

## ***N*-Aroyl-3,5-bis(benzylidene)-4-piperidones: A novel class of antimycobacterial agents**

Umashankar Das<sup>a</sup>, Swagatika Das<sup>a</sup>, Brian Bandy<sup>a</sup>, James P. Stables<sup>b</sup>, and Jonathan R. Dimmock<sup>a,\*</sup>

<sup>a</sup>College of Pharmacy and Nutrition, University of Saskatchewan, Saskatoon, Sask., Canada S7N 5C9

<sup>b</sup>National Institute of Neurological Disorders and Stroke, Rockville, MD 20852, USA

### **Abstract**

A number of 3,5-bis(benzylidene)-4-piperidones **1** and some *N*-4-(2-aminoethoxy)phenylcarbonyl analogs **3–6** display excellent in vitro antimycobacterial properties. In particular, **1c** and **6d** are potent antimycobacterials which are well tolerated in mice and are identified as important lead molecules. The nature of both the benzylidene aryl rings and the terminal basic groups which affect the antimycobacterial potencies and the absence of neurotoxic side effects were identified. Several representative compounds stimulated respiration in mitochondria isolated from rat liver and this effect was not caused by the swelling of these organelles. Various guidelines for the creation of further related novel antimycobacterial agents are provided.

### **Keywords**

3,5-Bis(benzylidene)-4-piperidones; Antimycobacterial; Neurotoxic; Mitochondria; Rodent toxicity; Piperidones; Antitubercular

## **1. Introduction**

Tuberculosis is an infection caused by *Mycobacterium tuberculosis* and is a major problem at the present time. Currently this microorganism is present in approximately two billion people.<sup>1</sup> There are about 8 million new cases each year and at least 2 million people die annually from this disease.<sup>2</sup> The rise in the incidence of mycobacterial infections is multifactorial including the pandemic of acquired immunodeficiency syndrome which lowers resistance to infection by *M. tuberculosis*.

At the present time, a number of drugs are available for the initial treatment of tuberculosis, principally rifampin, isoniazid, ethambutol and pyrazinamide. These drugs are often administered over a period of six months<sup>3</sup> and this prolonged treatment leads on occasions to the development of drug resistance.<sup>4</sup> Thus novel classes of antimycobacterials are urgently required which have modes of action that are divergent from those drugs used in current therapy.

\*Corresponding author. Tel.: +1 306 966 6331; fax: +1 306 966 6377; jr.dimmock@usask.ca.

A major interest in these laboratories is the synthesis and bioevaluations of Mannich bases (3-aminoketones) derived from  $\alpha,\beta$ -unsaturated ketones.<sup>5</sup> Various studies revealed that the mode of action of representative compounds includes alkylation of thiols<sup>6–8</sup> and interference with respiration in the electron transport chain in mitochondria.<sup>9–11</sup> Thus the way in which these compounds exert their bioactivities is likely different from that of contemporary antimycobacterial drugs, for example, ethambutol interferes with the incorporation of mycolic acids into the bacterial cell wall.<sup>12</sup> A number of acyclic Mannich bases possess antibacterial properties<sup>13–15</sup> and recently a series of these molecules were described which displayed antimycobacterial activity.<sup>16</sup> However a recurrent problem with many acyclic Mannich bases is their ability to cause neurotoxicity as well as death by respiratory depression.<sup>17,18</sup> Thus consideration was given to attaching arylidene groups to a cyclic Mannich base, namely 4-piperidone, leading to **1a** which was isolated as the hydrochloride salt. This compound showed excellent potency towards *M. tuberculosis* whereby the growth of the microorganism was inhibited completely at a concentration of 6.25  $\mu\text{g/mL}$ .<sup>19</sup> Intraperitoneal injection of **1a** hydrochloride into mice revealed that this compound lowered hepatic thiol concentrations.<sup>20</sup> Hence the interaction of the 1,5-diphenyl-3-oxo-1,4-pentadienyl group of **1a** with cellular thiols likely accounts, at least in part, for its antimycobacterial properties. Compound **1a** had been shown earlier to have cytotoxic properties<sup>21</sup> and recently a project was initiated to use **1a** as the lead molecule in the simultaneous development of both candidate antimycobacterial and cytotoxic agents. In both cases the hypothesis was made that the 1,5-diphenyl-3-oxo-1,4-pentadienyl group interacts at a primary binding site while groups attached to the piperidyl nitrogen atom could interact at a complementary binding site thereby increasing potencies. Conversely, steric repulsion between the *N*-substituents and various groups adjacent to the primary binding site could take place. This communication outlines the efficacy of these compounds as antimycobacterial agents; the synthesis and cytotoxic properties of these molecules have recently been disclosed.<sup>22</sup>

