

Scopolamine Administration Modulates Muscarinic, Nicotinic and NMDA Receptor Systems

Soheil Keihan Falsafi¹✉, Alev Deli¹✉, Harald Höger², Arnold Pollak¹, Gert Lubec^{1*}

1 Department of Pediatrics, Medical University of Vienna, Vienna, Austria, **2** Core Unit of Biomedical Research, Division of Laboratory Animal Science and Genetics, Medical University of Vienna, Vienna, Austria

Abstract

Studies on the effect of scopolamine on memory are abundant but so far only regulation of the muscarinic receptor (M1) has been reported. We hypothesized that levels of other cholinergic brain receptors as the nicotinic receptors and the N-methyl-D-aspartate (NMDA) receptor, known to be involved in memory formation, would be modified by scopolamine administration. C57BL/6J mice were used for the experiments and divided into four groups. Two groups were given scopolamine 1 mg/kg i.p. (the first group was trained and the second group untrained) in the multiple T-maze (MTM), a paradigm for evaluation of spatial memory. Likewise, vehicle-treated mice were trained or untrained thus serving as controls. Hippocampal levels of M1, nicotinic receptor alpha 4 (Nic4) and 7 (Nic7) and subunit NR1-containing complexes were determined by immunoblotting on blue native gel electrophoresis. Vehicle-treated trained mice learned the task and showed memory retrieval on day 8, while scopolamine-treatment led to significant impairment of performance in the MTM. At the day of retrieval, hippocampal levels for M1, Nic7 and NR1 were higher in the scopolamine treated groups than in vehicle-treated groups. The concerted action, i.e. the pattern of four brain receptor complexes regulated by the anticholinergic compound scopolamine, is shown. Insight into probable action mechanisms of scopolamine at the brain receptor complex level in the hippocampus is provided. Scopolamine treatment is a standard approach to test cognitive enhancers and other psychoactive compounds in pharmacological studies and therefore knowledge on mechanisms is of pivotal interest.

Citation: Falsafi SK, Deli A, Höger H, Pollak A, Lubec G (2012) Scopolamine Administration Modulates Muscarinic, Nicotinic and NMDA Receptor Systems. PLoS ONE 7(2): e32082. doi:10.1371/journal.pone.0032082

Editor: Georges Chapouthier, Université Pierre et Marie Curie, France

Received: November 2, 2011; **Accepted:** January 19, 2012; **Published:** February 23, 2012

Copyright: © 2012 Falsafi et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: The authors have no support or funding to report.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: gert.lubec@meduniwien.ac.at

✉ These authors contributed equally to this work.

Introduction

Scopolamine, a tropane alkaloid, is a very potent psychoactive drug that is used as a standard/reference drug for inducing amnesia in mammals. A characteristic feature of these alkaloids is that subjects do not recall memories of the time they were intoxicated, and the user loses all sense of reality. The effects are generally interpreted as a cholinergic deficit and related to the fact that acetylcholine is involved in memory functions. The use of scopolamine as a pharmacological model of 'cholinergic amnesia' became very popular after the cholinergic hypothesis of geriatric memory dysfunction was postulated [1]. This hypothesis assumes that the age-related decline in cognitive functions is predominantly related to the decrease of the integrity of cholinergic neurotransmission. Since scopolamine-induced amnesia was proposed to be due to blockade of cholinergic neurotransmission, this substance was used to model the cognitive deficits that could be observed in aging and dementia. Scopolamine appears to be a nonselective muscarinic receptor antagonist and it has been demonstrated that scopolamine has a high selectivity for the muscarinic receptor [2], although it has been reported that high doses of scopolamine are also blocking nicotinic receptors [3]. The scopolamine model is used extensively for preclinical testing of new substances designed to treat cognitive impairment [4–9].

First studies that investigated the central effects of scopolamine in animals were reported in the fifties [10,11]. Scopolamine was also used to test the hypothesis which stated that cholinergic neurotransmission is acting as an inhibitor of an "activating system", whereby scopolamine was used to show that a muscarinic antagonist induced behavioral disinhibition [12]. In later studies the effects of scopolamine were investigated in experiments that examined the effects on cognitive functions. In a passive avoidance test bilateral hippocampal scopolamine injections decreased the step-through latency after post-training administration [13]. The effects of intra-hippocampal scopolamine have also been investigated in reinforced T-maze alternation and visual discrimination [14]. In this study it was found that the visual discrimination performance was impaired at a dose of 35 µg whereas the delayed alternation performance was already impaired at a dose of 12 µg. Consequently, it was argued that working memory was more sensitive to blockade of the hippocampal cholinergic synapses than reference memory. The hippocampal cholinergic system has also been suggested to play a role in potentiation of odor by taste conditioning and odor aversion learning [15]. It was revealed that scopolamine delayed the extinction of the odor aversion conditioning. In a T-maze task scopolamine induced a reduction in alternation performance which was most pronounced after the first injection [16]. Scopolamine impaired the repeated acquisition

in a Morris water escape task indicating that scopolamine affected spatial working memory [17]. In a three-panel runway task, which assesses working- and reference memory performance, scopolamine increased the number of working memory errors whereas the reference memory performance was not affected [18]. It has been found that daily injections with scopolamine impaired the acquisition of an eight-arm radial maze [19]. Scopolamine disrupted acquisition performance of all rats in this radial maze task. Since this procedure forces rats to use spatial cues to guide their behavior, it was tentatively concluded that scopolamine affected spatial working memory. A further study revealed that scopolamine increased the number of errors of trained rats in an eight-arm radial maze without using a delay between choices [20]. The effects of scopolamine have also been investigated in the Morris water maze (MWM) to evaluate the role of the cholinergic system in spatial learning in rats and mice: the effects of scopolamine were impairment of the acquisition performance in this task [21].

A more detailed analysis of the effects of scopolamine in the MWM examined the effects of pre-training and pre-treatment [22–24]: Using a relative high dose of scopolamine, it was found that pre-training prevented the scopolamine-induced spatial learning impairment observed in naive group [22,24–26]. Scopolamine-treated C57BL/6J showed reduced learning ability [27] in the MWM and Harrison et al. [28] observed scopolamine-induced cognitive deficits in B6C3F1/J mice using the MWM. As shown by observations in a radial maze, Godding et al. [29] proposed that scopolamine side-effects (as sedation, impairment of coordinative and reactive skills, visual disturbances) may be responsible for the apparent decline in this spatial memory paradigm.

It was the aim of the current study to show hippocampal patterns of major brain receptors known to be involved in memory processes paralleling scopolamine-induced impaired memory retrieval.

Results

Basic neurological and physiological observational assessment

In this neurological observational battery spatial locomotion was decreased whereas startle response in scopolamine-treated mice was increased. Grip strength and wire manoeuvre were reduced in scopolamine-treated mice (table 1).

The Rota rod

In the rota rod scopolamine-treated mice showed shorter performance on the revolving rod indicating decreased motor coordination or strength (figure 1)

Multiple T-maze

In the MTM vehicle-treated mice learned the task expressed as latencies in seconds, in contrast to scopolamine-treated animals (figure 2A). Untrained mice did not learn the task but were spending the same time in the maze without any reward.

Vehicle-treated mice showed shorter pathlength as scopolamine-treated mice (figure 2B). As shown in figure 2C there were significantly more correct decisions in the vehicle-treated group.

The number of successful entries into the goal box was unequivocally higher in vehicle-treated mice (figure 2D).

At the probe trial on day 8 vehicle-treated mice showed significantly shorter latencies than the scopolamine-treated animals (figure 2E).

Determination of brain receptor complexes

A single band at about 480 kDa was representing the muscarinic receptor complex containing M1.

M1 was significantly increased in scopolamine-treated trained mice as compared to vehicle-treated trained mice.

Scopolamine-treated and untrained animals showed higher levels than vehicle-treated untrained animals (figure 3).

A single band was observed for nicotinic receptor alpha 7 (Nic7) between 480 and 720 kDa.

Nic7 was significantly increased in scopolamine-treated trained mice as compared to vehicle-treated trained mice.

Scopolamine-treated and untrained animals showed higher levels than vehicle-treated untrained animals.

Scopolamine-treated trained mice showed increased levels as compared to scopolamine-treated and untrained mice (figure 4).

As to the nicotinic alpha 4 receptor (Nic4) no significant differences were observed, although a trend was observed when scopolamine-treated and trained mice were compared to vehicle-treated trained mice, as well as for the comparison between scopolamine-treated untrained and vehicle-treated untrained mice (figure 5).

NMDA receptor subunit NR1 showed two bands indicating two different receptor complexes. The band between 480 and 720 kDa was higher in scopolamine-treated trained mice than in vehicle-treated trained mice.

The band at about 242 kDa was comparable between groups (figure 6A,B).

Loading controls showed comparable protein loading as shown in figure 7.

Correlation studies. There were no significant correlations between parameters from the MTM and brain receptor complex levels.

