

# NIH Public Access

**Author Manuscript** 

J Immunol. Author manuscript; available in PMC 2012 March 25.

# Published in final edited form as:

J Immunol. 2010 July 1; 185(1): 15–22. doi:10.4049/jimmunol.0903856.

# **Understanding Latent Tuberculosis: A Moving Target**

# Philana Ling Lin<sup>\*</sup> and JoAnne L. Flynn<sup>†</sup>

<sup>\*</sup>Department of Pediatrics, Children's Hospital of Pittsburgh of the University of Pittsburgh Medical Center, Pittsburgh, PA 15224

<sup>†</sup>Department of Microbiology and Molecular Genetics, University of Pittsburgh School of Medicine, Pittsburgh, PA 15261

# Abstract

Tuberculosis (TB) remains a threat to the health of people worldwide. Infection with *Mycobacterium tuberculosis* can result in active TB or, more commonly, latent infection. Latently infected persons, of which there are estimated to be ~2 billion in the world, represent an enormous reservoir of potential reactivation TB, which can spread to other people. The immunology of TB is complex and multifaceted. Identifying the immune mechanisms that lead to control of initial infection and prevent reactivation of latent infection is crucial to combating this disease.

In 1991, the World Health Organization recognized the resurgence of tuberculosis (TB) as a global health problem. Despite the initiation of directly observed therapy strategies, TB accounts for 1.8 million deaths in 2008, second only to HIV as an infectious cause of death. New reported cases of TB increased from 8.0 million in 1997 to 9.4 million in 2008 (1). Poverty, the advent of multidrug-resistant cases, imperfect diagnostic assays, poor access to health care, limited vaccine efficacy, and availability of new drugs remain obstacles. The HIV epidemic, especially in poverty-affected countries, is among the most important contributors to the resurgence of TB.

The epidemiology of TB is a key component to understanding the complexity of this global health burden. Mycobacterium tuberculosis, the etiologic agent of TB, is an acid-fast bacillus. Person-to-person transmission of *M. tuberculosis* occurs by aerosolized droplets generated by a person with active disease, which are inhaled into the large and small airways, where infection can be established. An estimated 30% of exposed individuals will have evidence of infection by tuberculin skin test (TST) (2). Among those infected, only 5-10% will develop clinical manifestations of active TB disease within 2 y postexposure (known as primary TB). The majority of infected individuals develop latent infection defined as having evidence of *M. tuberculosis* infection by immunologic tests (TST or IFN- $\gamma$ release assay [IGRA]) without clinical signs or symptoms of disease and a normal chest radiograph. This represents a state of equilibrium in which the host is able to control the infection but not completely eradicate the bacteria. Patients with latent infection are the largest reservoir for potential transmission. Although most patients with latent infection will not die of TB, the greatest danger is in reactivation (active TB after remote infection) cases and the subsequent silent spread to close contacts. The majority of active TB disease in lowprevalence regions, such as the United States, is attributed to reactivation TB (47-87%) (3-

Copyright © 2010 by The American Association of Immunologists, Inc.

Address correspondence and reprint requests to Dr. JoAnne L. Flynn, Department of Microbiology and Molecular Genetics, University of Pittsburgh School of Medicine, W1144 Biomedical Science Tower, Pittsburgh, PA 15261. joanne@pitt.edu.

Disclosures

The authors have no financial conflicts of interest.

6). In contrast, recent infection accounts for the majority of active TB cases in highly endemic areas (7). The risk of reactivation TB is estimated as 10% per lifetime. Impaired immunity, as in the case of HIV infection, increases the risk to 10% per year (8). Active TB, either from recent infection or reactivation, results in contagious spread of the pathogen. For every year that a single person with active TB is untreated, he or she will infect an average of 10–15 people (1). HIV-negative people are estimated to be sputum smear positive (i.e., with acid fast bacilli observed in sputum) 1–3 y prior to diagnosis in resource-poor settings (9, 10). By these estimates, a single person with active TB could infect as many as 45 other individuals. It is no wonder that this disease remains a global health threat for which the incidence has not been reduced despite tremendous efforts.

A growing amount of clinical and epidemiologic data has recently challenged the notion that *M. tuberculosis* infection exists only as a bimodal distinction of latent infection and active TB (11). In fact, a variable spectrum of latent infection likely exists just as active TB has varying degrees of severity. In this review, we discuss the immunologic establishment and maintenance of latent infection, limitations of the data and how they relate specifically to human infection, risk factors of reactivation, and potential mechanisms, and the changing paradigm of latent infection.

