

Insights into the structural stability of Bax from molecular dynamics simulations at high temperatures

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Abstract: Bax is a member of the Bcl-2 protein family that participates in mitochondrion-mediated apoptosis. In the early stages of the apoptotic pathway, this protein migrates from the cytosol to the outer mitochondrial membrane, where it is inserted and usually oligomerizes, making cytochrome c-compatible pores. Although several cellular and structural studies have been reported, a description of the stability of Bax at the molecular level remains elusive. This article reports molecular dynamics simulations of monomeric Bax at 300, 400, and 500 K, focusing on the most relevant structural changes and relating them to biological experimental results. Bax gradually loses its α -helices when it is submitted to high temperatures, yet it maintains its globular conformation. The resistance of Bax to adopt an extended conformation could be due to several interactions that were found to be responsible for maintaining the structural stability of this protein. Among these interactions, we found salt bridges, hydrophobic interactions, and hydrogen bonds. Remarkably, salt bridges were the most relevant to prevent the elongation of the structure. In addition, the analysis of our results suggests which conformational movements are implicated in the activation/oligomerization of Bax. This atomistic description might have important implications for understanding the functionality and stability of Bax *in vitro* as well as within the cellular environment.

Keywords: Bax; Bcl-2 family; thermal unfolding; MD simulations; apoptosis; protein stability; electrostatic interactions

Additional Supporting Information may be found in the online version of this article.

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Introduction

The rigorous regulation of proliferation and programmed cell death (apoptosis) is crucial for preserving the survival of multicellular organisms and preventing pathological conditions.^{1–4} The physical and dynamical behavior of a large family of proteins (Bcl-2 family) is of fundamental importance in apoptosis control by regulating mitochondrial permeability and also the release of Ca²⁺ from the endoplasmic reticulum. Interestingly, some members of this family promote apoptosis, whereas the rest have

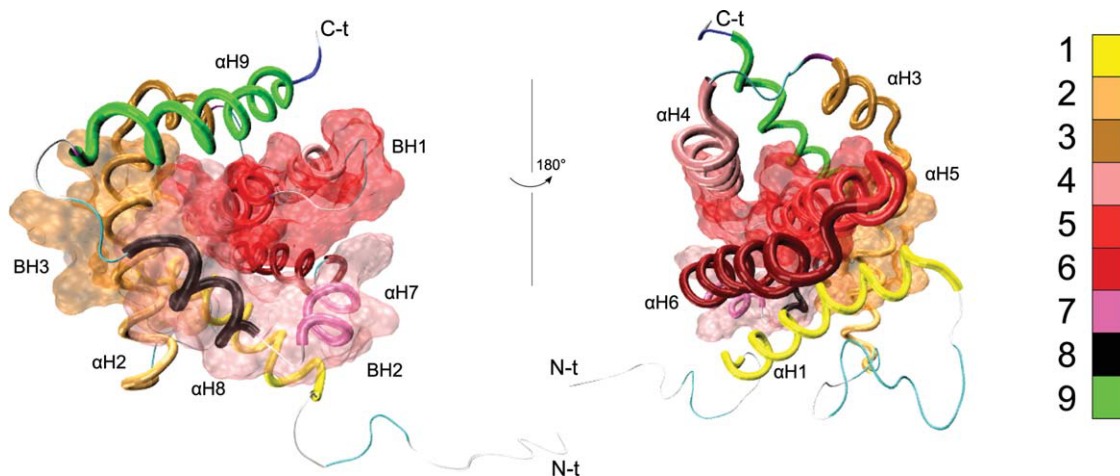


Figure 1. Bax 3D structure (PDB ID: 1F16). The nine α -helices are shown as tubes. The location of BH1-3 is indicated by the transparent surfaces. Labels indicate the N-terminal (N-t) and C-terminal (C-t) ends of the protein. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

antiapoptotic properties.^{5–7} Furthermore, recently reported clinical strategies targeting the Bcl-2 family have yielded promising results in cancer treatment.^{8,9} Bcl-2 family proteins have one to four Bcl-2 Homology (BH) domains and can be grouped into three subfamilies: (a) the proapoptotic multidomain proteins with three BH domains (BH1-3); (b) the antiapoptotic members, with either BH1-3 or BH1-4; and (c) the proapoptotic BH3 only proteins.⁹ Antiapoptotic members perform their function by sequestering those who are proapoptotic.¹⁰ Proapoptotic multidomain members promote the mitochondrial apoptotic pathway,¹⁰ whereas BH3 only members can either activate multidomain proapoptotic members¹¹ or inactivate antiapoptotic members.¹² Other biomolecules have been shown to interact with the Bcl-2 family members and regulate their activity, such as the antiapoptotic viral protein M11L,¹³ ceramide,¹⁴ and cardiolipin.¹⁵

Human Bax (Bcl-2-associated X, 192 amino acids) belongs to the proapoptotic multidomain subfamily (BH1-3), shows a large N-terminal loop (M1-T14), and contains a hydrophobic C-terminal helix (Q171-W188). Native Bax is a monomer located in the cytosol and plays an important role in apoptosis. For example, during apoptosis, Bax expression is upregulated^{16,17} and its structure is activated,¹⁸ which results in its migration toward the mitochondrial outer membrane (MOM).¹⁹ Afterward, Bax is inserted in the MOM and is capable of making oligomers^{2,20,21} to permeabilize the MOM.²² The three-dimensional (3D) structure of Bax has been determined by nuclear magnetic resonance, identifying nine alpha helices (α H1-9), joined by loops (Lp1-2, Lp2-3, Lp3-4, Lp4-5, Lp5-6, Lp6-7, Lp7-8, and Lp8-9) with some turns. With this 3D structure for Bax, it is possible to identify the characteristic functional domains BH1, BH2, and BH3, which are

