Analysis of NADH dehydrogenase proteins, ATPase subunit 9, cytochrome *b*, and ribosomal protein L14 encoded in the mitochondrial DNA of *Paramecium*

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ABSTRACT

The mitochondrial (mt) encoded ndh1, ndh3, ndh4, ndh5, rpl14, cyt b and atp9 gene products were identified by sequence comparisons with known proteins. Amino acid sequence comparisons between predicted Paramecium mt gene products and proteins in current databases were quantitated approximately by the means of similarity scores for pairs of aligned sequences. The comparisons show that the Paramecium gene products are very divergent from all others with the exception of those from a closely related ciliate, Tetrahymena. The similarity scores of comparisons between a Paramecium mt DNA encoded protein, cytochrome b for example, and the homologous protein from a group of organisms as diverse as other protozoans, vertebrates, fungi, plants, and prokaryotes were all about the same. The Paramecium gene products appear to be equally divergent from proteins representing a number of different kingdoms and organelles.

INTRODUCTION

The identities of a few Paramecium mt DNA encoded proteins have already been established by means of comparisons with sequences of known polypeptides (1, 2, 3). These include cytochrome c oxidase subunits one (COI) and two (COII), a part of NADH dehydrogenase subunit 1 (protein ND1 encoded by the gene locus ndh1), ribosomal proteins L2, S12, S14, and photosystem II protein G. Two other NADH dehygrogenase subunits found encoded in Paramecium mt DNA are somewhat unusual (2). ORF400 was identified by its similarity to homologous ORFs in chloroplasts (cp), in the protozoan kinetoplastid DNA of Leishmania tarentolae, and now in the bovine nuclear genome (4). These ORFs were shown to be an additional NADH dehydrogenase subunit (4) not encoded in animal or fungal mt genomes. The Paramecium gene corresponding to ndh2 has also been identified but appears to be lacking the 5' end found in most other mt and cp genomes (2). The amino acid sequence of the region that is encoded is very divergent compared to known ND2 sequences, even for a *Paramecium* gene product, but this region, of all the *Paramecium* mt genome, is the most similar to known ND2 genes.

In general, proteins encoded in the *Paramecium* mt genome have been found to be quite divergent compared to analogous gene products from other sources. Only about 26% of the amino acids in the *Paramecium* COI protein are identical to the aligned residues from other known sequences (1), about 23% for ribosomal protein L2 (2), and less than 20% for ND2 (2). On the other hand the *Paramecium* ORF400 shows 34% identity in alignments with the corresponding gene encoded in cp genomes and the *Paramecium* ribosomal protein S14 shows about 32% identity with the cp DNA encoded proteins (2).

In this communication we identify, by amino acid sequence comparisons, additional genes encoded in the *Paramecium* mt genome that have previously been found encoded in mt, cp, or nuclear genomes of different organisms. As with previously published *Paramecium* mt gene products, the sequences are divergent compared to those from a wide range of taxonomic groups. These results are consistent with the idea that *Paramecium* mt DNA branched from other groups early in evolution and evolved very rapidly.

MATERIALS AND METHODS

DNA nucleotide sequences of species 4 stock 51 *Paramecium aurelia* mt DNA, presented in the accompanying communication (5), were analyzed using the similarity search programs FASTP and FASTN (6, 7) utilized either on computers provided by BIONET National Computer Resource for Molecular Biology, or on an IBM-compatible computer with commercially available versions of the programs (IBI/Pustell Sequence Analysis, IBI, Inc., New Haven, Conn.). The programs and the amino acid replaceability matrix (the PAM-250 scoring matrix) used in FASTP are described in more detail elsewhere (2, 6).

The similarity scores tabulated in Figs. 2 and 9 were obtained from the version of FASTP (7) supported on the BIONET computer resource while the sequence alignments shown in the other figures were obtained from the IBI/Pustell version of the

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Cytochroi	ne i) 1 <i>a</i> 3 a	34	A.Q.	5.0	50	74 +	0.0	. (30	1 9 9
Paramecii Yeast	4 W :	ILNIFNYFKNLRVSFHEVFSLFG	FFTFNTIIVQLVSG	TNLAFSSVP NH	EPWLIPTVRC	DEEDIEDLYTDO	FFWLHERGVD	LIFIFSY	Hrru	IKLYLNVF	DLET
[284]	38		LCLVI*I*T*	IFN*Y**NI	*LAFSSVEHI	[NR*VHNG*I	LRY**AN*AS	FF*NVNFI	I*NAK	G**YGSY	(RSPR>
R.sphaer([264]	oid 51	25	LA*-*LVL*I*T*	IV**IDYT*	HVD*-AFASV	¥ NANKANGGA	NRYI*AN*AS	*F*LAV*)	****	G**YGSY	(KAPR>
Human [255]	33	** ^ ^	SLLGACL*L*ITT*	NH LF**Y*PDA	STAFSSIAHI	ITR*V-N-*GWI	IRY**AN*AS	IF * * CLFI	*IG1	:G**YGS*	r-#Y\$>
T.brucei [236]	29	*I-YGYG***-*	**IALQ**CG-*CL	FI AW*F**CCS	NWYFYLFLW	*-F*L-G-FV	IRSV*ICFTS		[*IFK	L SIT*II*	**THI>
Naize mt [205]	38	\$\$ ^^	CLAGICLVI*I*T*	¥F**NHYT*	HVD*-AFNSV	*HINR*YEGGW	Y LLRN*AN*AS	NFL ¥VVHI	*IF*	G##HASY	(SSPR>