Initially van der Waals bonding between aryl ring C and a possible complementary area on a binding site was considered leading to the decision to prepare **2a**. However at a concentration of 6.25  $\mu\text{g/mL}$ , this compound was inactive. Notwithstanding this result, the placement onto aryl ring C of a methoxy group, which has hydrophilic and lipophilic components which would allow hydrogen and hydrophobic bonding, respectively, to occur as found in **2b**, restored antimycobacterial properties *vide infra*. Thus the decision was made to increase the size of the group on ring C with a mixture of hydrophilic and hydrophobic components by preparing **3a**, **4a**, **5a**, **6a** and **7a**. In order to compare the relative efficiencies of the different *N*-acyl groups to increase antimycobacterial properties, various substituents were placed in aryl rings A and B of **1a** leading to **1b–d** which were subsequently converted to the corresponding amides in series **3–7**. In order to gain an appreciation of the possible contribution of the *N*-acyl group per se to antimycobacterial properties, the synthesis and bioevaluation of **8** were planned. This molecule contains most of the structural features of the *N*-aroyl substituent in series **4**. The recent reports of the promising antimycobacterial properties of a number of aromatic compounds containing 2-aminoethoxyphenyl groups<sup>23–25</sup> afforded a further reason for developing these compounds for assessment against *M. tuberculosis* (Fig. 1).

A further segment of the study involved assessing the compounds for murine neurotoxicity and survival while selected compounds were planned to be examined for any toxic symptoms in rats.

In summary, the objectives of the present study were to examine a variety of 3,5-bis(benzylidene)-4-piperidones and *N*-aroyl analogs as candidate antimycobacterials and to assess their toxicity in rodents. The aspiration was that from the biodata generated, a decision could be reached whether a major expansion in this area was warranted or not.

## 2. Results

All of the compounds in series **1–8** were evaluated against *M. tuberculosis* H<sub>37</sub>Rv. This strain was chosen since it is drug sensitive and is more virulent than *M. tuberculosis* H<sub>37</sub>Ra.<sup>27</sup> The data are presented in Table 1. The enones **1c**, **1d**, **3a** and **6d** were examined for their effect on respiration in rat liver mitochondria; these results are portrayed in Figure 2. Both **1c** and **3a** were assessed for their ability to cause swelling in mitochondria. All of the compounds in series **1–8** were examined for neurotoxicity and survival in mice while several compounds were assessed in rats.

## 3. Discussion

The initial evaluation of the compounds in series **1–8** against *M. tuberculosis* is presented in Table 1. A compound is considered to be active which causes at least 90% inhibition of the growth of the microorganism at a concentration of 6.25 µg/mL. This criterion was met by 75% of the compounds in series **1** and 69% of the amides **3–6**.

The 4-piperidones **1a–c** possess antimycobacterial properties and the minimum inhibitory concentration (MIC) figures, which are presented in Table 1, reveal that **1c** in particular is a promising lead molecule. The lack of inhibition of *M. tuberculosis* by **1d** is surprising and the evaluation was repeated in which case the inhibition was 4% at a concentration of 6.25 µg/mL, thereby confirming its inactivity. In contrast to **1a–c**, the 4-methyl analog **1d** has an electron-donating group in rings A and B which may have contributed to its lack of potency. Clearly, development of series **1** as candidate antimycobacterials is warranted whereby one or more electron-attracting groups are placed in the aryl rings. In addition, a null hypothesis should be evaluated by preparing analogs having negative Hammett  $\sigma$  values in the aryl rings; these compounds would be predicted to be weakly potent or possibly inactive antimycobacterials.

While benzylation of **1a** led to the inactive amide **2a**, the 4-methoxy analog **2b** displayed weak antimycobacterial properties. Expanding the hydrophilic–hydrophobic methoxy substituent in ring C led to the 1-[4-(2-aminoethoxy) phenylcarbonyl]-3,5-bis(benzylidene)-4-piperidones **3–6** which are a new cluster of antimycobacterials.