formed by residues F100-V121, W151-F165, and L59-D71, respectively (Fig. 1).²³

Several details in the structure of Bax have been recognized as being important for the functionality and stability of this protein. For instance, a computational effort has predicted that the amino acid sequence of human Bax has a highly conserved region formed by 10 relevant residues in BH1 (G103, N106, G108, R109, V111, and A112) and BH2 domains (W151, G157, and W158), as well as outside of them (V95). Interestingly, the structural analysis showed that many of these residues are protected by the C-terminus of α H9 (I175-W188) which could act as a regulating gate for the access to this region.²⁴ Also, the N-terminal domain of Bax might be implicated in mitochondrial targeting. The first 20 N-terminal residues, that form a loop with a turn (M1-T14; LpNt), and the first turn in α H1 (S15-M20) might hinder exposure of a mitochondrial targeting sequence (M20-R37).^{25,26} Previous studies determined that residues G12-A24 are exposed when Bax is treated with detergents, unfolded at high temperature and when it is activated.^{27–29} Also, the C-terminal regions of α H1 as well as α H2 (which contains the BH3 domain) and α H5, should be exposed during Bax activation.^{23,25} There are other Bax residues, namely K64, I66 and L70 (α H2) and S184 (α H9) that are associated with a function that would imply their location on the surface in active or oligomeric Bax. In particular, S184 determines the interaction of Bax with the mitochondria as a part of the activation mechanism.²⁹ Thus, there are residues that have been suggested to be close to the Bax homo-oligomer interface, such as A24 (α H1), R37, L47 (Lp1-2), D68, E69 (α H2), N73 (Lp2-3), M79 (α H3), A82 (Lp3-4), R134, R145 (α H6), and K189 or K190 (α H9).²⁷

Interactions of Bax with Bcl-2 family members are primarily mediated via their BH3 domain and a

hydrophobic groove formed by BH1-3.^{30–32} Ding *et al.* found that the interface of the Bcl-2/Bax heterodimer is likely to be formed by the BH3 region of Bax and the BH1-3 groove of Bcl-2.³³ The hydrophobic C-terminal helix (α H9) is located in this groove in the native conformation of Bax. Moreover, Lalier *et al.* identified D68, E69, and D71 as the residues involved in the binding of Bax to Bcl-2 and Bcl-xL.²⁹ Other reports propose that Bax possesses an additional site to interact with the BH3 domains of other proteins. This site includes the residues K21, Q28, Q32 (α H1), and E131, R134 (α H6).^{11,34} There are several proposed models for the molecular interactions of Bax with other proteins and the MOM.^{11,27–29,34–36}

Regarding the stability of Bax, several interactions have been detected that contribute to strengthening the conformation of this protein.^{28,29} Lalier *et al.* described a motif in Lp1-2 (E44-A46) that seems to establish bonds with residues in α H6 (I133 and M137). They also reported that the disruption of the native electrostatic interaction between D33 and K64 leads to exposure of normally hidden epitopes (BH3 and the C-terminal part of α H1 constituted by residues 24–33).²⁹ Additionally, Bax has been submitted to proteolysis at different experimental conditions.²⁸ At 4°C, this proteolysis yielded a fragment lacking 16 C-terminal residues as the major cleavage product, but fragments lacking 18, 19, and 21 C-terminal residues were also observed. This data led the authors to propose that α H9 is not tightly attached to the protein, at least at the cleavage points, whereas the N-terminus was resistant to proteolysis. Furthermore, they were not able to find any C-terminus proteolytic cleavage sites in the case of detergent-induced oligomers, but they observed cleavage after S4, M38, L45, and less frequently after A81 and A82. Moreover, they identified a similar degradation pattern at 70°C, whereas at 80°C, a mixture of N-terminal (cleaved after A46) and C-terminal (mainly cleaved after F176) truncated Bax was identified. The authors also reported that preincubation at 90°C led to a completely inaccessible C-terminus, but the region of 45 N-terminal amino acids was cleaved and Bax was no longer monomeric. These results indicate that Bax shows remarkable structural stability at high temperatures.²⁸ Another member of the Bcl-2 family (Bcl-x_L) has also shown important structural stability.³⁷

Molecular dynamics (MD) simulations are a powerful theoretical approach that can successfully complement and extend the experimental results and reproduce them with reasonable accuracy.^{38,39} In particular, MD simulations at high temperature conditions have been widely and successfully used to study the unfolding of several proteins.^{40–46} A recent work published in this fine journal used MD simulations at temperatures up to 600 K to study the

stability of an important protein, focusing on the role of glycosylation therein.⁴⁷

Bax has been the object of several biophysical, chemical, and computational studies. Nevertheless, the atomic-level description of Bax stability remains to be fully explored. In this work, we use MD simulations at different temperatures to study the conformational space of monomeric human Bax at 300, 400, and 500 K. Several important interactions for Bax stability (and possibly function) are described. Hydrophobic and electrostatic interactions, as well as hydrogen bonds were detected contributing to the stability of Bax under thermal stress. Our findings might contribute to advance our understanding of the functionality and stability of Bax.

Results and Discussion

Overall stability

The time course evolution of several Bax structural features at 300, 400, and 500 K during 20 ns is shown in Figure 2. Figure 2(a) depicts α -carbons root mean square deviation (RMSD), which is a measure of backbone conformational movement; LpNt contributes to the difference between initial and final structures at 300 K and, albeit to a lesser extent, Lp1-2 also contributes.

Equilibration for 300 and 400 K is observed after 10 ns of MD simulation. Initially, RMSD values increase in the MD simulations during the heating of the system. The values were approximately bounded between 0.3 and 0.6 nm at 300 and 400 K. RMSD values at 500 K show two stages: the first from 5 to 11 ns (RMSD between 0.8 and 1.0 nm) and the second from 11 to 20 ns (RMSD between 1.0 and 1.2 nm). Regarding the radius of gyration (Rg) values [Fig. 2(b)], the starting value is about 1.8 nm, which reflects an expanded polypeptide, mainly due to the initial elongation of LpNt (Fig. 1). During the MD simulations at the three different temperatures studied, the Rg values decrease during the equilibration period. The curves at 300 and 400 K reach values around 1.6 nm after 5 ns, whereas the Rg values at 500 K fluctuate from 1.6 to 1.8 nm. The Rg values of Bax decrease at 300 and 400 K during the equilibration stage mainly due to LpNt movement toward the globule, but the compactness of the conformation is maintained from 5 to 20 ns. In this sense, at 500 K, the Rg fluctuating values are evidence of movements of the LpNt toward and away from the globule, as well as conformational variation of the whole protein. The solvent accessible surface area (SASA) values start close to 170 nm² and decrease to values around 146 nm², showing small fluctuations at 300 and 400 K after 5 ns. In both curves, an increase in SASA is observed: at 300 K, this increase is of no more than 20 nm² (17–19 ns), whereas at 400 K, the value of SASA reaches more than

