

NOTE

SmD1 Is Required for Spliced Leader RNA Biogenesis

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The Sm-binding site of the kinetoplastid spliced leader RNA has been implicated in accurate spliced leader RNA maturation and *trans*-splicing competence. In *Trypanosoma brucei*, RNA interference-mediated knock-down of SmD1 caused defects in spliced leader RNA maturation, displaying aberrant 3'-end formation, partial formation of cap 4, and overaccumulation in the cytoplasm; U₂₈ pseudouridylation was unaffected.

Spliced leader RNA plays a central role in kinetoplastid gene expression, as it is *trans*-spliced onto every nucleus-derived mRNA. Spliced leader RNA maturation is a multistep process involving precursor 3' extension removal (16, 18), 5'-end methylation (2), pseudouridylation of U₂₈ (ψ_{28}) (8), and trafficking to the cytoplasm mediated by exportin 1 (21). The details of spliced leader RNA nuclear import have not yet been described, and the order and intracellular locations of the various modifications have not been defined unambiguously.

A key structural element of spliced leader RNA is the Sm-binding site (11, 16), RAU_{4,6}GR, that is also present on many of the small nuclear RNAs (snRNAs) involved in splicing. The Sm-binding site interacts with a heptameric complex of Sm proteins (20). The mammalian U1 snRNP is assembled via three RNA-free Sm subcomplexes, D1-D2, F-E-G, and D3-B, that form a ring around the single-stranded Sm-binding site (15). None of the subcomplexes can associate with the Sm-binding site in isolation (13). The importance of the spliced leader RNA Sm-binding site has been demonstrated by mutagenesis in *Leishmania tarentolae* (16) and *Leptomonas collosoma* (11), which affected spliced leader RNA maturation and abolished *trans*-splicing. Conversely, Sm-binding site mutants of *Leptomonas seymouri* were competent for *trans*-splicing, but only when delivered on a low-copy-number episome (9). Transcription studies in *Trypanosoma brucei* demonstrated that the Sm-binding site forms a boundary for cap 4 formation (10).

We challenged the role of SmD1 protein (12) in kinetoplastid spliced leader RNA biogenesis by performing a knockdown experiment with RNA interference (RNAi). Our working model of the spliced leader RNA maturation pathway (21) predicted that 5' and 3' processing of spliced leader RNA would be impaired by the absence of SmD1 and that spliced leader RNA would accumulate in the cytosol. Our results sup-

port posttranscriptional cap 4 modification of spliced leader RNA.

Loss of the SmD1 mRNA is lethal to *T. brucei*. An inducible-RNAi plasmid containing SmD1 gene sequence was transfected into *T. brucei*. Upon induction with tetracycline, clonal lines containing the SmD1 RNAi construction ceased division after 24 h, whereas the growth of control 29-13 and noninduced cells was unaffected (Fig. 1A). RNA blotting (Fig. 1B) revealed increasing levels of induced double-stranded RNA and demonstrated that the SmD1 mRNA disappeared at 24 h. Reprobing the blot for a different Sm protein transcript (12) showed no loss of SmG mRNA at 24 h, confirming the specificity of the RNAi effect. Ethidium bromide staining of rRNA bands controlled for the quality and quantity of each RNA sample loaded.

SmD1 is necessary for accurate 3'-end formation. In *L. tarentolae*, spliced leader RNA Sm-binding site mutants lose 3' processing, accumulating in the form of primary transcription products with staggered 3' ends (18). The Sm-protein complex has been implicated in nuclear import of trafficked snRNAs. Thus, the 3'-end phenotype and subcellular localization of spliced leader RNA in the SmD1 protein in the induced RNAi cell line were examined.

