The Rate of Establishment of Complex Adaptations

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Abstract

A central problem in evolutionary theory concerns the mechanisms by which adaptations requiring multiple mutations emerge in natural populations. We develop a series of expressions that clarify the scaling of the time to establishment of complex adaptations with population size, mutation rate, magnitude of the selective disadvantage of intermediate-state alleles, and the complexity of the adaptation. In general, even in the face of deleterious intermediate steps, the time to establishment is minimized in populations with very large size. Under a broad range of conditions, the time to establishment also scales by no more than the square of the mutation rate, regardless of the number of sites contributing to the adaptive change, demonstrating that the emergence of complex adaptations is only weakly constrained by the independent acquisition of mutations at the underlying sites. Mutator alleles with deleterious side effects have only moderate effects on the rate of adaptation in large populations but can cause a quantum decrease in the time to establishment of some adaptive alleles in small populations, although probably not at a high enough rate to offset the increased deleterious mutation load. Transient hypermutability, whereby a subset of gamete-producing cells mutate at an elevated rate in a nonheritable manner, may also elevate the rate of adaptation, although the effect is modest and appears to result from a simple increase in the rate of transitions between intermediate states rather than from the saltational production of doublet mutations. Taken together, these results illustrate the plausibility of the relatively rapid emergence of specific complex adaptations by conventional population genetic mechanisms and provide insight into the relative incidences of various paths of allelic adaptation in organisms with different population genetic features.

Key words: adaptation, adaptive evolution, evolutionary rate, genome evolution, molecular evolution, mutation rate, mutator, fixation time, transient hypermutability.

Introduction

Understanding the mechanistic origins of complex adaptations (here defined as character alterations requiring more than one novel mutation to yield a functional advantage) remains a central challenge for evolutionary biology (Hartl and Taubes 1998; Orr 2002; DePristo et al. 2005; Dean and Thornton 2007). Some have even guestioned whether conventional mutational mechanisms and current principles of population genetics are capable of explaining the emergence of complex adaptations on reasonable evolutionary time scales (e.g., Behe and Snoke 2004; Pigliucci 2008). Because mutations are rare events, the origin of complex adaptations is often expected to occur at very low rates in small populations, owing to the long cumulative time span necessary for the independent arrival and sequential fixation of multiple mutations. In contrast, whereas large populations provide more individual targets for mutational origin, should the intermediate steps toward a complex adaptation be disadvantageous, the increased efficiency of selection against intermediate mutants in large populations might inhibit adaptational advance.

A number of factors may facilitate the rate of emergence of complex adaptations in ways that defy these simple expectations. First, despite the short persistence times of deleterious intermediate-stage mutations in large populations, the steady input of new mutations results in the maintenance of a small stable reservoir of intermediate alleles poised to take the next step(s) in the path toward adaptation (Gillespie 1984). Likewise, even though nearly all neutral intermediate-step alleles are destined to be lost by drift, their accumulation by mutation pressure can provide a growing resource for secondary adaptive mutations. Second, owing to the reduced efficiency of natural selection, populations with small effective sizes are vulnerable to the accumulation of mutations with mild effects on the efficiency of DNA replication and repair loci (Lynch 2008). The predicted increase in the per capita mutation rate is consistent with the known gradient in the per-generation mutation rate from prokaryotes to multicellular eukaryotes (Lynch 2007) and could offset the decline in the number of individual mutational targets in species with relatively small population sizes. Third, because the principle of selectionmutation balance extends to the loci that define the mutation rate, even large populations will always contain a pool of individuals with mutation rates elevated above the population norm. Although maladapted at the individual level, this small segment of the population might be a major source of evolutionary novelties.

In the following sections, we attempt to incorporate the above-mentioned issues into a more comprehensive framework for understanding the population genetic environments in which complex adaptations are most likely to emerge by alternative routes. A series of analytical approximations, supported by computer simulations, demonstrate

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the existence of many plausible pathways by which complex adaptations can emerge much more rapidly than expected when mutations at independent sites are assumed to proceed to fixation in a sequential manner. Our results also reveal several scaling properties for the time to establishment of complex adaptations with respect to population size, mutation rate, and degree of adaptive complexity, providing the seeds for a general theory for the contexts in which adaptive evolution is most likely to proceed in organisms residing at different positions along these fundamental population genetic axes.

Background

The focus throughout will be on diploid sexual populations, with segregation of chromosomes occurring each generation, but complete linkage between the sites at the locus under consideration. For a variety of aspects of selection, mutation, and random genetic drift, stochastic computer simulations are used to estimate the mean time to establishment of a novel adaptation (defined as the first time at which all alleles in the population are of the adaptive type, starting with a population devoid of such an allele). A Wright-Fisher type of population structure is assumed such that random mating occurs in discrete generations, with the expected frequencies of gametes arising after each generation of selection and mutation being evaluated deterministically with sets of transition equations and then random genetic drift being imposed by multinomial sampling of the population at the expected set of genotypic frequencies. In most cases, it has been possible to obtain analytical approximations to provide mechanistic explanations for the results.

The initial focus will be on the simplest case in which the adaptation is a function of changes at two sites at a particular locus, with the fitness of genotypes at the locus being defined by the two alternative states at each site. These sites may represent a wide variety of molecular contexts: codons for two potentially interacting amino acids within a single protein-coding gene, two latent splice sites for a novel intron, a regulatory site and its associated coding region, or two entirely different interacting linked loci. Both sites will be assumed to be initially fixed in an ancestral state whose genotypic fitness is scaled to equal 1.0. The fitnesses of alternative genotypes involving the two sites will then be assumed to be additively determined such that alleles with single mutations at either site have a reduction in fitness equal to $s_1 \ge 0$, whereas alleles with mutations at both sites have an increment in fitness equal to $s_2 > 0$. Under this scheme, denoting derived mutations with uppercase letters, the fitnesses of (unordered) genotypes Ab/ab and aB/ab are $1 - s_1$, of Ab/Ab, aB/aB, and Ab/aB are $1 - 2s_1$, of Ab/AB and aB/AB are $1 - s_1 + s_2$, of ab/AB is $1 + s_2$, and of AB/AB is $1 + 2s_2$.