As shown in figure 8A and B there was a significant correlation ($R = -0.925$, $P = 0.008$) between M1 and Nic4 receptor complexes and a significant correlation ($R = 0.902$, $P = 0.014$) between Nic7 and NR1 (band 2).

Discussion

Blockade of muscarinic receptors by scopolamine, a muscarinic receptor antagonist, impairs learning and memory in mice [1,30] and inhibition of cholinergic neurotransmission by muscarinic receptor antagonists produces profound deficits in attention and memory.

Anagnostaras et al. examined different forms of memory in mice with a null mutation of the gene encoding the M1 receptor, the most densely distributed muscarinic receptor in the hippocampus and forebrain: Long-term potentiation (LTP) in response to theta burst stimulation in the hippocampus was reduced in mutant mice. M1 null mutant mice showed normal or enhanced memory for tasks that involved matching-to-sample problems, but they were severely impaired in non-matching-to-sample working memory as well as consolidation. Their results suggest that the M1 receptor is specifically involved in memory processes for which the cortex and hippocampus interact [31].

The M1 subtype is the most abundant of the muscarinic receptors in the forebrain and hippocampus [32,33] and Park et al. [34] provided further evidence for a key role of M1 receptors in memory and cognition.

Acetylcholine (ACh) activates two families of receptors that mediate its action in target tissues: nicotinic receptors, which function as ligand-gated cation channels that participate in rapid postsynaptic neurotransmission, and muscarinic receptors (mAChR), members of family A G-protein coupled receptors

Table 1. Observational battery average results of all mice treated with scopolamine or vehicle.

	Vehicle-treated	Scopolamine-treated	T-test
	mean \pm SD	mean \pm SD	
Body position	3.75 \pm 0.82	3.85 \pm 0.00	0.724
Palpebral closure	0.05 \pm 0.00	0 \pm 0.00	0.320
Locomotor activity	2.55 \pm 0.00	2.6 \pm 0.00	0.808
Bizarre behavior	0 \pm 0.00	0 \pm 0.00	
Exophthalmus	0 \pm 0.00	0 \pm 0.00	
Respiration rate	3.75 \pm 0.00	3.9 \pm 0.82	0.358
Tremor	0 \pm 0.00	0.025 \pm 0.00	0.320
Twitches	0 \pm 0.00	0 \pm 0.00	
Transfer arousal	3.65 \pm 0.41	3.45 \pm 0.55	0.132
Spatial locomotion*	3.2 \pm 1.03	2.463 \pm 0.88	0.000
Startle response*	3.65 \pm 1.26	4.45 \pm 1.79	0.015
Piloerection	4 \pm 0.00	3.975 \pm 0.00	0.320
Ataxic/Hypotonic/impaired gait	0 \pm 0.00	0 \pm 0.00	
Limb rotation	3.975 \pm 0.00	3.95 \pm 0.00	0.562
Pelvic elevation	4.1 \pm 0.82	4 \pm 0.82	0.699
Tail elevation	2.2 \pm 0.00	2.35 \pm 0.00	0.336
Finger approach	3.75 \pm 0.00	3.9 \pm 0.00	0.241
Finger withdrawal	3.65 \pm 0.82	3.8 \pm 0.00	0.452
Touch escape	3.9 \pm 1.79	4.2 \pm 1.10	0.412
Positional passivity	0 \pm 0.00	0 \pm 0.00	
Visual placing	2.3 \pm 0.41	2.5 \pm 1.60	0.299
Grip strength*	4.75 \pm 1.10	5.4 \pm 0.00	0.007
Body/Abdominal tone	5.55 \pm 0.00	5.6 \pm 0.00	0.788
Hypothermia	0 \pm 0.00	0 \pm 0.00	
Pinna reflex	2.75 \pm 0.00	5 \pm 2.42	0.000
Cornea	3.55 \pm 1.10	3.3 \pm 1.97	0.619
Toe pinch	5.5 \pm 2.34	5.1 \pm 1.03	0.713
Wire manoeuvre*	1.75 \pm 2.19	2.6 \pm 1.03	0.037
Skin color	4 \pm 0.00	4 \pm 0.00	
Diarrhea	0.025 \pm 0.00	0 \pm 0.00	0.320
Limb tone	4 \pm 0.00	4 \pm 0.00	
Lacrimation/Salivation	0 \pm 0.00	0 \pm 0.00	
Provoked biting	2.1 \pm 0.55	2.1 \pm 0.00	1.000
Vocalization	0 \pm 0.00	0 \pm 0.00	
Tail pinch	2.225 \pm 0.82	2.1 \pm 1.51	0.597
Righting reflex	1 \pm 0.00	1 \pm 0.00	
Negative geotaxis	0 \pm 0.00	0 \pm 0.00	
Cliff avoidance	0.95 \pm 0.00	0.9 \pm 0.00	0.402
Vestibular drop	0 \pm 0.00	0 \pm 0.00	
Proprioception	1 \pm 0.00	1 \pm 0.00	

*significant.

doi:10.1371/journal.pone.0032082.t001

(GPCRs), that play a key role in modulating the activity of many circuits within the CNS. These two classes of receptor families were originally named for their specific activation by nicotine and muscarine, respectively, but have been extensively characterized since that time on a molecular basis. The diversity and complexity of muscarinic cholinergic signaling is facilitated in part by five distinct receptor subtypes, M1–M5, the genes for which were

cloned in the mid to late 1980s [35–37]. These intronless genes encode muscarinic receptor proteins that have the typical structural features of the seven transmembrane helix GPCR superfamily, the largest family of cell-surface receptors and key regulators of a wide variety of physiological processes [38]. *In situ* hybridization experiments following the cloning of mAChR subtype genes revealed that individual subtypes were expressed



Figure 1. Results in the Rota rod. Significant differences in rota rod performance, in scopolamine-treated vs vehicle-treated groups ($P \leq 0.05$) are shown. Numbers are representing seconds of remaining on the revolving rod. doi:10.1371/journal.pone.0032082.g001

in partially overlapping tissues, with some regions, including the hippocampus, expressing all five mAChR subtypes [39,40]. Several of the muscarinic receptor subtypes M1–M5 might underlie the cognitive effects of scopolamine. Evidence for a role in mnemonic processes in both rodents and humans is strongest for the postsynaptic muscarinic M1 receptor [41–46]. This receptor is predominantly located in brain regions thought to be important for learning and memory such as cortex and hippocampus; the presence of the M1 receptor in the periphery is relatively limited [47,48]. Hence, M1 antagonists are considered an interesting option with regards to finding novel pharmacological alternatives to induce cognitive impairment which are not so much hampered by issues of nonselectivity or peripheral side-effect [9,41].

Nicotinic cholinergic receptors are a class of ligand-gated ion channels that are assembled from five subunits out of at least 17 identified subunits and are differentially expressed in both the central and peripheral nervous systems [49–52]. Neuronal nAChRs have a pentameric structure and are comprised of either α ($\alpha 7$ – $\alpha 10$) subunits or a combination of α ($\alpha 2$ – $\alpha 6$) and β ($\beta 2$ – $\beta 4$) subunits [53–56]. In the central nervous system, the $\alpha 4\beta 2^*$ which includes subclasses differentiated by the inclusion of $\alpha 3$, $\alpha 5$ or $\alpha 6$ subtypes [56,57] and $\alpha 7$ nAChRs are the two predominant nAChR subtypes [58,59], but they have diverse functional properties [60–63].

Herein, in a paradigm of spatial memory, increased levels of a M1 receptor complex were observed following scopolamine treatment in both, trained and untrained mice. As M1 receptors are colocalized with NMDA receptors in hippocampal pyramidal neurons, and co-activation with NMDA receptors results in amplified NMDA currents [64], we also determined a key subunit of the NMDA receptor complex, NR1. Increased NR1 complex levels have been shown to appear in spatial memory formation [65] and indeed, NR1 complex levels were increased in scopolamine-treated trained animals as compared to vehicle-treated trained animals. A possible M1 – NR1 interaction is also supported by the fact that the disruptive effect of scopolamine was intensified when NMDA receptor antagonists were co-adminis-

tered at doses that had no effect on the maze performance by themselves, while the selective AMPA antagonist YM90K failed to affect the disruptive effect of scopolamine [66]. This proposed interaction is further strengthened by recent data revealing that the central cholinergic system modulates the excitatory neurotransmission by using excitatory amino acids as neurotransmitters [67] and that ACh stimulation of muscarinic receptors selectively potentiates responses to NMDA [66,68].

Muscarinic M1 receptors couple to Gq-proteins that subsequently activate several signaling cascades via phospholipase C [47,69,70], which in turn can influence Ca^{2+} and K^+ currents [70], raise cyclic AMP levels [69], and can stimulate other receptor systems including NMDA receptors, i.e. currents produced by hippocampal CA1 pyramidal neurons [64,71,72]. Moreover, M1 receptors and NR1 receptor subunits were found to be colocalized at glutamatergic synapses, suggestive of a direct interaction between another receptor system. A link between M1 receptor signaling and long-term potentiation (LTP), a mechanism which is thought to underly learning and memory processes, has also been put forward [73–79].

