sánchez J, Rollinson S, Gibbs JR, Schymick JC, Laaksovirta H, Van Swieten JC, Myllykangas L, et al. 2011. A hexanucleotide repeat expansion in *C9ORF72* is the cause of chromosome 9p21-linked ALS-FTD. *Neuron* doi: 10.1016/j.neuron.2011.09.010.
- Richter H. 1918. Eine besondere Art von Stirnhirnschwund mit Verblödung. *Z Neurol* **38**: 127–159.
- Ritson GP, Custer SK, Freibaum BD, Guinto JB, Geffel D, Moore J, Tang W, Winton MJ, Neumann M, Trojanowski JQ, et al. 2010. TDP-43 mediates degeneration in a novel *Drosophila* model of disease caused by mutations in VCP/p97. *J Neurosci* **30**: 7729–7739.
- Roberson ED, Scarce-Levie K, Palop JJ, Yan F, Cheng IH, Wu T, Gerstein H, Yu GQ, Mucke L. 2007. Reducing endogenous tau ameliorates amyloid  $\beta$ -induced deficits in an Alzheimer's disease mouse model. *Science* **316**: 750–754.
- Rossor MN, Fox NC, Mummery CM, Schott JM, Warren JD. 2010. The diagnosis of young-onset dementia. *Lancet Neurol* **9**: 793–806.
- Sampathu DM, Neumann M, Kwong LK, Chou TT, Misceniyi M, Truax A, Bruce J, Grossman M, Trojanowski JQ, Lee VMY. 2006. Pathological heterogeneity of frontotemporal lobar degeneration with ubiquitin-positive inclusions delineated by ubiquitin immunohistochemistry and novel monoclonal antibodies. *Am J Pathol* **169**: 1343–1352.
- Sandberg MK, Al-Doujaaily H, Sharps B, Clarke AR, Collinge J. 2011. Prion propagation and toxicity *in vivo* occur in two distinct mechanistic phases. *Nature* **470**: 540–542.
- Santacruz K, Lewis J, Spire T, Paulson J, KKKotilinek L, Ingelsson M, Guimares A, DeTure M, Ramsden M, McGowan E, et al. 2005. Tau suppression in a neurodegenerative mouse model improves memory function. *Science* **309**: 476–481.
- Schmitt-John T, Drepper C, Musmann A, Hahn P, Kuhlmann M, Thiel C, Hafner M, Lengeling A, Heimann P, Jones JM, et al. 2005. Mutation of *Vps54* causes motor neuron disease and defective spermiogenesis in the wobbler mouse. *Nat Genet* **37**: 1213–1215.
- Schneider C. 1927. Über Picksche Krankheit. *Mtschr Psychiat Neurol* **65**: 230–275.
- Schneider C. 1929. Weitere Beiträge zur Lehre von der Pickschen Krankheit. *Z Neurol* **120**: 340–384.
- Seelaar H, Klijnsma KY, de Koning I, Van der Lugt A, Chiu WZ, Azmani A, Rozemuller AJM, Van Swieten JC. 2010. Frequency of ubiquitin and FUS-positive, TDP-43-negative frontotemporal lobar degeneration. *J Neurol* **257**: 747–753.
- Sharp AJ, Hansen S, Selzer RR, Cheng Z, Regan R, Hurst JA, Stewart H, Price SM, Blair E, Hennekam RC, et al. 2006. Discovery of previously unidentified genomic disorders from the duplication architecture of the human genome. *Nat Genet* **38**: 1038–1042.
- Shaw CE. 2010. Capturing VCP: Another molecular piece in the ALS jigsaw puzzle. *Neuron* **68**: 812–814.
- Shaw-Smith C, Pittman AM, Willatt L, Martin H, Rickman L, Gribble S, Curley R, Cumming S, Dunn C, Kalaitzopoulos D, et al. 2006. Microdeletion encompassing *MAPT* at chromosome 17q21.3 is associated with developmental delay and learning disability. *Nat Genet* **38**: 1032–1037.
- Skibinski G, Parkinson NJ, Brown JM, Charkrabarti L, Lloyd SL, Hummerich H, Nielsen JE, Hodges JR, Spillantini MG, Thusgaard T, et al. 2005. Mutations in the endosomal ESCRTIII-complex subunit *CHMP2B* in frontotemporal dementia. *Nat Genet* **37**: 806–808.
- Snowden JS, Goulding PJ, Neary D. 1989. Semantic dementia: A form of circumscribed atrophy. *Behav Neurol* **2**: 167–182.
- Spillantini MG, Crowther RA, Goedert M. 1996. Comparison of the neurofibrillary pathology in Alzheimer's disease and familial presenile dementia with tangles. *Acta Neuropathol* **92**: 42–48.
- Spillantini MG, Goedert M, Crowther RA, Murrell JR, Farlow MR, Ghetti B. 1997. Familial multiple system tauopathy with presenile dementia: A disease with abundant neuronal and glial tau filaments. *Proc Natl Acad Sci* **94**: 4113–4118.
- Spillantini MG, Bird TD, Ghetti B. 1998a. Frontotemporal dementia and parkinsonism linked to chromosome 17: A new group of tauopathies. *Brain Pathol* **8**: 387–402.