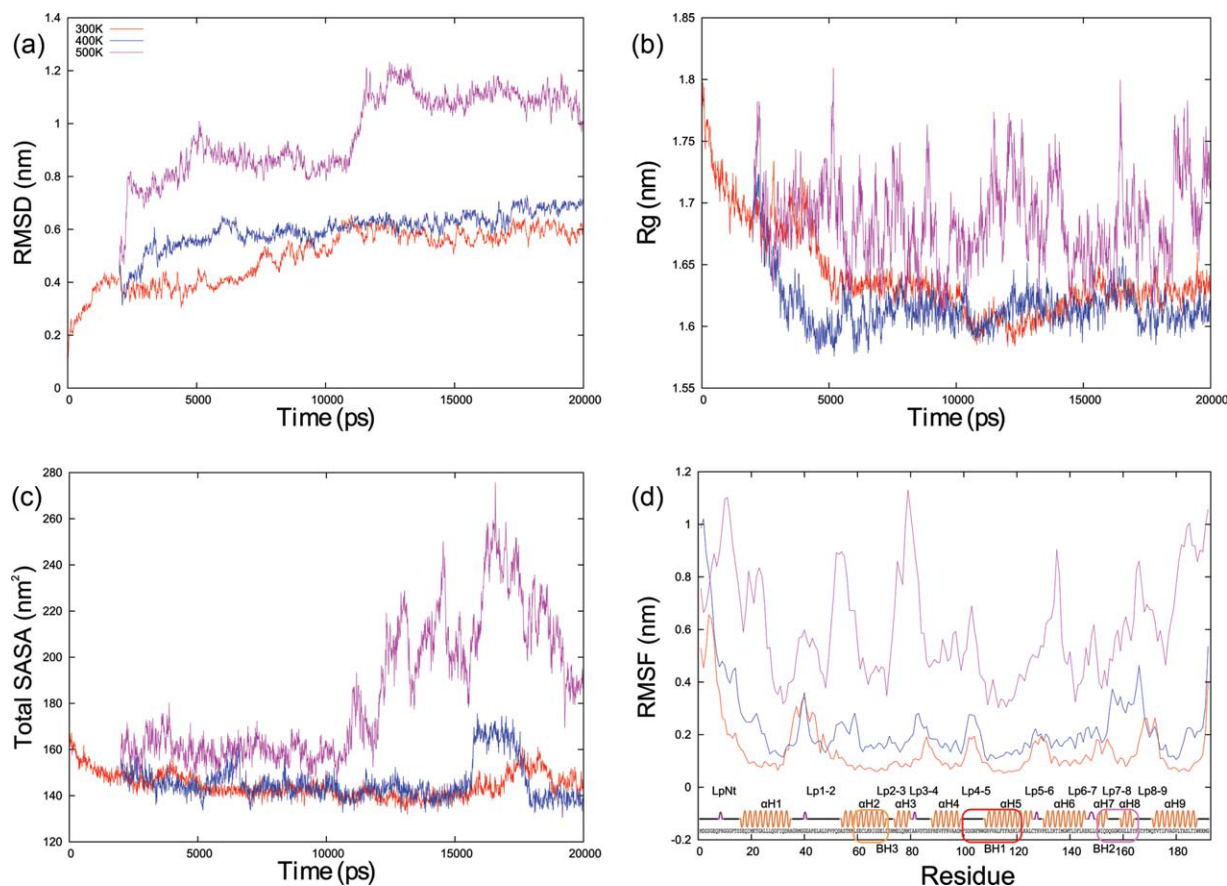


Figure 2. Measurements from the MD simulations. Time evolution of (a) α -carbons RMSD, (b) radius of gyration, and (c) solvent accessible surface area of the protein at different temperatures. (d) The root mean square fluctuations of α -carbons coordinates are shown as a function of residue number at different temperatures; a representation of the secondary structure with labels for the main domains is included. The color coding defined in (a) is the same in all plots. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

160 nm² (16–18 ns). In both cases, SASA recovers its original value by the end of the MD simulations. At 500 K, the SASA values display two phases. In the first one (5–11 ns), the SASA values do not vary noticeably (around 160 nm²), whereas a second phase involves large increases in the SASA values, showing four peaks: 195.9 nm² (11.2 ns), 236.3 nm² (13.0 ns), 250.1 nm² (15.5 ns), and 277.7 nm² (16.5 ns). After 16.5 ns, the SASA values decrease to about 185.9 nm² [Fig. 2(c)]. Concerning RMSF values, they are smaller at all temperatures in those regions that belong to α -helices, and larger in loops and turns. It also is evident that LpNt is the most mobile protein region. At 300 and 400 K, RMSF oscillates around 0.2 nm, whereas at 500 K, the RMSF values increase from 0.4 to 1.1 nm, showing seven peaks larger than 0.8 nm. Interestingly, one of these peaks is located at the C-terminus of Bax, which reflects an important conformational change in this region. It is important to mention that RMSF values from BH3 remain low compared with those of the rest of the protein in all MD simulations. The curves obtained follow closely those published by other groups, despite their using of a Bax from a

different species,⁴⁰ or performing the MD simulations with a different force field than the one employed in this work.⁴⁴

Secondary structure evolution

Secondary structure changes in Bax were evaluated by means of the MD simulation results. Figure 3(a) depicts the secondary structure content in the first 2 ns at 300 K. The last conformation obtained after these 2 ns was considered as the starting point for the next 18 ns of thermal unfolding evaluations. Figure 3(b) shows that at 300 K most of the α -helices are maintained, with a slight reduction of helicity in α H1, α H8, and α H9. Moreover, the protein loses some α -helices at 400 K, particularly α H2 and α H8 [Fig. 3(c)]. However, the α -helix structure reduction is more noticeable at 500 K, where almost all α -helices are absent, except for α H5, which forms the hydrophobic core and appears to be remarkably stable against thermal unfolding. It is worth mentioning that at 500 K, α H6 remains curled, adopting the less stable 3_{10} -helix conformation intermittently [Fig. 3(d)]. Additionally, throughout the MD simulations, the hydrogen bond network is reorganized