An examination of the biodata in series **3–6** was made in order to determine the optimal substitutions in aryl rings A and B and also the preferred terminal amino functions. In regard to the aryl substituents, all of the unsubstituted compounds (**3a**, **4a**, **5a**, **6a**) and 4-methyl analogs (**3d**, **4d**, **5d**, **6d**) were active whereas the 4-chloro and 4-nitro derivatives displayed

activity in only one and two compounds, respectively. This result may be influenced by the electronic and steric properties of the R<sup>1</sup> groups since the Hammett  $\sigma$  values of the hydrogen atom and the 4-methyl, 4-nitro and 4-chloro substituents are 0.00, -0.17, 0.78 and 0.23, respectively, while the molecular refractivity (MR) figures are 1.03, 5.65, 7.36 and 6.03, respectively.<sup>28</sup> Thus amplification of the project should include the placement of small electron donating substituents in rings A and B, for example, the 4-hydroxy group ( $\sigma_p = -0.66$ ; MR = 5.42).<sup>28</sup>

In regard to the terminal basic group, the criterion for activity was met by three of the four members in series **3**, **5** and **6** suggesting that there is considerable tolerance for the size of the terminal basic group at a binding site. The observation that only **4a**, **d** displayed significant antimycobacterial properties indicates that in developing these compounds as candidate antimycobacterials, using the diethylamino substituent as the terminal basic group should be avoided. Neither the quaternary ammonium salts **7a–d** nor the aromatic acid **8** displayed noteworthy antimycobacterial properties. Since *M. tuberculosis* contains a thick cell wall, drug penetration of this microorganism is likely to be more facile with hydrophobic molecules. In fact a number of studies have reported positive correlation between log *P* values and antimycobacterial properties.<sup>4,29</sup> Hence the log *P* figures of the compounds in series **1–8** were calculated and are presented in Table 1. While no definite conclusions were drawn between the antimycobacterial potencies and hydrophobicity, two observations may be noted. First, those molecules causing greater than 90% inhibition at 6.25  $\mu\text{g/mL}$  have log *P* values of ~5–8 indicating that these compounds are markedly hydrophobic. It is conceivable that this property, inter alia, contributes to the bioactivity observed. Second, the compounds in series **7** and **8** which are virtually bereft of antimycobacterial potencies have substantially lower hydrophobic properties than the other compounds listed in Table 1.

Various acyclic Mannich bases interfere with respiration in mitochondria.<sup>30</sup> Thus an investigation was undertaken to determine if one of the ways in which bioactivity of these compounds is mediated involves interfering with respiration in mitochondria and if so, whether the magnitude of the responses correlated with antimycobacterial potencies. Representative compounds were chosen which have different potencies namely **1c**, **3a** and **6d**, possessing MIC values of 0.2, 1.56 and 0.78  $\mu\text{g/mL}$ , respectively, as well as **1d** which is bereft of antimycobacterial properties in the bioassay employed. These compounds were also chosen in view of their being well tolerated in mice *vide infra*. The effects of these compounds on rat liver mitochondria are portrayed in Figure 2. At the concentrations utilized, stimulation of respiration was observed. Thus interference with mitochondrial respiration is one way whereby antimycobacterial properties are mediated and, as mentioned previously, it is markedly divergent from the mechanisms of action of contemporary antimycobacterial drugs. The percentage stimulation is in order of **1c** > **3a** > **1d**, **6d**. Since the relative antimycobacterial potencies are **1c** > **6d** > **3a** > **1d**, there is no correlations between this observation and the percentage of stimulation of respiration.

Stimulation of respiration in mitochondria may be caused by different biochemical mechanisms including the swelling of these organelles. Concentrations of 5, 10 and 100  $\mu\text{M}$

of **1c** and **3a** did not demonstrate this effect. Hence the increases in respiration noted are caused in other ways which may include uncoupling of oxidative phosphorylation.