10 120 130 140 150 160 170 + 180 +190 200 FSFLVFQVVVFFGLVLCCTHLSEITLTIAANIFHTFFNFKGKAYWFLFTDKQLNTDTLIRLAYA<u>H</u>YVSAFYLSFLGLL<u>H</u>GIDIHYDWKNEPF Paramecium: EASWKSG ٧Y Yeast *GY**GON*HWGA*VIT*L*SAI*VGNDIVS*-*WGGFSVSN [284] 111 R.sphaeroides 126 *IT*IV*NVIY*LNNGTA*N*Y**PWGQN*FWGA*VITGL*GAIPGIPSIQA*-*LGGPAVDNA**N*FFSL [264] 'IAA*VAI*IWAF*TTGN*N*-> Human 111 *-T*NI*IILL*ATNATA*N*Y**PWGQN*FWGA*VIT*LLSAIVIGTDLVQ*-IWGGYSVDSP**T*F-FT*FILP*IIAA*AT**LLFL*ETGS*N*L> [255] T.brucei I*-*ILF**IIIII*I*Y**P**NN*YWG**YFS**IA*YPILGILC**-IWGSEFI*DF**LK*HYL*VLLP*I*LIILI**LFCL**FNSSDA*> [236] Naize mt : 117 *FV*CL**VI**LNI*TA*I*Y*PPWGQN*FWGA*VITSLASAIPVVGDŤVT*-*WGGFSVDNA**N*FFSL*HLLPLI*AGAS***LAAL^{*}*G-S*N*L> [205]

210 220 230 240 250 260 270 280 290 300 Paramecium: YDGLSSENLWNDEALSNELTNFFVLLVFITLAFFLLFEEPEALSYEIFNWGDIGLSTDVRFYGVAPHWYFRPFNAWLIACPFHKTGIFGLLFFFVTLYYQ Yeast [284] 211 GITGNLDRIPNHSYFIKD**TV*-*FNL*-**L*VFY-S*NT*GODNYIP*N-P*V*PA roides FW KDLFA Y P 225 -T*VEVRRTSKAD*EKDT*PPY**I*ALVL*G**AVVAM*NY*GHDNYIQAN-P***PAHI--*-*E***L**Y*-ILR-A* R.sphaeroides GTV -AADVW-VVILVDG*TFD> [264] Human 218 GITSH*DKITFHPYYTIK-DALGL**FLLS*NTLT**-S*DL*GDDNYTLAN-P*N*PPH---IK*E***LAYTI-*RSV*NKLG*VLA**LSILI*AMI> [255] T.brucei 203 C*RFAFYCERLSFCMWFY*RDN*LAFSILLCNNYYI*INW-YFYFHEES*VIYD-TLKTS-DKIL*E*F*LYLFGF*K*I*DKFN*L*LNYILLFS*F> [236] KD I SI p Naize mt [205] 217 --*VH***---*K-IAS-YPY*Y**VGRVAS***VI*FA*NV*GHDNYI-PANPMP*PPHI--*-*E***L*IH*I*RSI*DKAG*VAAIAPV*IS*>

 310
 320
 330
 340
 350
 360
 370
 380
 390

 Paramecium:
 PNLHGVSDQNSYGKKTLTISSTVLAKKNTATPFSISIDSNLYHQITYFFFINCCLYTPSFLPYGRFFNQIGGNWGFLFSYFYVFCYLAFTD

 Yeast
 [284]
 303
 LV*P-FT*RSVVRGN*FKVL*KFFF-FIFVFN*VLLGQIGAC*-VEVPYVL*GQIA*FIYFA*--*LIIVPVISTIENVL**I>

 R.sphaeroides
 W

 [264]
 328
 AKFF**IANFG-AIAVMALAPWLDTS*VRSGAYRPKFRNWFFLVLDFVVLTWVGANPTEY-**DWISLIASTY*-*AY-FLVILPL*GA*E>

 Human
 [255]
 305
 *I**-M*K*Q*NNFRP*SQ*LYW*LAADLLILTW*GGGPVS*-PF*IIGQVASV**FTTI*>

Figure 1. Amino acid sequence alignments of cytochrome b encoded in *Paramecium* mt (top line, complete sequence), yeast mt (8), *Rhodopseudomonas sphaeroides* (9), human mt (10), *Trypanosoma brucei* mt (11), and maize mt (12) DNAs. Below each organism name is the FASTP similarity score in brackets; the next number, outside of the brackets, is the number of the first amino acid aligned on that line. Letter(s) above a sequence line represents amino acid 'inserts' at that point in the sequence which are necessary for alignment purposes; a dash represents the absence of an amino acid, and an asterisk represents identity with the top *Paramecium* reference sequence. The carets below a sequence represent scores > +1 in the FASTP scoring for conservative amino acid replacements (6). The 4 invariant histidines (+) are at positions 72, 86, 173, and 187 in the *Paramecium* sequence.

program (6). The slight differences in scoring were insignificant for short proteins but for longer ones the BIONET scores were slightly different from the IBI/Pustell scores. The small differences probably reflects the uncertainty inherent in the algorithm and either score was considered valid. For more

A			
Cyt	ocn	rome	

		1 IDE	NTITY R	ELATIVE	TO:	SCORES RELATIVE TO:					
ORGANISM	SIZE	Para	T.bru	R.sph	Yst	Para	T.bru	R.sph	Yst		
Paramecium	391	100	22	25	21	2255	21Ø	219	245		
Yeast	385	21	24	49	100	245	474	862	2Ø89		
A.nidulans	387	21	28	55	62	232	471	855	1411		
N.crassa	385	2Ø	25	48	42	186	4Ø1	786	1300		
Human	38Ø	24	26	47	5Ø	215	457	853	1155		
Mouse	381	22	26	47	5Ø	2Ø7	452	855	1170		
D.yakuba	378	21	27	48	54	192	453	863	1190		
T.bru	363	22	100	25	24	21Ø	2352	415	474		
Maize mt	388	21	26	55	51	176	421	916	1155		
R.sph	437	25	27	100	49	219	412	2441	862		
LivCpb6	215		25	35	32		24Ø	448	381		

Figure 2. Cytochrome b similarity comparisons. The % of the amino acids that are identical at corresponding positions for each paired alignment, and the FASTP program similarity score obtained for each alignment are given. The proteins that are not shown in the alignments in Fig. 1 are encoded in *Aspergillus nidulans* mt (13), *Neurospora crassa* mt (14), mouse mt (15), *Drosophila yakuba* (16), and tobacco cp (cytochrome b6, ref. 17) DNAs. Abbreviations are T.bru for *Trypanosoma brucei* and R.sph for *Rhodopseudomonas sphaeroides*. No valid alignment was obtained between the *Paramecium* and liverwort protein sequences.

accurate comparisons, however, all scores considered should be from the same program.