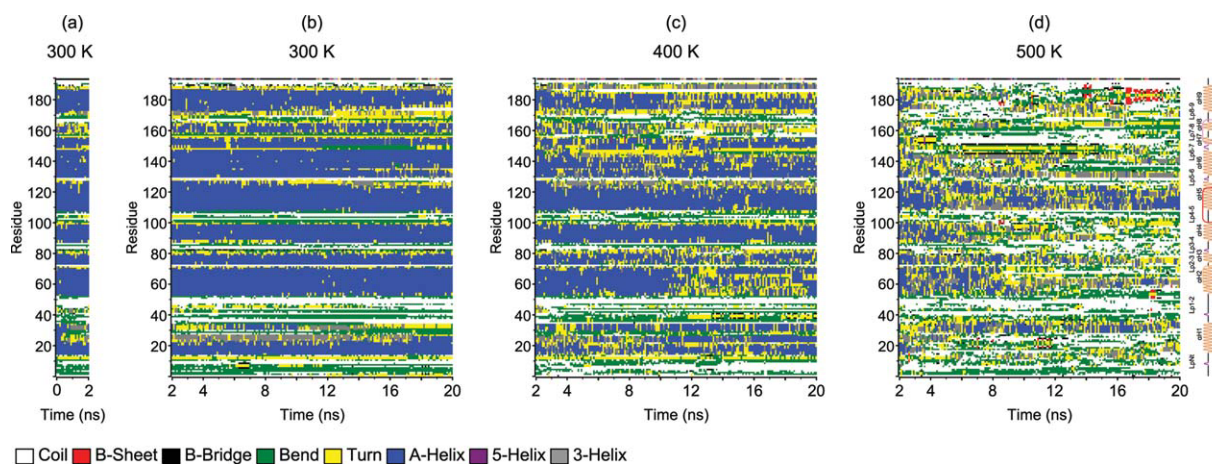


Figure 3. Time evolution of the secondary structural elements of the protein in the MD simulations. The representation of the secondary structure from Figure 2(d) is reproduced on the right. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

(data not shown), preventing the elongation of the polypeptide under heating conditions. These results are correlated with the evolution of R_g values [Fig. 2(b)]; these findings indicate that the structure tends to remain compact.

Electrostatic interactions keep Lp1-2 close to Lp5-6

It is known that electrostatic interactions are important for maintaining 3D structure of proteins both *in vitro*⁴⁸ and in MD simulations.⁴⁹ In the native structure of Bax, there is an electrostatic interaction between R134 (α H6) and E41 (Lp1-2). After 5 ns at 300 K [Fig. 4(a-i)], these two residues move away from each other, making a new electrostatic interaction between E44 (Lp1-2) and K128 (Lp5-6). Figure 4(b-i-iv) and (c-i-iv) show these four residues (E41, E44, K128, and R134) at small distances from each other [Fig. 4(b-i), inset], which helps to keep Lp1-2 close to Lp5-6. This proximity is also observed in all NMR models obtained by Suzuki *et al.*, despite the large mobility shown by E41 (Supporting Information Fig. S1), and the electrostatic interactions between these residues seem to be favored at high temperatures. An interaction of one or both of the acidic residues in Lp1-2 with one or both of the basic residues close to Lp5-6 appears constantly in all the MD simulations, where the minimum distance between the elements of one of these pairs is below 0.2 nm and is close to the van der Waals distance (0.16 nm) 80% of the time at 300 K, 90% of the time at 400 K, and 70% of the time at 500 K (Supporting Information Fig. S2). The persistence of these electrostatic interactions is remarkable at the end of the MD simulation at 500 K, when most of the helicity is lost [Fig. 4(c-iv), left inset]. These interactions appear even at 600 and 700 K (results to be published); these findings suggest that they play an important role in keeping Lp1-2 close to Lp5-6 and

therefore contribute to the structural stability of Bax at high temperatures.

Residue D33 remains close to BH3

Cartron *et al.* found that point-mutated Bax (D33A) was unable to interact with tBid, which suggests that D33 (α H1) is an important residue for Bax function.²⁵ Our MD simulation results show that D33 is close to two basic residues in BH3 (K64 and R65). The minimum distance between D33 and either K64 or R65 is close to the van der Waals distance (0.16 nm) 15% of the time at 300 K and 30% of the time at 400 and 500 K (Supporting Information Fig. S3). These electrostatic interactions contribute to the proximity of α H1 and α H2 and hence to the structural stability of Bax. The disruption of a similar “ionic lock” has been identified as important in the activation process of a protein with transmembrane helices.⁵⁰ The previously mentioned residues involved in these electrostatic interactions are shown in Figure 4. Figure 4(b-iii) shows that at 15 ns in the MD simulation at 400 K, α H1 and α H2 have a tendency to move away from each other. The electrostatic interactions between D33 and K64, R65 or both help to prevent this separation [Fig. 4(b-iii), inset]. These interactions are also observed in the MD simulation at 500 K [Fig. 4(c-iv), right inset].

Unfolding of α H4 protects the hydrophobic core

During the first 4 ns of the MD simulation at 500 K, all α -helices tend to unfold, as can be seen in Figure 3(d). There is a large jump in the RMSD plot for this MD simulation [Fig. 2(a), 11–12 ns], which may be related to the loss of the great majority of helicity, as can be seen in the snapshot at 15 ns [Fig. 4(c-iii)]. When α H4 unfolds and becomes an unstructured coil (15 ns), it remains close to the core of the protein and protects α H5, and this domain folds back into a helix. Likewise, unfolded α H4