A major factor in considering whether to develop antimycobacterial drugs is their tolerability in animals. In particular, an investigation was conducted to ascertain whether neurotoxicity and/or lethality were found in these cyclic Mannich bases. All of the compounds in series **1–8** were administered intraperitoneally to mice using doses of 30, 100 and 300 mg/kg and the animals were observed after 0.5 and 4 h. These data are presented in Table 1. The following observations were made regarding structure–toxicity relationships. First, in terms of survival, mice tolerated the maximum dose of 300 mg/kg of each compound in series **1–6** and **8**. For series **7**, deaths were noted using 100 mg/kg (**7a** and **c**) and 300 mg/kg (**7d**); this observation coupled with their relatively low inhibitory properties towards *M. tuberculosis* indicate that these quaternary ammonium compounds do not serve as prototypes for further development as candidate antimycobacterials. Second, neurotoxicity was absent in approximately half of the compounds. No neurological deficit was observed when a dose of 30 mg/kg was administered to mice except in the case of **4a** (in 2/4 animals after 0.5 h). In general, neurotoxicity was more pronounced at the end of 0.5 than 4 h. Third, neurotoxicity was absent in series **1** and bearing in mind the promising antimycobacterial properties of **1a–c**, expansion of this series is clearly warranted. Fourth, an assessment was made of the optimal aryl substituents and terminal basic group in regard to the absence of neurotoxicity of the compounds in series **3–6**. Three compounds containing a 4-chloro group (**3b**, **4b**, **6b**), two with a 4-methyl substituent (**4d** and **6d**) and in one unsubstituted analog (**3a**) did not display neurotoxicity. Thus 4-chloro and 4-methyl groups (and other halogens and alkyl substituents) are recommended for placement in the aryl rings A and B when developing these compounds. Two 4-piperidones in each of the series **3**, **4** and **6** did not display neurotoxicity revealing that the dimethylamino, diethylamino and 4-morpholinyl groups are preferable to the 1-piperidino substituent.

Fifth, a dose of 50 mg/kg of **3a**, **4a** and **6b** was injected intraperitoneally into rats and a similar result to the murine screens was observed, that is, only **4a** showed neurological deficit. However when 50 mg/kg of **4a** was administered to rats by the oral route, no neurotoxicity was observed indicating that the appropriate route of administration may eliminate the side effect of neurotoxicity. In addition, **3a**, **3c**, **6b** and **7c** were given orally to rats using a dose of 30 mg/kg and no neurotoxicity was noted. The observation with rats was generally made 0.25, 0.5, 1, 2 and 4 h after administering the compound.

## 4. Conclusions

There is a crucial need to develop agents to combat tuberculosis whose structures and mode of action are divergent from contemporary drugs. This study has demonstrated that some 3,5-bis(benzylidene)-4-piperidones **1** have excellent antimycobacterial properties and are well tolerated in mice. The 1-[4-(2-aminoethoxy) phenylcarbonyl]-3,5-bis(benzylidene)-4-piperidones **3–6** are a new group of potent antimycobacterial agents representative of which stimulate respiration in mitochondria. In general these compounds are well tolerated in mice. Various guidelines have been formulated for expansion of the project based on the

correlations noted between variations in the structures and the bioactivities observed which are summarized in Figure 3.

## 5. Experimental

### 5.1. Synthesis of compounds 1–8

The preparation of the compounds **1–8** has been described previously.<sup>22</sup>

### 5.2. Antimycobacterial evaluations

All of the compounds were evaluated initially against *M. tuberculosis* H<sub>37</sub>Rv (ATCC 27294) using concentrations of 12.5 µg/mL (**1a–c**) and 6.25 µg/mL for the remaining compounds. The methodology for conducting this assay, as well as the determination of the MIC values, was undertaken in BACTEC 12B medium using the broth microdilution assay.<sup>27</sup> The MIC values are the lowest concentrations which cause a 90% reduction in fluorescence relative to controls.

### 5.3. Evaluation of various compounds on respiration and swelling in rat liver mitochondria

Rats were euthanized by isoflurane anesthesia and then decapitated. A literature procedure was used for isolating the hepatic mitochondria by differential centrifugation.<sup>31,32</sup> A homogenization buffer of 250 mM sucrose, 10 mM Hepes, 1 mM EDTA, pH 7.2 and a wash buffer of 250 mM sucrose, 10 mM Hepes, 0.3 mM EGTA, pH 7.2 were employed. The final mitochondrial pellet was resuspended in 1–2 mL of 250 mM sucrose, 10 mM Hepes, pH 7.2 to yield 50–80 mg protein/mL. The mitochondrial preparations displayed respiratory control ratios of 5–7.

Mitochondrial oxygen consumption was measured polarographically using a respirometer system (Hansatech Instruments Ltd, Norfolk, UK). Fresh mitochondria (0.5 mg protein/mL) were incubated at 30 °C in a respiratory buffer of 125 mM sucrose, 65 mM potassium chloride, 10 mM Hepes, 5 mM potassium phosphate, 1 mM magnesium chloride, pH 7.2 containing 5 mM succinate as respiratory substrate.