Based upon participation in the cholinergic system and the fact that Nic4 and Nic7 have been described to be key elements in memory formation we decided to determine Nic4 and Nic7 receptor complex levels in order to show involvement in scopolamine-induced memory impairment. And indeed, Nic7 was increased in scopolamine-treated trained mice as compared to vehicle-treated trained mice and also scopolamine-treated untrained animals showed higher Nic7 receptor complex levels than vehicle-treated and untrained mice.

A significant difference between scopolamine-treated trained and untrained animals was shown, providing evidence for the notion that scopolamine is modulating even this brain receptor subtype.

No significant correlation was observed between Nic7, M1 and NR1 and the observed correlations between M1 and Nic4 is of no obvious meaning in this context as Nic4 was not significantly modified by scopolamine treatment and so is the interpretation for the correlation between Nic7 and NR1 band 2, that was not regulated by scopolamine either.

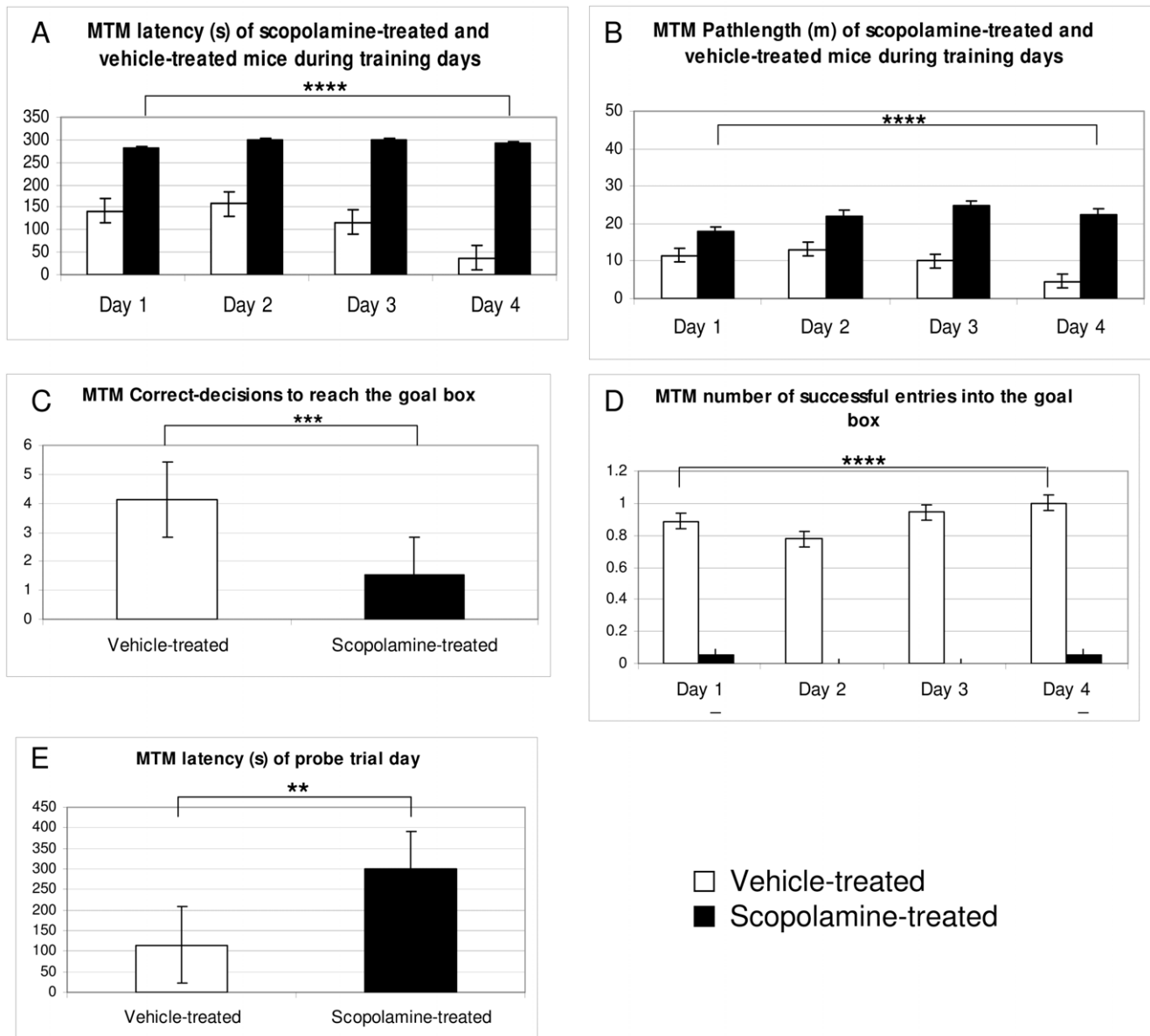


Figure 2. Results in the MTM. Significantly higher latencies and longer pathlengths were observed in the scopolamine-treated animals (A,B). Correct decisions to reach the goal box were higher in vehicle-treated and trained mice (C) and the number of successful entries into the goal box during training days was significantly higher in the vehicle-treated and trained mice (D). At the probe trial latencies were significantly lower in the vehicle-treated and trained mice (E). doi:10.1371/journal.pone.0032082.g002

There was no significant correlation between any receptor complex level with parameters from the MTM.

Taken together, we have shown the concerted action of hippocampal M1, NR1, Nic4 and Nic7 receptor complexes. The innovative result is that we have not only revealed the change of a single receptor subunit but a pattern of receptor complexes and this is of importance as the receptor complexes rather than simply subunits are functional. Moreover, it was shown that hippocampal levels of these four brain receptors, known to be essential for memory formation, were modified by scopolamine, showing scopolamine-dependent expression or levels and interplay of brain receptors along with impairment in memory retrieval, as the probe trial was carried out on day 8 following 4 days of learning. Receptor complex levels of GluR1 and GluR2 [80], M1 and Nic7 [81] have been already shown to parallel memory training in

paradigms of spatial memory. In addition, it may well be that neurological deficits shown in the neurological observational battery and on rota rod may represent altered muscarinic and nicotinic innervation and may even have been affecting behavior and performance in the multiple T-maze.

The broad array of deficits produced by anticholinergics such as scopolamine, atropine or more selective ligands could result from action at multiple receptor subtypes.

Materials and Methods

Ethics statement

Experiments were done under license from the Federal Ministry of Education, Science and Culture, which includes an ethical evaluation of the project (Approval number: BMWF-66.009/

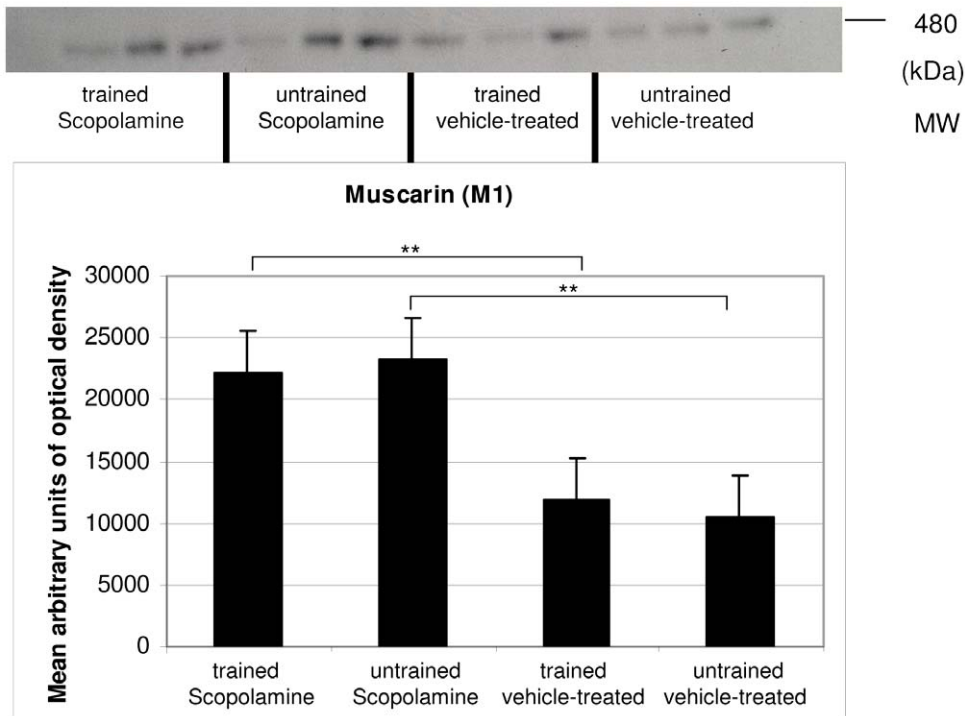


Figure 3. Western blot results of M1. The M1 receptor complex levels were significantly higher in scopolamine-treated groups. doi:10.1371/journal.pone.0032082.g003