Since the *Paramecium* mt gene products are quite divergent, identities were based on a number of considerations including: (1) similarity scores, (2) an appropriate size for the gene between possible initiation codons ATG, ATA, ATT, ATC, GTG and stop codons TAG and TAA (ref. 5), (3) regions in the sequence alignments that were conserved in almost all of the possible pairwise comparisons, and (4) specific amino acids, such as histidines, that had previously been observed to be invariant.

RESULTS AND DISCUSSION

Cytochrome b

The *Paramecium cyt b* gene product is identified by the aligned sequence comparisons shown in Fig. 1 and is further characterized by the relative comparisons summarized in Fig. 2. This gene product is typical of most of the *Paramecium* genes in terms of its extreme divergence from most of the other known cytochrome b proteins. This particular gene product is analyzed in more detail because many cytochrome b sequences from a wide variety of organisms and organelles are available. Since the *Paramecium* gene product is so divergent, it was difficult to identify. We rely on FASTP analysis which scores conserved amino acid replacements (shown as carets below the sequence line in Fig. 1) as well as identities at corresponding positions



Figure 3. Amino acid sequence alignments with ND1 encoded in bovine mt (22), Marchantia polymorpha cp (Liverwort Cp, ref. 23), and Aspergillus nidulans mt (24) DNAs. The + marks above two regions of the Paramecium sequence identifies the homologous segments noted in ref. 21. Other symbols are given in the Fig. 1 legend.

ND1							
ORGANISM	Para	Bovine	LivCp	Human	n Mouse	e Dros	o N.cras
SIZE	261	318	369	318	315	324	371
%ID vs Para	100	32	33	32	31	34	35
<pre>%ID vs Bovine</pre>	32	100	38	78	79	48	42
SCORE vs Para	1452	457	454	428	442	428	428
SCORE vs Bovine	457	1677	533	14Ø3	1449	861	685
ND3							
ORGANISM	Para	LivCp	TobCp	Podo	Human	Mouse	Starfish
SIZE	12Ø	12Ø	12Ø	13Ø	115	114	116
%ID vs Para	100	25	22	24	21	22	28
%ID vs LivCp	25	100	72	35	31	28	28
*ID vs Mouse	22	28	32	28	64	100	49
SCORE vs Para	7Ø7	198	17Ø	169	125	133	175
SCORE vs Mouse	133	215	233	189	464	623	290
SCORE vs LivCp	198	7Ø7	526	227	255	215	248
ND4							
ORGANISM	Para	T.bru	LivCp	Droso	A.nid	Human	Starfish
SIZE	474	439	499	446	488	459	46Ø
ID vs Para	100	25	23	27	22	23	22
ID vs Human	23	21	27	41	28	100	43
SCORE vs Para	2645	475	418	466	424	359	321
SCORE vs Human	359	258	521	1Ø49	577	2378	12Ø1
ND5							
OPCANTSM	Dara	PrinMt	LivCo	TobCr	N.cm	as Bovi	ine T.bru
ST7F	570	591	693	710	715	600	5 590
tID vs Para	100	29	27	26	28	26	5 25
NID vs LivCn	27	41	100	56	30	31	L 26
TD vs PrirMt	29	100	41	41	49	36	5 28
SCORE vs Para	3127	734	698	653	707	682	2 635
SCORE vs LivCp	693	827	3854	2226	953	722	2 532
SCORE vs PrinMt	734	311Ø	827	807	12Ø3	790	5 437

Figure 4. NADH dehydrogenase similarity comparisons. Selected sequence alignments for proteins ND1, ND3, ND4, and ND5 from a variety of organisms are summarized by the% of the amino acids that are identical at corresponding positions for each paired alignment (%ID vs --), and by the FASTP program similarity score obtained for each alignment (SCORE vs). The proteins that are not shown in the alignments in Figs. 3, 5, 6, 7 are ND1 encoded in human mt (10), mouse mt (15), Drosophila yakuba mt (Droso, ref. 16), and Neurospora crassa mt (N.cras ref. 21) DNAs; ND3 encoded in starfish mt DNA (25); ND4 encoded in Trypanosoma brucei mt (T.bru, hypothetical protein 8, ref. 26), Drosophila yakuba mt (16), Aspergillus mt (A.nid, ref. 27), human mt (10), and starfish mt (25) DNAs; ND5 encoded in primrose mt (PrirMt, ref. 28), tobacco cp (TobCp, ref. 17), bovine mt (22), and T. brucei mt (hypothetical protein C-590, ref. 26) DNAs. Para is for Paramecium, and LivCp is for liverwort cp.

in aligned sequences. In addition, there are, in cytochrome b, four invariant histidine residues that have been identified and are believed to bind two heme groups (18). These histidines are present in the *Paramecium* gene product as shown in Fig. 1.

In a comparison of sequences scored relative to that of the Paramecium gene product, Fig. 2 shows uniformly low scores and 20-25% identity. There is no significant difference between similarity scores of the Paramecium gene product compared to the analogous protein encoded in the mt DNAs of the protozoan, T. brucei (210), the vertebrate, mouse (207), and the plant (176). If the gene product from any other organism is used as the reference sequence, the comparative similarity scores are higher than those compared to the Paramecium gene product sequence. Shown in Fig. 2, for example, are scores relative to the yeast gene product. All scores are above 1000 (50-62% identity) in comparisons with the protein encoded in vertebrate, insect, fungal, and plant mt DNAs. The highest scores in this comparison are with the other fungal genes, as expected. The gene product from another protozoan, T. brucei, has low similarity scores in most comparisons including the comparison with the Paramecium gene product which shows only 22% identity.