Throughout, we assume mutation rates (u) and effective population sizes (N) that are well within the limits of existing biological observations (Lynch 2007). Because the mutations arising at any particular locus are specific to two individual sites, the rates of production of alternative allelic

states are expected to be very low, typically $< 10^{-6}$ per site per generation, and the back-mutation rate is assumed to equal zero.

Results

Constant Mutation Rate

Prior to evaluating the consequences of background variation in the mutation rate, we consider the classical case in which the mutation rate is assumed to be invariant among individuals. For the special situation in which the intermediate state at the adaptive locus is neutral ($s_1 = 0$), some fairly simple analytical approximations for the time to origin and complete fixation of the adaptive allele (hereafter, the time to establishment or \overline{t}_{e}) are attainable. Several aspects of this behavior have been described by previous authors for different ranges of population size (e.g., Gillespie 1984; Stephan 1996; Christiansen et al. 1998; Higgs 1998; Innan and Stephan 2001; Carter and Wagner 2002; Komarova et al. 2003; Iwasa et al. 2004, 2005; Weinreich and Chao 2005; Durrett and Schmidt 2008; Weissman et al. 2009), but the following appears to provide a unifying approach to the problem.

First, as noted by prior authors, if the population is sufficiently small in size, the evolutionary dynamics will proceed as a two-step process, with a first-step mutant almost always becoming fixed prior to the arrival of any second-step mutation. In the initial stages, 4Nu first-step mutations arise per generation (because either A or B mutations can arise in the 2N gametes), each with fixation probability 1/(2N), so the mean arrival time of the first first-step mutation destined to fix is 2u generations. Because the average time to fixation of a neutral allele is 4N generations under the Wright-Fisher model, and the rate of origin of second-step mutations is $\leq 2Nu$ per generation (because there are no more than 2N first-step mutations in the population), there is a negligible chance of a second-step mutation arising prior to fixation of a first-step mutation if $(4N)(2Nu) \ll 1$ or equivalently $N \ll 1/\sqrt{8u}$. Letting

$$\phi_2 = \frac{1 - e^{-4Ns_2p}}{1 - e^{-4Ns_2}} \tag{1}$$

denote the probability of fixation of a beneficial (secondstep) mutation initiating at frequency p (Kimura 1962), then for mutations that fix sequentially, the rate of appearance of the first double mutant destined to fix is the reciprocal of the sum of the average arrival times of the two mutations,

$$r_{\rm s} = \frac{2u}{1 + (1/N\phi_2)}.$$
 (2a)

The initial frequency of the mutant allele is equal to 1/(2N), in which case the probability of fixation can be approximated by $2s_2$ provided $4Ns_2 \gg 1$. The mean time to complete establishment of the double mutant is then

$$\overline{t}_{\rm e} = \frac{1}{r_{\rm s}},\tag{2b}$$

ignoring the time for the second mutation to fix, which is negligible compared with the arrival times of mutations when N is small.

Second, for $N > 1/\sqrt{8u}$, there is a significant chance that a beneficial second-step mutation will arise on a descendant of a first-step mutation prior to fixation of the latter. Although all but a tiny fraction (1/2N) of first-step mutations are destined to be lost by drift, such a process can occasionally rescue such mutants, propelling them to fixation by positive selection. This type of pathway, in which a beneficial second-step mutation goes to fixation prior to the population ever achieving a pure first-step state, has been called stochastic tunneling by Komarova et al. (2003) and Iwasa et al. (2004) in the context of cancer development, and a similar phenomenon has been described in the context of duplicate-gene evolution (Lynch et al. 2001; Lynch 2005). The rate of tunneling with a neutral intermediate has been worked out by Komarova et al. (2003) and Iwasa et al. (2004) in an application of the Moran model, where allele frequency changes occur by a random birth-death process involving single individuals in each unit of time. With some modifications, their results are readily extended to the current case. After accounting for diploidy and the 2-fold reduction in the rate of drift with the Wright-Fisher model, the rate of appearance of the first double mutant destined to fixation by tunneling becomes

$$r_{\rm t} \simeq (1 - {\rm e}^{-4Nu}) \sqrt{u \phi_2}. \tag{3a}$$

Two modifications appear in equation (3a) relative to previous derivations: 1) In prior applications, the leading term in this expression has been 4Nu rather than (1 e^{-4Nu}), and this has constrained the generality of tunneling theory to situations in which 4Nu < 1 because the probability that a given generational cohort can ultimately fix in a population cannot exceed 1.0 (Weissman et al. 2009). This limitation is dealt with by the leading term in equation (3a), which is well approximated by 4Nu when $4Nu \ll 1$, but asymptotically approaches one as $4Nu \rightarrow \infty$. 2) In prior applications, the initial frequency of a mutant allele has been assumed to equal 1/(2N) (as noted above), but in sufficiently large populations (2Nu > 1), more than one mutant allele appears per generation, so a more appropriate estimate of the probability of fixation of a secondary mutation is obtained by solving equation (1) with $p = \frac{1+(2N-1)u}{(2N)}$ (one mutation has definitely arisen, but a fraction u of all other members of a cohort will also have accumulated mutations at the same time).

Accounting for both the sequential and the tunneling paths and including the final time required for the fixation of a beneficial allele (\bar{t}_{f}), the mean number of generations until the establishment of the double mutant generalizes to

$$\overline{t}_{e} \simeq \frac{1}{r_{s} + r_{t}} + \overline{t}_{f},$$
 (3b)

where the final fixation time can be obtained by a deterministic approximation (derived from equation 5.3.13 in Crow and Kimura 1970), with the beneficial allele increasing from frequency [1 + (2N - 1)u]/(2N) to 1 - (1/2N),

$$\overline{t}_{\rm f} \simeq rac{2}{s_2} \ln\left(rac{2N}{1+2Nu}
ight).$$
 (3c)



FIG. 1. Mean number of generations until establishment of a double mutant as a function of the effective population size, with the intermediate states having a selective disadvantage of s_1 . Data points are based on 50-300 independent simulations with a constant mutation rate of $u = 10^{-8}$ per site involved in the adaptation. The double mutant is assumed to have a selective advantage of $s_2 = 0.02$. Simulations were not run for conditions in which the time to establishment was on the order of 10⁹ generations or greater. The continuous lines give the analytical approximations described in the text. (The slight underestimates of the time to establishment at very large N and very small s₁ relative to the simulation results is simply a consequence of the simulations having been initiated under states in which first-step mutations are initially absent, whereas the theory assumes an equilibrium frequency at the outset. It can be shown that the time to attain a fraction x of the equilibrium frequency of the deleterious first-step mutation is $\sim -[\ln(1-x)]/(s_1-u)$ generations. With very small s_1 and large N, the time to reach the initial equilibrium state makes a nonnegligible contribution to \overline{t}_{e} [with $s_{1} = 0.0001$, as in the lowest curve, it takes about 2,000 generations for the frequency of first-step alleles to approximate the equilibrium, and this accounts for the discrepancy in the figure]).