Mitochondrial swelling was observed using fresh mitochondria incubated at 30 °C in a respiratory buffer composed of 125 mM sucrose, 65 mM potassium chloride, 10 mM Hepes, 5 mM potassium phosphate, 1 mM magnesium chloride, pH 7.2 and 5 mM succinate. The mitochondrial swelling was followed by the loss of absorbance at 520 nm.<sup>33</sup>

### 5.4. Toxicity and neurotoxicity screens

The toxicity and neurotoxicity tests were carried out by previously published method.<sup>34</sup> Specific details of the toxicity and neurological deficit are as follows. Mortalities in mice were caused by three compounds (number of animals died/number of animals tested, dose in mg/kg, time of observation in h), namely: **7a** (8/8, 100, 0.5; 4/4, 300, 0.5), **7c** (2/8, 100, 0.5; 1/8, 100, 0.5–4; 4/4, 300, 0.5) and **7d** (2/2, 300, 4). Neurotoxicity in mice is presented in Table 1. In the case of rats injected intraperitoneally with 50 mg/kg of **4a**, neurological deficit was noted in the following number of animals/total number of rats tested (time in h in parentheses), namely 3/8 (0.25), 5/8 (0.5), 6/8 (1), 4/8 (2), 2/8 (4), 0/8 (6) and 0/8 (24). The



ataxia was mild but all animals were reluctant to move unless stimulated by a tail pinch up to and including 6 h after the administration of **4a**. All animals behaved in a normal fashion after 24 h. The following compounds (dose in mg/kg in parentheses) were administered to rats by the oral route and the animals examined after 0.25, 0.5, 1, 2 and 4 h, namely **3a** (30), **3c** (30), **4a** (50), **6b** (30) and **7c** (30). No neurotoxicity was observed. Four animals were employed in all cases, except in the case of **3c** after 0.5 h, only three rats were observed. The animals were handled, fed and housed following the procedures outlined in the National Research Council Publication 'Guide for the Care and use of Laboratory Animals'. Euthanasia of the mice and rats used in this study was consistent with the policies of the Institute of Laboratory Resources which describe the humane care of laboratory animals.

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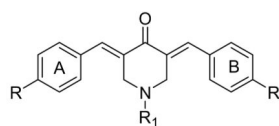
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## References and notes

1. Nayyar A, Jain R. *Curr Med Chem.* 2005; 12:1873–1886. [PubMed: 16101507]
2. Gadad AK, Noolvi MN, Karpoomath RV. *Bioorg Med Chem.* 2004; 12:5651–5659. [PubMed: 15465343]
3. Bass JB Jr, Farer LS, Hopewell PC, O'Brien R, Jacobs RF, Ruben F, Snider DE Jr, Thornton G. *Am J Respir Crit Care Med.* 1994; 149:1359–1374. [PubMed: 8173779]
4. Sriram D, Yogeewari P, Reddy SP. *Bioorg Med Chem Lett.* 2006; 16:2113–2116. [PubMed: 16464574]
5. Dimmock JR, Kumar P. *Curr Med Chem.* 1997; 4:1–22.
6. Mutus B, Wagner JD, Talpas CJ, Dimmock JR, Phillips OA, Reid RS. *Anal Biochem.* 1989; 177:237–243. [PubMed: 2729541]
7. Dimmock JR, Raghavan SK, Logan BM, Bigam GE. *Eur J Med Chem.* 1983; 18:248–254.
8. Pati HN, Das U, Sharma RK, Dimmock JR. *Mini-Rev Med Chem.* 2007; 7:131–139. [PubMed: 17305587]
9. Dimmock JR, Shyam K, Hamon NW, Patil SA, Smith PJ. *Neoplasma.* 1985; 32:85–91. [PubMed: 3982563]
10. Dimmock JR, Hamon NW, de Gooijer CA, Grant GF, Jonnalagadda SS, Hancock DS. *Pharmazie.* 1984; 44:560–562.
11. Hamon NW, Kirkpatrick DL, Chow EWK, Dimmock JR. *J Pharm Sci.* 1982; 71:25–29. [PubMed: 6276528]
12. Martin, AR. Wilson and Gisvold's Textbook of Organic Medicinal and Pharmaceutical Chemistry. 10. Delgado, JN., Remers, WA., editors. Lippincott-Raven Publishers; Philadelphia: 1998. p. 207
13. Khachatourians GG, Holmlund PK, Dimmock JR. *J Pharm Sci.* 1984; 73:803–808. [PubMed: 6204038]
14. Dimmock JR, Nyathi CB, Smith PJ. *J Pharm Sci.* 1979; 68:1216–1221. [PubMed: 512849]
15. Lorand T, Koesis B, Sohar P, Nagy G, Kispal G, Krane HG, Schmitt H, Weckert E. *Eur J Med Chem.* 2001; 36:705–717. [PubMed: 11672880]
16. Dimmock JR, Kandepu NM, Das U, Zello GA, Nienaber KH. *Pharmazie.* 2004; 59:502–505. [PubMed: 15296085]
17. Dimmock JR, Patil SA, Shyam K. *Pharmazie.* 1991; 46:538–539. [PubMed: 1784618]