# **Establishing latent infection**

#### From the beginning

Infection first occurs when the bacilli enter the alveoli and are phagocytized by alveolar macrophage and resident dendritic cells. Dendritic cells carrying bacilli and Ags travel from the distal airways to the draining mediastinal lymph nodes, where they initiate T cell responses. Lymphocytes and macrophages then migrate to the primary site of infection to form a granuloma. In most animal models of TB, bacterial growth increases logarithmically and then reaches a plateau coincident with T cell response initiation, when granulomas are observed histologically (12). The granuloma, the hallmark of TB, is a focal collection of inflammatory cells that have specific architectural structures in humans. Granulomas are thought to represent an immunologic and physical barrier to contain infection and prevent dissemination. Maintenance of the granulomas is a dynamic process of continual immunologic control of bacterial replication.

#### The innate immune response

In the United States, close-contact investigations revealed that an estimated 20–30% of close contacts had latent infection, and another 1% had active TB (2, 13). This indicates that 70–80% of exposed individuals do not become infected. Although difficult to prove, these data suggest that some humans are resistant to infection and that the innate immune response likely plays an important role in clinical outcome.

Host resistance against mycobacterial infection begins with the innate immune response involving interaction of the bacillus with macrophages and dendritic cells. TLRs have been recognized as important pattern recognition receptors for *M. tuberculosis* infection. Increased susceptibility to *M. tuberculosis* in MyD88-deficient mice first indicated that TLRs were important in the initial host response (14–16), although the exact TLRs involved are controversial. However, MyD88-dependent IL-1R expression is also critical for resistance to *M. tuberculosis* (17). In vitro and in vivo studies in murine models have implicated TLR2, TLR4, and TLR9 as important in the response to *M. tuberculosis* (18–24), although other studies have not supported these findings (25, 26). Genetic polymorphisms of TLR genes have been associated with increased risk of *M. tuberculosis* infection or disease (27–29). Human in vitro studies have shown that TLR activation induces vitamin D-dependent production of antimicrobial peptides that have mycobactericidal activity (30, 31).

#### Transition to control of infection

Little is known about the transition between controlling acute infection and establishment of latent infection, in large part due to lack of appropriate animal models. In nonhuman primates, clinical outcome is both dose dependent (32) and host dependent (33). Cynomologus macaques infected with a low dose of *M. tuberculosis* are the only animal model in which both active disease and latent infection have been described and characterized (34); gross signs of disease are observed as early as 4 wk postinfection (35). By 6 wk postinfection, greater production of mycobacteria-specific IFN-γ in PBMCs was observed among monkeys who later developed active disease compared with latent monkeys (34). Human contact studies, in which humans exposed to a case of active TB are followed, have also found that high responses to ESAT6 (an *M. tuberculosis* protein) correlated to active disease outcome (2-y follow-up) compared with those without disease (36). In our experience, increases in IFN- $\gamma$  often reflect increased bacterial burden, suggesting that control of bacterial replication early post-infection plays an important role in establishing latent infection. The amount of gross disease seen at 8 wk post-infection could not predict which monkeys would later develop latent or active disease. Yet the overall bacterial burden at 8 wk is significantly more than latently infected monkeys (P.L. Lin and J.L. Flynn, unpublished observations), suggesting that the host response of latent monkeys reduces the initial bacterial burden over time.

### Adaptive immune responses in establishing and maintaining latency

#### T cells

The cell-mediated immune system is critical to overcoming acute *M. tuberculosis* infection. Cells found within the granuloma include CD4 and CD8 T cells, B cells, macrophages, neutrophils, fibroblasts, and multinucleated giant cells. Murine models have shown that CD4 T cells are required for control of acute and chronic infection (reviewed in Ref. 37). CD4 T cells are major producers of IFN- $\gamma$ , contribute to TNF production, and are important for optimal CD8 T cell function. This has been confirmed in patients with HIV in whom the risk of TB increases with decreasing CD4 T cell counts (38), and in nonhuman primates with SIV, in which the initial reduction in CD4 T cell levels was associated with time to reactivation of latent infection (39). Furthermore, the prevalence of extrapulmonary TB (an indicator of severe disease) was inversely proportional to CD4 T cell count (40).

Although initially controversial, CD8T cells play an important role in the immune response to TB. In some murine acute infection studies, mice lacking functional MHC class I had higher bacterial burden compared with wild-type controls (reviewed in Ref. 41). In mice with minimal bacterial loads due to antibiotic treatment, depletion of CD8 T cells resulted in exacerbation of infection (42). CD8 T cells can also produce IFN- $\gamma$  (though less than CD4 T cells, at least in mice) (43) and TNF, but are best known for their cytotoxic capacity against infected cells. CD8 T cells can secrete perforin that allows pore formation into the cellular membrane of an infected cells and delivery of granule-associated proteins, such as granzymes, resulting in apoptosis. Perforin from CD8 T cells has been shown to play an important protective role during acute infection in mice (44). Human CD8 T cells also produce IFN- $\gamma$  (46), but their role in human TB remains unclear. In rhesus macaques, CD8 depletion resulted in impaired Bacillus Calmette-Guérin–induced immune response during acute *M. tuberculosis* infection (47), suggesting that CD8 T cells play an important role in the protective response.