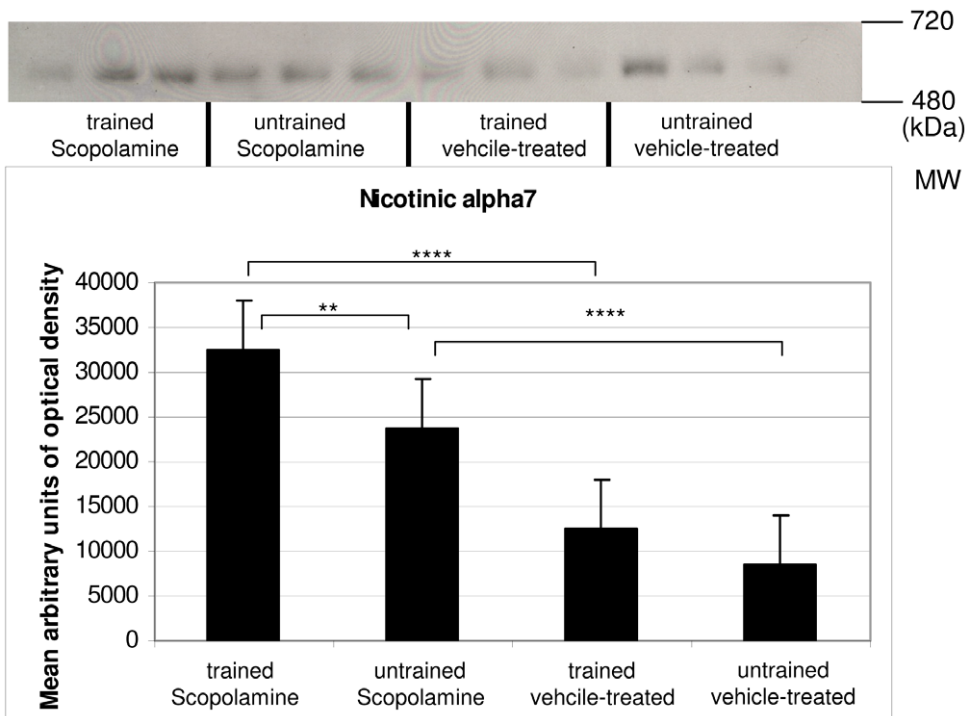


Figure 4. Western blot results of Nic 7. Nic7 complex levels were significantly higher in scopolamine-treated groups. Significant differences were also observed when scopolamine-treated trained and untrained mice were compared. doi:10.1371/journal.pone.0032082.g004

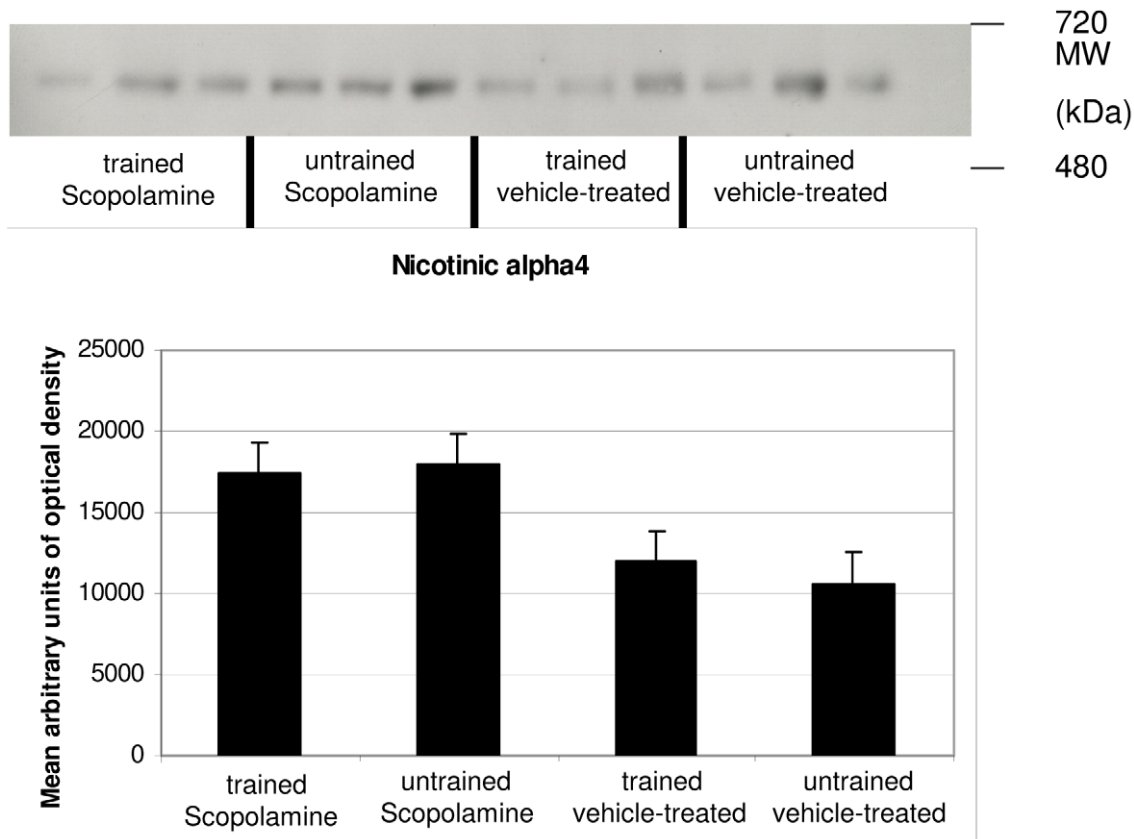


Figure 5. Western blot results of Nic 4. Although a trend was suggested no significant differences between groups were observed. doi:10.1371/journal.pone.0032082.g005

0240-II/10b/2009). Housing and maintenance of animals were in compliance with European and national regulations.

Animals

C57BL/6J mice (6 per group, total $n = 24$, male, aged 10–12 weeks) were used for the study. Mice were obtained from JANVIER SAS laboratory (France) and maintained in cages made of Makrolon and filled with wood chips in the core unit of Biomedical Research, Division of Laboratory Animal Science and Genetics, Medical University of Vienna. All mice were bred and maintained in polycarbonate cages Type II (207 9 140 9 265 mm, Ehret, Austria) and filled with autoclaved wood chips (Ligncell select, Rettenmaier, Austria). An autoclaved Altromin standard rodent diet (Altromin, Germany) and water were available ad libitum. Room temperature was $22 \pm 1^\circ\text{C}$ and relative humidity was $50 \pm 10\%$. The light/dark rhythm was 14:10. Ventilation with 100% fresh air resulted in an air change rate of 15 times per hour. The room was illuminated with artificial light at an intensity of about 200 lx in 2 m from 5 a.m. to 7 p.m. The MTM was performed between 8 a.m. and 1 p.m.

Experimental design

This study examines the C57BL/6J mouse strain which is the most commonly used strain in behavioral studies. The advantage of inbred strains is that they possess clearly defined genomes: for each strain, resulting from several generations of brother x sister mating, all the subjects have identical genes, excepting for the sex-related genes.

Mice were treated with Scopolamine hydrochloride (Sigma-Aldrich, St. Louis, MO), 1 mg/kg (30 min before all trials, including the probe trial on day 8, ip) [27,28,82,83]. Four groups (6 animals per group) were used. The first group obtained scopolamine and was tested in the MTM with reward in the goal box. (scopolamine-treated and trained). The second group was scopolamine-treated in the same way but had no reward in the goal box. (scopolamine-treated and untrained). The third group was given a sodium chloride solution and was tested in the MTM with reward in the goal box (vehicle-treated and trained) and the fourth group was given a sodium chloride solution without any reward in the goal box (untrained and untrained).

Behavioral studies

Behavioral studies as observational assessment and rota rod were carried out 1 week prior to testing in the MTM.

Basic neurological and physiological observational assessment (OB). The procedure was following the set up by Irwin [84]. A battery of tests was applied to reveal defects in gait or posture, changes in muscle tone, grip strength, visual acuity and temperature. To complete the assessment, vitally important reflexes were scored. Throughout the manipulations incidences of abnormal behaviour, fear, irritability, aggression, excitability are monitored.

