

Published in final edited form as:

*Ann Hum Genet.* 2012 March ; 76(2): 121–127. doi:10.1111/j.1469-1809.2011.00698.x.

## Genome-wide association of serum uric acid concentration: replication of sequence variants in an island population of the Adriatic coast of Croatia

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### Summary

A genome-wide association study of serum uric acid levels was performed in a relatively isolated population of European descent from an island of the Adriatic coast of Croatia. The study sample included 532 unrelated and 768 related individuals from 235 pedigrees. Inflation due to relatedness was controlled by using genomic control. Genetic association was assessed with 2,241,249 SNPs in 1300 samples after adjusting for age and gender. Our study replicated four previously reported serum uric acid loci (*SLC2A9*, *ABCG2*, *RREB1*, and *SLC22A12*). The strongest association was found with a SNP in *SLC2A9* (rs13129697,  $P=2.33 \times 10^{-19}$ ), which exhibited significant gender-specific effects, 35.76  $\mu\text{mol/L}$  ( $P=2.11 \times 10^{-19}$ ) in females and 19.58  $\mu\text{mol/L}$  ( $P=5.40 \times 10^{-5}$ ) in males. Within this region of high linkage disequilibrium, we also detected a strong association with a non-synonymous SNP, rs16890979 ( $P=2.24 \times 10^{-17}$ ), a putative causal variant for serum uric acid variation. In addition, we identified several novel loci suggestive of association with uric acid levels (*SEMA5A*, *TMEM18*, *SLC28A2*, and *ODZ2*), although the  $P$ -values ( $P < 5 \times 10^{-6}$ ) did not reach the threshold of genome-wide significance. Together, these findings provide further confirmation of previously reported uric acid-related genetic variants and highlight suggestive new loci for additional investigation.

### Keywords

Serum uric acid; genome-wide association; Adriatic island population

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**Authors' Contributions** The study was conceived and designed by RD, PR and RC. Recruitment of subjects, data and sample collection, data cleaning was conducted by DH-A, NN, DR, SM, ZD and PR. Genotyping was performed by GS, SRI and HC. Statistical analysis was conducted by GZ and RK. The draft manuscript was prepared by RK, GZ, PR, RC and RD. RK and GZ have contributed equally. All authors read and approved the manuscript.

**Conflict of Interest Statement** The authors declare no conflict of interest. Supplementary information is available at the AHG's website.

## Introduction

Genome-wide association studies (GWAS) have uncovered over 30 common sequence variants influencing serum uric acid (SUA) concentration and gout (Hindorff et al. 2011). Among these, the most significant findings are the single nucleotide polymorphisms (SNPs) located within the solute carrier family 2 member 9 (*SLC2A9*) gene on chromosome 4, which have been consistently replicated across multiple populations (Li et al. 2007; Dehghan et al. 2008; Doring et al. 2008; McArdle et al. 2008; Vitart et al. 2008; Wallace et al. 2008; Kolz et al. 2009; Yang et al. 2010; Charles et al. 2011). Additional GWAS and meta-analyses have identified variants in several other genes including *PDZK1*, *GCKR*, *ABCG2*, *RREB1*, *LRRC16A*, *SLC17A1*, *SLC17A3*, *SLC22A11*, *SLC22A12* that have reached genome-wide significance levels (Dehghan et al. 2008; Kolz et al. 2009; Yang et al. 2010). We conducted a GWAS of metabolic traits, including SUA concentration, in a relatively isolated population from the Adriatic coast of Croatia. This study strongly replicated the *SLC2A9* findings and identified several suggestive novel loci that may represent genuine effects. In addition, our study also replicated associations of SNPs in *ABCG2*, *RREB1* and *SLC22A12*, although the signals did not reach genome-wide significance.

## Materials and Methods

### Subjects

The study population has been described previously (Zhang et al. 2010; Karns et al. 2011). Briefly, participants were derived from the middle Dalmatian island of Hvar on the eastern Adriatic coast of Croatia. The population is primarily of Slavic descent, which had emigrated from the mainland before the 18<sup>th</sup> century and remained relatively isolated since that time (Rudan et al. 1992). Phenotypic measures and blood samples were collected in two field surveys conducted in May 2007 and May 2008, with no consideration of disease status or medication. Blood samples were collected following an overnight fast, and SUA levels were measured using the enzymatic color method. In total, 1,395 related and unrelated subjects aged >20 years with SUA measures were included in the current study. Descriptive statistics of quantitative traits (age, body mass index, fasting plasma glucose, blood pressure) and prevalence of four metabolic disorders (type 2 diabetes, hypertension, gout and metabolic syndrome) are provided in the Supplementary Table. Data on type 2 diabetes, gout and hypertension were collected through self-reports, medical review and clinical diagnostic measures. The study was approved by the Ethics Committee of the Institute for Anthropological Research in Zagreb, Croatia and the Institutional Review Board of the University of Cincinnati.