#### Macrophage activation

Macrophage activation is the key to mycobacterial killing. IFN- $\gamma$  (primarily from T cells) seems to be an essential cytokine for macrophage activation. In mice, the generation of reactive nitrogen intermediates by NO synthase 2 (NOS2) is important during early and chronic infection (48–50), and macrophage activation is synonymous with the production of NOS2 in this model. Although NOS2 expression and function has been observed in human samples (51, 52), its role in human TB warrants further investigation. In humans, alterations in the NOS2A gene have been associated with increased susceptibility to TB (28).

#### Cytokines

Cytokines play a critical role during primary and latent infection. IL-12 is important in the Th1 response, and mice deficient in IL-12 had poor survival and increased bacterial burden compared with controls (53). In humans, genetic defects in the IL-12/IL-23/IFN- $\gamma$  axis are associated with severe disseminated mycobacterial disease (54). Murine studies have shown that IFN- $\gamma$  produced by T cells is critical for early protection and essential for inducing NOS2 (reviewed in Refs. 37, 55). Humans also produce IFN- $\gamma$  in response to mycobacterial Ags, which is the basis for the diagnostic IGRA. In humans, IFN- $\gamma$  also induces autophagy (56) as a mechanism of reducing mycobacterial burden. Genetic defects in IFN- $\gamma$ R increase the susceptibility to TB as well as other nontuberculous mycobacteria (reviewed in Ref. 57).

Lack of functional TNF was long recognized as being critical to controlling acute and chronic murine infection, presumably from the poor granuloma formation observed in that model as well as deficient macrophage activation (58–60). More recently, zebrafish and nonhuman primate models have shown that although TNF is important for overcoming acute infection and preventing reactivation, granuloma formation overall is normal in the absence of TNF (61, 62). In humans, genetic heterogeneity of the TNFR has been associated with increased susceptibility to active TB in Africa (63). The increased incidence of TB among patients treated with anti-TNF agents for inflammatory diseases underscores the importance of TNF (64). Although many of these cases were presumed to be from reactivation TB, there is justified concern over the risk of fulminant acute TB as these drugs become available in high TB endemic areas.

TNF affects expression of adhesion molecules (65) and chemokines (66–68), some of which were found to play important roles in early infection. TNF is also a mediator of apoptosis, which is believed to be detrimental to the survival of mycobacteria within macrophages. In human alveolar macrophages, *M. tuberculosis*-induced TNF production leads to apoptosis as a means of reducing intracellular bacterial burden (69); more virulent strains appear to induce less TNF expression. An attenuated *M. tuberculosis* strain that increases apoptosis induced stronger CD8 T cell responses and provided increased protection against virulent challenge in animal models, indicating that apoptosis is associated with a better outcome of infection (70).

# Immunologic distinctions of active disease and latent infection in humans: the challenges

Human studies, arguably the most relevant, can be challenging. In TB, the time and frequency of exposure, strain, inoculum, severity of disease, and presence of coinfections or other complications can all affect the interpretation of results, yet are difficult, if not impossible, to control. Many studies have examined the immunologic characteristics of active disease and latent infection in humans, although the results can make it difficult to assign cause or effect to any specific factor. *M. tuberculosis*-specific production of IFN- $\gamma$  in blood (IGRAs or ELISPOTs) has been used to detect differences between active and latent

cases with variable success (71–74). In general, active TB cases have been associated with greater production of IFN- $\gamma$  compared with latently infected patients (73, 75), although a large degree of variability between groups results in poor predictive value. Additionally, the prevalence of TB in the area partly determines the predictive value of these assays (i.e., the cutoff for positive and negative results) (76).

An equilibrium of pro- and antiapoptotic regulators likely contributes to clinical outcome. Studies among household contacts and community controls have shown that increased IFN-y to ESAT6 or CFP10 Ags was associated with active disease outcome (36, 77), similar to that observed in nonhuman primates (34). Whole blood mRNA expression of death domain complex (proapoptotic) genes was followed among cohorts with active TB, household contacts, and community controls (78). Contacts with greater response to ESAT6 (presumably at greater risk for developing active TB) (36) had higher expression of TNF and Fas, similar to active TB cases. This is consistent with other studies among active cases of TB, in which severe cases were associated with a greater ratio of TNF to TNFR1 (79, 80). The authors suggest these parameters in household contacts are a measure of active infection rather than latency. Death receptor inhibitors FLIPs and FLIPI were also upregulated in patients with active TB (78). But if increased apoptotic markers are associated with active TB outcome (not confirmed in this study but based on previous studies of a similar cohort), these data contradict the presumption that early apoptotic function is associated with good outcome. Thus, a more complex balance of inflammation may be required for control of infection.