Rota rod (RR). The rota rod (Rota Rod “Economex”, Columbus Instruments, Ohio, USA) tests balance and coordination and comprises a rotating drum which is accelerated from 4 to 40 rpm over the course of 5 min. The time at which each animal falls from the drum will be recorded automatically when it contact a plate which will stop the timer. Each animal

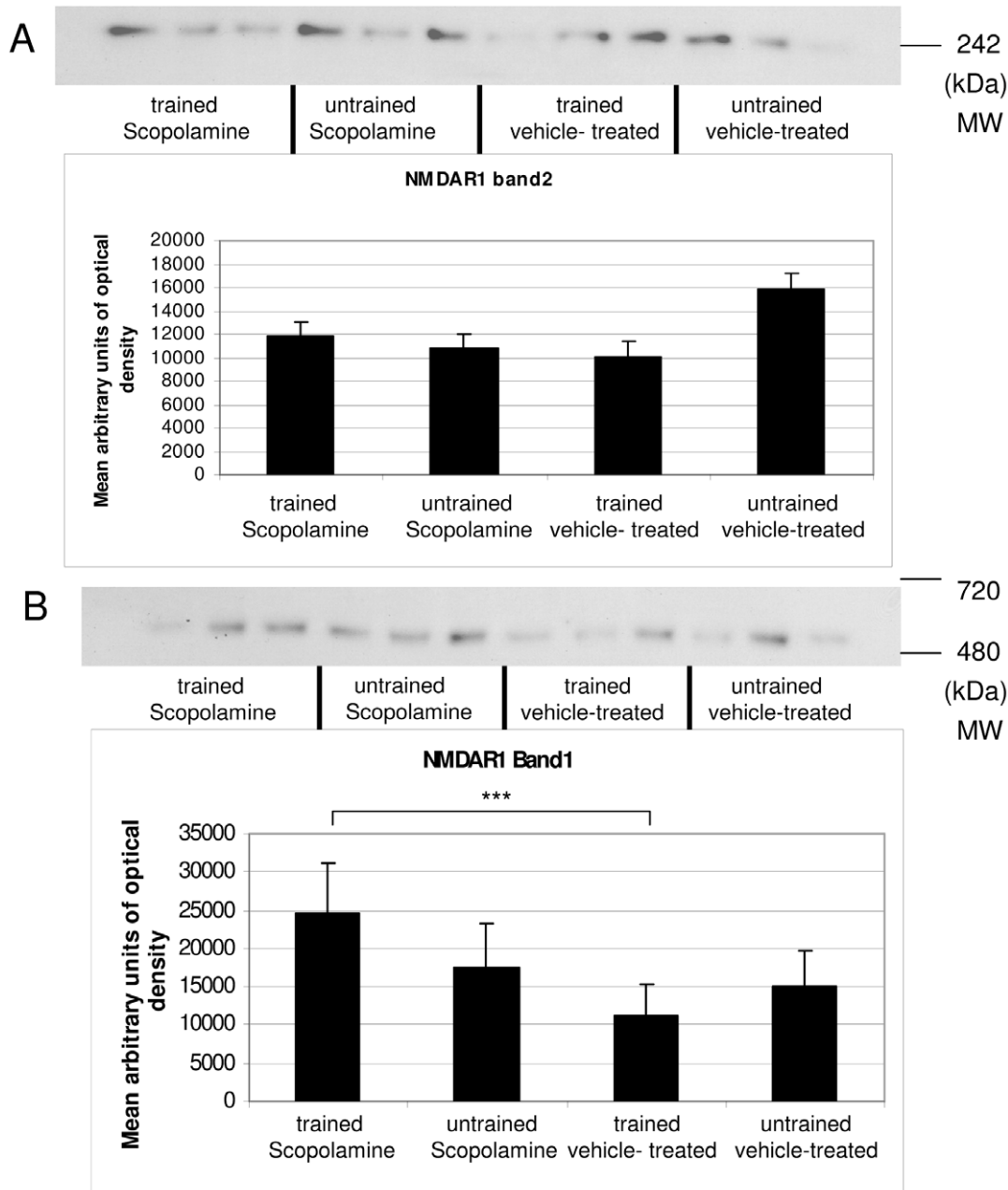


Figure 6. Western blot results of NR1. NR1 containing NMDA complex levels (band 1) were significantly increased in the scopolamine-treated trained group vs the vehicle-treated and trained panel (A). Band 2 (B) was not significantly different between groups.
doi:10.1371/journal.pone.0032082.g006

received three pre-training trials. Afterwards, each mouse received three more consecutive trials and the longest time on the drum was used for analysis [85].

Multiple T-maze (MTM). In this spatial learning task, animals learn to find the goal box based on their memory of previously visited arms [80,86,87]. The MTM is constructed of wood and consists of a wooden platform with seven choice points and the dimensions 150 cm×130 cm×15 cm and a path width of 8 cm (figure 9). Prior to testing, mice were deprived of food for 16 h to motivate food searching. Mice were placed in a start box in a black cylindrical start chamber. Each trial started with them leaving the start box and was completed when mice had reached the goal box or, if failed, after 5 min. Upon arriving in the goal

box, mice were allowed to consume a small piece of a food pellet as provided reward and transferred to their home cage. Immediately after each trial, the entire maze was cleaned with 1% incidin solution. After testing, animals were given food as per body weight (120 g/kg) into the home cage, representing the amount to maintain their body weight but keep them hungry for the following day for MTM tests. Mice were trained with 3 trials per day for 4 days. Trials were carried out using 20 min intervals. Trials were recorded using a computerized tracking/image analyzer system (video camcorder: 1/3 in. SSAMHR EX VIEWHAD coupled with computational tracking system: TiBeSplit). The system provided the following parameters, correct or wrong decisions (wrong means a path ending), path

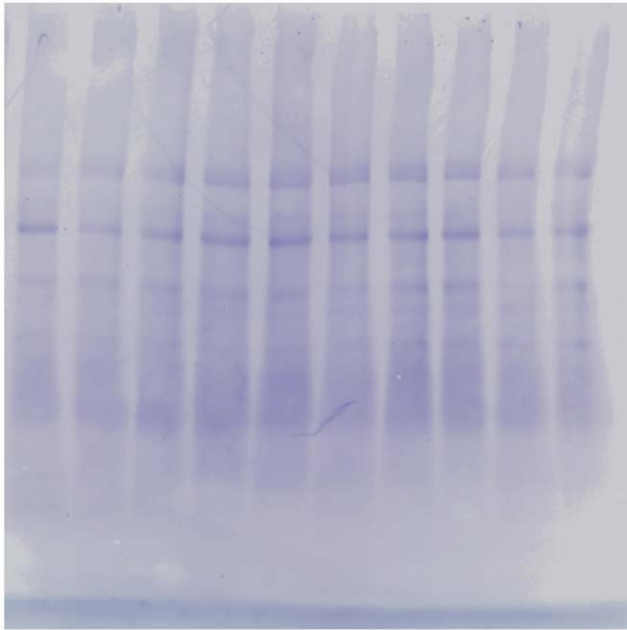


Figure 7. Loading control. The membrane used for immunoblotting was stained by Coomassie blue R-350. Adequate loading was shown. doi:10.1371/journal.pone.0032082.g007

length and latency to reach the goal box. On the eighth experimental day (memory retention), subjects were undergoing a probe trial for 5 min. Mice were allowed to explore the maze and path length, time to reach the goal and correct and wrong decisions were recorded.

Yoked controls were placed into the MTM to remain the same time as their trained mates, but without food provided. Since animals were exposed to the same spatial cues, but without food,

mice did not develop an association between the extra-maze cues and the location of the food.

After completion of each cognitive test, mice were deeply anaesthetized (CO_2) and sacrificed by neck dislocation. Hippocampi were rapidly dissected and stored at -80°C for further proteomic and biochemical analysis.

Protein studies

Sample preparation. 12 hippocampi of trained and untrained mice each were homogenized in ice-cold homogenization buffer [10 mM HEPES, pH 7.5, 300 mM sucrose, one complete protease inhibitor tablet (Roche Molecular Biochemicals, Mannheim, Germany) per 50 mL] by Ultra-Turrax (IKA, Staufen, Germany). The homogenate was centrifuged for 10 min at $1,000 \times g$ and the pellet was discarded. The supernatant was centrifuged at $50,000 \times g$ for 30 min in an ultracentrifuge (Beckman Coulter Optima-L-90K). Subsequently, the pellet was homogenized in 5 mL washing buffer (homogenization buffer without sucrose), kept on ice for 30 min and centrifuged at $50,000 \times g$ for 30 min. All individual 24 samples were used for the gel experiments, using sucrose gradient ultracentrifugation for membrane fractionation. The plasma membrane purification procedures from the pellet were carried out as described previously, with slight modifications [65,88,89]. Sucrose density gradient centrifugation solutions of 700 μL each of 69, 54, 45, 41, and 37% (w/v) were formed. Membrane pellets in 500 μL were resuspended in homogenization buffer, layered on top of the tubes that were filled with homogenization buffer. Samples were ultracentrifuged at 4°C at $70,000 \times g$ for 3 h. After centrifugation the 41% fraction from the sucrose interface was collected, diluted 10 times with homogenization buffer, and then ultracentrifuged at 4°C at $100,000 \times g$ for 30 min. After discarding the supernatant, the pellet was stored at -80°C until use.