As the fixation time is relatively fast, this final term is of negligible importance when the arrival times of mutations are limiting (4Nu < 1).

Despite its approximate nature, equation (3b) yields predictions that are in close agreement with simulated data over the full range of population sizes (fig. 1). With neutral intermediates, the time to establishment of the double mutant declines monotonically with population size, although there are three approximate domains of scaling behavior. 1) At small population sizes ($N \ll 1/\sqrt{8u}$), establishment of the double mutant almost always proceeds by sequential fixation, with a slight reduction in \overline{t}_{e} as population size increases and selection is more effective at promoting the second-step mutation. 2) At intermediate population sizes, tunneling becomes an increasingly large contributor to the establishment process, with \overline{t}_{e} scaling inversely with N in accordance with equation (3a) provided $N \ll 1/4u$. 3) At very large population sizes, multiple adaptive mutations arise each generation and the time to establishment of the adaptive mutation is limited only by the essentially

deterministic expansion of the aggregate pool to frequency 1.0.

We now turn to the situation in which first-step alleles are disadvantageous. For populations sufficiently small that fixation proceeds only in a sequential manner, the rate of appearance of double mutants destined to fix is

$$r_{\rm s} = \frac{2Nu}{(1/2\phi_1) + (1/\phi_{12})},\tag{4a}$$

where ϕ_1 , the probability of fixation of a newly arisen (deleterious) first-step mutation, is obtained by substituting $-s_1$ for s_2 in equation (1) and ϕ_{12} is obtained by applying ($s_1 + s_2$) in place of s_2 . Both fixation probabilities are evaluated with initial frequency p = 1/(2N). At population sizes large enough to prevent fixation of first-step mutations ($N \gg 1/4s_1$), the latter are expected to rapidly approach the low frequency maintained under selection–mutation balance ($2u/s_1$ in this case), providing a launching pad for beneficial second-step alleles. Following the logic leading to the development of equation (3a), earlier results of Komarova et al. (2003) and Iwasa et al. (2004, 2005) can then be modified to obtain an analytical approximation for the rate of tunneling,

$$r_{\rm t} \simeq 1 - {\rm e}^{-4Nu^2\phi_2/s_1}$$
, (4b)

where ϕ_2 is evaluated with p = 1/(2N). The structure of this equation can be understood as follows. In each generation, there are an expected $4Nu/s_1$ copies of the firststep mutation segregating in the population, a fraction uof which are converted to second-step mutations, which fix with probability ϕ_2 . As above, use of the exponential approximation accounts for the fact that no more than one tunneling event can occur per cohort, although with $4Nu^2\phi_2/s_1\phi_2 \ll 1$, which will be satisfied in all but enormous populations, the preceding expression is adequately approximated by $r_t \simeq 4Nu^2\phi_2/s_1$. Provided $N \gg 1/4s_2$, ϕ_2 is close enough to $2s_2$ that r_t further simplifies to \simeq $8Nu^2s_2/s_1$.

The predicted mean time to establishment with deleterious intermediates, which is given by equation (3b) using equations (4a, b) and (3c) to define r_s , r_t , and \overline{t}_f , respectively, fits the simulated data quite closely throughout the entire range of population sizes and selection coefficients (fig. 1). Again, it can be seen that there are three predominant domains of behavior with respect to population size. 1) At small population sizes, establishment of the double mutant almost always proceeds by sequential fixation, but after a slight decline in \overline{t}_{e} with initial increases in population size, there is an increase in \overline{t}_{e} . This increase, which is quite dramatic when first-step mutations are highly deleterious, arises in populations that are large enough that there is a very low probability of fixing a deleterious first-step mutation but small enough that the rate of tunneling is also very low. 2) For population sizes in excess of $4/s_1$, there is essentially no chance of establishment by sequential fixation, and the process of adaptive evolution is governed entirely by tunneling, the rate of which scales nearly linearly with population size. 3) At enormous population sizes, there is again

a slowdown in the decrease in \overline{t}_e with increasing N due to the saturating effect of multiple tunneling mutations being produced per generation and the added constraint on the time to fixation of such mutations.

Mutator Alleles

All the preceding results (and indeed, almost all prior applications of population genetic theory) assume that all members of the population have identical mutation rates. However, a substantial number of loci produce proteins with roles involved in DNA replication and repair, and like all other genes, they are subject to mutation. As a consequence, variation in the mutation rate must exist among individuals, the most extreme examples being manifest in the numerous human genetic disorders caused by mutations at DNA repair loci (including Fanconi anemia, xeroderma pigmentosum, Bloom's syndrome, Werner syndrome, and Lynch syndrome). This raises the possibility that a small (maladapted) fraction of the population may be a primary source of mutations with long-term adaptive advantages.

To gain a qualitative understanding of this matter, we evaluate the situation in which a single diallelic locus governs the mutation rate. Letting M and m designate alternative alleles at the mutator locus, the per-locus mutation rates for genotypes mm, Mm, and MM are, respectively, denoted as u_0 , u_1 , and u_2 , where the subscript represents the number of mutator alleles in the genotype. Under this scheme, the gametic fractions of mutator (M) alleles produced by mm, Mm, and MM genotypes are u_0 , $(1 + u_1)/2$, and 1.0, respectively. Back mutations (M \rightarrow m) are assumed to occur at a negligible rate because whereas there are multiple ways to deactivate an allele, restoration requires a very specific back mutation. To convert the per-locus mutation rates relevant to the inactivation of m alleles to per-site rates at the adaptive locus, u_0 , u_1 , and u_2 are multiplied by a factor k, here generally assumed to equal 0.01. The definition of a mutator allele used here is general, in that it applies to any allelic variant that magnifies the mutation rate, no matter how small.