Recent murine data showed that removal of regulatory CD4 T cells (CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup>) decreased bacterial burden (81). In humans, the picture is more complex. An increased frequency of regulatory T cells was observed among active TB cases compared with controls, but this likely reflects a response to excessive inflammation (82, 83). In a separate study among TB contacts, whole blood measurement of FoxP3 mRNA was greater among contacts that never developed a positive TST or IGRA compared with those who did convert to both assays (84). In monkeys prior to infection, a higher frequency of regulatory T cells (CD4<sup>+</sup>FoxP3<sup>+</sup>) was observed in blood among those who would later develop latent infection compared with those that would develop active TB. Soon after infection, a rapid decrease in regulatory T cells in the blood occurred followed by an increase of regulatory T cells in the airways, suggesting migration to the lungs. These cells then rebound in the blood and continue to increase in the subset of monkeys that developed active disease (85). These data suggest that regulatory T cells may play a protective role in early infection, possibly by reducing early pulmonary inflammation, but increase during active disease in response to increased inflammation. In humans with active TB, PBMC expression of a marker of T cell exhaustion, PD-1, was upregulated by IFN- $\gamma$ , suggesting that this may be a potential mechanism of impaired immune response that results in active disease (86). However, in mice, there are few data so far supporting that PD-1 expression plays a major role in control of chronic TB (87).

The recent emphasis in multifunctional T cells (producing multiple cytokines) has led to interesting findings in memory T cell responses. Using tetramers, Caccamo et al. (88) observed that latently infected patients had a much higher proportion of Ag-specific CD8 T cells producing both IFN- $\gamma$  and IL-2 compared with active TB cases. Latent infection was also associated with CD8 T cells with a terminally differentiated phenotype compared with active TB cases with predominantly central and effector memory T cell phenotypes (88). These studies were consistent with murine studies suggesting that vaccine-induced protection occurred via multifunctional T cells (89). However, Sutherland et al. (90) found higher frequencies of *M. tuberculosis*-specific CD4 T cells producing two or more of the cytokines TNF, IL-2, and IFN- $\gamma$  among TB cases compared with contacts. Children with TB

Lin and Flynn

disease had a higher frequency of specific effector memory CD4 T cells expressing IFN- $\gamma$ ,TNF, and/or IL-2 in response to *M. tuberculosis* compared with latent infection (91). These last two studies contradict the notion that multifunctional T cells are protective, but may instead increase as bacterial load increases. Recently, IL-17–producing cells have been shown to mediate vaccine-induced protection in mice (92). Other murine studies have suggested that IL-17 is produced by the  $\gamma\delta$  T cells in the lungs early in infection, suggesting that this cytokine could have different roles at different stages of infection (93). In humans, an increased frequency of IL-17<sup>+</sup> T cells was associated with latent infection (90). In another human study, more severe disease was associated with reduced IL-17<sup>+</sup> T cells (94). Thus, the relative contribution of multifunctional T cells as well as Th17 cells still remains to be determined in human *M. tuberculosis* infection and even in animal models.

In short, limited data prevents us from accurately interpreting which immunologic responses are protective or contribute to the establishment of latency in humans. Presumably proinflammatory Th1 cytokines, such as IFN- $\gamma$  and TNF, would promote bacterial killing necessary to establish latent infection, yet these are seen more in active disease patients, probably due to increased antigenic stimulation from higher bacterial loads. Likewise, multifunctional cytokine-producing T cells, shown to be protective in other infectious models, have not been clearly established as a surrogate marker for protection, yet human vaccine studies are actively using this as a primary endpoint. Anti-inflammatory factors are presumably necessary to limit pathology and inflammation during initial infection, active disease, or even during latency. The anti-inflammatory factors may include regulatory T cells, IL-10, TGF- $\beta$ , alternatively activated macrophages, and even Th2 T cell responses. The reports of immune reconstitution disease (immune reconstitution inflammatory syndrome, IRIS) in HIV<sup>+</sup> patients following initiation of antiretroviral therapy leading to unmasking of subclinical M. tuberculosis infection or even reactivation of latent infection (95, 96) suggests that increased inflammation during the subclinical or latent phase is not beneficial to control of infection. The available data support that a complex balance between inflammation and anti-inflammatory factors is crucial to optimal control of all phases of M. tuberculosis infection (Fig. 1). This balance likely varies during the course of infection and disease, from individual to individual, and even within individual granulomas in an infected lung. The complexity of the host immune response interactions makes it difficult to assign a positive or negative role to each response, as the real answer likely lies in the context of the response, and more specifically, in the context of each individual granuloma, in which the balance of pro- and anti-inflammatory responses is reached and maintained and where the host-pathogen standoff takes place.