### Genotyping

Genome-wide SNP genotyping was performed using the Affymetrix Human SNP Array 5.0 following the manufacturer's protocol. Genotype calls were determined using the CRLMM algorithm (Carvalho et al. 2007, 2010) among chips that passed the prescribed Dynamic Model genotyping QC call rate (> 0.86). Following further QC filtering of the genotype data (MAF >0.02, HWE  $P > 0.0001$ , call rate >95%) using the check.marker function implemented in GenABEL (Aulchenko et al. 2007), we obtained a cleaned data set of 344,512 SNPs in 1300 samples (563 males and 737 females). From this cleaned data set, we performed genotype imputation using MACH (Li et al. 2009) and the reference haplotype data from the Phase II CEU HapMap (International HapMap Consortium 2007). The same QC procedures were performed on the imputed data, yielding a final genotype data set of 2,241,249 SNPs in 1300 samples.

## Statistical Analysis

All statistical analyses were performed in R v2.11; genome-wide association analysis was performed using the GenABEL package (v.1.6). Single-locus tests adjusted for age and sex were conducted using the qtscore routine. Since our samples included 532 unrelated as well as 768 related individuals from 235 families, genomic control (GC) was applied to correct for inflation due to inclusion of related individuals (Devlin & Roeder 1999; Devlin et al. 2001). The inflation factor ( $\lambda$ ) was estimated using the median method (Bacanu et al. 2002), and  $P$ -values based on the adjusted test statistics (1 d.f. assuming additive effects) were reported. Association signals of significant regions were plotted using LocusZoom (Pruim et al. 2010).

## Results

SUA levels were normally distributed in both males ( $N=563$ ) and females ( $N=737$ ) and the mean levels were significantly higher in males ( $361.0 \pm 79.19 \mu\text{mol/L}$ ) than females ( $265.4 \pm 77.65 \mu\text{mol/L}$ ), though Bartlett's and Fligner's tests revealed no significant gender-based differences in SUA variance. Regression analysis indicated that SUA levels were significantly associated with age in both genders and the association was more significant in females. SUA change per year in females was  $1.797 \mu\text{mol/L}$  ( $P=2.2 \times 10^{-24}$ ,  $r^2=13.1\%$ ) and in males was  $0.786 \mu\text{mol/L}$  ( $P=0.00019$ ,  $r^2=2.28\%$ ) (Supplementary Figure S1).

As anticipated from the relatedness among the samples, the test statistics were inflated compared to the null distribution with an estimated inflation factor  $\lambda=1.20$ . As shown in the quantile-quantile (QQ) plot (Supplementary Figure S2), after adjustment for this inflation factor and exclusion of significant SNPs in the *SLC2A9* region the test statistics fit well with the expected values, indicating appropriate control of false positive rate.

A Manhattan plot of the genome-wide association signals (Figure 1) shows the strongest association around the *SLC2A9* gene, a well-established uric acid-associated gene. The significant region spans roughly 650kb and covers the *SLC2A9* and the *WDR1* genes (Figure 2). This region is delimited by two recombination hot spots with local recombination rate  $>25\text{cM/Mb}$ . One-hundred and sixteen SNPs with  $P$ -value less than the genome-wide significant level ( $5 \times 10^{-8}$ ) were identified within this region. The strongest signal was found on the imputed SNP rs13129697 ( $P=2.33 \times 10^{-19}$ ); the minor allele was associated with an average SUA decrease of  $28.99 \mu\text{mol/L}$ . Consistent with previous studies, the effect size estimate showed substantial gender difference with  $35.76 \mu\text{mol/L}$  ( $P=2.11 \times 10^{-19}$ ) in females and  $19.58 \mu\text{mol/L}$  ( $P=5.40 \times 10^{-5}$ ) in males. This pattern was similar across all of the 116 SNPs that reached genome-wide significance. Of interest is a non-synonymous SNP, rs16890979 ( $P=2.24 \times 10^{-17}$ ), also reported previously with genome-wide significance (Dehghan et al. 2008; McArdle et al. 2008). Re-analysis of the region, conditional on either rs13129697 or rs16890979 failed to completely abolish the signals of the other SNPs with the smallest conditional  $P \sim 1.7 \times 10^{-3}$  (data not shown) suggesting the possibility of multiple functional variants within the region.