The similarities relative to the purple bacterium, R. sphaeroides, are interesting because previous studies based on rRNA sequence comparisons suggest a close evolutionary link between mt and purple bacterium DNA (19) while cp DNA is more closely related to cyanobacteria (20). However, when cytochrome b from R. sphaeroides is compared with the cytochrome b encoded in Paramecium mt DNA and cytochrome b6 encoded in liverwort cp DNA, the liverwort protein has a higher measure of similarity. These results probably reflect a rapid rate of divergence of Paramecium and T. brucei mt genes.

NADH Dehydrogenase Proteins

The ND1 protein alignments shown in Fig. 3, are in agreement with features of a comparison in a previous study (21). Burger and Werner noted regions of high similarity which are also relatively conserved in the *Paramecium* gene as shown in Fig. 3. These conserved regions are the most polar in an otherwise hydrophobic protein and may be the functional domains. The sequence comparisons, summarized in Fig. 4, show that a variety



Figure 5. Amino acid sequence alignments with ND3 encoded in liverwort cp (23), tobacco cp (17), *Podospora anserina* mt (29), mouse mt (15), and human mt (10) DNAs. The + marks above the *Paramecium* sequence denote conserved regions.

of organisms have a uniform and moderately low (31-35%) identity) degree of similarity with the *Paramcium* gene product. When comparisons are made, for example, relative to the bovine ND1 protein, high degrees of similarity (79% identity) are seen with other vertebrate genes, as expected. Genes from non-

vertebrates show smaller similarity scores but even the liverwort cp gene has a slightly higher similarity score than *Paramecium*.

The *ndh3* gene sequence was previously published as *Paramecium* ORF2 (1), but has since been identified as encoding the ND3 protein after the liverwort and tobacco cp *ndh3* gene

ND4 64 144 Paramecium: NFAVYLYFSFKKPEASKAÖLDFFNLTFKŸILKGSFFLNLVLALFCALFTFDLNFSAKNLLYPNEYIWDSGDFFFYKNGALKFSLNLYGLILVFLCLLTGF L.tarentolae M**F**VICN*IL*F-LIVTLI-FINYS*C-LA*Q-F-N**-YINIYLN*I-[565] -N*W-FIYFN***VF**IF*LSR> Liverwort Cp **IP*-L*STGNKIIRWYT*GV-CL*--E-**-*ITYI**--YHYQ--*NDHLIQLKED*N*I*FIN*HWRL*IDG**IG*I-*LTG*ITT*ATL> [471] 19 Bovine S *LI*FTS**LN*QFGDN*LN*LLFFSDS*STP*LILTNW*LP*W*MASQ> [429] 32 110 130 140 150 169 179 186 Paramecium: VAISTVDŇĽÝSEDKLKFYLIFFQFFLAVLGFIKCSDLIAFFFFYEVLŇĽGSVLVVFFGŠÝSKKSIHAVIÝFVAVTQLGSLFVLLACLYIÝSLTNSTNFFY L.tarentolae 67[°]KLV*YSKYF*--IL*SYIF***DVT*II--L--ID*FNC*NILF*S*FFPICF*SL*FNFNNRF*F*IF*L*IFSS*S*INCIĬĬĬIFHFNIL*LQS*ID> [565] Liverwort Cp 0 1#3[°]A*-WP*TR---NPR*-**FLNLANYSGQI*LFASQ*ILL***NW*LEL*-P*YLLLANWGG*RRLY*ATK*ILY*AA****I*IGG*ÏN*NSNEF*D**L> [471] R Bovine 82 HHL*K-E**TRKKLFITN**SL*L**-INT*TA-NE**L*YILF*ATLVPTLIIITW*NQTER-LN*GL**LFY*LA***PL***NTVG*L**LN> [429] 210 220 230 240 274 254 266 284 344 204 Paramecium: IKTFVFSKTQANTIYSLLFVGFGIKFPINPLHYWLTKTHVEASTGFSIYLSGFLYKTALFGFYRLTNLIÖVELDTTFFLAVLYAGVIDSSLNNWSOTDLK .tarentolae 164 *CI*-D*LYLGLYVWI***IN*S**Y****F*Y**PEL***VN*EL**L*ASVV *KFLF*SFNQ*SING*IDS**L*LTFLAIT [565] Liverwort Cp TVY 201 *NK-KYPLELEII**LSFLIAYAY*L**I*F*T**PD**G**HYSTCHL*A*I*L*NGAY*LI*I-*NELLPHAHS**APLVIV*A*OAA*TSL**RN**> [471] ¥ H C Bovine 179 LQYW*QPVHNSSNYFW*ÄNNA*NY*N*LYG**L**P*A****PIAG*NY*AAV*L*LGGY*NL*I*LILNPHT*YP*INLS*VGNINT**ICLR-***** [429] 310 320 330 340 350 360 370 380 390 400 Paramecium: KLVAYCTIQENNLIAIFFLKGDSSLIAYGFLFTINHALNSTLNFFLVECIYSRYKSRSTLVYNGVFFSFNNLALAIIFNVLFFSGILGTLKFVCEFFVFN L.tarentolae SIW 266[°]*II*TWSYIHTGIGL*LLWHN*IIFLGLLIFCNLS*IIS*AF**MM*GYM*DN*GY*IF*MLI-S**ĞΊS*F*G*FLFNID*-PF-ML*FYIDI*LLYG> [565] Liverwort Co ^rrri**ssysh*gfy-ligigsitn*lngai*qn*s*g*igasl***agis*D*--T*-***LDqn-ggig*-snpk**-T**T*csnas*alp-gnsg*i> [471] 303 Bovine 284 \$*!**\$\$Y\$H*A*YIŸILIQTPW*YNGATA*N-*A*G*T*\$NL*C*AN\$N*E*IH**TNILAR*LQTLLPLN*TWULLAS*TNLALPP*INLIG*L**Y-> [429] 410 420 430 440 450 460 470 Paramecium: LTLHVSWPIGVIFVVVVSAIGLIGFSKNWFNAIFCAPSKDVGPDALDLSKKELVIIFLCFAGLIFLTFLPFLNI L.tarentolae 366 -LISL*FIYICC*YII*L*VF*SSIYWYMCLSFYSFIWL*KYL-R***TINDI*LY*ITSIIV*LFFY*IY*L> [565] Liverwort Cp 396 AE*NI-F-L***> [471] Bovine 383 NS-TF**S-NIT-IILN-GVNNV-IT-ALY-SLYNLINTOR*KYTYHI--NNISPS*TRENA*NS*HI**L*LL [429]

Figure 6. Amino acid sequence alignments with ND4 encoded in *Leishmania tarentolae* mt (30), liverwort cp (23), and bovine mt (22) DNAs. The + marks above the *Paramecium* sequence denotes a region discussed in the text.