#### The bacillus

There has been a major increase in our understanding of the virulence factors associated with *M. tuberculosis*, and many of these virulence factors appear to modulate host responses. This is an important area of study and will be enlightening with respect to host-pathogen interactions and outcome of infection. For the purposes of this review, we focus on the broader idea that the infecting strain can play an important role in the outcome of infection or severity of disease. For many years, it was believed that *M. tuberculosis* was relatively stable in its genomic composition. However, recent evidence demonstrates that the genome of *M. tuberculosis* has much more plasticity than previously appreciated and that there are major differences among strains and isolates that may contribute to virulence and outcome of infection. There are also data that certain clades of *M. tuberculosis* are associated with populations from specific geographic regions and appear to have coevolved with those populations (97). The Beijing strain has emerged in the past decade as a major cause of TB and accounts for ~50% of strains from east Asia. There are data that Beijing strains are more drug resistant, and some, but not all, studies have demonstrated that more

severe disease is associated with Beijing strain infections (reviewed in Ref. 98). In guinea pigs, some Beijing strains were found to be more virulent than non-Beijing strains, but this was not universally true (99). A small study in the Gambia indicated that contacts infected with *Mycobacterium africanum*, a member of the *M. tuberculosis* complex, were less likely to progress to active TB (i.e., more likely to develop latency) than those infected with *M. tuberculosis* strains, including the Beijing strain (100). In one study, strains of a Euro-American lineage were more likely to cause pulmonary disease rather than meningeal TB compared with other strains (101). In this same study, Beijing strains were associated with individuals with polymorphisms in the TLR2 gene, which was previously shown to be involved in susceptibility to TB (102). Thus, the infecting strain has a role in influencing infection outcome, with the final outcome likely dependent on contributions from both host and bacillus.

#### Latent infection: the spectrum

The classic teaching of TB is that it exists strictly as active disease or latent infection without overlap. There is now growing evidence that, like active TB (which can manifest on a continuum of severity), there is likely a spectrum of latent infection. A simple review of IGRA results shows that although the overall mean of IFN- $\gamma$  production is greater in active disease than among latent patients, there is a tremendous degree of variability within the latent as well as active groups, impairing the predictive value of the assay itself (72). Barry et al. (11) recently published a review suggesting that the paradigm of latent infection is changing.

The diagnosis of latent infection is made on the basis of a positive TST or blood test (IGRA) without symptoms and signs (x-ray) of disease, with no further workup required, which limits the ability to detect subclinical disease. In a study of 601 cases of culture-positive TB cases, 9% had normal chest x-rays, and many were asymptomatic. Five percent of these subclinical cases were non-HIV infected, and 22% were HIV infected, a known risk factor for atypical chest x-ray findings (40, 103). These data suggest that subclinical disease occurs, but at a relatively low prevalence. However, subclinical disease (sputum positive despite normal chest x-ray and lack of signs or symptoms of disease with positive TST or blood test) is dramatically higher in HIV<sup>+</sup> patients who live in a high TB endemic area (10, 104, 105). This has resulted in sputum as a standard screening practice for TB in areas with high endemic rates of both HIV and TB (38). A thorough and systematic investigation of subclinical rates of TB among immune competent individuals has not been performed. Recent advances in medical imaging have anecdotally noted metabolically active lesions in humans with latent infection, suggesting that latency can be a dynamic process (106, 107). In the nonhuman primate model, ~5% of infected monkeys develop subclinical infection (34), and preliminary data suggests that they may be more susceptible to reactivation disease (P.L. Lin and J.L. Flynn, unpublished observation).

It stands to reason that where a person lies on the spectrum of latent infection predicts their relative risk of reactivation (Fig. 1). For instance, some humans with unrecognized subclinical TB would otherwise be called latently infected and likely have a high rate of reactivation, whereas patients without subclinical disease are likely to have a lower risk. Conversely, some humans are unlikely to ever reactivate despite immune suppression; this may be due to actual clearance of the infection or to bacilli existing in a truly dormant state, and distinguishing these states is impossible in humans given current technologies. Biomarkers that can discriminate the position on the spectrum of latency are urgently needed to identify those infected individuals for whom immune interventions (such as anti-TNF therapy) may be most risky and those for whom antibiotic therapy is most beneficial.

# **Risk factors for reactivation**

The current treatment for latent infection consists of 9 mo of isoniazid (108), a drug for which the efficacy is dependent on mycolic acid synthesis during active replication. It has been shown that *M. tuberculosis* can persist for decades within a human host (109–111), and therefore it is logical to think of latent infection as a dynamic process of bacterial persistence and immunologic control. Based on epidemiologic studies, known risk factors for reactivation of latent infection include: HIV, malnutrition, tobacco smoke, indoor air pollution, alcoholism, silicosis, insulin dependent diabetes, renal failure, malignancy, and immune suppressive treatment, such as glucocorticoids (112–114).