Mutator alleles are expected to confer a selective disadvantage (resulting, e.g., from elevated rates of somatic mutation and/or the associated load of linked deleterious germline mutations; Dawson 1999; Johnson 1999; Lynch 2008) such that the fitnesses of the mm, Mm, and MM genotypes are 1, $1 - h_M s_M$, and $1 - s_M$, respectively, where s_M is the selective disadvantage of a mutator homozygote and h_M is a measure of dominance of the mutator allele (0.5 implying additivity). The overall fitness of an individual is then the product of the fitnesses at the mutator and adaptive loci.

If the frequencies of the mutator alleles are set to their expectations (\hat{q}) in the absence of selection on the adaptive locus, this sets up an immediate discontinuity in the mutation rate with respect to population size. For situations in which the heterozygous disadvantage of the mutator allele is smaller than the power of random genetic drift, 1/(2N), selection is incapable of preventing fixation of the mutator allele. With the initial frequency of the M allele being 1.0, all



FIG. 2. Mean number of generations until establishment of an advantageous double mutant as a function of the effective population size in populations containing mutator alleles. The lines unaccompanied by data points give the theoretical expectations for situations in which there is a constant mutation rate per site at the adaptive locus, $u = 10^{-8}$ (from fig. 1). The lines accompanied by data points are the theoretical expectations for situations in which the mutation rate can evolve, obtained using equations (5a-c) and (3b), with the data points derived from computer simulations. The genotypic mutation rates are equal to 10^{-6} , 10^{-5} , and 10^{-4} per locus for the mm, Mm, and MM genotypes, respectively, and in all cases, $s_2 = 0.02$, k = 0.01, and $h_{\rm m} = 0.1$. Solid points and lines: the intermediate (first-step) mutation at the adaptive locus is neutral ($s_1 = 0.0$); open points and dotted lines: the first-step mutation has a 0.1% selective disadvantage $(s_1 = 0.001)$. The lower left sets of lines give the expectations under the assumptions that the mutator allele has fixed. The vertical lines denote the critical threshold population size $N_c = 1/(4h_M s_M)$, above which the mutator allele is expected to be maintained at selectionmutation balance in the early phase of the adaptive process.

the preceding expressions can then be used to estimate \overline{t}_e after substituting u_2k for u. In contrast, when the power of drift is weaker than the selective disadvantage, the mutator allele will be maintained at a low frequency by selection– mutation balance, the expected value of which is obtained by solving equation (6) in Lynch (2008). The approximate effective population size at which this transition in behavior occurs is $N_c \simeq 1/(4h_M s_M)$, assuming $s_M \gg u_0$, which is generally likely to be the case.

For situations in which the M/m polymorphism is maintained, the estimation of \overline{t}_e must take into account the relative probabilities of mutations arising at the adaptive locus on different mutation-rate backgrounds, as outlined in the Appendix. Again, the derived analytical approximations, equations (A1a-c), yield results that are in close accordance with those obtained by stochastic simulations for the full range of population sizes (fig. 2). Several notable points can be gained from these observations.

First, as noted above, there is an abrupt transition in the behavior of the establishment time at population sizes near $N_c = 1/(4h_M s_M)$, with the same scaling with population size above and below this point, but a stepwise reduction

in the establishment time below the critical population size necessary for preventing fixation of the mutator allele. Second, for population sizes larger than N_c , mutator alleles must have very minor heterozygous effects if they are to enhance the overall rate of adaptive evolution. With the mutation rates applied in figure 2, mutator alleles with $h_{\rm M} = 0.1$ and $s_{\rm M} = 0.01$ (i.e., a 0.1% reduction in fitness in heterozygotes) have an equilibrium frequency of 0.001 and essentially no influence on \overline{t}_{e} relative to the case with a constant background mutation rate. Third, for mutators with very weak heterozygous fitness effects, there can be a substantial decline in \overline{t}_{e} at intermediate population sizes, where because the establishment time is otherwise quite long, numerous opportunities exist for successful tunneling events. Fourth, at very large population sizes, the effects of mutator alleles become asymptotically negligible due to the fact that double mutants arise frequently even in nonmutator backgrounds. Taken together, all these factors imply an overall dampening of the influence of population size on the time to establishment of a complex adaptive allele when the mutation rate is an evolvable feature.

Transient Hypermutability

A major limitation of mutator alleles is that they reduce the net selective advantage of the linked beneficial mutations that they produce (and hence the fixation probability of the latter) via the indirect consequences of the additional background rain of linked and unlinked deleterious mutations that they produce in both the germline and the soma. Both these complications were implicitly incorporated into the theory in the preceding sections. To produce scenarios that avoid these long-term costs, previous theoretical attempts to highlight the prospects of mutator alleles in adaptive evolution have assumed a high rate of reversion to nonmutators following the promotion of an adaptation (Taddei et al. 1997).

An alternative mechanism that can yield elevated mutation rates with no heritable long-term disadvantage is transient hypermutability, whereby a subset of germline replication events experience elevated levels of mutagenicity associated with an aberrant DNA polymerase/damage repair apparatus or reaction (Ninio 1991; Drake 2007). Stochastic variation associated with transcription or translation errors, erroneous protein folding, nucleotide pool imbalance, and/or metabolic limitations will inevitably generate intercellular variation in mutation rates, but unlike the situation with mutator alleles, the mutagenic conditions will not be heritable. Thus, transient hypermutability might provide a mechanism for a substantial elevation in the rates of origin of complex adaptations by concentrating the production of multiple mutations within single replication events (Drake et al. 2005; Drake 2007). Observations on elevated frequencies of doublet and triplet mutations relative to random expectations in reporter constructs in microbes and mice and in de novo human disease alleles have encouraged this view (Drake et al. 2005; Drake 2007; Wang et al. 2007; Chen et al. 2008, 2009), and the unexpectedly high clustering of mutations seen in some phylogenetic analyses



FIG. 3. Mean number of generations until establishment of an advantageous double mutant as a function of the effective population size in the presence of transient hypermutability. In all cases, the genotypic mutation rates are equal to 10^{-6} , 10^{-5} , and 10^{-4} per locus for the mm, Mm, and MM genotypes, respectively, and $s_2 = 0.02$, k = 0.01, $h_M = 0.1$, $s_M = 0.01$, and $f_m = 0.01$. Results for $f_m = 0.001$ and x = 1000 are given by crosshairs for comparison with those for $f_m = 0.01$ and x = 100, with both cases implying $f_m x = 1$. The lines give the theoretical expectations; for each set of s_1 , the top line gives the results assuming no transient hypermutability ($f_m = 0$). For parameters leading to mean times to establishment substantially in excess of 10^8 generations, simulation results could not be readily obtained, but it is apparent that the theoretical expressions work well through all domains of analysis.