Blue native-polyacrylamide gel electrophoresis (BN-PAGE). Membrane pellets from the 41% sucrose gradient

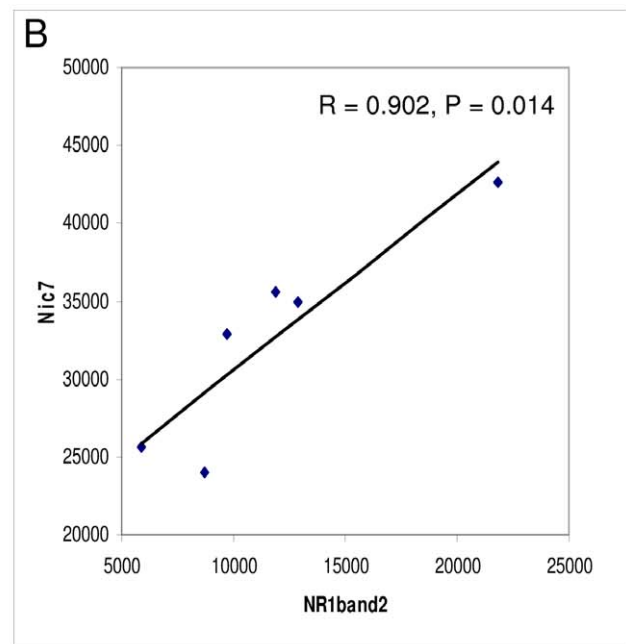
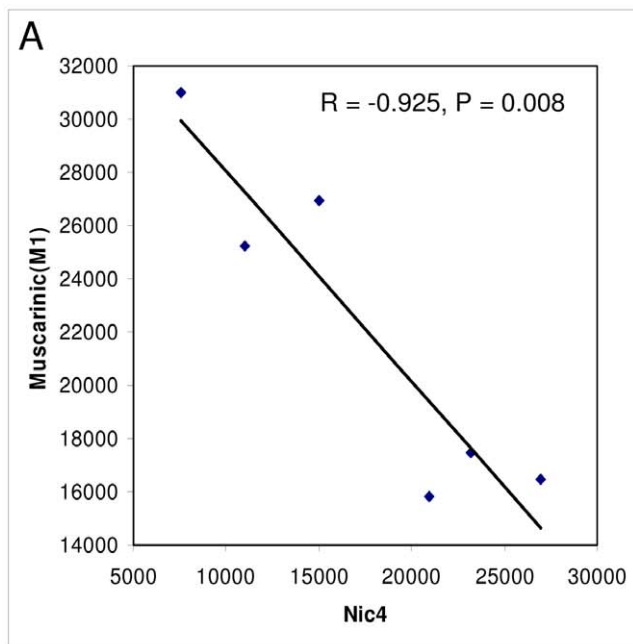


Figure 8. Significant correlations. (A) The significant correlation between Nic4 and M1 complex levels as well as (B) the significant correlation between NR1 (band 2) and Nic7 is demonstrated. doi:10.1371/journal.pone.0032082.g008

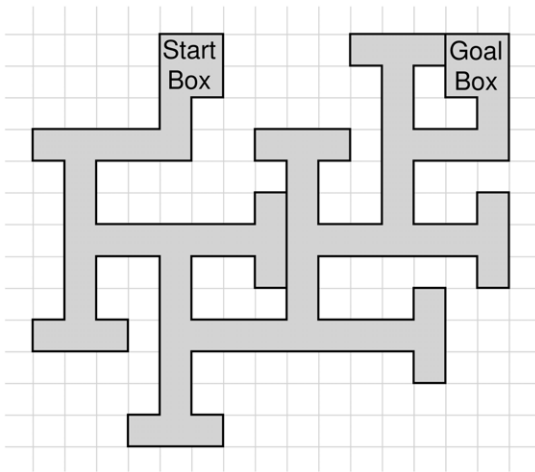


Figure 9. Scheme of multiple T-maze. The multiple T-maze is a landmark paradigm testing spatial memory. Animals with food deprivation search for food that is provided in the goal box. doi:10.1371/journal.pone.0032082.g009

ultracentrifugation fraction were solubilized in extraction buffer [1.5 M 6-aminocaproic acid, 300 mM Bis-Tris, pH 7.0] and 10% DDM (n-dodecyl β -D-maltoside) [to achieve final 1% DDM concentration] with vortexing every 10 min for 1 h. Following solubilization, samples were cleared by centrifugation at $20,000 \times g$ for 60 min at 4°C . The protein content was estimated using the BCA protein assay kit (Pierce, Rockford, IL, USA). 50 μg of the membrane protein preparation were applied onto gels. 16 μL BN PAGE loading buffer [5% (w/v) Coomassie G250 in 750 mM 6-aminocaproic acid] were mixed with 100 μL of the membrane protein preparation and loaded onto the gel. BN-PAGE was performed in a PROTEAN II xi Cell (BioRad, Germany) using 4% stacking and 5–18% separating gel. The BN-PAGE gel buffer contained 500 mM 6-aminocaproic acid, 50 mM Bis-Tris, pH 7.0; the cathode buffer 50 mM Tricine, 15 mM Bis-Tris, 0.05% (w/v) Coomassie G250, pH 7.0; and the anode buffer

References

- Bartus RT, Dean RL, 3rd, Beer B, Lipka AS (1982) The cholinergic hypothesis of geriatric memory dysfunction. *Science* 217: 408–414.
- Hulme EC, Birdsall NJ, Burgen AS, Mehta P (1978) The binding of antagonists to brain muscarinic receptors. *Mol Pharmacol* 14: 737–750.
- Schmeller T, Sporer F, Sauerwein M, Wink M (1995) Binding of tropane alkaloids to nicotinic and muscarinic acetylcholine receptors. *Pharmazie* 50: 493–495.
- Barak S, Weiner I (2009) Towards an animal model of an antipsychotic drug-resistant cognitive impairment in schizophrenia: scopolamine induces abnormally persistent latent inhibition, which can be reversed by cognitive enhancers but not by antipsychotic drugs. *Int J Neuropsychopharmacol* 12: 227–241.
- Buccafusco JJ, Terry AV, Jr., Webster SJ, Martin D, Hohnadel EJ, et al. (2008) The scopolamine-reversal paradigm in rats and monkeys: the importance of computer-assisted operant-conditioning memory tasks for screening drug candidates. *Psychopharmacology (Berl)* 199: 481–494.
- Cunha GM, Canas PM, Melo CS, Hockemeyer J, Muller CE, et al. (2008) Adenosine A2A receptor blockade prevents memory dysfunction caused by beta-amyloid peptides but not by scopolamine or MK-801. *Exp Neurol* 210: 776–781.
- Loiseau F, Dekeyne A, Millan MJ (2008) Pro-cognitive effects of 5-HT6 receptor antagonists in the social recognition procedure in rats: implication of the frontal cortex. *Psychopharmacology (Berl)* 196: 93–104.
- Vaisman N, Pelled D (2009) n-3 phosphatidylserine attenuated scopolamine-induced amnesia in middle-aged rats. *Prog Neuropsychopharmacol Biol Psychiatry* 33: 952–959.
- Klinkenberg I, Blokland A (2011) A comparison of scopolamine and biperiden as a rodent model for cholinergic cognitive impairment. *Psychopharmacology (Berl)* 215: 549–566.
- Boren JJ, Navarro AP (1959) The action of atropine, benactyzine, and scopolamine upon fixed-interval and fixed-ratio behavior. *J Exp Anal Behav* 2: 107–115.
- Hearst E (1959) Effects of scopolamine on discriminated responding in the rat. *J Pharmacol Exp Ther* 126: 349–358.
- Carlton PL (1963) Cholinergic mechanisms in the control of behavior by the brain. *Psychol Rev* 70: 19–39.
- Wiener NI, Messer J (1973) Scopolamine-induced impairment of long-term retention in rats. *Behav Biol* 9: 227–234.
- Brito GN, Davis BJ, Stopp LC, Stanton ME (1983) Memory and the septo-hippocampal cholinergic system in the rat. *Psychopharmacology (Berl)* 81: 315–320.
- Bermudez-Rattoni F, Coburn KL, Fernandez J, Chavez AF, Garcia J (1987) Potentiation of odor by taste and odor aversions in rats are regulated by cholinergic activity of dorsal hippocampus. *Pharmacol Biochem Behav* 26: 553–559.
- Messer WS, Jr., Thomas GJ, Hoss W (1987) Selectivity of pirenzepine in the central nervous system. II. Differential effects of pirenzepine and scopolamine on performance of a representational memory task. *Brain Res* 407: 37–45.
- Blokland A, Honig W, Raaijmakers WG (1992) Effects of intra-hippocampal scopolamine injections in a repeated spatial acquisition task in the rat. *Psychopharmacology (Berl)* 109: 373–376.
- Ohno M, Watanabe S (1996) D-cycloserine, a glycine site agonist, reverses working memory failure by hippocampal muscarinic receptor blockade in rats. *Eur J Pharmacol* 318: 267–271.
- Watts J, Stevens R, Robinson C (1981) Effects of scopolamine on radial maze performance in rats. *Physiol Behav* 26: 845–851.
- Eckerman DA, Gordon WA, Edwards JD, MacPhail RC, Gage MI (1980) Effects of scopolamine, pentobarbital, and amphetamine on radial arm maze performance in the rat. *Pharmacol Biochem Behav* 12: 595–602.
- Buresova O, Bolhuis JJ, Bures J (1986) Differential effects of cholinergic blockade on performance of rats in the water tank navigation task and in a radial water maze. *Behav Neurosci* 100: 476–482.
- Decker MW, Gill TM, McGaugh JL (1990) Concurrent muscarinic and beta-adrenergic blockade in rats impairs place-learning in a water maze and retention of inhibitory avoidance. *Brain Res* 513: 81–85.