In addition to the well-established *SLC2A9* region, five other potentially significant regions (Table 1) were identified with at least one SNP above a threshold of  $P < 5 \times 10^{-6}$ . The most salient of these is a suggestive novel locus in an intergenic region on chromosome 5; the most significant variant, rs200113 ( $P=7.02 \times 10^{-8}$ ), is located  $\sim 400$  kb downstream of the *SEMA5A* gene. The remaining four regions are located within or near the genes *TMEM18*, *SLC28A2*, *ODZ2* and *ABCG2*, respectively. *ABCG2* is a confirmed uric acid associated gene (Dehghan et al. 2008; Kolz et al. 2009; Yang et al. 2010) and the variant showing the highest signal ( $P=5.14 \times 10^{-6}$ ) is a non-synonymous SNP (rs2231142, NP\_004818.2, Gln141Lys) and the mutant allele associated with an average increase of  $27.40 \mu\text{mol/L}$ .

(Supplementary Figure S3). This SNP showed significant gender-specific effects, with 31.11  $\mu\text{mol/L}$  in males compared to 22.97  $\mu\text{mol/L}$  in females. Reanalysis conditional on this SNP explained all the association across the region, which is highly suggestive of this missense variant being the functional SNP at the *ABCG2* locus.

To compare our findings with previous GWA studies, we analyzed 30 serum urate- or uric acid-associated SNPs listed in the GWA catalog (six of the 36 reported SNPs were missing from our imputed and cleaned data set) (Table 2). In addition to the aforementioned *SLC2A9* and *ABCG2* loci, we replicated associations with nominal significance ( $P < 0.05$ ) at two additional loci, *RREB1* (rs675209,  $P = 0.0032$ ) and *SLC22A12* (rs17300741,  $P = 0.0034$ ). The effects of all significant SNPs were in the same direction as those reported in previous GWA studies.

## Discussion

We present the results of a genome-wide association study of SUA in an isolated island population based on 2,241,249 imputed and genotyped SNPs in 1300 samples. Our purpose was to replicate previously reported loci and uncover novel SUA-related loci, taking advantage of population attributes of limited admixture and homogeneous environmental exposures. We have used GC to provide correction for inflation due to relatedness while maximizing power to detect associations by including all samples. Previous study indicated that GC is a valid and powerful method for the analysis of pedigree based quantitative trait loci (Amin et al. 2007).

The most significant associations emerged from multiple SNPs in and around *SLC2A9* on chromosome 4, a widely replicated SUA-associated region. The SNP with the strongest signal, rs13129697, is located in intron 7 of the gene. Of particular interest, however, was the association of rs16890979 (Val253Ile), a non-synonymous imputed SNP that has been reported in previous GWAS (Dehghan et al. 2008; McArdle et al. 2008).

We performed a comparative analysis of previously reported per-allele effect sizes of the significant SNPs in *SLC2A9* and found that, in general, our effect sizes are somewhat higher than those reported in previous GWAS (Dehghan et al. 2008; Doring et al. 2008; McArdle et al. 2008; Kolz et al. 2009; Yang et al. 2010; Zemunik et al. 2009) (Supplementary Figure S4). Across studies, *SLC2A9* variant effect sizes in females are markedly elevated compared to males. In our population we found males had significantly higher mean SUA concentrations, though female SUA concentration was more strongly associated with age. Sex-specific effects of *SLC2A9* variants were more extensively examined by Doring et al. (2008), who showed that in addition to genotypic effects, *SLC2A9* expression levels were stronger in females. Together these observations suggest that the *SLC2A9* variants may play a more significant role influencing uric acid concentrations in females which could be due to physiological and vascular differences between males and females and due to decreased uricosuric-related estrogen action following menopause (Adamopoulos et al. 1977; Puig et al. 1991). In addition, they suggest a potential gene-environment interaction that may be related to the gender-specific effects of the *SLC2A9* variants.

In addition to reconfirming the *SLC2A9* locus, we provide replications for three previously reported GWAS loci (*ABCG2*, *RREB1*, and *SLC22A12*), though the  $P$ -values do not reach strict GWAS significance. We report significant gender-specific effects of a non-synonymous variant in *ABCG2*, similar to those previously reported by Kolz et al. (2009). In addition to the replicated regions, we observed suggestive association signals ( $P < 5 \times 10^{-6}$ ) at several novel loci. The most significant was a SNP (rs200116) located downstream of *SEMA5A*, which encodes the semaphorin-5A protein. SNPs in its vicinity were significantly

associated with Parkinson's disease and autism in separate GWAS (Maraganore et al. 2005; Weiss et al. 2009). While elevated uric acid is correlated with lower risk of developing Parkinson's disease (Davis et al. 1996; de Lau et al. 2005; Weisskopf et al. 2007; Alonso et al. 2007), apart from a hyperuricosuric subtype of autism (Page et al. 2000) no link between autism and uric acid has been reported.