18. Dimmock JR, Jonnalagadda SS, Phillips OA, Erciyas E, Shyam K, Semple HA. *J Pharm Sci.* 1992; 81:436–440. [PubMed: 1403675]
19. Jha A, Dimmock JR. *Pharmazie.* 2006; 61:562–563. [PubMed: 16826979]
20. Dimmock JR, Arora VK, Wonko SL, Hamon NW, Quail JW, Jia Z, Warrington RC, Fang WD, Lee JS. *Drug Des Deliv.* 1990; 6:183–194. [PubMed: 2076179]
21. Dimmock JR, Padmanilyam MP, Puthucode RN, Nazarali AJ, Motaganahalli NL, Zello GA, Quail JW, Oloo EO, Kraatz HB, Prisciak JS, Allen TM, Santos CL, Balzarini J, De Clercq E, Manavathu EK. *J Med Chem.* 2001; 44:586–593. [PubMed: 11170648]
22. Das U, Alcorn J, Shrivastav A, Sharma RK, De Clercq E, Balzarini J, Dimmock JR. *Eur J Med Chem.* 2007; 42:71–80. [PubMed: 16996657]
23. Das SK, Panda G, Chaturvedi V, Manju YS, Gaikwad AK, Sinha S. *Bioorg Med Chem Lett.* 2007; 17:5586–5589. [PubMed: 17764933]
24. Panda G, Parai MK, Das SK, Shagufta, Sinha M, Chaturvedi V, Srivastava AK, Manju YS, Gaikwad AN, Sinha S. *Eur J Med Chem.* 2007; 42:410–419. [PubMed: 17112639]
25. Panda G, Shagufta, Mishra JK, Chaturvedi V, Srivastava AK, Srivastava R, Srivastava BS. *Bioorg Med Chem.* 2004; 12:5269–5276. [PubMed: 15388155]
26. ACD/Labs release 10.00, version 10.02. <http://www.acdlabs.com>
27. Collins L, Franzblau SG. *Antimicrob Agents Chemother.* 1997; 41:1004–1009. [PubMed: 9145860]
28. Chu, KC. *Burger's Medicinal Chemistry, Part I.* 4. Wolff, ME., editor. John Wiley and Sons; New York: 1980. p. 401
29. Sivakumar PM, Seenivasan SP, Kumar V, Doble M. *Bioorg Med Chem Lett.* 2007; 17:1695–1700. [PubMed: 17276682]
30. Dimmock JR, Shyam K, Hamon NW, Logan BM, Raghavan SK, Harwood DJ, Smith PJ. *J Pharm Sci.* 1983; 72:887–894. [PubMed: 6620142]
31. Johnson D, Lardy H. *Methods Enzymol.* 1967; 10:94–96.
32. Schneider WC. *J Biol Chem.* 1948; 176:259–266. [PubMed: 18886166]
33. Kowaltowski AJ, Castilho RF, Grijalba MT, Bechara EJH, Vercesi AE. *J Biol Chem.* 1996; 271:2929–2934. [PubMed: 8621682]
34. Stables, JP., Kupferberg, HJ. *Molecular and Cellular Targets for Antiepileptic Drugs.* Vanzini, GA, Tanganelli, P., Avoli, M., editors. John Libby and Company Ltd; London: 1997. p. 191-198.