(Bazykin et al. 2004; Davis et al. 2009; Donmez et al. 2009) might also be a consequence of such effects.

Although there may be a near continuum of cell-specific mutation rates within an individual organism (or population of cells), the data are also compatible with a distinct subset of up to 3% of replication events experiencing an up to 200-fold inflation in the mutation rate. Thus, to evaluate the degree to which transient mutation magnifies the rate of origin of complex adaptations, we assume a population in which a random fraction (f_m) of replicons mutates x times faster than the baseline rate. The expressions for the time to establishment of an adaptive mutation then need to modified in two ways. First, with a baseline mutation rate of u_{i} the expected mutation rate becomes $[1 + (x - 1)f_m]u$, or $\sim (1 + f_m x)u$ for $x \gg 1$. All the preceding expressions for rates and times to fixation then apply after multiplying each mutation rate by $(1 + f_m x)$. Second, the existence of hypermutable cells introduces a new pathway for the production of adaptations-the immediate appearance of an adaptive allele by simultaneous mutations at multiple sites within a single gamete (Appendix). However, even with a substantial increase in the baseline mutation rate in a subset of cells, the probability of the instantaneous production of a double mutant is still a very low probability event, raising the question as to whether this mode of production actually elevates the rate of production of evolutionary adaptations.

Both the simulation and the theoretical results indicate that the mean time to establishment of a complex adaptation scales with population size in essentially the same way in the presence and absence of transient hypermutability (fig. 3). In small populations, transient hypermutability has very small effects because there are still insufficient numbers of mutational targets to avoid the usual path of sequential fixation, whereas at larger population sizes, the rate of tunneling proceeds in the same way as described above after accounting for the increased mutation rate. As essentially the same theoretical results are obtained by ignoring the path of double mutation (Appendix), it appears that transient hypermutability primarily influences the rate of adaptation by simply magnifying the mutation rate and elevating the rate of passage of secondary mutations through the tunneling pathway, rather than by the simultaneous production of double mutants.

The latter point can be seen in the following way. From the preceding expressions, the rate of tunneling is a function of $f_m x$, whereas the rate of production of double mutants is a function of $f_m x^2$. Thus, if transient hypermutability primarily influences the rate of establishment of adaptations by elevating the rate of origin of nonsynchronous mutations, the mean time to establishment should be independent of f_m and x so long as their product remains constant. The results in figure 3 demonstrate that this is the case. For both $(f_m = 0.01, x = 100)$ and $(f_m = 0.001, x = 1000)$, the mean time to establishment behaves as though the mutation rate is doubled (in both cases, $1 + f_m x = 2$), with no need to account for the concentrated incidence of double mutants in the small subset of transient mutators.

Increasing Complexity

Having established the basic behavior of the two-site model, we now consider the situation with larger numbers of sites. Although such complexity introduces numerous higher order effects that thwart the development of a general analytical treatment, some results for limiting conditions are readily obtained. Consider first the situation in which changes at *d* sites are essential to the novel adaptation, with all alleles carrying changes at fewer than *d* sites being neutral. In the first step toward adaptation, there are *d* sites at which changes may occur, whereas there are d - 1 second-step sites, and so on. Therefore, confining our attention to the situation in which the mutation rate is a constant *u* per site, for populations that are sufficiently small that the only route to adaptation involves sequential fixation, which requires $N \ll 1/\sqrt{8(d-1)u}$,

$$\overline{t}_{e,s} \simeq \frac{1}{u} \left(\sum_{i=2}^{d} \frac{1}{i} + \frac{1}{2N\phi_2} \right)$$
(5a)

(Gokhale et al. 2009). Although increasing the number of steps involved in adaptation increases the mean time to establishment, the effect is fairly weak at small population sizes because of the elevated number of paths toward the final adaptation. For d = 2, 3, 4, 5, and 10, the summation in the preceding expression is 0.50, 0.83, 1.08, 1.28, and 1.93,



FIG. 4. Mean number of generations until establishment for complex adaptive alleles involving d = 2, 3, 4, and 5 sites (denoted in the right margin) for the case in which the intermediate states are neutral. The lower solid line gives the theoretical results for d = 2. A constant mutation rate of $u = 10^{-8}$ per site is assumed, and the adaptive allele is assumed to have a selective advantage of $s_2 = 0.02$. The data are based on simulations of 25–100 replicates, and the curved lines are the approximate fits described in the text.

respectively, implying a less than 4-fold increase in \overline{t}_{e} over this 5-fold range of complexity.

The limiting behavior as $N \rightarrow \infty$ can also be inferred for neutral intermediates by treating the population deterministically and noting that fixation of the *d*-step mutation will occur before fixation has occurred at any intermediate step. The expected frequency of *d*-step alleles at time *t* is $\simeq (ut)^d$ for $ut \ll 1$ and taking $1/(4Ns_2)$ to be the frequency at which the probability of fixation is essentially 1.0,

$$\overline{t}_{e,t} \simeq \frac{1}{u(4Ns_2)^{1/d}} + \overline{t}_{f}, \tag{5b}$$

where the latter term again accounts for the time to fixation.

These expressions point to two striking aspects of the rate of origin of complex adaptations via neutral intermediates. First, regardless of the complexity of the adaptation, the time to establishment is inversely proportional to the rate at which mutations arise at single sites, not the low probability of multiple independent mutations (u^d) . Second, with increasing d, the response of \overline{t}_e to population size becomes progressively flatter (fig. 4).