HIV infection and treatment with TNF inhibitors are the most well-described risk factors for reactivation. TNF inhibitors were first introduced more than a decade ago for treatment of inflammatory diseases. An increased incidence of TB (presumed to be reactivation of latent infection) was noted among patients on TNF inhibitors (reviewed in Refs. 64, 115). Reactivation could also be induced by TNF neutralization in nonhuman primates with true latent infection; unlike the murine model, granuloma structure was completely normal, as confirmed in the human literature (62). Studies in humans treated with TNF inhibitors showed that cells in blood had decreased T cell activation, IFN-y production and proliferation, and decreased CD8 memory T cells with reduced granulysin, though these data did not correlate to reactivation cases (116-118). However, in monkeys, we found appropriate levels of IFN- $\gamma$  within mediastinal lymph nodes, suggesting that immunologic factors in the blood do not necessarily correlate with regional disease. TNF neutralization altered chemokine receptor expression, impaired cellular recruitment (i.e., T cells) to disease sites, and resulted in a disproportionate degree of extrapulmonary disease (62). More importantly, although a high rate of reactivation (~65%) was noted in latently infected monkeys, not all monkeys reactivated following short-term anti-TNF treatment (62), indicating that TNF is an important but not always a critical factor in maintaining latent infection and that the spectrum of latent infection likely plays an important role on the overall risk of reactivation.

HIV remains the most common risk of reactivation TB. The immune suppression by HIV has been an overwhelming factor in the resurgence of TB as a global health threat. The risk of reactivation among HIV patients is almost 10-fold higher than non-HIV patients (113). Prior to the HIV epidemic, 85% of cases of TB were limited to only pulmonary involvement (119). In contrast, a disproportionately high rate of disseminated, extrapulmonary disease was observed in advanced HIV-infected patients with TB (5). The greatest risk of TB was associated with CD4 T cell counts <200 cells/ml regardless of antiretroviral therapy (38, 95), suggesting that CD4 T cells are critical to maintaining latent infection. Even under optimal conditions when CD4 T cell counts improve, the risk of TB is still increased (120, 121), indicating that HIV infection induces additional immune deficiencies independent of CD4 count. Latently infected monkeys were infected with SIV, and all monkeys develop reactivation TB early or late after SIV infection (39). Early reactivation was associated with more severe T cell depletion soon after SIV infection with poor recovery of T cells thereafter, which agrees with human data. The immunologic interactions reported between HIV and M. tuberculosis from various studies include: HIV-induced loss of mycobacterial specific CD4 T cells, M. tuberculosis-induced increases in HIV load in serum and macrophages, shift from Th1 to Th2 response via alterations in IL-10, regulatory T cells, IL-12, IL-4, and TNF, loss of granuloma integrity, and alterations in apoptotic mechanisms (reviewed in Ref. 122).

In summary, the immunology of TB is complex, and understanding latent *M. tuberculosis* infection is even more challenging due to the difficulties in obtaining human data from the

site of infection. Only recently have animal models (nonhuman primates) that adequately reflect human latent infection been described. The more sophisticated analysis of human samples, in conjunction with targeted studies in the nonhuman primate model of latent infection, should provide more answers in the near future regarding the various manifestations and consequences of latent TB. This will lead to improved therapy and perhaps even preventive strategies to reduce reactivation.

## Acknowledgments

We thank the intellectual and scientific contributions of the members of the Gates Grand Challenge 11 program, especially Douglas Young, Clifton Barry, III, and Robert Wilkinson, in developing the idea of the spectrum of latency. We also thank the members of the Flynn and Lin laboratories for helpful discussions and scientific data.

This work was supported by National Institutes of Health Grants HL075845, HL092883, AI50732, AI37859, and HL71241 (to J.L.F), AI063101 (to P.L.L.), the Otis Childs Foundation (to P.L.L.), and the Bill and Melinda Gates Foundation (to J.L.F. and P.L.L.).