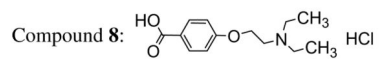


a: R= H; b: R= Cl; c: R= NO<sub>2</sub>; d: R= CH<sub>3</sub>

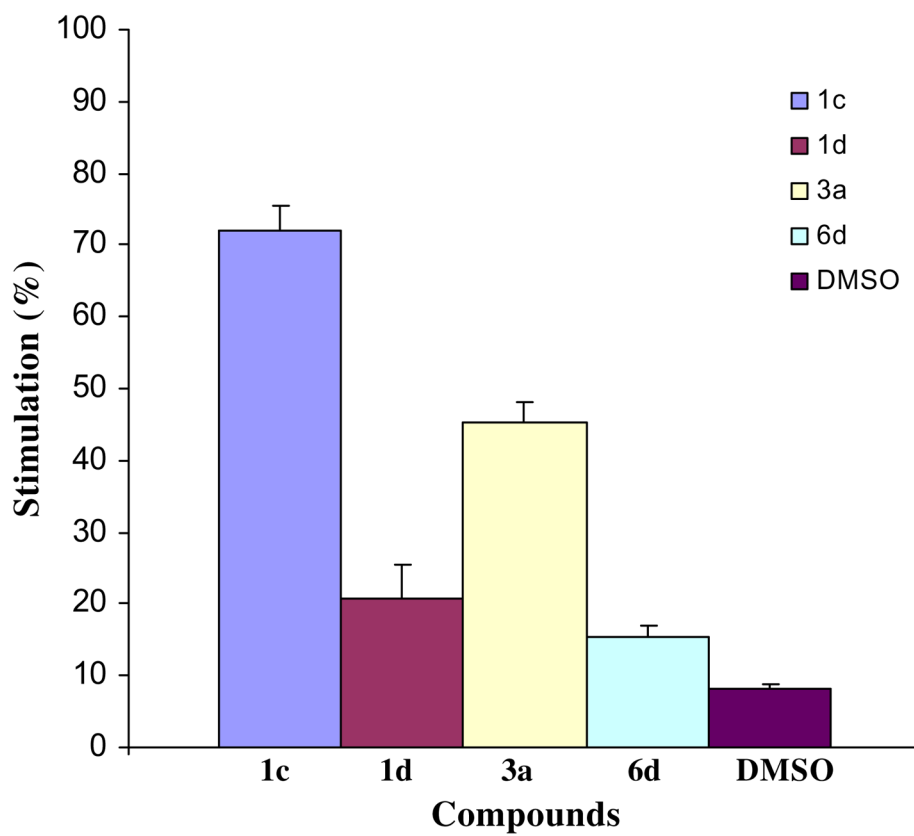
Compounds	R <sub>1</sub>
1a-d <sup>a</sup>	H
2a	
2b	
3a-d <sup>b</sup>	
4a-d <sup>b</sup>	
5a-d <sup>b</sup>	
6a-d <sup>b</sup>	
7a-d	

<sup>a</sup>1a was prepared as the hydrochloride salt.

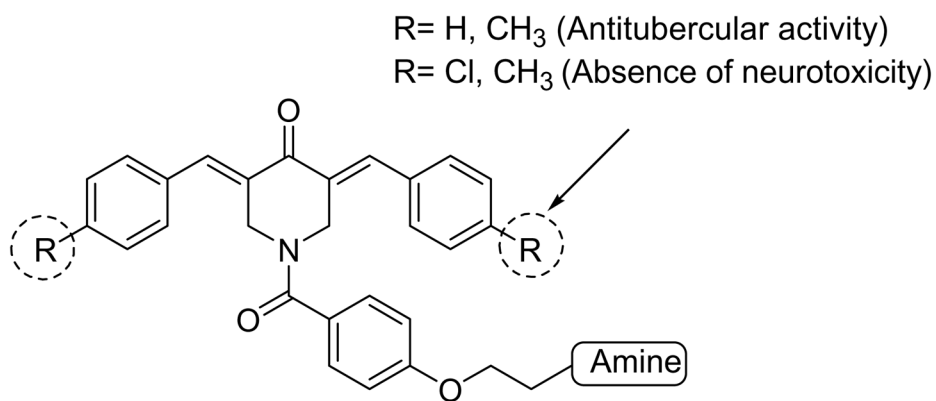
<sup>b</sup>Prepared as hydrochloride salts.