NO5 10 20 30 40 50 60 70 80 90 100 Paramecium: NFSFFFSFYALSVIFSLLFKHFLSSKGVFLLNTTSIGLFVAYSLSNLNLFFIKNKLIAIHLFRWFPLSAGYLVNFSFYIDTVAYSFTLLTLTIGVFVNLY 50 N.crassa GF 1 *YLSIIILPL*GS*VAGF*GRKYGVS*A-Q*-I*CLSVIITTG*AI*AF*EVN*IPVT*N****ID-*EV*NILWG*QF*SLTVANLIPV*I*SSL*HI*> [837] Liverwort Cp ⁹^w*vpl*p*l-^*illgg*-ff*pn*ikkrr*ssfisin*lnin*lsfhf*wqqitgsp**rylswv*yknfyleigyll*pltsinlv*vt*va*n*n!*> [769] L.tarentolae Ĩ[®]**N***N*FFNFGFICGI*--*IGRHIĽ*N*SVV-LCI*LVN*-TIFSC*CLSICIYGYCFYDFLLINLDFFINLV**CNGF-*I*I*YLID*VF*IVF*> [739] Paramecium: TYSYFRYEPHISRLISLINAFIASHILLYNSGNLYYFFFGWELIGITSFFLINFWGERAPTFKSAFKAFSFNKFSDSAVLIALILIYANVHDLNFEAILN 299 N.crassa [837] 188 SI**NSHD**NQ*FF*YLSL*TFN*****TAN*YLLN*V***GV*VC*YL*VS**FT*IAANQ*SNS**LT*RVG*CFLT*GNFVYLVTLGN*DYATVFS> Liverwort Cp [769] 111 SD**WF*DEGYIKFFCYLSL*T***LG**L*P**Q*YI-F***V*NC*YL**G**FT*PSAANACQ***VT*RIG*FGL*LGILGF*WITGSFD*Q-Q*-> **^**^ ^ ^ ^ ^ ^ L.tarentolae [739] 199 AFY*NYFDNLLA*FFHIFWW*VLC*NFFIL*YDFLTAYC****L*LF****************FYAL*FG****FIS*VG*--**--*L*VF*-I-TFLNNG-YC> ***** 210 220 230 240 250 260 270 280 290 300 Paramecium: YSHLYSENKLGSTPQINSWNLISFCLLFAAFYKSAQFGFHYWLPDSHEAPYPASALIHSATLYSAGYFLIHRFYPILELSLYFKLYTALYGALTALAGGL N.crassa N.CPASSA [837] 200 LAP-*--IN--*--D*-A-TI*GI***IG*WA**S*V*L*****NA**G*T*V****A**N*T***Y*L**SS*LI*Y*STVL*LCLWL**I*TYFSS*> L.tarentolae 310 320 330 340 350 360 370 380 390 400 Paramecium: SAVFQTDLKKILAYSTISHCGFLIFLCSFGNFKLVIVYLFYHGFFKAISFLCVGNLIRFSKSYQDLRRHGSFFKYLPAEFFFLVFSLLNLSGLPFFFGFY N.crassa HSHEPIV Liverwort Cp [769] 308 I*LA*K****G*****M*QL*YNNLALGI*SY*AGLFH*IT*AYS**LL**GS*SV*GHPNKS*NNIF**GLRQ*N*ITAITFL*GT*S*C*I*P*AC*W> CY L.tarento]ae [739] 291 CVFYNF*V*RYV*F***CQIS*SN*-*CL-SLD*YVGC**F*N*Y**TL*IVL*VW*H*FFGL**V*CY--**T*FCGCILL*I*AI**SCS*W*LC***> 410 420 439 440 450 460 470 480 490 500 Paramecium: SKTLLFNISDVLYFRDAIFCNILLSCITGLFYSFNILYSFFDSKKARKSIYAGVISEYLRSYYYSNTTNASNIAIFLLIYSSCLLCAYLINFYLLSLST [837] 391 **DFILESAYGQF*SG*VYIIATIGA*FTTL**VKV**LT*LANPNGYIHF*RHF*-L*E*L*V*VSY*-GKE-E-*Y*P-KH--NSKEIN*L-PR*V*G> Liverwort Cp [769] 416 **DE-ILYNSW*H*-PILGSIAFFTAGLTA**N*R-I*FLT*E-GDF*GHFFD-DYK-K*S*ISI-WGSLEF*KEQ*K*DKK*T*YE*NN*NLFP*IIL*> L.tarentolae L.tarentolae CL [739] 392 C*D**LLNLTSFF*ILEFL*VC*FFIFFTVI*NYFL*FFC**KCFCLVDTLF-LLFDFECCLV*CTFCLVNCF-VL*FF*LD-F*VVFIFSS*C*-FW-> 540 550 560 570 VIIAIFFVILVLFSYYQKKTTAEVSLAGFFDFFLGGFFF 510 520 530 Paramecium: ATDFYLVYVKTFSFTLAPLSEAALLNYSFF N.crassa TH [837] 485 EGG*F.*S-LPLVILA*FSIF-FGFITKDI*IGLGSN**IDNS**PIHEIMIDT*FAYPTL*K-L*-P*I*> Liverwort Co [769] 511 IPTVFIGFIGI-L*DENKNNVD-S*S*VLTLS*NSFNYSNSEK*LEFLFNAIPS**I-A**GIL-IA*Y> L.tarentolae [739] 490 -S-***Y*ASF*DIA*FTYFIIDIIKF-YILSGYIFY*FNIDCINFF-WRYFLFITM-**LF*IFTTWY*>

Figure 7. Amino acid sequence alignments with ND5 encoded in N. crassa (31), liverwort cp (23), and Leishmania tarentolae mt (30) DNAs. The + marks above the Paramecium sequence denote conserved regions noted in ref. 28.