50 mM Bis-Tris, pH 7.0. The voltage was set to 50 V for 1 h, 75 V for 6 h, and was increased sequentially to 400 V (maximum current 15 mA/gel, maximum voltage 500 V) until the dye front reached the bottom of the gel [65,89]. Native high molecular mass markers were obtained from Invitrogen (Carlsbad, CA, USA).

Western blots. Membrane proteins were transferred from BN-PAGE and BN/SDS-PAGE to PVDF membranes. After blocking of membranes for 1 h with 10% non-fat dry milk in 0.1% TBST (100 mM Tris-HCL, 150 mM NaCl, pH 7.5, 0.1% Tween 20), membranes were incubated with diluted primary antibodies rabbit anti-mouse Muscarinic M1 (1:3,000, Abcam, ab75178; Cambridge, UK), rabbit anti-mouse Nicotinic Acetylcholine Receptor alpha 4 (1:5,000, Abcam, Cambridge, ab41170, UK) rabbit anti-mouse Nicotinic Acetylcholine Receptor alpha 7 (1:25,000, Abcam, Cambridge, ab 23832, UK), rabbit anti-mouse NMDAR1 (1:5,000, Abcam, Cambridge, ab 28669, UK) and detected with horseradish peroxidase-conjugated anti-rabbit IgG (Abcam, Cambridge, UK). Membranes were developed with the ECL Plus Western Blotting Detection System (GE Healthcare, Buckinghamshire, UK). Arbitrary optical densities of immunoreactive bands were measured by the Image J software program (<http://rsb.info.nih.gov/ij/>) [80]. Loading controls were carried out by staining membranes by Coomassie blue as given in a previous publication [90].

Statistical calculations

Results from the MTM were analyzed by ANOVA. The level of probability was considered significant at $P \leq 0.05$. Data from Western blotting were handled by unpaired Student's t test and data are given as means \pm SD. Pearson correlations were calculated for relations between receptor systems. Calculations were performed using SPSS for windows 15.0 [80].

Author Contributions

Conceived and designed the experiments: GL. Performed the experiments: SKF AD. Analyzed the data: SKF. Contributed reagents/materials/analysis tools: HH AP. Wrote the paper: SKF GL.