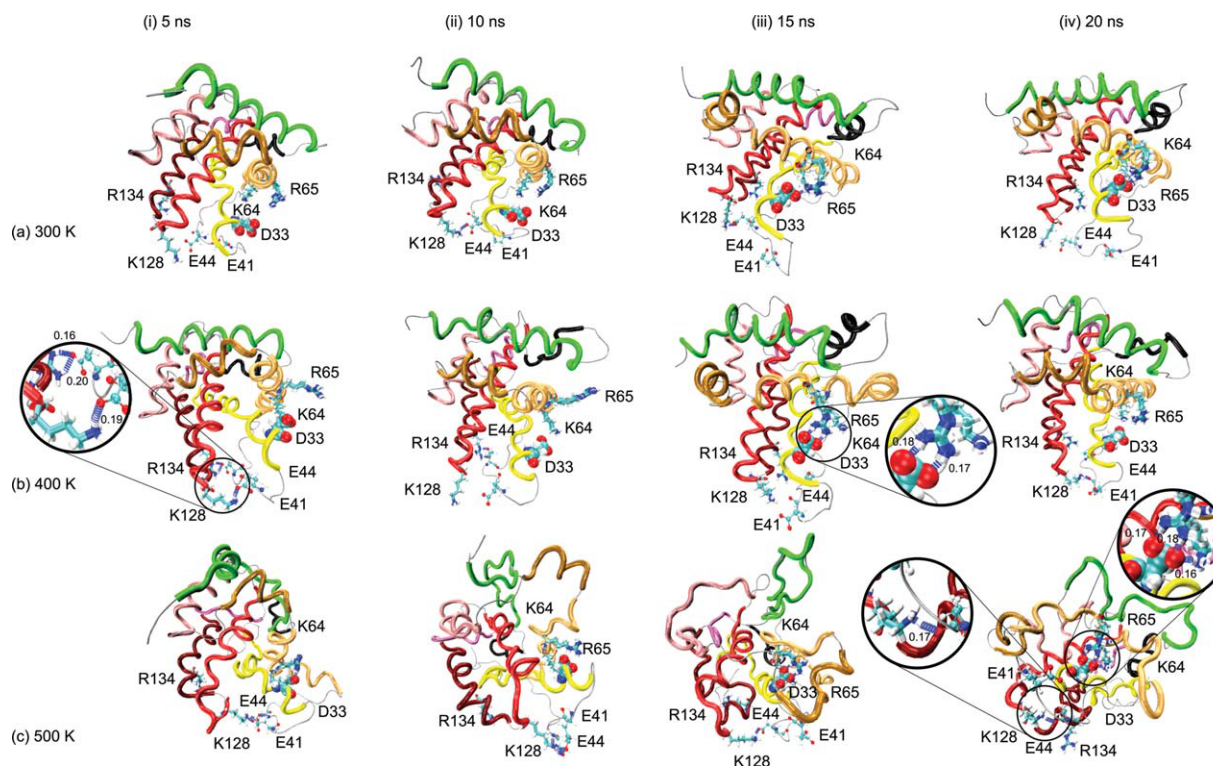


Figure 4. Snapshots of the protein structure at different time points at different temperatures. Important residues and hydrogen bonds are shown where they are involved. The insets show distances in nm. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

prevents α H7 from moving away from the core of the protein [Fig. 4(c-iii)]. With the refolding of α H5 and the holding of α H7 close to the core, α H6 regains helicity [20 ns, Fig. 4(c-iv)]. After becoming a coil, α H4 protects the hydrophobic core. This concerted mechanism helps BH1, BH2, and BH3 to remain close to each other. These conformational changes are depicted in Figure 4(c) and can also be followed in Figure 3(d), where the loss of α H4 at 500 K is closely followed by a significant recovery of α H5 and a small recovery of α H6. The changes in the trend in SASA at 500 K can also be related to this mechanism, where a growing trend starting after 10 ns, which could be caused by the unfolding, is replaced by a decreasing trend after reaching 16 ns, which could be explained by the recovery of structure [Fig. 2(c)].

Exposure of region immunodetected by antibody 6A7 (residues 12–24) at high temperatures

There is evidence that Bax activation exposes an epitope (G12–A24).^{27–29} The SASA values for these residues in the MD simulations show little change, as they are around 8 nm² at 300 K and remain below 11.2 nm² even at 500 K (Supporting Information Fig. S4). Searching for structural changes involved in the exposure of these residues, it was observed that the GGG motif close to the N-terminus of α H1

(G10–G12) is very flexible and allows M1–G10 (LpNt) to interact with α H6. In the MD simulations performed, favorable electrostatic interactions were detected between D2 (LpNt) and R145 (α H6) at 300 and 500 K. In addition, electrostatic interactions between D2 and R9 (both LpNt) at 300 and 500 K and between D6 and R9 at 400 K were found to help to bend LpNt (Supporting Information Figs. S5 and S6). These D2 interactions explain its local RMSF minima at 300 K and at 500 K [Fig. 2(d)].

Other groups have proposed that residues M1–M20 (LpNt) play an inhibitory role for Bax mitochondrial localization.^{26,29} Our results show that LpNt protects the 6A7 epitope on one side during the course of the MD simulations. On the opposite side, we found that residues P49–S60 and the 6A7 epitope make a cavity that allows the latter to be in contact with the solvent in the native conformation and during the MD simulation at 300 K; the small size of this cavity shown during the MD simulations might prevent the corresponding antibody from reaching the epitope. This access to solvent but not to larger molecules explains the small change in SASA for this domain in the MD simulations. These residues (P49–S60) start in Lp1–2 and include the N-terminal part of α H2 (Fig. 5). Their proximity to the epitope [Fig. 5(a)] is partially stabilized by the electrostatic interaction between K21 (epitope) and Q52 (Lp1–2). This interaction is lost at 400 K and