High-resolution RNA blots were hybridized initially with a stem-loop I probe (5'-CTACTGGGAGCTTCTCATCA; Fig. 2A), which yielded a complex pattern of bands ranging from 3'-extended products to transcripts below mature spliced leader RNA size. In the uninduced RNA population, mature spliced leader RNA was identified as the band at 142 nucleotides. Based on 5'-end determination by primer extension (data not shown and Fig. 2C), higher-molecular-weight bands represent intermediates at various stages of 3' extension (21). In RNA from tetracycline-induced cells, 3'-extended forms of spliced leader RNA were detected, as were additional longer forms. A major spliced leader RNA band was present at 141 nucleotides with greater relative abundance than the uninduced RNA sample, and minor products were present at single-nucleotide intervals above and below the major band. Two lower-molecular-weight bands migrating at approximately 115 to 117 nucleotides were prominent in the induced RNA.

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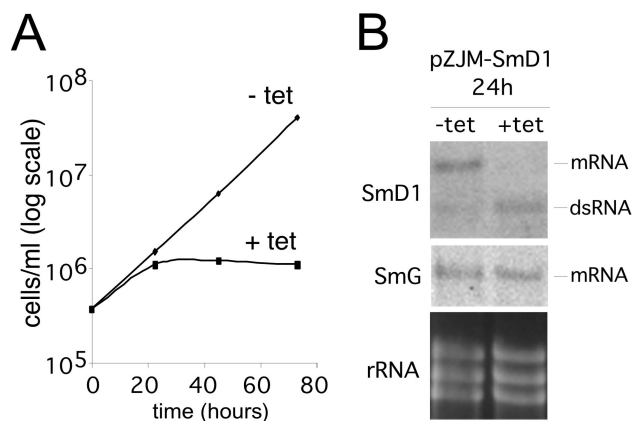


FIG. 1. SmD1 mRNA is required for growth of *T. brucei*. (A) Comparative growth curves of induced and uninduced *T. brucei* (clonal line E) containing a tetracycline (tet)-regulated RNAi construction directed against SmD1 mRNA. (B) SmD1 mRNA is abolished by RNAi at 24 h postinduction. Total-cell RNA from induced and noninduced cultures in separate experiments was size separated on a 1% agarose-formaldehyde gel, blotted, and probed for the presence of SmD1 mRNA. As a control, the gel was stained with ethidium bromide to visualize rRNA bands and the blot was probed for SmG mRNA.

Hybridization of the blot for the spliceosomal U4 snRNA (21), which contains an Sm-binding site, showed a twofold reduction in induced cells, consistent with a stability defect in the absence of SmD1 protein; no 3'-extended forms of U4 snRNA were detected. Spliceosomal U2 snRNA, which also contains an Sm-binding site, was detected by the oligonucleotide 5'-TATCAGGAGTTACTCTGATAAGAACA, revealing equivalent levels in both uninduced and induced cell lines and no longer forms. The unaltered U2 snRNA level is consistent with normal core U2 snRNP assembly in U2 Sm-binding site mutants (6). The relative abundances of U2 snRNA and small ribosomal RNAs visualized by ethidium bromide staining (data not shown) indicated that spliced leader RNA transcript was hyperaccumulating in the induced lines.

The accumulation of spliced leader RNA relative to U2 snRNA may be due to the hundredfold difference in gene copy number (4, 19) coupled with different rates of transcription (14), variable half-lives, and loss of consumption of spliced leader RNA in the *trans*-splicing reaction. As a substrate in *trans*-splicing, spliced leader RNA has a shorter half-life than the other spliceosomal components, estimated to be between 3 min (7) and 1.4 h (5) in *T. brucei*.

SmD1 depletion results in cytoplasmic accumulation of spliced leader RNA. The distribution of spliced leader RNA was analyzed by fluorescence in situ hybridization in the induced SmD1 RNAi cell line. In the absence of the SmD1 mRNA, spliced leader RNA signal became more intense throughout individual cells in an increasing percentage of the population over time (14% of cells at 24 h and 25% at 48 h), consistent with the increased abundance of spliced leader RNA observed in RNA blots. Higher magnification showed that spliced leader RNA was overaccumulating in the cytoplasm (Fig. 2B). The relative abundance and distribution of U3 snoRNA, which does not contain an Sm-binding site, were unchanged in the induced cells. All directly compared images were captured at the same shutter speed and processed in

parallel. We have demonstrated previously that nuclear spliced leader RNA signal is not due to DNA or cytosolic signal to mRNA or autofluorescence (21).