23. Saucier D, Hargreaves EL, Boon F, Vanderwolf CH, Cain DP (1996) Detailed behavioral analysis of water maze acquisition under systemic NMDA or muscarinic antagonism: nonspatial pretraining eliminates spatial learning deficits. *Behav Neurosci* 110: 103–116.
24. Riekkinen P, Jr., Sirvio J, Valjakka A, Pitkanen A, Partanen J, et al. (1990) The effects of concurrent manipulations of cholinergic and noradrenergic systems on neocortical EEG and spatial learning. *Behav Neural Biol* 54: 204–210.
25. Cain DP (1998) Testing the NMDA, long-term potentiation, and cholinergic hypotheses of spatial learning. *Neurosci Biobehav Rev* 22: 181–193.
26. Lieben CK, Blokland A, Sik A, Sung E, van Nieuwenhuizen P, et al. (2005) The selective 5-HT6 receptor antagonist Ro4368554 restores memory performance in cholinergic and serotonergic models of memory deficiency in the rat. *Neuropsychopharmacology* 30: 2169–2179.
27. Thouvarcq R, Caston J, Protais P (2007) Cholinergic system, rearing environment and trajectory learning during aging in mice. *Physiol Behav* 90: 155–164.
28. Harrison FE, Hossaini AH, Dawes SM, Weaver S, May JM (2009) Ascorbic acid attenuates scopolamine-induced spatial learning deficits in the water maze. *Behav Brain Res* 205: 550–558.
29. Godding PR, Rush JR, Beatty WW (1982) Scopolamine does not disrupt spatial working memory in rats. *Pharmacol Biochem Behav* 16: 919–923.
30. Remer UD, Oertel R, Kirch W (2005) Pharmacokinetics and pharmacodynamics in clinical use of scopolamine. *Ther Drug Monit* 27: 655–665.
31. Anagnostaras SG, Murphy GG, Hamilton SE, Mitchell SL, Rahnema NP, et al. (2003) Selective cognitive dysfunction in acetylcholine M1 muscarinic receptor mutant mice. *Nat Neurosci* 6: 51–58.
32. Levey AI, Kitt CA, Simonds WF, Price DL, Brann MR (1991) Identification and localization of muscarinic acetylcholine receptor proteins in brain with subtype-specific antibodies. *J Neurosci* 11: 3218–3226.
33. Wei J, Walton EA, Milici A, Buccafusco JJ (1994) m1–m5 muscarinic receptor distribution in rat CNS by RT-PCR and HPLC. *J Neurochem* 63: 815–821.
34. Park SJ, Kim DH, Lee IK, Jung WY, Park DH, et al. (2010) The ameliorating effect of the extract of the flower of *Prunella vulgaris* var. *lilacina* on drug-induced memory impairments in mice. *Food Chem Toxicol* 48: 1671–1676.
35. Bonner TI, Buckley NJ, Young AC, Brann MR (1987) Identification of a family of muscarinic acetylcholine receptor genes. *Science* 237: 527–532.
36. Peralta EG, Ashkenazi A, Winslow JW, Smith DH, Ramachandran J, et al. (1987) Distinct primary structures, ligand-binding properties and tissue-specific expression of four human muscarinic acetylcholine receptors. *Embo J* 6: 3923–3929.
37. Bonner TI, Young AC, Brann MR, Buckley NJ (1988) Cloning and expression of the human and rat m5 muscarinic acetylcholine receptor genes. *Neuron* 1: 403–410.
38. Lefkowitz RJ (2007) Seven transmembrane receptors: something old, something new. *Acta Physiol (Oxf)* 190: 9–19.
39. Buckley NJ, Bonner TI, Brann MR (1988) Localization of a family of muscarinic receptor mRNAs in rat brain. *J Neurosci* 8: 4646–4652.
40. Weiner DM, Levey AI, Brann MR (1990) Expression of muscarinic acetylcholine and dopamine receptor mRNAs in rat basal ganglia. *Proc Natl Acad Sci U S A* 87: 7050–7054.
41. Conn PJ, Jones CK, Lindsley CW (2009) Subtype-selective allosteric modulators of muscarinic receptors for the treatment of CNS disorders. *Trends Pharmacol Sci* 30: 148–155.
42. Fornari RV, Moreira KM, Oliveira MG (2000) Effects of the selective M1 muscarinic receptor antagonist dicyclomine on emotional memory. *Learn Mem* 7: 287–292.
43. Kimura Y, Ohue M, Kitaura T, Kihira K (1999) Amnesic effects of the anticholinergic drugs, trihexyphenidyl and biperiden: differences in binding properties to the brain muscarinic receptor. *Brain Res* 834: 6–12.
44. Soares JC, Fornari RV, Oliveira MG (2006) Role of muscarinic M1 receptors in inhibitory avoidance and contextual fear conditioning. *Neurobiol Learn Mem* 86: 188–196.
45. Roldan G, Bolanos-Badillo E, Gonzalez-Sanchez H, Quirarte GL, Prado-Alcala RA (1997) Selective M1 muscarinic receptor antagonists disrupt memory consolidation of inhibitory avoidance in rats. *Neurosci Lett* 230: 93–96.
46. Wezenberg E, Verkes RJ, Sabbe BG, Ruigt GS, Hulstijn W (2005) Modulation of memory and visuospatial processes by biperiden and rivastigmine in elderly healthy subjects. *Psychopharmacology (Berl)* 181: 582–594.
47. Caulfield MP (1993) Muscarinic receptors—characterization, coupling and function. *Pharmacol Ther* 58: 319–379.
48. Volpicelli LA, Levey AI (2004) Muscarinic acetylcholine receptor subtypes in cerebral cortex and hippocampus. *Prog Brain Res* 145: 59–66.
49. Le Novere N, Grutter T, Changeux JP (2002) Models of the extracellular domain of the nicotinic receptors and of agonist- and Ca²⁺-binding sites. *Proc Natl Acad Sci U S A* 99: 3210–3215.
50. Rush R, Kuryatov A, Nelson ME, Lindstrom J (2002) First and second transmembrane segments of alpha3, alpha4, beta2, and beta4 nicotinic acetylcholine receptor subunits influence the efficacy and potency of nicotine. *Mol Pharmacol* 61: 1416–1422.
51. Lena C, Changeux JP (1998) Allosteric nicotinic receptors, human pathologies. *J Physiol Paris* 92: 63–74.
52. Cordero-Erausquin M, Marubio LM, Klink R, Changeux JP (2000) Nicotinic receptor function: new perspectives from knockout mice. *Trends Pharmacol Sci* 21: 211–217.
53. McGehee DS (1999) Molecular diversity of neuronal nicotinic acetylcholine receptors. *Ann N Y Acad Sci* 868: 565–577.
54. Hogg RC, Raggenbass M, Bertrand D (2003) Nicotinic acetylcholine receptors: from structure to brain function. *Rev Physiol Biochem Pharmacol* 147: 1–46.
55. Jones S, Sudweeks S, Yakel JL (1999) Nicotinic receptors in the brain: correlating physiology with function. *Trends Neurosci* 22: 555–561.
56. Salminen O, Murphy KL, McIntosh JM, Drago J, Marks MJ, et al. (2004) Subunit composition and pharmacology of two classes of striatal presynaptic nicotinic acetylcholine receptors mediating dopamine release in mice. *Mol Pharmacol* 65: 1526–1535.
57. Salminen O, Drapeau JA, McIntosh JM, Collins AC, Marks MJ, et al. (2007) Pharmacology of alpha-conotoxin MII-sensitive subtypes of nicotinic acetylcholine receptors isolated by breeding of null mutant mice. *Mol Pharmacol* 71: 1563–1571.
58. Marks MJ, Collins AC (1982) Characterization of nicotine binding in mouse brain and comparison with the binding of alpha-bungarotoxin and quinuclidinyl benzilate. *Mol Pharmacol* 22: 554–564.
59. Whiteaker P, Davies AR, Marks MJ, Blagbrough IS, Potter BV, et al. (1999) An autoradiographic study of the distribution of binding sites for the novel alpha7-selective nicotinic radioligand [3H]-methyllycaconitine in the mouse brain. *Eur J Neurosci* 11: 2689–2696.
60. Broide RS, Leslie FM (1999) The alpha7 nicotinic acetylcholine receptor in neuronal plasticity. *Mol Neurobiol* 20: 1–16.
61. Wonnacott S (1997) Presynaptic nicotinic ACh receptors. *Trends Neurosci* 20: 92–98.
62. Perry DC, Xiao Y, Nguyen HN, Musachio JL, Davila-Garcia MI, et al. (2002) Measuring nicotinic receptors with characteristics of alpha4beta2, alpha3beta2 and alpha3beta4 subtypes in rat tissues by autoradiography. *J Neurochem* 82: 468–481.
63. Kenney JW, Gould TJ (2008) Modulation of hippocampus-dependent learning and synaptic plasticity by nicotine. *Mol Neurobiol* 38: 101–121.
64. Marino MJ, Rouse ST, Levey AI, Potter LT, Conn PJ (1998) Activation of the genetically defined m1 muscarinic receptor potentiates N-methyl-D-aspartate (NMDA) receptor currents in hippocampal pyramidal cells. *Proc Natl Acad Sci U S A* 95: 11465–11470.
65. Ghafari M, Patil SS, Hoger H, Pollak A, Lubec G (2011) NMDA-complexes linked to spatial memory performance in the Barnes maze in CD1 mice. *Behav Brain Res* 221: 142–148.
66. Li HB, Matsumoto K, Tohda M, Yamamoto M, Watanabe H (1996) NMDA-but not AMPA-receptor antagonists augment scopolamine-induced spatial cognitive deficit of rats in a radial maze task. *Brain Res* 725: 268–271.
67. Mrzljak L, Levey AI, Goldman-Rakic PS (1993) Association of m1 and m2 muscarinic receptor proteins with asymmetric synapses in the primate cerebral cortex: morphological evidence for cholinergic modulation of excitatory neurotransmission. *Proc Natl Acad Sci U S A* 90: 5194–5198.
68. Harvey J, Balasubramanian R, Collingridge GL (1993) Carbachol can potentiate N-methyl-D-aspartate responses in the rat hippocampus by a staurosporine and thapsigargin-insensitive mechanism. *Neurosci Lett* 162: 165–168.
69. Jones SV (1993) Muscarinic receptor subtypes: modulation of ion channels. *Life Sci* 52: 457–464.
70. Liu L, Zhao R, Bai Y, Stanish LF, Evans JE, et al. (2006) M1 muscarinic receptors inhibit L-type Ca²⁺ current and M-current by divergent signal transduction cascades. *J Neurosci* 26: 11588–11598.
71. Calabresi P, Centonze D, Gubellini P, Pisani A, Bernardi G (1998) Endogenous ACh enhances striatal NMDA-responses via M1-like muscarinic receptors and PKC activation. *Eur J Neurosci* 10: 2887–2895.
72. Ma L, Seager MA, Wittmann M, Jacobson M, Bickel D, et al. (2009) Selective activation of the M1 muscarinic acetylcholine receptor achieved by allosteric potentiation. *Proc Natl Acad Sci U S A* 106: 15950–15955.
73. Boddeke EW, Enz A, Shapiro G (1992) SDZ ENS 163, a selective muscarinic M1 receptor agonist, facilitates the induction of long-term potentiation in rat hippocampal slices. *Eur J Pharmacol* 222: 21–25.
74. Burgard EC, Sarvey JM (1990) Muscarinic receptor activation facilitates the induction of long-term potentiation (LTP) in the rat dentate gyrus. *Neurosci Lett* 116: 34–39.
75. Calabresi P, Centonze D, Gubellini P, Bernardi G (1999) Activation of M1-like muscarinic receptors is required for the induction of corticostriatal LTP. *Neuropharmacology* 38: 323–326.
76. Doralp S, Leung LS (2008) Cholinergic modulation of hippocampal CA1 basal-dendritic long-term potentiation. *Neurobiol Learn Mem* 90: 382–388.
77. Kamsler A, McHugh TJ, Gerber D, Huang SY, Tonegawa S (2010) Presynaptic m1 muscarinic receptors are necessary for mGluR long-term depression in the hippocampus. *Proc Natl Acad Sci U S A* 107: 1618–1623.
78. Hopseian SV, Anwyl R, Rowan MJ (2004) Endogenous acetylcholine lowers the threshold for long-term potentiation induction in the CA1 area through muscarinic receptor activation: in vivo study. *Eur J Neurosci* 20: 1267–1275.
79. Shinoc T, Matsui M, Taketo MM, Manabe T (2005) Modulation of synaptic plasticity by physiological activation of M1 muscarinic acetylcholine receptors in the mouse hippocampus. *J Neurosci* 25: 11194–11200.
80. Ghafari M, Falsafi SK, Hoeger H, Lubec G (2011) Hippocampal levels of GluR1 and GluR2 complexes are modulated by training in the multiple T-Maze in C57BL/6j mice. *Brain Struct Funct*; in press, DOI:10.1007/s00429-00011-00335-00428.

81. Sase A, Khan D, Hoyer H, Lubec G (2011) Intraperitoneal injection of saline modulates hippocampal brain receptor complex levels but does not impair performance in the Morris Water Maze. *Amino Acids*; in press, DOI:10.1007/s00726-00011-01130-00729.
82. Robinson L, Harbaran D, Riedel G (2004) Visual acuity in the water maze: sensitivity to muscarinic receptor blockade in rats and mice. *Behav Brain Res* 151: 277–286.
83. Thouvarcq R, Protais P, Jouen F, Caston J (2001) Influence of cholinergic system on motor learning during aging in mice. *Behav Brain Res* 118: 209–218.
84. Irwin S (1968) Comprehensive observational assessment: Ia. A systematic, quantitative procedure for assessing the behavioral and physiologic state of the mouse. *Psychopharmacologia* 13: 222–257.
85. Rogers DC, Jones DN, Nelson PR, Jones CM, Quilter CA, et al. (1999) Use of SHIRPA and discriminant analysis to characterise marked differences in the behavioural phenotype of six inbred mouse strains. *Behav Brain Res* 105: 207–217.
86. Patil SS, Sunyer B, Hoyer H, Lubec G (2009) Evaluation of spatial memory of C57BL/6J and CD1 mice in the Barnes maze, the Multiple T-maze and in the Morris water maze. *Behav Brain Res* 198: 58–68.
87. Zheng JF, Patil SS, Chen WQ, An W, He JQ, et al. (2009) Hippocampal protein levels related to spatial memory are different in the Barnes maze and in the multiple T-maze. *J Proteome Res* 8: 4479–4486.
88. Chen P, Li X, Sun Y, Liu Z, Cao R, et al. (2006) Proteomic analysis of rat hippocampal plasma membrane: characterization of potential neuronal-specific plasma membrane proteins. *J Neurochem* 98: 1126–1140.
89. Kang SU, Fuchs K, Sieghart W, Lubec G (2008) Gel-based mass spectrometric analysis of recombinant GABA(A) receptor subunits representing strongly hydrophobic transmembrane proteins. *J Proteome Res* 7: 3498–3506.
90. Welinger C, Ekblad L (2010) Coomassie Staining as Loading Control in Western Blot Analysis. *J Proteome Res* 10: 1416–1419.