M. Goedert et al.

- Spillantini MG, Murrell JR, Goedert M, Farlow MR, Klug A, Ghetti B. 1998b. Mutation in the tau gene in familial multiple system tauopathy with presenile dementia. *Proc Natl Acad Sci* **95**: 7737–7741.
- Spillantini MG, Crowther RA, Kamphorst W, Heutink P, Van Swieten JC. 1998c. Tau pathology in two Dutch families with mutations in the microtubule-binding region of tau. *Am J Pathol* **153**: 1359–1363.
- Spina S, Murrell JR, Huey ED, Wassermann EM, Pietrini P, Grafman J, Ghetti B. 2007. Corticobasal syndrome associated with the A90D progranulin mutation. *J Neuropathol Exp Neurol* **66**: 892–900.
- Sreedharan J, Blair IP, Tripathi VB, Hu X, Vance C, Rogelj B, Ackerley S, Durnall JC, Williams KL, Buratti E, et al. 2008. TDP-43 mutations in familial and sporadic amyotrophic lateral sclerosis. *Science* **319**: 1668–1672.
- Stefansson H, Helgason A, Thorleifsson, Steintorsdottir V, Masson G, Barnard J, Baker A, Jonasdottir A, Ingason A, Gudnadottir VG, et al. 2005. A common inversion under selection in Europeans. *Nat Genet* **37**: 129–137.
- Steele JC, Richardson JC, Olszewski J. 1964. Progressive supranuclear palsy. *Arch Neurol* **10**: 333–359.
- Stertz G. 1926. Über die Pickische Atrophie. *Z Neurol* **101**: 729–749.
- Strittmatter WJ, Roses AD. 1995. Apolipoprotein E and Alzheimer disease. *Proc Natl Acad Sci* **92**: 4725–4727.
- Sun Z, Diaz Z, Fang X, Hart MP, Chesi A, Shorter J, Gitler AD. 2011. Molecular determinants and genetic modifiers of aggregation and toxicity for the ALS disease protein FUS/TLS. *PLoS Biol* **9**: e1000614.
- Tang W, Lu Y, Tian QY, Zhang Y, Guo FJ, Liu GY, Syed NM, Lai Y, Lin EA, Kong L, et al. 2011. The growth factor progranulin binds to TNF receptors and is therapeutic against inflammatory arthritis in mice. *Science* **332**: 478–484.
- Taniguchi S, Suzuki N, Masuda M, Hisanaga SI, Iwatsubo T, Goedert M, Hasegawa M. 2005. Inhibition of heparin-induced tau filament formation by phenothiazines, polyphenols and porphyrins. *J Biol Chem* **280**: 7614–7623.
- Ticozzi N, Silani V, LeClerc AL, Keagle P, Gellera C, Ratti A, Taroni F, Kwiatkowski TJ, McKenna-Yasek DN, Sapp PC, et al. 2009. Analysis of FUS gene mutation in familial amyotrophic lateral sclerosis within an Italian cohort. *Neurology* **73**: 1180–1185.
- Tollervey JR, Curk T, Rogelj B, Riese M, Cereda M, Kayikci M, König J, Hortobágyi T, Nishimura AL, Zupunski V, et al. 2011. Characterizing the RNA targets and position-dependent splicing regulation by TDP-43. *Nat Neurosci* **14**: 452–458.
- Vance C, Al-Chalabi A, Ruddy D, Smith BN, Hu X, Sreedharan J, Siddique T, Schelhaas HJ, Kusters B, Troost D, et al. 2006. Familial amyotrophic lateral sclerosis with frontotemporal dementia is linked to a locus on chromosome 9p13.p2–21.3. *Brain* **129**: 868–876.
- Vance C, Rogelj B, Hortobágyi T, De Vos KJ, Nishimura AL, Sreedharan J, Hu X, Smith B, Ruddy D, Wright P, et al. 2009. Mutations in FUS, an RNA processing protein, cause familial amyotrophic sclerosis type 6. *Science* **323**: 1208–1211.
- Van Deerlin VM, Sleiman PMA, Martinez-Lage M, Chen-Plotkin A, Wang LS, Graff-Radford NR, Dickson DW, Rademakers R, Boeve BF, Grossman M, et al. 2010. Common variants at 7p21 are associated with frontotemporal lobar degeneration with TDP-43 inclusions. *Nat Genet* **42**: 234–239.
- Van der Flier WM, Pijnenburg YAL, Fox NC, Scheltens P. 2011. Early-onset versus late-onset Alzheimer's disease: The case of the missing APOE ε4 allele. *Lancet Neurol* **10**: 280–288.
- Van Mansvelt J. 1954. *Pick's disease*. Mjvd Loeff, Enschede.
- Van Swieten JC, Heutink P. 2008. Mutations in progranulin (GRN) within the spectrum of clinical and pathological phenotypes of frontotemporal dementia. *Lancet Neurol* **7**: 965–974.
- Verhaart WJC. 1930. Over de ziekte van Pick. *Nederl Tijdschr Geneesk* **74**: 5586–5598.
