# NRl<sup>1990</sup> Oxford University Press <sup>1447</sup> Ribosomal protein L4 of Saccharomyces cerevisiae: the gene and its protein

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# ABSTRACT

The sequence of a gene for ribosomal protein L4 of Saccharomyces cerevisiae has been determined. Unlike most ribosomal protein genes of S. cerevisiae this gene has no intron. The single open reading frame predicts that L4 is highly homologous to mammalian ribosomal protein L7a. There appear to be two genes for L4, both of which are active.

### **INTRODUCTION**

As part of an ongoing effort to determine the structure of the ribosome of the yeast Saccharomyces cerevisiae, we and others have cloned the genes for a number of the ribosomal proteins.(Reviewed in 1) Our hope is that the genetic approaches available in the yeast system will provide opportunities for greater insight into the structure and function of the eukaryotic ribosome. We present here the sequence of the gene for ribosomal protein LA (formerly called RP6), the third largest protein of the 60S subunit (2), and the predicted amino acid sequence of the protein that is derived from it. L4 is highly homologous to mammalian ribosomal protein L7a.(3,4) There are two genes for L4, both of which appear to be active. The gene studied in this paper is termed RPL4A.

# MATERIALS AND METHODS

Strains and Plasmids. Saccharomyces cerevisiae, strain W303(MATa/ $\alpha$ , ade2-1, his3-11,15, leu2-3,112, trp1-1,  $u\tau a3 - 1$ , can $1 - 100$ ) is a homozygous diploid obtained from R. Rothstein (Columbia University). A haploid MATa strain was derived from a spore of W303. Yeast strains were grown in synthetic medium and plasmids carrying the URA3 gene were maintained by growth in media lacking uracil. Recombinant DNA techniques and preparation of nucleic acids were carried out essentially as described in (5). Plasmid pYERP6, containing a 9.2 Kb EcoRI/EcoRI fragment with the RPL4A gene, was derived from a lambda clone (A83) described previously (2). Escherichia  $\text{coli}$  strain DH5 $\alpha$  was used throughout for the transformation and propagation of the plasmids.

DNA Preparation, Labeling and Sequencing. Plasmid DNA preparations, restriction enzyme digestions, and ligation

reactions were performed as described (5). DNA was labeled by nick translation using  $(\alpha^{-32}P)dCTP$  (Amersham Corp., 3000) Ci/mmol.) and DNA polymerase. Oligonucleotide probes were 5'-end labeled with  $(\gamma^{-32}P)ATP$  (Amersham Corp., 3000) Ci/mmol) and T4 polynucleotide kinase. DNA fragments were subcloned in the vector pGEM blue (Promega) and their sequence determined by the dideoxy chain termination method (6) applied to DNA minipreps (7).

DNA Blot Analysis. Restriction enzyme digested DNA was blotted to Nytran (Schleicher & Schuell) and probed with nick translated DNA fragments or with kinased oligonucleotides.

#### RESULTS AND DISCUSSION

#### Subcloning and Sequencing of RPL4A

RPL4A was originally cloned as a 9.2 kb EcoRI fragment in a lambda vector. The identity of the product of the gene as ribosomal protein LA was established by 2D gel analysis of the in vitro translation of mRNA hybridizing to the DNA fragment (2). Further characterization led to the restriction map shown in Figure 1A. Several of the fragments  $(A-E$  in Figure 1B) were subcloned into pGEM vectors. The location and orientation of the transcript (Figure iB) was determined originally by probing northern blots of yeast RNA with RNA probes transcribed from these subclones (data not shown). Dideoxy sequencing (6) was carried out using either primers from within the vector sequences or primers synthesized according to preliminary sequence data as it was obtained.

The sequence of the RPL4A gene is shown in Figure 2. There is a single open reading frame coding for 256 amino acids, suggesting that the gene is uninterrupted by an intron. This view is confirmed by the lack of the canonical sequences (8) characteristic of the <sup>5</sup>' splice site (GTATGT) or of the lariat site (TACTAAC), either within or upstream of the open reading frame.

The sequences upstream of the open reading frame are also characteristic of a ribosomal protein, with its tripartite promoter.  $(9,10,11)$  There is a putative TATA sequence at  $-95$  (Figure

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Figure 1: A. Restriction map of the 9.2 EcoRI fragment that contains RPL4A. B. Restriction map and subclones  $A - E$ . The 3.5 Kb Sall-Sall and 1.75 Kb Sall-EcoRI fragments were isolated from the lambda clone  $A83,(2)$  subcloned into fragments A-E that were then sequenced. The solid bar represents the open reading frame predicted from the nucleotide sequence.  $BI = Ball$ ,  $BII = BgIII$ ,  $E = EcoRI$ ,  $H = HindIII$ ,  $S = Sall$ ,  $Sp = Spel$ ,  $X = Xbal$ .