Using a branching process approach, Durrett et al. (2008) concluded that the time to establishment scales inversely with $Nu^{1+(1/2)+\dots+(1/2)^{d-1}}$, but as can be seen in figure 4, the domain of this scaling is extremely restrictive for biologically realistic parameters, as for any particular d, the scaling first increases and then decreases with increasing N, and for d > 3 is never as steep as 1/N, becoming progressively flatter with increasing d. Although we have been unable to obtain a direct analytical approximation for the full behavior of this multidimensional diffusion process, the following ad



FIG. 5. Mean number of generations until establishment for complex adaptive alleles involving d = 2, 3, 4, and 5 sites (denoted in the right margin) for the case in which the intermediate states are deleterious ($s_1 = 0.001$ for all intermediate-state alleles) relative to the starting state. Because of the very extended times to establishment in this case, the mutation rate employed in simulations was quite high ($u = 10^{-6}$); mutator alleles and transient hypermutability are ignored. The adaptive allele is assumed to have a selective advantage of $s_2 = 0.02$, and each data point is based on simulations of 25–100 replicates. The smooth lines are derived from the analytical approximations described in the text.

hoc expression appears to provide a reasonable first-order approximation of the transition of \overline{t}_e from low to high N,

$$\overline{t}_{e} \simeq \theta \overline{t}_{e,s} + (1 - \theta) \overline{t}_{et}$$
 (5c)

···· 1/4

where

$$\theta = \mathrm{e}^{-\alpha(4\mathrm{N}u)^{1/u}}.$$
 (5d)

The most unsatisfactory aspect of this expression is the lack of an explicit definition of α , which governs the rate of decline of \overline{t}_e with increasing N, but for the parameters utilized in figure 4, $\alpha \simeq 6.0$, 3.0, and 2.2 for d = 3, 4, and 5, respectively. Schweinsberg (2008) provides a detailed evaluation of the purely neutral model, yielding approximate scalings for first-arrival times for various ranges of N, although the approximation domains become diminishingly small with increasing d, for example, with nine domains of behavior with respect to N recognized when d = 3.

The scaling of the time to establishment with the number of sites is substantially altered when the intermediate states are deleterious, where \overline{t}_e increases dramatically with increasing *d* at moderate to large population sizes (fig. 5). Such behavior results from the fact that when the power of drift is weak relative to the strength of selection against deleterious intermediate-step alleles, the frequencies of the latter hover near the selection–mutation balance expectation. In this situation, each newly arisen first-step allele is eliminated quite rapidly either by drift or by selection (the expected half-life being no more than $1/s_1$ generations; Garcia-Dorado et al. 2003). In order for tunneling to yield an adaptive allele in this short period of time, a descendant of a

first-step allele must accumulate an additional d-1 mutations, the probability of which becomes diminishingly small with larger d. In contrast, at the smallest population sizes, there is only a small increase in $\overline{t}_{\rm e}$ with increasing complexity because deleterious intermediate-state alleles are capable of drifting to fixation in an effectively neutral manner, thereby providing a progressive stepwise path to the final adaptive state.

For the special situation in which all intermediate-state alleles are equally deleterious (fig. 5), a good analytical approximation to the time to establishment can be obtained as follows. Sequential fixation follows the path in which 1) the first deleterious mutation, which can occur at d sites, drifts to fixation; 2) the next d - 2 mutations drift to fixation in a neutral manner because they are selectively equivalent to the first-step allele; and 3) the final mutation is fixed by positive selection. The mean time to sequential fixation is then

$$\overline{t}_{e,s} \simeq \frac{1}{u} \left(\frac{1}{2Nd\phi_1} + \sum_{i=2}^{d-1} \frac{1}{i} + \frac{1}{2N\phi_{12}} \right),$$
 (6a)

and the rate of sequential fixation is the reciprocal of $\overline{t}_{e,s}$. Fixation via tunneling requires that, prior to being eliminated by selection, a deleterious first-step mutant allele procures all the downstream mutations necessary for progression to fixation by positive selection. Letting the mean time that a deleterious first-step mutation remains segregating prior to elimination by selection be $\sim 1/s_1$ generations, and assuming that each such mutation is generally present in just a single copy most of the time, the rate of tunneling is

$$r_{\rm e,t} \simeq 4Nd! (u/s_1)^d (s_2/s_1).$$
 (6b)

Finally, under the assumption that final-step beneficial mutations generally arise as single copies in any generation, the final fixation time is

$$\overline{t}_{\rm f} \simeq \frac{2\ln(2N)}{s_2}.$$
 (6c)

When these expressions are applied to equation (3b), they are in excellent agreement with simulation data for nearly the full range of population sizes.

The Effect of Recombination

Although recombination has been ignored in the previous analyses, so long as the mutations underlying the adaptations under consideration reside within gene-sized stretches of DNA, the following arguments suggest that recombination will usually either have negligible effects or accelerate the rate of adaptation. First, recombination will have no impact at population sizes where adaptation proceeds entirely by sequential fixation, simply because multiple polymorphic sites are never present simultaneously. Second, in populations of moderate size, recombination between alternative intermediate-state alleles can be a major source of origin of adaptive alleles, although too high a recombination rate will impede the maintenance of an adaptive allele when it is at low frequency. Third, at very large population sizes, where evolution proceeds in a nearly deterministic fashion, multisite mutants are created so frequently that recombination between intermediate-state alleles becomes a negligible source of origin of the final adaptive allele, although the progression of such alleles to fixation will be inhibited if the recombination rate exceeds the selective advantage.