**Figure 1.**  
Structures of the compounds in series 1–8.



**Figure 2.** Effects of 10  $\mu$ M of **1c**, **1d**, **3a** and **6d** as well as solvent control (dimethylsulfoxide 4  $\mu$ L) on respiration in rat liver mitochondria.



Amine = -N(CH<sub>3</sub>)<sub>2</sub>, -N $\square$ , -N $\square$ O (Antitubercular activity)

Amine = -N(CH<sub>3</sub>)<sub>2</sub>, -N(C<sub>2</sub>H<sub>5</sub>)<sub>2</sub>, -N $\square$ O (Absence of neurotoxicity)

**Figure 3.**  
 Optimal aromatic ring substituents and amino groups for displaying antimycobacterial activity and absence of neurotoxicity.

Table 1

Evaluation of the compounds in series 1–8 against *M. tuberculosis* H<sub>37</sub>Rv in vitro and for neurotoxicity and survival in mice

Compound	Antimycobacterial evaluation <sup>a</sup>		log <i>P</i> <sup>b</sup>	Neurotoxicity <sup>c</sup>			
	% Inhibition 6.25 µg/mL	MIC (µg/mL)		0.5 h		4 h	
				100 mg/kg	300 mg/kg	100 mg/kg	300 mg/kg
1a	99	3.13	5.7	—	—	—	—
1b	99	6.25	6.8	—	—	—	—
1c	100	0.2	5.3	—	—	—	—
1d	0	—	6.6	—	—	—	—
2a	0	—	5.5	1/8	3/4	1/4	1/2
2b	47	—	6.0	2/8	—	—	—
3a	98	1.56	5.9	—	—	—	—
3b	100	1.56	7.0	—	—	—	—
3c	27	—	5.5	—	1/4	—	2/2
3d	100	3.13	6.8	1/8	—	—	—
4a	99	—	7.0	1/8	1/4	—	—
4b	37	—	8.0	—	—	—	—
4c	34	—	6.5	1/8	1/4	1/4	1/2
4d	100	3.13	7.9	—	—	—	—
5a	100	6.25	7.1	3/8	2/4	1/4	1/2
5b	86	—	8.1	3/8	3/4	1/4	1/2
5c	100	3.13	6.6	1/8	1/4	—	1/2
5d	100	3.13	8.0	1/8	—	—	—
6a	100	6.25	5.4	—	1/4	—	—
6b	0	—	6.4	—	—	—	—
6c	100	6.25	5.0	—	2/4	—	—
6d	100	0.78	6.3	—	—	—	—
7a	0	—	2.2	<i>d</i>	<i>d</i>	<i>d</i>	<i>d</i>
7b	23	—	3.2	—	—	—	—
7c	0	—	1.8	7/8	4/4	1/2	<i>e</i>
7d	81	—	3.1	—	1/4	—	<i>f</i>

Compound	Antimycobacterial evaluation <sup>a</sup>		log <i>P</i> <sup>b</sup>	Neurotoxicity <sup>c</sup>	
	% Inhibition	MIC (µg/mL)		0.5 h	4 h
<b>8</b>	12	—	2.9	—	—

<sup>a</sup>The hydrochloride salt of **1a** was employed in this assay. A concentration of 12.5 µg/mL was used to obtain the % inhibition figures for **1a-c**. A concentration of 6.25 µg/mL of a reference drug rifampin caused 100% inhibition of the growth of *M. tuberculosis* H37Rv. Rifampin has a MIC value of 0.07 (0.015-0.125) µg/mL. The single line—indicates that no attempt was made to determine the MIC value.

<sup>b</sup>The log *P* values were calculated using the algorithm and database statistics of ACD.<sup>26</sup>

<sup>c</sup>Compounds **1a, b** were evaluated as the hydrochloride salts. The figures refer to the number of animals demonstrating neurotoxicity/total number of animals dosed. The single line—indicates an absence of neurotoxicity.

<sup>d</sup>After 0.5 h, 8/8 and 4/4 mice receiving 100 and 300 mg/kg, respectively, of **7a** were dead.

<sup>e</sup>After 0.5 h, 2/8 mice receiving 100 mg/kg of **7c** were dead and another animal died between 0.5 and 4 h. A dose of 300 mg/kg was fatal to 4/4 mice when observed 0.5 h after the administration of the compound.

<sup>f</sup>After 4 h, 2/2 animals receiving a dose of 300 mg/kg were dead.