- Vidal R, Delisle MB, Ghetti B. 2004. Neurodegeneration caused by proteins with an aberrant carboxyl-terminus. *J Neuropathol Exp Neurol* **63**: 787–800.
- Von Braunmühl A. 1932. Pickische Krankheit und amyotrophische Lateralsklerose. *Allg Z Psychiat* **96**: 364–366.
- Von Braunmühl A, Leonhard K. 1934. Über ein Schwesternpaar mit Pickischer Krankheit. *Z Neurol* **150**: 209–241.
- Watts GDJ, Wymer J, Kovach MJ, Mehta SG, Mumm S, Darvish D, Pestronk A, Whyte MP, Kimonis VE. 2004. Inclusion body myopathy associated with Paget disease of bone and frontotemporal dementia is caused by mutant valosin-containing protein. *Nat Genet* **36**: 377–381.
- Wernicke C. 1874. *Der aphasische Symptomencomplex: Eine psychologische Studie auf anatomischer Basis*. Max Cohn & Weigert, Breslau.
- Wetzel MK, Naska S, Laliberte CL, Rymar VV, Fujitani M, Biernaskie JA, Cole CJ, Lerch JP, Spring S, Wang SH, et al. 2008. p73 regulates neurodegeneration and phospho-tau accumulation in aging and Alzheimer's disease. *Neuron* **59**: 708–721.
- Whitwell JL, Jack CR, Senjem ML, Josephs KA. 2006. Patterns of atrophy in pathologically confirmed FTL D with and without motor neuron degeneration. *Neurology* **66**: 102–104.
- Whitwell JL, Przybelski SA, Weigand SD, Ivnik RJ, Vemuri P, Gunter JL, Senjem ML, Shiung MM, Boeve BF, Knopman DS, et al. 2009a. Distinct anatomical subtypes of the behavioural variant of frontotemporal dementia: A cluster analysis study. *Brain* **132**: 2932–2946.
- Whitwell JL, Jack CR, Boeve BF, Senjem ML, Baker M, Rademakers R, Ivnik RJ, Knopman DS, Wszolek ZK, Petersen RC, et al. 2009b. Voxel-based morphometry patterns of atrophy in FTL D with mutations in MAPT or PGRN. *Neurology* **72**: 813–820.
- Whitwell JL, Avula R, Senjem ML, Kantarci K, Weigand SD, Samikoglu A, Edmonson HA, Vemuri P, Knopman DS, Boeve BF, et al. 2010a. Gray and white matter water diffusion in the syndromic variants of frontotemporal dementia. *Neurology* **74**: 1279–1287.
- Whitwell JL, Jack CR, Boeve BF, Parisi JE, Ahlskog JE, Drubach DA, Senjem ML, Knopman DS, Petersen RC, Dickson DW, et al. 2010b. Imaging correlates of pathology in corticobasal syndrome. *Neurology* **75**: 1879–1887.

- Wilhelmsen KC, Lynch T, Pavlou E, Higgins M, Nygaard TG. 1994. Localization of disinhibition-dementia-parkinsonism-amyotrophy complex to 17q21–22. *Am J Hum Genet* **55**: 1159–1164.
- Williams DR, Lees AJ. 2009. Progressive supranuclear palsy: Clinicopathological concepts and diagnostic challenges. *Lancet Neurol* **8**: 270–279.
- Williams DR, Holton JL, Strand C, Pittman A, de Silva R, Lees AJ, Revesz T. 2007. Pathological tau burden and distribution distinguishes progressive supranuclear palsy-parkinsonism from Richardson's syndrome. *Brain* **130**: 1566–1576.
- Wischik CM, Novak M, Thogersen HC, Edwards PC, Runswick MJ, Jakes R, Walker JE, Milstein C, Roth M, Klug A. 1988. Isolation of a fragment of tau derived from the core of the paired helical filament of Alzheimer disease. *Proc Natl Acad Sci* **85**: 4506–4510.
- Wittmann CW, Wszolek ZF, Shulman JM, Salvaterra PM, Lewis J, Hutton M, Feany MB. 2001. Tauopathy in *Drosophila*: Neurodegeneration without neurofibrillary tangles. *Science* **293**: 711–714.
- Yokoseki A, Shiga A, Tan CE, Tagawa A, Kaneko H, Koyama A, Eguchi H, Tsujino A, Ikeuchi T, Kakita A, et al. 2008. TDP-43 mutation in familial amyotrophic lateral sclerosis. *Ann Neurol* **63**: 538–542.
- Yu CE, Bird TD, Bekris LM, Montine TJ, Leverenz JB, Steinbart E, Galloway NM, Feldman H, Woltjer R, Miller CA, et al. 2010. The spectrum of mutations in progranulin. *Arch Neurol* **67**: 161–170.
- Zhou J, Greicius MD, Gennatas ED, Growdon ME, Jang JY, Rabinovici GD, Kramer JH, Weiner M, Miller BL, Seeley WW. 2010. Divergent network connectivity changes in behavioural variant frontotemporal dementia and Alzheimer's disease. *Brain* **133**: 1352–1367.