Weinreich and Chao (2005) demonstrated that recombinational breakdown of adaptive alleles will be of negligible significance if adaptive mutants destined to fixation are able to attain a sufficiently high frequency for selection to operate in an effectively deterministic fashion prior to a recombination event. This requires a recombination rate between sites smaller than $-s_2/[1.27 + \ln(s_2)]$, which for s₂ = 0.001, 0.01, and 0.1 is 0.00018, 0.0030, and 0.097, respectively. High-density genetic maps indicate that the recombination rate per site ranges from $\sim 10^{-6}$ / nucleotide site in microbial eukaryotes with small genomes to $\sim 10^{-9}$ in the bloated genomes of animals and land plants (Lynch 2007). To an order of magnitude, genes in the former group are $\sim 10^3$ bp in length, whereas those in multicellular organisms range from $\sim 10^3$ to $\sim 10^6$ bp (including introns). Considering a full range of potential distances between adaptive sites within genes and a minimum of \sim 10, this implies recombination rates between selected sites <0.001 in essentially all situations, ranging down to 10^{-8} for closely spaced sites in multicellular species. These observations suggest that at the within-gene level recombination will generally be a very weak force opposing the establishment of complex adaptive alleles.

Discussion

The preceding results illustrate that the mean times to establishment of complex adaptations often exhibit a simple scaling with basic population genetic parameters. As variables such as N and u vary by orders of magnitude across phylogenetic lineages in predictable ways (Lynch 2007), this opens the possibility of developing a general framework for understanding the likelihood of alternative paths to adaptive evolution in lineages varying with respect to population genetic parameters.

Considering first the situation in which the mutation rate is assumed to be immune to evolutionary change and the intermediate states are neutral, the mean time to establishment of the adaptive allele (\overline{t}_e) monotonically declines with increasing population size, with the steepness of this decline becoming more relaxed when larger numbers of sites are involved in the adaptation, scaling with $N^{-1/d}$ at very large N(fig. 4). If the intermediate states are deleterious, \overline{t}_e is maximized at an intermediate population size, where both the rates of sequential fixation and tunneling are low (figs. 1 and 5), and the log of \overline{t}_e increases linearly with increasing complexity at moderate N and at accelerating rate at very large N (due to the diminishingly low probability of a multisite mutant arising in a deleterious lineage otherwise destined to rapid purging).

In general, the mean time to establishment of a novel adaptive allele scales inversely with the 0.5-2.0 power of

the mutation rate regardless of the level of complexity of the adaptation, provided there are not multiple deleterious states (in which case \overline{t}_{e} scales with 1/u to $1/u^{d}$ depending on the population size). Thus, with increasing numbers of sites necessary for an adaptation, the increase in \overline{t}_{e} is often relatively modest. A primary reason for this behavior is that a multistage route to adaptation increases the number of paths to the final adaptive state by magnifying the numbers of possible mutations in the early stages of the process. At large d, this increase in the number of possible paths comes close to compensating for the increase in the number of steps required to reach the final adaptive state. Remarkably, as can be seen from equation (5b) in very large populations with neutral intermediates, as $d \rightarrow \infty$, the time to establishment converges on the reciprocal of the per-site mutation rate, becoming independent of the number of mutations required for the adaptation.

This scaling of \overline{t}_{e} with population size will be altered when mutator alleles are allowed to enter the population, for under the assumption that the latter have mild deleterious effects, such alleles will only rise to high frequencies in small populations. Thus, allowing for genetic variation in the mutation rate, even with neutral intermediates, \overline{t}_{e} may be maximized at an intermediate population size (fig. 2), although the gradient in change of \overline{t}_{e} with population size will become progressively relaxed when mutator alleles have milder effects on fitness and can even rise to moderate frequencies in intermediate-sized populations. Transient hypermutability, a purely physiological effect in a subset of cells that has no heritable component, appears to increase the rate of adaptation only to the extent that it magnifies the average population-level mutation rate, without altering the scaling with population size noted above.

Overall, our results suggest that segregating mutator alleles and transient hypermutability are unlikely to make major contributions to the origins of adaptations in natural populations unless the former have much milder associative deleterious effects or the latter have much higher incidences or magnifying effects than assumed in the preceding analyses. Although fixed mutator alleles theoretically have an especially high likelihood of spawning adaptive alleles in small populations, one could argue that the latter situation is unlikely to be biologically sustainable, as any population that is capable of fixing alleles with mildly deleterious effects will typically be on a path to extinction (Lynch et al. 1995).

Noting that realistic population sizes, mutation rates, and selection coefficients have been applied throughout, these results suggest that quite complex alleles, with multiple neutral or deleterious intermediate states, can readily emerge in populations on time scales of 10^3-10^8 generations. Thus, for microbes with generation lengths of hours to days and very large population sizes, the mean time to establishment can easily be on the order of a few weeks to several months depending on the complexity of the final allelic state. Even multicellular species, with effective population sizes in the vicinity of 10^6 (Lynch 2007), are capable of establishing fairly complex adaptations on time scales of a few tens of millions

of generations, the exact time span depending on the magnitude of the selective (dis)advantages of the intermediate and end states.

For several reasons, these conclusions should be viewed as guite conservative. First, in the case of neutral intermediates, it has been assumed throughout that the base population is completely devoid of site-specific mutational variants that might contribute to the final adaptation, although firststep alleles were incorporated into the theory for the case of deleterious intermediates. Owing to the recurrent input of mutations, such variants can be expected to be present in most settings. Second, the adaptations that we have envisioned have been assumed to be products of mutations at specific nucleotide sites. If multiple combinations of sites can be deployed for construction of the novel adaptation (e.g., multiple pairs of amino acid-encoding sites for a protein adaptation), the mutation rates to various states would have to be magnified accordingly (by simply multiplying the mutation rates at each stage by the number of relevant sites). Finally, although the results presented herein have focused on the mean time to establishment of an adaptive allele, for any specific set of conditions, the distribution of establishment times is typically very broad, approaching negative exponential, with high probabilities of establishment at times far below the mean (e.g., Durrett et al. 2009), and the standard deviation of t_e among replicate runs being approximately equal to \overline{t}_{e} .