ATPase, s	abun	it 9	1								+++++			+++	+++			
				10		21			30		48		50		68		78	
Parameciu	n: X	LLVI	AD.	ŤĹV	LGLC	NLPIS	5444	LGVG	IĹFA	GYNIA	VSRNP	DEAETI	FNGT	LNGFA	LVÉTI	VFNSF	FFGVIV	YFI
reast mt																		
[132]	1	VXX		TIG	A*15	I IGLI	.G×6	1*18	****	ALING	*****	SIKU¥¥	* PRA		*S*A)	CLIC	WYSFLL	~
N.crassa	nucl	ear																
[119]	9		vs	N×G	N=SA	AIGĽ	IC XC	I # I #		ALLNG	*****	NL RGQĽ	*SYA	ILsss	F##A]	GLEDL	HVALHA	κ»>
Bovine nu	clea	r												_				
[105]	5		***	FIG	A*AA	TĂĈĂ	NG SG	V#I#	TV×G	SLI¥G	*****	erkőář	*SYA	ILTE	*S*AI	IGĽFCĽ	WVAFLI	[*>
Maize at																		
[97]	1	NL:	GA	SIC	****	TIALI	1G##	VVVV AzIz	NĂTR	SSINS	*****	SL *KQS	*GYA	IL***	*T*A]	ASFAP	WWAFLI	S*Y>
E.coli																		
[84]	7 8	**Y	(- <i>N</i> /	ANN	₩ *- L	- ^^-	IG**	I*I*	**GG	KFLEG	AA*Q*1	LIPLĬ	RTQF	FIVNG	**DA]	I PHIAY	GLILY	₩>

Figure 8. Amino acid sequence alignments with ATPase subunit 9 encoded in yeast mt (YstMt, ref. 32, sometimes called oli1 gene product), *N. crassa* (nuclear gene, ref. 33), bovine (nuclear gene, ref. 33), maize mt (34), and *E. coli* (lipid-binding protein, c chain, ref. 35) DNAs.

sequences were published. As shown in the alignments in Fig. 5 and the comparisons in Fig. 4, the cp genes have the highest degree of similarity compared to the Paramecium gene (score 198, Fig. 4) which aided in the identification. Further confirmation is provided by the appropriate size of the Paramecium gene product, the two regions of conserved identities noted in Fig. 5, and the number of conserved amino acid replacements noted in the figure. The first block of conserved amino acids noted in Fig. 5 is within a region that is absolutely conserved in a previously published comparison (25) of the ND3 protein from six animal species. The similarity scores of the genes from fungal and animal sources, relative to the Paramecium protein, are only slightly lower than those of the cp genes and the difference is probably not significant. When the sequences are scored relative to the mouse gene product, a close similarity among the animal mt proteins is noted (>50% identity, Fig. 4). Relative to the cp sequence, another cp gene product has high similarity (72% identity) and the animal mt genes show much less similarity (28-30%), as expected. The Paramecium gene product has the lowest similarity scores in any of the comparisons (21-28% identity).

The alignments of the ND4 protein, Fig. 6, show few long stretches of amino acid identities with the Paramecium sequence, but the overall similarities over the long polypeptide shown in Fig. 4 are clear. One moderately conserved region, indicated in Fig. 6, contains significant similarity with the ND4 protein from a protozoan flagellate, L. tarentolae. The similarity scores shown in Fig. 4 include comparisons with another closely related flagellate, T. brucei, whose genes are highly similar to those of L. tarentolae. The scores, relative to Paramecium, are higher for the liverwort cp gene than for the human or starfish gene, probably reflecting only a high degree of divergence of the Paramecium gene. Similarities relative to the human ND4 show >40% identity with that encoded in other animal mt DNAs. The human mt ND4 has a greater similarity score compared to the liverwort cp gene product than compared to that encoded in the protozoan mt DNA.

There are three highly conserved domains shown in the *Paramecium* ND5 alignments, Fig. 7, which have been noted in a previous comparison (28) of the gene product from primrose mt DNA (*Oenothera*), *N. crassa*, and mouse. These domains contain long stretches of identities aligned with the *Paramecium* sequence, but overall (Fig. 4), *Paramecium* ND5 has only 25-29% identity in any of the comparisons ranging from the mt gene from the protozoan, *T. brucei*, to the cp gene from

AIP9							
ORGANISM	Para	YstMt	N.cras	Bovine	MzeMt	E.coli	LivCp
SIZE	75	76	81	75	74	79	81
Organelle	Mt	Mt	Nuclear	Nuclear	Mt		Ср
%ID vs Para	100	32	28	25	25	26	
%ID vs LivCp		3Ø	28	32	26	31	100
%ID vs N.cras	28	56	100	58	5Ø	3Ø	28
SCORE vs Para	384	135	119	105	97	73	
SCORE vs LivCp		112	113	102	87	128	357
SCORE vs N.cras	119	210	375	215	194	93	113
Ribosomal protei	n L14						
ORGANISM	Para	T.pyr	E.coli	B.stear	Myco	LivCp	TobCp
SIZE	119	119	123	122	122	122	123
<pre>\$ID vs Para</pre>	100	42	36	31	36	28	29
<pre>\$ID vs T.pyr</pre>	42	100	34	35	36	32	27
%ID vs E.coli	36	34	100	68	53	58	54
SCORE vs Para	622	264	196	194	163	137	119
SCORE vs T.pyr	264	600	202	206	196	177	145
SCORE vs E.coli	196	2Ø2	581	419	326	38Ø	368