#### Abbreviations used in this paper

IGRA	IFN-γ release assay
NOS2	NO synthase 2
ТВ	tuberculosis
TST	tuberculin skin test

## References

- 1. World Health Organization. Geneva, Switzerland: WHO Press; 2009. WHO Report 2009: Global Tuberculosis Control. Epidemiology, Strategy, Financing.
- Jereb J, Etkind SC, Joglar OT, Moore M, Taylor Z. Tuberculosis contact investigations: outcomes in selected areas of the United States, 1999. Int. J. Tuberc. Lung Dis. 2003; 7 Suppl 3(12):S384–S390. [PubMed: 14677827]
- Weis SE, Pogoda JM, Yang Z, Cave MD, Wallace C, Kelley M, Barnes PF. Transmission dynamics of tuberculosis in Tarrant county, Texas. Am. J. Respir. Crit. Care Med. 2002; 166:36–42. [PubMed: 12091168]
- Barnes PF, Yang Z, Preston-Martin S, Pogoda JM, Jones BE, Otaya M, Eisenach KD, Knowles L, Harvey S, Cave MD. Patterns of tuberculosis transmission in Central Los Angeles. JAMA. 1997; 278:1159–1163. [PubMed: 9326475]
- Small PM, Hopewell PC, Singh SP, Paz A, Parsonnet J, Ruston DC, Schecter GF, Daley CL, Schoolnik GK. The epidemiology of tuberculosis in San Francisco. A population-based study using conventional and molecular methods. N. Engl. J. Med. 1994; 330:1703–1709. [PubMed: 7910661]
- Jasmer RM, Hahn JA, Small PM, Daley CL, Behr MA, Moss AR, Creasman JM, Schecter GF, Paz EA, Hopewell PC. A molecular epidemiologic analysis of tuberculosis trends in San Francisco, 1991–1997. Ann. Intern. Med. 1999; 130:971–978. [PubMed: 10383367]
- Verver S, Warren RM, Munch Z, Vynnycky E, van Helden PD, Richardson M, van der Spuy GD, Enarson DA, Borgdorff MW, Behr MA, Beyers N. Transmission of tuberculosis in a high incidence urban community in South Africa. Int. J. Epidemiol. 2004; 33:351–357. [PubMed: 15082639]
- Selwyn PA, Alcabes P, Hartel D, Buono D, Schoenbaum EE, Klein RS, Davenny K, Friedland GH. Clinical manifestations and predictors of disease progression in drug users with human immunodeficiency virus infection. N. Engl. J. Med. 1992; 327:1697–1703. [PubMed: 1359411]
- Borgdorff MW. New measurable indicator for tuberculosis case detection. Emerg. Infect. Dis. 2004; 10:1523–1528. [PubMed: 15498151]
- 10. Corbett EL, Charalambous S, Moloi VM, Fielding K, Grant AD, Dye C, De Cock KM, Hayes RJ, Williams BG, Churchyard GJ. Human immunodeficiency virus and the prevalence of undiagnosed

tuberculosis in African gold miners. Am. J. Respir. Crit. Care Med. 2004; 170:673–679. [PubMed: 15191919]

