

A polymorphism in exon 2 of the human LDL-receptor gene (LDLR)

A.K.Soutar

MRC Lipoprotein Team, Hammersmith Hospital, Duane Road, London W12 0HS, UK

Description of Probes: The synthetic oligonucleotides for amplification of exon 2 of the LDL-receptor gene were based on those described by Leitersdorf *et al.* (1). The allele-specific oligonucleotides for detection of the polymorphism were: 5' TCT TTC ACA TCT GT (AKS-32) and 5' ACA GAT GCG AAA GA (AKS-31). The polymorphic base is underlined.

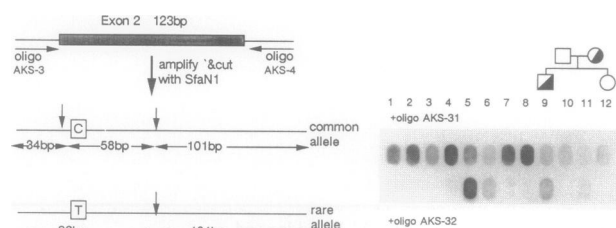
Polymorphism: The polymorphism was first identified as a sequence difference between the published sequence for the LDL-receptor DNA (2) and the sequence of the LDL-receptor cDNA obtained from a patient with homozygous familial hypercholesterolaemia (3). The polymorphic site is substitution of C for T in the third base of the codon for amino acid residue Cys₆ and introduces a recognition site for the restriction enzyme SfaNI. Since the fragments are small it is more convenient to detect the polymorphism by differential oligonucleotide hybridisation.

Protocol: Human genomic DNA was amplified (4) with oligonucleotides flanking exon 2 of the LDL-receptor gene, and the product applied in duplicate to a nylon membrane using standard slot blotting techniques (5). Prehybridisation and hybridisation were carried out each for 1 hour at 29°C in 5×SSPE, 5×Denhardt's and 0.5% (w/v) SDS (5); for hybridisation the ³²P end-labelled oligonucleotide was included at 1×10⁶ cpm/ml. The blots were washed briefly, then twice for 10 min at 22°C. Under these conditions the unmatched oligonucleotide does not hybridise with the amplified fragment (see Figure).

Frequency: In 40 normolipemic individuals from London, UK (non-whites excluded), C-(SfaNI site present) 0.835; T-(SfaNI site absent) 0.165. The less common variant is that originally described for the human LDL-receptor gene (2).

Mendelian Inheritance: Observed in one family.

References: 1) Leitersdorf, E., Tobin, E.J., Davignon, J. and Hobbs, H.H. (1990) *J. Clin. Invest.* **85**, 1014–1023. 2) Yamamoto, T., Davis, C.G., Brown, M.S., Schneider, W.J., Casey, M.L., Goldstein, J.L. and Russell, D.W. (1984) *Cell* **39**, 27–38. 3) Soutar, A.K., Knight, B.L. and Patel, D.D. (1989) *Proc. Natl. Acad. Sci. USA* **86**, 4166–4170. 4) Saiki, R.K., Bugawan, T.L., Horn, G.T., Mullis, K.B. and Erlich, H.A. (1986) *Nature* **324**, 163–166. 5) Maniatis, T., Fritsch, E.F. and Sambrook, J. (1982) *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, p. 122.



A to G polymorphism in ELN gene

G.Tromp, A.Christiano¹, N.Goldstein², Z.Indik², C.Boyd³, J.Rosenbloom², S.Deak, D.Prockop and H.Kuivaniemi*
¹Department of Biochemistry and Molecular Biology, and
²Department of Dermatology, Jefferson Medical College, Philadelphia, PA 19107, ³Center for Oral Health Research, University of Pennsylvania, Philadelphia, PA 19104, and ³Department of Surgery, UMDNJ-RWJMS, New Brunswick, NJ 08903, USA

Source and Description of Sequence Polymorphism: Sequencing of human elastin cDNA clones revealed a nucleotide variant that converted the codon -AGT- (serine) at amino acid position 422 (1) to -GGT-, (glycine) (2). To determine the frequency of this variant, the PCR was carried out with primers below. Digestion of PCR products with BstNI yielded fragments of 8, 11, 17, 89, 111, 167 and 262 bp for the serine allele, and fragments of 8, 11, 17, 18, 89, 111, 167 and 244 bp for the glycine allele. Digestion of the PCR products with RmaI yielded fragments of 235 and 430 bp for the serine allele and undigested full-length fragment of 665 bp for the glycine allele. BstNI and RmaI digestion of PCR products was used to study 129 unrelated individuals: 37 U.S. Americans, 30 Finns, 14 Swedes, 14 Xhosas, 11 Afrikaners, 7 Canadians, 4 Britons, 3 Italians, 2 Dutch, 2 Greeks, 2 Irish, 2 Japanese and 1 German. Heterozygous individuals were present in all nationalities. All experiments were carried out twice with both restriction endonucleases using two different PCR products from the same individual. Water blanks were negative.

5'-CGCTCTAGACAAGGCCTGGGGAAATTTACATCC-3'
(HEL15 primer in IVS19)

5'-CGCAAGCTTCTGGAGGCCTGGGAGCCAGTTTG-3'
(HEL16 primer in IVS21)

Frequency: Estimated from 258 chromosomes: Serine allele (A) 0.415 and Glycine allele (G) 0.585

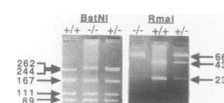
A/A	Expected: 0.172	Observed: 0.163
A/G	0.486	0.504
G/G	0.343	0.333

Chromosomal Location: 7q11.1–21.1 (3).

Other Comments: Amplification conditions 1 min 30 sec at 94°C, 1 min at 62°C and 1 min at 72°C for 50 cycles.

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References: 1) Indik *et al.* (1987) *Proc. Natl. Acad. Sci. USA* **84**, 5680–5684. 2) Christiano (1991) Ph.D. Thesis, Rutgers Univ, NJ, USA. 3) Fazio *et al.* (1991) *Am. J. Hum. Genet.* **48**, 696–703.



* To whom correspondence should be addressed