In summary, our study replicated four previously reported SUA associated loci (*SLC2A9*, *ABCG2*, *RREB1* and *SLC22A12*) with different levels of significance, and detected suggestive associations at several novel loci (*SEMA5A*, *TMEM18*, *SLC28A2*, *ODZ2*) that did not reach the threshold of genome-wide significance. However, due to the moderate sample size and the lack of a replication cohort the observed associations at these novel loci are preliminary and require further exploration and confirmation in other populations.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

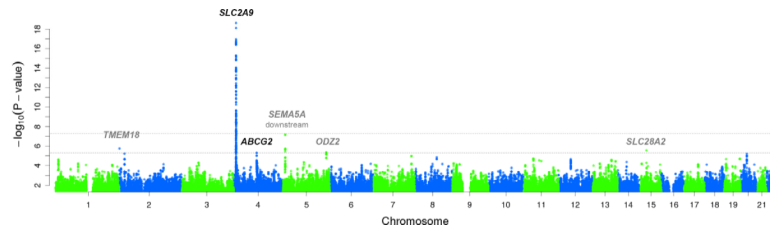
The study was supported by grants from the National Institutes of Health, USA (R01 DK069845 and P30 ES006096) and from the Croatian Ministry of Science, Education and Sports (196-1962766-2751, 196-1962766-2747 and 196-0342282-0291). RK was supported by a training grant fellowship from the National Institutes of Environmental Health Sciences, USA (T32 ES010957).

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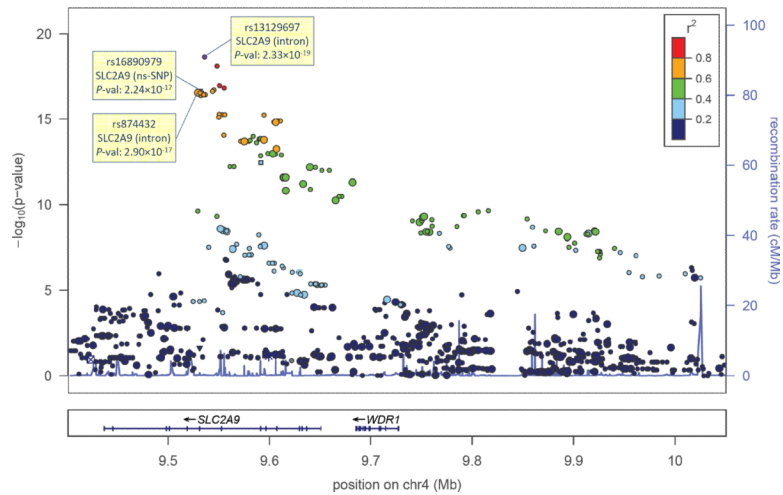
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**Figure 1.** Manhattan plot of GWA single-locus  $P$ -values. The two horizontal dash lines indicate significant thresholds at  $5 \times 10^{-8}$  and  $5 \times 10^{-6}$ . Six regions that reach suggestive genome-wide significance ( $P < 5 \times 10^{-6}$ ) are highlighted with names of nearby genes. Gene names in black are previously reported uric acid associated genes.





**Figure 2.**

LocusZoom plot of the *SLC2A9* region. GC adjusted single-locus  $P$ -values are plotted against SNP physical positions (NCBI build 36). Pairwise linkage disequilibrium ( $r^2$ ) from the most significant SNP (rs13129697) is color-coded. Size of each dot indicates whether the SNP is a genotyped (large) or imputed (small). The light blue curve shows the local recombination rate based on HapMap Phase II data. rs16890979 is the non-synonymous SNP and rs874432 is the most significant genotyped SNP.