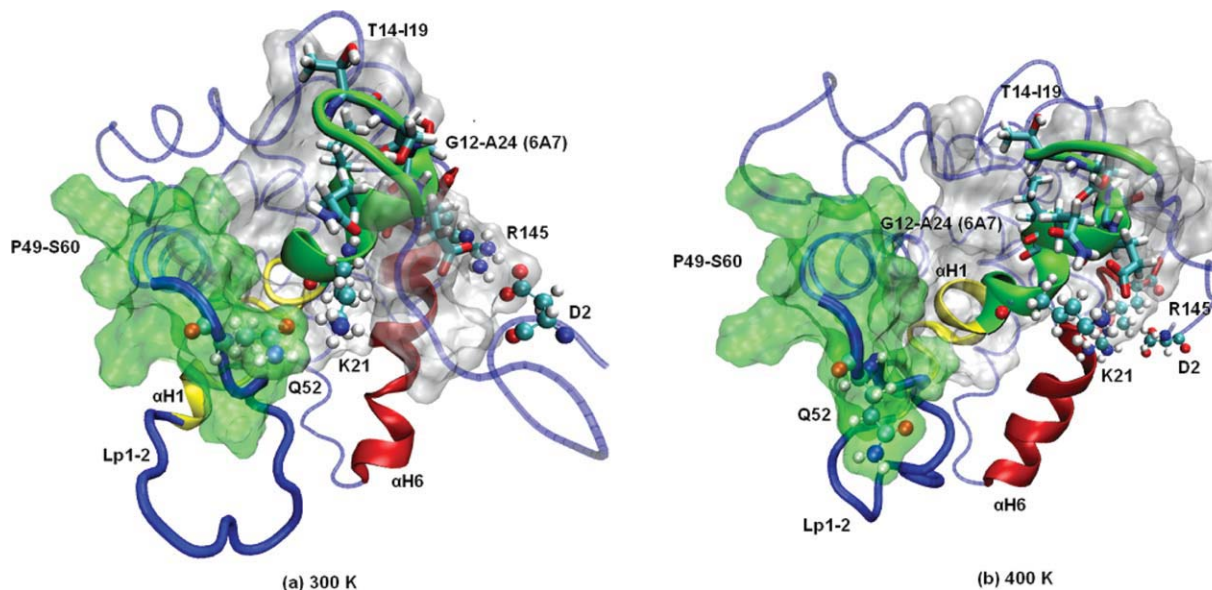


Figure 5. Exposure of the epitope 6A7 under heating conditions. Final structures for the MD simulations at 300 K (a) and 400 K (b) are shown. Residues K21, Q52, D2, and R145 are shown in CPK. The epitope is shown in green; it starts in LpNt and includes the first residues in α H1, and the rest of α H1 is shown in yellow. The side chains of the region that interacts with the antibody 6A7 (T14-I19) are shown in Licorice. Residues P49-S60 are shown with transparent surfaces in green. α H6 is shown in red. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

the 6A7 epitope is exposed [Fig. 5(b)]. At 300 K, Lp1-2 shows some mobility but remains close to the core for most of the MD simulation [Fig. 5(a)]. At 400 K, Lp1-2 adopts a different conformation and points away from the core [Fig. 5(b)]. These movements allow the residues P49-S60 to move away from the epitope, leaving it exposed. At 500 K, the epitope is uncovered by P49-S60 moving away from the core early in the MD simulation. It is known that the free energy of antigen-antibody complex formation represents a balance between enthalpic and entropic forces. Analyzing only the 6A7 epitope, a period ranging from 12 to 18 ns exhibited a robust increase in SAS up to 50% at 400 K. As expected, free energy solvation showed a sustained decrease (from -15 to 38 kJ/M) starting at 12 ns until 17 ns approximately, followed by a recovery. These patterns were presented at 300 K but modest and shifted in time scale (Data not shown). Therefore, an apoptosis-compatible energetic barrier should be overcome at physiological temperature.

Another factor that prevents α H1 from moving away from the core of the protein is the formation of several hydrogen bonds between LpNt (residues M1-T14) and the core of the protein. The structural analysis also shows an increment in the number of residue pairs closer than 0.35 nm. These pairs are likely to have a favorable interaction, which contributes to keep these residues close to the core (Supporting Information Fig. S7).

These MD simulations results reveal that Bax can show the 6A7 antibody epitope at high temperatures. This finding is in agreement with the experimental results in the literature.²⁸

Hydrophobic groove covering by α H9

The hydrophobic groove formed by BH1-3, which is occupied by α H9 in the native conformation, has been shown to be important in the interaction of Bax with other members of the Bcl-2 family.^{30,31} Therefore, the exposure of this groove could be involved in the oligomerization process of Bax. This hydrophobic groove is protected constantly by α H9 despite the great conformational changes induced by heating conditions in the MD simulations.

These MD simulations provide evidence that electrostatic interactions involving K189 and K190 are important for protecting the hydrophobic groove by α H9 under heating conditions. K189 and K190 tend to point in opposite directions due to steric hindrance effects and their equal positive charges. With this spatial disposition, these residues form a positively charged “anchor” that is trapped in an ionic “cage” formed by four negatively charged residues (E69, D86, E90, and D98) in the MD simulations at high temperatures. For example, at 400 K, D98 interacts with K190 [Fig. 6(a)], whereas at 500 K, K190 is close to D86 [5 ns, Fig. 6(b)] and such interactions continue to arise. At 10 ns, K189 replaces K190 in the proximity of D86 and also interacts with E90 [Fig. 6(c)]. At 15 ns, another combination emerges with K190 interacting with D98 and K189 with E69 [Fig. 6(d)]. The electrostatic interactions involving K189 and K190 could account for their local RMSF minima at 400 and 500 K [Fig. 2(d)], and they seem to contribute to the structural stability of Bax at high temperatures (Supporting Information Fig. S1).

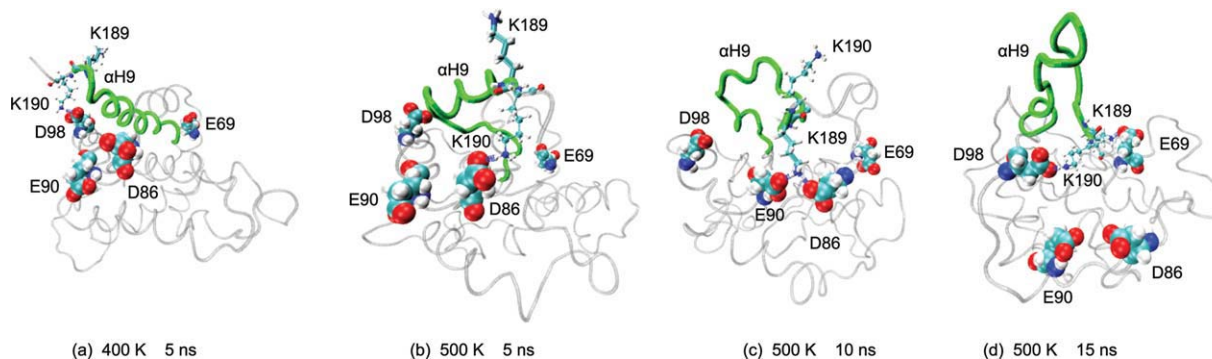


Figure 6. Electrostatic interactions that prevent α H9 from moving away from the protein core. K189 is shown in Licorice and K190 in CPK. α H9 is shown in green. Acidic residues that form the ionic “cage” are depicted in the VDW representation. In all cases, the minimum distance between the lysines and the acid residues is close to 1.6 Å, which is the van der Waals distance for a nonbonded O—H pair. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

Moreover, there is evidence that hydrogen bonds involving the sidechain of S184 contribute to protect the hydrophobic groove by α H9.²³ In the native structure, S184 forms a hydrogen bond with the backbone carbonyl of V95. In the MD simulation at 300 K, the sidechain of S184 forms a hydrogen bond with the sidechain of D98, which keeps the rotamer of S184 pointing to the core of the protein. At 400 K, within the first 5 ns, the hydrogen bond of S184 with V95 is reformed, whereas D98 contributes to the stability of Bax in a different manner, by interacting with K190 as described above. At 10 ns, S184 forms a hydrogen bond with N106 (Lp4-5), but the helix turn of α H9 that contains S184 partially unfolds and leaves S184 exposed to the solvent. Furthermore, the electrostatic interaction between D98 and K190 is maintained and a hydrogen bond appears between D98 and T186. At 15 ns, S184 is still exposed to the solvent, but it forms a hydrogen bond with the backbone carbonyl of V180. D98 remains close to either K190 or T186 for the rest of the MD simulation. At 20 ns, S184 is close to R109 but is mostly accessible to the solvent (data not shown). At 500 K, S184 is completely exposed, thus no longer forming hydrogen bonds with other residues (Supporting Information Fig. S8). The exposure of S184 could be the cause of a large increase in the mobility of α H9 at 500 K [Fig. 2(d)], but it is not enough to separate this helix from the hydrophobic groove.

