A CCA/CCG neutral dimorphism in the codon for Pro 626 of the human protein S gene $PS\alpha$ (PROS1)

C.M.Diepstraten, J.K.Ploos van Amstel*, P.H.Reitsma and R.M.Bertina

Haemostasis and Thrombosis Research Unit, University Hospital, PO Box 9600, 2300 RC Leiden, The Netherlands

Exon 15 of the human protein S gene PS α (PROS1) was amplified using the polymerase chain reaction. Sequence analysis showed an A/G neutral dimorphism at the third position of the codon for Pro⁶²⁶ of the PS α gene. The dimorphism can be screened with allele specific oligonucleotides (see figure).

Primer Sequences: PS 15 A (CAAGATGCTAAAAGTCTTGG) and PS 15 B (GATAGCAAGAGAAGTAAGAATTTC) were used in the PCR. PS 626 A (CTCATGTCCATCAGTTTGG) and PS 626 G (CTCATGTCCGTCAGTTTGG) were used as allele specific oligonucleotides in the hybridization.

PCR conditions: Amplifications, 5'-end labelling of PS 626 A and PS 626 G and the oligonucleotide hybridization of the PCR fragments were performed essentially as described (2).

Chromosomal Localization: The human protein S locus that consists of the active $PS\alpha$ gene and the pseudogene $PS\beta$ is localized on chromosome $3p11.1 \rightarrow 3q11.2$ (3, 4).

Frequency:

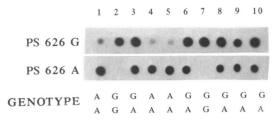
CCG allele (A1) 0.48

CCA allele (A2) 0.52

studied in 28 unrelated European Caucasians.

Mendelian Inheritance: Demonstrated in two families.

References: 1) Ploos van Amstel, J.K., Reitsma, P.H., Van der Logt, C.P.E. and Bertina, R.M. (1990) Biochemistry 29, 7853–7861. 2) Bertina, R.M., Ploos van Amstel, J.K., van Wijngaarden, A., Coenen, J., Leemhuis, M.P., Deutz-Terlouw, P.P., van der Linden, I.K. and Reitsma, P.H. (1990) Blood 76, 538–548. 3) Ploos van Amstel, J.K., Van der Zanden, A.L., Bakker, E., Reitsma, P.H. and Bertina, R.M. (1978) Thromb. Haemostas. 58, 982–987. 4) Watkins, P.C., Eddy, R., Fukushima, Y., Byers, M.G., Cohen, E.H., Dackowski, W.R., Wydro, R.M. and Shows, T.B. (1988) Blood 71, 238–241.



^{*} To whom correspondence should be addressed

A *Mae*III polymorphism within intron A of the insulin (INS) gene detectable by PCR

P.R.Hoban* and A.M.Kelsey1

CRC Department of Cancer Genetics, Paterson Institute for Cancer Research, Christie Hospital and Holt Radium Institute, Wilmslow Road, Manchester M20 9BX and ¹Department of Pathology, Royal Manchester Childrens Hospital (RMCH), Hospital Road, Pendlebury, Manchester M27 1HA, UK

Source/Description: Oligonucleotide primers were used to amplify a 350 bp genomic fragment from within intron A of the insulin gene. A sequencing variant from within this region had previously been described (1).

PCR Primers:

5'GCTGCATCAGAAGAGGCCATC 3' 5'ACACTAGGTAGAGAGCTTCCA 3'

Method: 1 μ g of genomic DNA was amplified using 0.5 μ g of each primer in a total volume of 100 μ l containing $1 \times Taq$ polymerase buffer (BCL) and 200 μ M dNTPs. Samples were heated to 97°C for 5 min after which 2 units of Taq polymerase (BCL) were added. Reactions were cycled 30 times at 94°C for 1 min, 58°C for 1 min and 74°C for 1 min. Amplified products and restriction enzyme digests (10 μ l of amplified product in a total volume of 50 μ l) were analysed by electrophoresis through a 3% agarose gel.

Polymorphism: Digestion with MaeIII reveals a two allele system, allele R1 is 350 bp and allele R2 is 197 bp and 153 bp.

Frequency: Determined from a population of 50 unrelated individuals

Allele R1: 0.37 Allele R2: 0.63

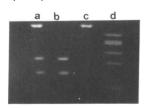
The frequency of heterozygotes is 42% (21 out of 50 individuals).

Mendelian Inheritance: Codominant segregation was observed in three families (14 individuals).

Chromosomal Localisation: The insulin gene has been localised to 11p15.5 (2).

Acknowledgements: This work was supported by RMCH from the Pendlebury leukaemia Research fund.

References: 1) Ulrich, A. et al. (1980) Science 209, 612-615. 2) Bell, G.I. et al. (1982) Nature 295, 31-35.



The figure shows amplification products from three individuals digested with MaeIII (a-c). Size marker (d) is $\Phi X 174$ HaeIII digest.

^{*} To whom correspondence should be addressed