Figure 9. ATPase subunit 9 and ribosomal protein L14 similarity comparisons. Selected paired sequence alignments are summarized as described in the Fig. 4 legend. The proteins that are not shown in the alignments in Figs. 8 and 10 are ATPase9 encoded in liverwort cp DNA (LivCp, *atpH* gene product ref. 23), and L14 encoded in *Bacillus stearothermophilus* (B.stear, ref. 36), *Mycoplasma capricolum* (Myco, ref. 37), and tobacco cp (*atpH* ref. 17) DNAs. Other abbreviations are: Para, Paramecium; YstMt, yeast mt DNA (32); N.cras, *N. crassa* (33); MzeMt, maize mt DNA (34); T.pyr, *Tetrahymena pyriformis* (T.pyr, ref. 38).

liverwort. Even the primrose mt protein sequence has ca. 40% identity in an alignment with the cp ND5 polypeptide and 49% identity with the mt ND5 from *N crassa*.

ATPase, Subunit 9

Identification of the ATPase subunit 9 gene (EC 3.6.1.34) in Paramecium mt DNA is interesting because this gene is found in different organelles in different organisms. The mt proton translocating complex (ATPase) contains a hydrophobic membrane component, denoted F_0 , and an F1 component consisting of relatively hydrophilic subunits. Most of the subunits are encoded by nuclear genes and imported into the mitochondrion from the cytoplasm. Subunit 9, of the F_0 component, is encoded in the nucleus of N. crassa and cow (33), but is mt DNA encoded in yeast (32), maize (34), and Paramecium. Alignments with the Paramecium gene, Fig. 8. exhibit two regions that are highly conserved regardless of the organism or organelle in the comparison. The scores given in Fig. 9 show that the *Paramecium* gene product has the greatest similarity with the mt DNA encoded yeast polypeptide (32%) identity), but also has comparable similarity with the nuclear encoded N. crassa protein (28% identity). Although the NADH dehydrogenase subunits encoded in cp DNA are as similar to the Paramecium mt DNA encoded proteins as the animal or fungal NADH dehydogenase subunits are, the same is not true for the ATPase 9 protein sequence. The FASTP simililarity program does not yield a meaningful alignment or score in comparisons between the Paramecium mt DNA and liverwort cp DNA encoded gene products, although all other sequences tested could be aligned with the cp protein (atpH gene product, ATPase subunit III, refs. 17 and 23). Relative to the N. crassa gene product, the similarity scores are about the same in comparisons with the mt DNA encoded yeast gene product (56% identity), the nuclear DNA encoded bovine gene (58% identity), and even the mt DNA encoded gene in maize (50%). These similarities suggest a common origin for these genes which are now encoded in different organelles. There is a significantly lower similarity score in the comparison with the Paramecium gene product.

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Figure 10. Amino acid sequence alignments with ribosomal protein L14 encoded in T. pyriformis mt (38), E. coli (39), and liverwort cp (23) DNAs.

Ribosomal Protein L14

In the ribosomal L14 alignments shown in Fig. 10 and the comparisons provided in Fig. 9, the high degree of similarity (42% identity) of gene products from two ciliates, *Paramecium* and *Tetrahymena*, is seen. The fact that both genomes even have the gene is significant since such ribosomal proteins, corresponding to ones found in prokaryotes, have been previously identified only in plant mt and cp DNA. The gene's location in the linear mt DNA of *Paramecium* and *Tetrahymena* is also approximately equivalent.

The prokaryote L14 proteins have 31-36% identity with the Paramecium protein, and the plant cp gene products have 28-29%. It can be seen that similarities scored relative to L14 from Tetrahymena are almost identical to those relative to Paramecium's ribosomal protein. When comparisons are made relative to E. coli's L14, we see a high degree of similarity with the prokaryote and with the cp gene products, 53-68% identity, and a lower degree of similarity with the two protozoan mt proteins, 34 and 36% identity. Therefore, even though the ciliate gene products have a relatively high % identity compared to the plant cp and prokaryote proteins, the protozoan polypeptides are still, not surprisingly, the most divergent of those known. A similar situation exits with the 'chloroplast-like' genes that have been found in the Paramecium mt genome (2). The Paramecium psbg gene product has 44-48% identity compared to corresponding prokaryote and cp gene products. Genes encoding ribosomal proteins L2, S12, and S14 have also been previously identified in the Paramecium genome (2). Comparisons of these Paramecium gene products with corresponding proteins encoded in prokaryote and cp DNA show approximately 30% identity for L2, 30% for S14, and 40-45% for S12.

CONCLUSIONS

In all paired comparisons of these proteins, the *Paramecium* gene products are shown to be among the most divergent. Proteins from the protozoan kinetoplastids *Leishmania* and *Trypanosoma* are also very divergent from other known polypeptides including those of *Paramecium*. There is probably no significant difference between most scores of sequence alignments with any one *Paramecium* gene product and the corresponding protein from any of the organisms included in this study. The scores are low and the degree of error is significant compared to score differences between comparisons involving different organisms. However, proteins encoded in the mt DNA of a ciliate, *Tetrahymena*, show significantly greater similarities with the homologous *Paramecium* proteins (Fig. 10). Also, some comparisons seem to show unusual similarities between the

Paramecium mt and plant cp genomes (2), ribosomal proteins for example (Fig. 9). But, other *Paramecium* mt gene products, such as ATPase 9, show little or no similarity with the corresponding cp DNA encoded proteins (Fig. 9). From these results, it appears that the genes encoded in the *Paramecium* mt genome are equally divergent from those encoded in the DNAs from a number of different kingdoms.