Brock *et al.* found that the release of α H9 is essential for the activation of Bax⁵¹; thus, the interactions reported in this section that help to maintain the helix tucked into the hydrophobic groove at high temperatures would contribute to the structural stability of Bax.

Hydrophobic interaction between W107 and BH2 residues

W107 belongs to BH1 (F100-V121) and is close to BH2 (W151-F165) in the native conformation. BH2

includes most of α H7, Lp7-8, and α H8. In the MD simulation at 300 K, an aromatic–aromatic interaction between W107 and Y164 is suggested. This kind of interaction has been reported as a mechanism of tertiary structure stabilization.⁵² Also, in this MD simulation, W107 buries itself in the core, approaching W158 in Lp7-8, which helps to keep BH1 close to BH2 [Fig. 7(a)]. The SASA value for W107 increases in the MD simulations under heating conditions (Supporting Information Fig. S9). At 400 K, W107 is partially exposed and the hydrophobic and aromatic residues in BH2 are not as close as they were at 300 K because BH2 and Lp7-8 elongate, although they remain close to BH1 [Fig. 7(b)]. At 500 K, W107 is mostly exposed, although it is close to Y164 and F165, which favors aromatic–aromatic interactions; however, BH2 shows a loss of helicity during this MD simulation [Fig. 7(c)]. W107 is at the beginning of the RMSF valley that corresponds to α H5 in all the MD simulations [Fig. 2(d)]. The observed hydrophobic and aromatic–aromatic interactions of W107 with residues in α H7 (W151, I152), Lp7-8 (W158), and α H8 (L161, L162, Y164, and F165) are in agreement with structural studies reported elsewhere.⁵³

Normal mode analysis

Normal mode analysis (NMA) identifies potential conformational changes in proteins.⁵⁴ In this work, NMA was performed with the Elastic Network Model, which is implemented with the “rotation-translation-block” approximation in the web interface Elnémo.⁵⁵ Previous reports in the literature suggest that the consideration of the first 10 nontrivial modes should be sufficient for a reasonable analysis of a protein with a number of degrees of freedom similar to Bax.^{56,57} NMA was performed on the native structure of Bax and the final structures of each MD simulation at different temperatures. A common pattern among the modes considered was an important mobility in the loops, particularly in

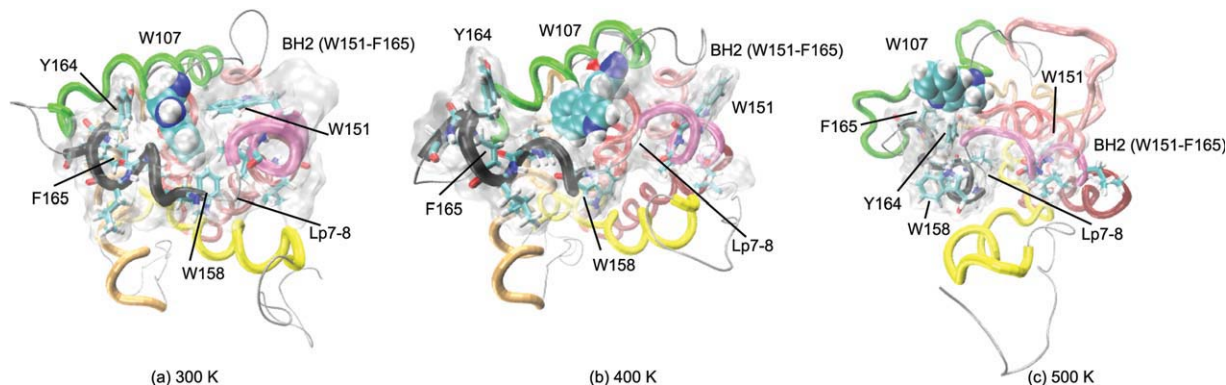


Figure 7. Exposure of W107. Final structures (20 ns) are shown from the MD simulations at 300 K (left), 400 K (middle), and 500 K (right). W107 is shown in VDW and the SASA of the region formed by residues L148-F165 in transparent gray. The color code for the helices is the same as in Figure 1. At 300 K, W107 remains buried and its aromatic rings are perpendicular to Lp7-8. At 400 K, W107 is more exposed, and its aromatic rings are almost parallel to Lp7-8. At 500 K, W107 is mostly exposed. The hydrophobic and aromatic residues in BH2 (W151, I152, W158, L161, L161, Y164, and F165) are shown in Licorice. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

LpNt and Lp1-2 as well as α H9. Modes with low collectivity are likely to suggest domains that would have significant mobility in a conformational change. For the native structure, modes 9 and 11 show little collectivity; this finding is explained by the large mobility of LpNt. The same is true for the three low collectivity modes in the NMA of the final structure at 300 K (modes 7–9), whereas the rest of the modes show high mobility in Lp1-2 and Lp5-6, as does the low collectivity mode obtained at 400 K (mode 9). Modes 14–16 at 400 K also show a high mobility of BH2. The NMA of the final structure at 500 K shows one low collectivity mode with a huge displacement of E41 and the residues around it in Lp1-2. Several of the other modes for this structure show increased mobility levels for α H4, which can be explained by its unfolding at 500 K [Fig. 4(c)]. Also, a large mobility observed in Lp1-2 and Lp5-6 in the NMA suggests a coherent motion of Lp1-2 and H5-6 which could favor the separation of these domains. This would lead to a major conformational change and expose the transmembrane domain, which would be important for the functionality of Bax (Supporting Information Fig. S10).

Other relevant residues

Mancinelli and coworkers performed 10 ns of MD simulation at 300 K and 5 ns at 400 K using a united-atom force field. Their analysis aimed to identify the protein regions and the motions of Bax that are involved in the conformational change that targets Bax into the MOM.⁴⁴ However, the specific interactions that contribute to the stability of Bax and their nature are not investigated as is attempted in this work.