Table 1

Summary of the significant SNPs and their regions

Chr	Significant Region				Most significant (index) SNP of the region									
	Start	End	Gene(s)	rs ID	Position	A1	A2	MAF	CEU MAF	P-value	Eff-All	Eff-M	Eff-F	
4	9401384	10050458	<i>SLC2A9, WDRI</i>	rs13129697	9536065	T	G	0.36	0.29	$2.33 \times 10^{-19}$	-28.99	-19.58	-35.76	
5	8654931	8780372	<i>SEMA5A (downstream)</i>	rs200113	8705870	T	C	0.1	0.17	$7.02 \times 10^{-08}$	28.59	33.34	23.59	
2	628483	730659	<i>TMEM18</i>	rs12999373	653483	G	A	0.27	0.22	$1.79 \times 10^{-06}$	-17.65	-11.67	-21.26	
15	43262339	43621220	<i>SLC28A2</i>	rs765787	43287339	A	G	0.15	0.16	$2.81 \times 10^{-06}$	20.9	26.93	14.45	
5	1.67E+08	1.67E+08	<i>ODZ2</i>	rs13358864	1.67E+08	T	A	0.1	0.13	$4.68 \times 10^{-06}$	-24.09	-24.25	-22.8	
4	89151747	89313430	<i>ABCG2</i>	rs2231142	89271347	G	T	0.08	0.11	$5.14 \times 10^{-06}$	27.4	31.11	22.97	

Note: Position is based on NCBI Genome Build 36; A1 and A2 are the major and minor alleles respectively; Eff-All, Eff-M, Eff-F are effect sizes ( $\mu\text{mol/L}$ ) of the minor alleles in total, male and female samples. A significant region was selected starting with an "index" SNP ( $P < 5 \times 10^{-6}$ ) and was progressively extended to adjacent ( $< 50\text{kb}$ ) significant SNPs ( $P < 5 \times 10^{-3}$ ). The *ABCG2* region was included in the table although the  $P$ -value of the index SNP (rs2231142,  $5.14 \times 10^{-06}$ ), a non-synonymous variant, did not strictly reach the standard threshold ( $P < 5 \times 10^{-6}$ ).

Table 2

Genome-wide replication of previously reported GWA SNPs

SNP	Chr	Position	A1	A2	MAF	P-value	Eff	Gene
rs1967017	1	144435002	C	T	0.48	0.451555	2.46	PDZK1
rs12129861	1	144437046	A	G	0.49	0.405643	2.75	
rs780094	2	27594741	C	T	0.49	0.492119	-2.27	GCKR
rs780093	2	27596107	C	T	0.49	0.492119	-2.27	
rs6442522	3	15415560	C	T	0.47	0.462695	-2.38	NR
rs16890979	4	9531265	C	T	0.32	2.24E-17	-28.29	
rs734553	4	9532102	T	G	0.31	4.10E-17	-28.21	SLC2A9, WDR1
rs13129697	4	9536065	T	G	0.36	2.33E-19	-28.99	
rs737267	4	9543842	G	T	0.32	2.42E-17	-28.21	
rs6855911	4	9545008	A	G	0.32	1.95E-17	-28.25	
rs7442295	4	9575478	A	G	0.28	1.98E-14	-27.09	
rs3775948	4	9604280	C	G	0.30	1.46E-15	-27.48	
rs717615	4	9713768	A	G	0.44	0.003137	-9.57	
rs2199936	4	89264355	G	A	0.08	9.68E-06	26.87	
rs2231142	4	89271347	G	T	0.08	5.14E-06	27.40	
rs3776331	5	142421987	C	T	0.29	0.699763	-1.35	
rs675209	6	7047083	C	T	0.20	0.003237	11.88	RREB1
rs742132	6	25715550	A	G	0.32	0.837566	0.71	LRRC16A, SLC17A3
rs1165196	6	25921129	A	G	0.46	0.154939	-4.55	
rs1183201	6	25931423	T	A	0.47	0.138613	-4.73	
rs1165205	6	25978521	A	T	0.47	0.136142	-4.74	ARID1B
rs9478751	6	157482742	A	G	0.08	0.6207	-3.17	
rs2244967	10	49894772	T	C	0.22	0.895854	0.50	C10orf72
rs17300741	11	64088038	A	G	0.48	0.420491	-2.59	SLC22A12
rs2078267	11	64090690	C	T	0.49	0.490532	-2.22	
rs505802	11	64113648	T	C	0.34	0.003435	10.03	
rs1106766	12	56095723	C	T	0.14	0.082384	-8.31	R3HDM2, INHBC

SNP	Chr	Position	A1	A2	MAF	P-value	EF	Gene
rs4771450	13	102767492	A	G	0.32	0.50946	2.22	NR
rs6085920	20	7128056	A	T	0.10	0.701849	2.10	NR
rs8139900	22	24724724	A	G	0.46	0.959965	-0.16	MYO18B

Note: Replicated regions reaching nominal significance are shown in bold