2A), the T-rich region from  $-162$  to  $-213$ , and at  $-227$  to  $-240$  a potential RPG box, the upstream activating sequence for transcription of genes for ribosomal proteins, translation factors, RNA polymerase subunits and many others. (Reviewed in 1)

The codon usage for IA, like that for other ribosomal proteins of yeast, is characteristic of an abundant protein. The predicted protein is highly basic, the basic amino acids outnumbering the acidic ones by 49 to 23, also a characteristic of ribosomal proteins. The predicted molecular weight and charge of L4 are entirely consistent with its migration on a two dimensional polyacrylamide gel. (12)

#### Yeast IA is Homologous to Mammalian L7a

Comparison of the sequence of L4 with several databanks led to the finding that it is highly homologous to the mammalian ribosomal protein L7a, sequenced in human (3) and mouse (4).(See Figure 3) (Note that the mammalian ribosomal proteins L7 and L7a are entirely distinct (13)). Mammalian L7a is slightly longer than yeast LA at both the N and C terminal ends, but the rest of the sequence is homologous throughout. This finding confirms the extraordinary conservation of the sequence of ribosomal proteins through the evolution of the eukaryotes (reviewed in 1, 14). However, LA does not have substantial identity with any of the ribosomal proteins of E. coli.

A particularly interesting aspect of the sequence is the relatively hydrophobic region of amino acids 156-177, rather uncharacteristic of ribosomal proteins. This region is 95% identical between human and yeast, suggesting that it has an important function.

#### There are Two Functional Genes for L4

Many of the proteins of the yeast translation apparatus, including a majority of the ribosomal proteins, are encoded by two genes. (1) To ask if IA were in this category, <sup>a</sup> genomic Southernwas probed with an oligonucleotide from just outside the coding region (Figure 4A) and with another from within the coding region (Figure 4B). In the former case we detect only one band, in the latter two, even under stringent washing conditions, suggesting that there are two genes.

#### SEQUENCE OF RPL4A OF S. CEREVISIAE



Figure 2: Nucleotide sequence of the RPL4A gene and predicted amino acid sequence of yeast ribosomal protein L4 (EMBL Accession # X17204). The nucleotides are numbered with respect to the translation initiation codon, designated  $+1.$ 

To ask about the function of RPLAA, it was disrupted by insertion of a 1.1 kb HindIII fragment carrying the  $URA3$  gene into a BalI site downstream of codon 52 in the plasmid pYEPLAA. (See Figure 2) The 3.5 kb Sall fragment derived from the resulting construct was transformed into <sup>a</sup> diploid cell, where it could recombine with one of the two authentic RPL4A genes (15). Selection on -Ura medium yielded transformants that have the expected 1.1kb insert in one of their two RPL4A genes. (Figure 5) On sporulation of this diploid, and subsequent dissection of the tetrads, we observed four viable spores, two of which gave rise to significantly smaller colonies. Figure <sup>5</sup> shows that two of the spores had no wild type copy of RPL4A. These were Ura<sup>+</sup> and grew slowly, with a doubling time  $30\%$ longer than wild type cells. Therefore RPL4A must be functional.

Two dimensional polyacrylamide gel analysis of the ribosomal proteins of the Ura<sup>+</sup> cells showed that the ratio of L4 to other



COXPARISON 0P NUXNN L7a WITH 8.CZRBVISIAZ L4A

Figure 3: Amino acid sequence homology between Saccharomyces L4 and Homo L7a (HUML7a) (3). The predicted amino acid sequences of LA and L7a were compared (18). Identical amino acid residues are indicated by two dots, conservative amino acid substitutions by one dot.



Figure 4: Copy number analysis of genes for L4. A Southern blot of yeast genomic DNA (20 ug) digested either with EcoR1, Sal1, Kpn1, Pst1 or BglI, was hybridized to two kinased oligonucleotides derived from the RPL4A gene. One oligonucleotide (A) was from the coding region  $(+611$  to  $+630)$ ; the other (B) was from a noncoding region  $(-146 \text{ to } -127)$ . The numbers on the left indicate the size of the lambda HindIlI markers. Note that restriction enzyme sites for SalI, KpnI, and BglII lie between the two probes from RPLAA; therefore the sizes of the fragments in A and B are not the same. The bands representing the RPL4A gene in B are indicated by. asterisks.

ribosomal proteins was near wildtype (data not shown). This result demonstrates that the other copy of the L4 gene, RPL4B, is also functional. These results are similar to those obtained with a number of other ribosomal protein genes in yeast, e.g. RP51 (16) which has two active genes each of which is capable of



Figure 5: Gene disruption of RPL4A. A: Genomic Southern blot of the parental diploid strain (P), a transformed diploid transformed with RPL4A into which URA3 had been inserted (T), and four spores of a tetrad derived from the transformed diploid  $(A - D)$ . DNA samples were digested with EcoRI and probed with a nick translated 0.77 kb BglII-XbaI fragment that is upstream of the coding region (see Figure 1). The insertion of the 1.1 kb URA3 gene increases the size of the EcoRI fragment from 9.2 to 10.3 kb. Its presence in the 10.3 kb fragment was confirmed by reprobing the blot with URA3 sequences (data not shown). The Ura phenotype and the doubling time (Td) of each strain is shown (see text).

supporting growth, albeit at a reduced rate. It is curious that S. cerevisiae has this duplication of ribosomal protein genes whereas mammalian organisms have numerous pseudogenes but only a single functional gene (17).

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