There are other residues that have been proposed to be close to the Bax surface when it forms a complex with a member of the Bcl-2 family, such as

A24, Q28, Q32, R37, L47, I66, D68, L70, N73, M79, A82, and E131.^{27,29,34} The behavior of these residues is briefly discussed below. Because Q32 and I66 remain hidden in these MD simulations, their exposure might be important in the activation mechanism of Bax. R37, L47, D68, and E131 remain exposed, and A24 remains close to the surface in our MD simulations. The exposure of R37 seems to be stabilized by electrostatic interactions. N73, M79, and A82 are part of the surface of Bax, but their SASA values increase at 400 and 500 K. Q28 is fully exposed until α H1 starts to unfold, when it faces toward the core; however, when α H1 is fully unfolded, Q28 is exposed again. L70 approaches the surface at 10 ns in the MD simulation at 500 K (data not shown).

A putative functional region of Bax was also predicted, and it includes several residues (V95, G103, N106, G108, R109, V111, A112, W151, G157, and W158).²⁴ Among these residues, Gurudutta *et al.* suggested that G108, R109, and W151 are important in the oligomerization of Bcl-2 family proteins by interacting with the BH3 domain of another Bcl-2 family member.⁵⁸ Although we find that G108 is fully exposed in these MD simulations, R109 is found to be partially covered by α H4 when this α -helix unfolds at 500 K, whereas W151 switches from exposed to hidden at 500 K. The hiding of R109 and W151 at high temperatures as observed in these MD simulations would hinder the oligomerization of Bax and thus contribute to its structural stability. As for V111 and A112 (both in α H5), they are hidden in all the MD simulations and have hydrophobic interactions with V180 (α H9) and I66 (α H2) in the final MD simulation structure at 400 K and additionally with A183 (α H9) in the final MD simulation structure at 500 K (data not shown). These residues are located in the transmembrane

domain of Bax,²³ and the hydrophobic interactions that hinder their exposure might prevent Bax from activating. G103, N106 (both Lp4-5), and G157 (Lp7-8) are exposed in these MD simulations, interacting mostly with the solvent. Their contribution to the structural stability of Bax seems to be limited to the hydrogen bonds involving their backbone atoms. To the best of our knowledge, the roles of these particular residues in the function or stability of Bax have not been addressed in the literature. The contributions of V95 and W158 to the structural stability of Bax were discussed above.

Conclusions

Stable conformations for Bax are reached in all MD simulations treated at different temperatures, and most of the original α -helices are lost at high temperatures. The stability of the 3D structure is preserved by electrostatic interactions under heating conditions. Hydrophobic interactions and hydrogen bonds are also found to play a role in maintaining the 3D structure of Bax in the MD simulations. There are some domains hidden in the native structure that become exposed at high temperatures, such as the 6A7 epitope (G12-A24).

The 3D structure of Bax appears to have several recovery mechanisms to prevent its domains from moving away from each other when subjected to high temperatures to maintain the protein compactness. Electrostatic interactions arise between acidic residues in Lp1-2 (E41 and E44) and basic residues in α H5 (K128) and α H6 (R134) and are important to keep Lp1-2, α H5, and α H6 together when Bax is subjected to high temperatures. At 500 K, α H4 unfolds into a coil but remains close to the globule and has interactions with BH1, BH2, and BH3 that help to keep these domains close to each other. Electrostatic interactions of K189 and K190 with acidic residues in other domains emerge and are important to keep α H9 covering the hydrophobic groove under heating conditions. The contributions of several electrostatic interactions to the stability of Bax under thermal stress might have their evolutionary origin in the importance of preventing this protein from aimlessly activating and thus triggering apoptosis.

Materials and Methods

The starting structure (native) for MD simulations of the Bax monomer was obtained from the first NMR model located in the Protein Data Bank (PDB) by Suzuki *et al.* (PDB ID: 1F16).²³ All MD simulations were performed with GROMACS 4,⁵⁹ using the OPLS-AA force field.⁶⁰ The leap-frog algorithm for integrating Newton equations was used, and periodic boundary conditions were applied. The protein was solvated in a rectangular box of modified SPC,⁶¹ with a minimum distance of 1 nm from the protein to the edge of the box. To obtain a neutral total

charge in the system, 3 Na⁺ counterions were added. The total size of the system was of 38,647 atoms, including 11,889 water molecules. All bonds were constrained using LINCS.⁶² During energy minimization, the steepest descents algorithm was used and the convergence was reached in 200 steps. Further equilibration of the system was accomplished in 5000 steps (10 ps) of MD with restricted protein atoms and NVT conditions with a box size of 9.44551 \times 7.45274 \times 7.68113 nm. Afterward, 2 ns of MD were performed at 300 K without any atom fixing and under NPT conditions, which were used for the following MD simulations as well, so that the size of the box could fluctuate to keep the pressure at a constant value. Using the latter trajectory as a starting point, another 18 ns of production MD were obtained at 300, 400, and 500 K. MD simulations were performed with a time step of 2 fs, and the coordinates for the whole system were saved every 250 steps. For all MD simulations, neighbor lists were updated every 10 steps. The PME algorithm⁶³⁻⁶⁵ was used for the electrostatic interactions with a cut-off of 1 nm, and a reciprocal grid of 54 \times 42 \times 44 cells was used with 4th order B-spline interpolation. A single cut-off of 1 nm was used for the calculation of van der Waals interactions. Temperature and pressure coupling were performed with the Nosé-Hoover algorithm^{66,67} and the Parrinello-Rahman algorithm,^{68,69} respectively. After stabilization of the system, in all cases, the potential energy was conserved during the MD simulations. The analysis tools included in GROMACS were used to calculate the RMSD, RMSF, Rg, and SASA. The evolution of the secondary structure was followed using the DSSP program.⁷⁰ Normal mode analysis was performed using the web-server Elnémo,⁵⁵ where the only change with respect to default settings was in the number of lowest frequency normal modes to be computed which was set to 10 (default = 5). The minimum perturbation (DQMIN), maximum perturbation (DQMAX), and step size between DQMIN and DQMAX were left at their default values of -100, 100, and 20, respectively. The advanced options were also left in their default settings (NRBL = auto and CUTOFF = 8). Graphical representations of the protein were obtained using VMD.⁷¹

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