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REFERENCES

- 1. Pritchard, A.E., Seilhamer, J.J., and Cummings, D.J. (1986) Gene 44, 243-253.
- Pritchard, A.E., Venuti, S.E., Ghalambor, M.A., Sable, C.L., and Cummings, D.J. (1989) Gene 78, 121-134.
- Mahalingam, R., Seilhamer, J.J., Pritchard, A.E., and Cummings, D.J. (1986) Gene 49, 129–138.
- Fearnley, I.M., Runswick, M.J., and Walker, J.E. (1989) EMBO J. 8, 665–672.
- Pritchard, A.E., Seilhamer, J.J., Mahalingam, M.A., Sable, C.L., Venuti, S.E., and Cummings, D.J. (1989) Nucl. Acids Res. 18, 173-180. (accompanying paper)
- 6. Lipman, D.J. and Pearson, W.R. (1985) Science 227, 1435-1441.
- 7. Pearson, W.R. and Lipman, D.J. (1988) Proc. Natl. Acad. Sci. USA 85, 2444-2448.
- 8. Lazowska, J., Jacq, C., and Slonimski, P.P. (1981) Cell 27, 12-14.
- 9. Gabellini, N., Sebald, W. (1986) Eur. J. Biochem. 154, 569-579.
- Anderson, S., Bankier, A.T., Barrell, B.G., de Bruijn, M.H.L., Coulson, A.R., Drouin, J., Eperon, I.C., Nierlich, D.P., Roe, B.ASanger, F., Schreier, P.H., Smith, A.J.H., Staden, R., and Young, I.G. (1981) Nature 290, 457-465.
- Benne, R., De Vries, B.F., Van den Burg, J., and Klaver, B. (1983) Nucl. Acids Res. 11, 6925-6941.
- 12. Dawson, A.J., Jones, V.P., and Leaver, C.J. (1984) EMBO J. 3, 2107-2113.
- 13. Waring, R.B., Davies, R.W., Lee, S., Grisi, E., Berks, M.M., and Scazzocchio, C. (1981) Cell 27, 4-11.
- 14. Citterich, M.H., Morelli, G., and Macino, G. (1983) EMBO J. 2, 1235-1242.
- Bibb, M.J., Van Etten, R.A., Wright, C.T., Walberg, M.W., and Clayton, D.A. (1981) Cell 26, 167–180.
- 16. Clary, D.O., Wolstenholme, D.R. (1985) J. Mol. Evol. 22, 252-271.
- Shinozaki, K., Ohme, M., Tanaka, M., Wakasugi, T., Hayashida, N., Matsubayashi, T., Zaita, N., Chunwongse, J., Obokata, J., Yamaguc Shinozaki, K., Ohto, C., Torazawa, K., Meng, B.Y., Sugita, M., De H., Kamogashira, T., Yamada, K., Kusuda, J., Takaiwa, F., Kato, A Tohdoh,

N., Shimada, H., and Sugiura, M. (1986) EMBO J. 5, 2043-2049.

- 18. Howell, N. and Gilbert, K. (1988) J. Mol. Biol. 203, 607-618.
- Yang, D., Oyaizu, Y., Oyaizu, H., Olsen, G.J., and Woese, C.R. (1985) Proc. Natl. Acad. Sci. USA 82, 4443-4447.
- Margulis, L. (1970) Origins of Eukaryotic Cells. Yale University Press, New Haven.
- 21. Burger, G. and Werner, S. (1985) J. Mol. Biol. 186, 231-242.
- Anderson, S., de Bruijn, M.H.L., Coulson, A.R., Eperon, I.C., Sanger, F., and Young, I.G. (1982) J. Mol. Biol. 156, 683-717.
- Ohyama, K., Fukuzawa, H., Kohchi, T., Shirai, H., Sano, T., Sano, S., Umesono, K., Shiki, Y., Takeuchi, M., Chang, Z., Aota, S. Inokuchi, H. and Ozaki, H. (1986) Nature 322, 572-574.
- Brown, T.A., Davies, R.W., Ray, J.A., Waring, R.B., and Scazzocchio, C. (1983) EMBO J. 3, 427–435.
- 25. Himeno, H., Masaki, H., Kawai, T., Ohta, T., Kumagai, I., Miura, K., and Watanabe, K. (1987) Gene 56, 219-230.
- Hensgens, L.A.M., Brakenhoff, J., De, Vries, B.F., Sloof, P., Tromp, M.C., Van, Boom, J.H., Benne, R. (1984) Nucl. Acids Res. 12, 7327-7344.
- 27. Lazarus, C.M., and Kuntzel, H. (1984) submitted to the NBRF-PIR Protein Sequence Database.
- Wissinger, B., Hiesel R., Schuster, W., and Brennicke A. (1988) Mol. Gen. Genet. 212, 56–65.
- Cummings, D.J., and Domenico, J.M. (1988) J. Mol. Biol. 204, 815-839.
 de la Cruz, V.F., Neckelmann, N. and Simpson, L. (1984) J. Biol. Chem. 259, 15136-15147
- 31. Nelson, M. and Macino G. (1987) Mol. Gen. Genet. 206, 307-317.
- 32. Macino, G. and Tzagoloff, A. (1979) J. Biol. Chem. 254, 4617-4623.
- Sebald, W., Hoppe, J. and Wachter, E. (1979) In Quagliariello, E., et al.(eds.). Function and Molecular Aspects of Biomembrane Transport. Elsevier/North-Holland, Amsterdam, pp. 63-74.
- Dewey, R.E., Schuster, A.M., Levings, C.S., III, and Timothy, D.H. (1985) Proc. Nat. Acad. Sci. USA 82, 1015-1019.
- Jans, D.A., Fimmel, A.L., Langman, L., James, L.B., Downie, J.A., Senior, A.E., Ash, G.R., Gibson, F., and Cox, G.B. (1983) Biochem. J. 211, 717-726.
- Kimura, M., Kimura, J., and Ashman, K. (1985) Eur. J. Biochem. 150, 491-497.
- Ohkubo, S., Muto, A., Kawauchi, Y., Yamao, F., and Osawa, S., (1987) Mol. Gen. Genet. 210, 314–322.
- 38. Suyama, Y. and Jenney, F. (1989) Nucl. Acids Res. 17, 803.
- Ceretti, D.P., Dean, D., Davis, G.R., Bedwell, D.M., and Nomura, M. (1983) Nucl. Acids Res. 11, 2599-2616.