These results demonstrate overaccumulation of spliced leader RNA in SmD1 *T. brucei* knockdown lines, consistent with the molecular phenotype of the Sm-binding site mutations in the *L. tarentolae* spliced leader RNA (16). Substrate spliced leader RNA overaccumulation in the cytoplasm indicated that the SmD1 protein is necessary for spliced leader RNA maturation prior to nuclear import; additional Sm-dependent spliced leader RNA modifications may be required as import signals.

Exon pseudouridylation occurs independent of Sm-binding site interaction. Although ψ_{28} of the spliced leader does not appear to be critical for *trans*-splicing (17) or association with polysomes (21), it may modulate or enhance mRNA interactions with the splicing or translation machinery. To determine whether pseudouridylation of spliced leader RNA is SmD1 dependent, total-cell RNA samples from induced and uninduced SmD1 RNAi lines were subjected to carboxymethyl cellulose treatment (1, 8). Irreversible carboxymethyl cellulose modification of ψ_{28} was visualized by primer extension with the oligonucleotide TbSL stem-loop I (Fig. 2C). As expected, RNA from uninduced cells yielded a termination at position 29 of the exon, indicative of ψ_{28} . RNA from 48-h-induced cells also showed reverse transcriptase termination at this position, indicating that pseudouridylation was unimpaired in the absence of SmD1. An internal control for the integrity of RNA and efficacy of RNAi induction was apparent in the form of extension products terminating at the 5' end of carboxymethyl cellulose-treated spliced leader RNA. In uninduced RNA, a fully modified cap 4 termination product was visible, whereas in induced RNA cap 0/cap 1 termination products were evident.

The localization of SLA1 RNA (8) combined with temporal and Sm-independent acquisition of the pseudouridylation of spliced leader RNA supports modification of ψ_{28} in the nucleus, most likely in the nucleolus, prior to cytosolic trafficking. However, the presence of SLA1 RNA in the nucleolus may be necessitated by its own processing requirements and does not preclude nucleoplasmic or cytosolic spliced leader RNA pseudouridylation. Furthermore, inhibition of cap 4 methylation in induced cell lines mirrored results obtained in Sm-binding site mutagenesis studies of spliced leader RNA in *L. tarentolae* and *Leptomonas collosoma*, in contrast to the cotranscriptional cap 4 acquisition model (10).

SmD1 defects define steps in the spliced leader RNA processing model. Our working model for biogenesis of kinetoplastid spliced leader RNA is as follows. Spliced leader RNA is transcribed as a 3'-extended precursor (18) and receives an m⁷G cap cotranscriptionally (10). The acquisition of ψ_{28} on spliced leader RNA is directed by SLA1 RNA (8) in the nucleolus. The m⁷G cap of nascent spliced leader RNA is recognized by homologues of cap-binding complex and exported to the cytoplasm via a nuclear export complex containing exportin 1 (21). Sm-independent methylations to spliced leader RNA occur posttranscriptionally in the nucleus or cytoplasm. Spliced leader RNA is bound by Sm proteins in the cytosol, an interaction required for accurate 3'-end trimming and further Sm-dependent 5'-end methylation. Localization of these SmD1-dependent events has not been determined experimentally, but

tent with the absence of polyribosome formation on mRNAs lacking mature cap 4 structures (22). The identification and experimental analysis of nuclear import factors, 5' cap structure of nuclear import-blocked spliced leader RNA, and the cap 4 methyltransferases will clarify the roles of Sm proteins and subsequent modifications in spliced leader RNA nuclear import.

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