A highly informative aspect of the preceding results is what they reveal about the potential mechanisms for the origin and establishment of novel adaptations in populations experiencing different intensities of random genetic drift and mutation. Consider the situation in which populations are large enough that tunneling is the primary mode of establishment of a complex adaptation. As just two examples, note that for any set of specific fitness effects of the intermediate- and final-state alleles: 1) for a two-step model with neutral intermediates, the rate of tunneling is proportional to $\sim Nu^{3/2}$ (eq. 3a); and 2) for situations in which there are d deleterious intermediate states, the rate of tunneling is proportional to Nu^d (eqs. 4b and 6b). As a first-order approximation, Nu averages \sim 0.001 in vertebrates and land plants, \sim 0.01 in smaller eukaryotes, and approaches 0.1 in some prokaryotes (Lynch 2007). For these same groups, for base substitution mutations, $u \simeq 10^{-8}$, 10^{-9} , and 10^{-10} , respectively (Lynch 2007). Thus, for twostep adaptations involving neutral intermediates, the rate of establishment of an adaptive allele scales 1:3:10 for vertebrates, small eukaryotes, and prokaryotes, whereas the scaling is approximately 1:1:1 for a two-step adaptation involving a deleterious intermediate. For situations in which there are two deleterious intermediate states, the scaling is 100:10:1, and with four deleterious intermediate states, it is 10.000:100:1.

Ultimately, the paths most frequently taken in the origins of evolutionary novelties will also depend on the rates at which neutral versus deleterious intermediate mutations arise. However, assuming that the distributions of selection coefficients for de novo mutations are not radically different among organisms, the preceding observations strongly suggest that the paths to adaptation may deviate strongly among organisms from different domains of life. Relative to multicellular eukaryotes, prokaryotes are expected to acquire adaptive alleles by paths involving neutral intermediates several times more rapidly than eukaryotes on a pergeneration basis. In contrast, when there are multiple deleterious intermediate steps, multicellular species have much greater expected rates of acquisition of adaptive alleles on a per-generation basis. Of course, because the generation lengths of multicellular species can easily be $10^3 - 10^5$ times greater than those for microbes, on an absolute time scale, rates of microbial adaptation may be comparable to or even greater than those for multicellular species even when intermediate allelic states are deleterious. However, the message to be gained from the preceding results is that the elevated power of both random genetic drift and mutation may enable the acquisition of complex adaptations in multicellular species at rates that are not greatly different from those achievable in enormous microbial populations.

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Appendix

Mutator Alleles

Because mutator alleles have an intrinsic selective disadvantage, when the first-step mutation confers no selective advantage (which is the case in all situations evaluated here), the path of sequential fixation will almost always begin on a gamete containing a nonmutator allele. The rate of origin of the first first-step mutation to fix will then be $4N\overline{u}_{\rm m}k(1-\widehat{q})\phi_1$, where $\overline{u}_{\rm m}=\widehat{q}u_1+(1-\widehat{q})u_0$ is the average background mutation rate for a m allele. Conditional on fixation of the first-step mutation, the rate of fixation of the second-step mutation is then $2N\overline{u}k\phi_{12}$, where $\overline{u} = (1 - \widehat{q})^2 u_0 + 2\widehat{q}(1 - \widehat{q})u_1 + \widehat{q}^2 u_2$ is the average background mutation rate experienced by the first-step mutation, assuming that the polymorphism at the background mutator locus remains at approximate selection-mutation equilibrium after the descendants of the first-step mutation drift to fixation. The rate of sequential fixation is then

$$r_{\rm s} \simeq \frac{2N\kappa}{[2\overline{u}_{\rm m}(1-\widehat{q})\phi_1]^{-1} + [\overline{u}\phi_{12}]^{-1}},\tag{A1a}$$

with ϕ_{\perp} in both cases being evaluated with p = 1/(2N).

The rate of tunneling must allow for the fact that firststep mutations arise linked to the m and M alleles at the mutator locus at the rates $4N\overline{u}_{m}k(1-\widehat{q})$ and $4N\overline{u}_{M}k\widehat{q}$, respectively. In the first case, assuming a neutral intermediate, tunneling then proceeds at rate $\sqrt{\overline{u}_{m}k}\phi_{2}$; in the second case, it proceeds at the approximate rate $\sqrt{u}_{\rm M}k \phi_{\rm 2M}$, with the selective advantage of the adaptive mutation being discounted by the selective disadvantage of the linked mutator allele, that is, as $s_2 - s_{\rm M}$. The total rate of tunneling with neutral intermediates is then approximated by an expansion of equation (3a),

$$\begin{split} r_{t} &\simeq (1 - e^{-4N\overline{u}k})(2k)^{0.5}\{[(1 - \widehat{q})\overline{u}_{m}s_{2}(1 + 2N\overline{u}_{m}k)] \\ &+ [\widehat{q}\overline{u}_{M}(s_{2} - s_{M})(1 + 2N\overline{u}_{M}k)]\}^{0.5}, \end{split} \tag{A1b}$$

As in the case of equation (3a), this expression attempts to correct for the constraints on the fixation probabilities of simultaneously segregating mutations, in this case weighting the probabilities for each mutation type according to the allele at the linked mutator locus. When first-step alleles are deleterious, expansion of equation (4b) leads to

$$\begin{split} r_{t} &\simeq (8Nk^{2})\{(s_{2}/s_{1})(1-\widehat{q})\overline{u}_{m}^{2} \\ &+ [(s_{2}-s_{M})/(s_{1}+h_{M}s_{M})]\widehat{q}\overline{u}_{M}^{2}\}. \end{split} \tag{A1c}$$

No attempt is made to correct for the parallel origins of tunneling mutations here because of their rarity (resulting from the fact that first-step mutations are kept at low frequency by selection), and the fixation probabilities are evaluated with p = 1/(2N). The mean time to establishment is again approximated by substituting r_s , r_t , and \overline{t}_f into equation (3b).

Transient Hypermutability

In the context of the material covered above, a fully general expression for the rate of establishment of double mutants by hypermutable cells is

$$\begin{split} r_{\rm d} &= 2N f_{\rm m}(xk)^2 \{ \widehat{q}^2 u_2^2 \phi_{2\rm M} + \widehat{q} \, (1 - \widehat{q}) u_1^2 [\phi_{2\rm M} + \phi_2] \\ &+ (1 - \widehat{q})^2 u_0^2 \phi_2 \}. \end{split} \tag{A2a}$$

Incorporating this third path toward adaptation into equation (3b),

$$\overline{t}_{\rm e} \simeq rac{1}{r_{
m s}+r_{
m t}+r_{
m d}}+\overline{t}_{
m f}.$$
 (A2b)

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