- Barry CE 3rd, Boshoff HI, Dartois V, Dick T, Ehrt S, Flynn J, Schnappinger D, Wilkinson RJ, Young D. The spectrum of latent tuberculosis: rethinking the biology and intervention strategies. Nat. Rev. Microbiol. 2009; 7:845–855. [PubMed: 19855401]
- Lazarevic V, Nolt D, Flynn JL. Long-term control of *Mycobacterium tuberculosis* infection is mediated by dynamic immune responses. J. Immunol. 2005; 175:1107–1117. [PubMed: 16002712]
- Marks SM, Taylor Z, Qualls NL, Shrestha-Kuwahara RJ, Wilce MA, Nguyen CH. Outcomes of contact investigations of infectious tuberculosis patients. Am. J. Respir. Crit. Care Med. 2000; 162:2033–2038. [PubMed: 11112109]
- Scanga CA, Bafica A, Feng CG, Cheever AW, Hieny S, Sher A. MyD88-deficient mice display a profound loss in resistance to *Mycobacterium tuberculosis* associated with partially impaired Th1 cytokine and nitric oxide synthase 2 expression. Infect. Immun. 2004; 72:2400–2404. [PubMed: 15039368]
- Reiling N, Ehlers S, Hölscher C. MyDths and un-TOLLed truths: sensor, instructive and effector immunity to tuberculosis. Immunol. Lett. 2008; 116:15–23. [PubMed: 18191460]
- Fremond CM, Yeremeev V, Nicolle DM, Jacobs M, Quesniaux VF, Ryffel B. Fatal *Mycobacterium tuberculosis* infection despite adaptive immune response in the absence of MyD88. J. Clin. Invest. 2004; 114:1790–1799. [PubMed: 15599404]
- Mayer-Barber KD, Barber DL, Shenderov K, White SD, Wilson MS, Cheever A, Kugler D, Hieny S, Caspar P, Núñez G, et al. Caspase-1 independent IL-1beta production is critical for host resistance to *mycobacterium tuberculosis* and does not require TLR signaling in vivo. J. Immunol. 2010; 184:3326–3330. [PubMed: 20200276]
- Means TK, Jones BW, Schromm AB, Shurtleff BA, Smith JA, Keane J, Golenbock DT, Vogel SN, Fenton MJ. Differential effects of a Toll-like receptor antagonist on *Mycobacterium tuberculosis*induced macrophage responses. J. Immunol. 2001; 166:4074–4082. [PubMed: 11238656]
- Kamath AB, Alt J, Debbabi H, Behar SM. Toll-like receptor 4- defective C3H/HeJ mice are not more susceptible than other C3H substrains to infection with *Mycobacterium tuberculosis*. Infect. Immun. 2003; 71:4112–4118. [PubMed: 12819102]
- Abel B, Thieblemont N, Quesniaux VJ, Brown N, Mpagi J, Miyake K, Bihl F, Ryffel B. Toll-like receptor 4 expression is required to control chronic *Mycobacterium tuberculosis* infection in mice. J. Immunol. 2002; 169:3155–3162. [PubMed: 12218133]
- Ito T, Schaller M, Hogaboam CM, Standiford TJ, Chensue SW, Kunkel SL. TLR9 activation is a key event for the maintenance of a mycobacterial antigen-elicited pulmonary granulomatous response. Eur. J. Immunol. 2007; 37:2847–2855. [PubMed: 17853411]
- 22. Pompei L, Jang S, Zamlynny B, Ravikumar S, McBride A, Hickman SP, Salgame P. Disparity in IL-12 release in dendritic cells and macrophages in response to *Mycobacterium tuberculosis* is due to use of distinct TLRs. J. Immunol. 2007; 178:5192–5199. [PubMed: 17404302]
- Bafica A, Scanga CA, Feng CG, Leifer C, Cheever A, Sher A. TLR9 regulates Th1 responses and cooperates with TLR2 in mediating optimal resistance to *Mycobacterium tuberculosis*. J. Exp. Med. 2005; 202:1715–1724. [PubMed: 16365150]
- 24. Kleinnijenhuis J, Joosten LA, van de Veerdonk FL, Savage N, van Crevel R, Kullberg BJ, van der Ven A, Ottenhoff TH, Dinarello CA, van der Meer JW, Netea MG. Transcriptional and inflammasome-mediated pathways for the induction of IL-1beta production by *Mycobacterium tuberculosis*. Eur. J. Immunol. 2009; 39:1914–1922. [PubMed: 19544485]
- 25. Hölscher C, Reiling N, Schaible UE, Hölscher A, Bathmann C, Korbel D, Lenz I, Sonntag T, Kröger S, Akira S, et al. Containment of aerogenic *Mycobacterium tuberculosis* infection in mice does not require MyD88 adaptor function for TLR2,-4 and-9. Eur. J. Immunol. 2008; 38:680–694. [PubMed: 18266299]
- Shim TS, Turner OC, Orme IM. Toll-like receptor 4 plays no role in susceptibility of mice to Mycobacterium tuberculosis infection. Tuberculosis (Edinb.). 2003; 83:367–371. [PubMed: 14623167]

- Davila S, Hibberd ML, Hari Dass R, Wong HE, Sahiratmadja E, Bonnard C, Alisjahbana B, Szeszko JS, Balabanova Y, Drobniewski F, et al. Genetic association and expression studies indicate a role of toll-like receptor 8 in pulmonary tuberculosis. PLoS Genet. 2008; 4:e1000218. [PubMed: 18927625]
- Velez DR, Hulme WF, Myers JL, Weinberg JB, Levesque MC, Stryjewski ME, Abbate E, Estevan R, Patillo SG, Gilbert JR, et al. NOS2A, TLR4, and IFNGR1 interactions influence pulmonary tuberculosis susceptibility in African-Americans. Hum. Genet. 2009; 126:643–653. [PubMed: 19575238]
- Velez DR, Wejse C, Stryjewski ME, Abbate E, Hulme WF, Myers JL, Estevan R, Patillo SG, Olesen R, Tacconelli A, et al. Variants in toll-like receptors 2 and 9 influence susceptibility to pulmonary tuberculosis in Caucasians, African-Americans, West Africans. Hum. Genet. 2010; 127:65–73. [PubMed: 19771452]
- Liu PT, Stenger S, Li H, Wenzel L, Tan BH, Krutzik SR, Ochoa MT, Schauber J, Wu K, Meinken C, et al. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. Science. 2006; 311:1770–1773. [PubMed: 16497887]
- 31. Liu PT, Schenk M, Walker VP, Dempsey PW, Kanchanapoomi M, Wheelwright M, Vazirnia A, Zhang X, Steinmeyer A, Zügel U, et al. Convergence of IL-1beta and VDR activation pathways in human TLR2/1- induced antimicrobial responses. PLoS ONE. 2009; 4:e5810. [PubMed: 19503839]
- 32. Sharpe SA, Eschelbach E, Basaraba RJ, Gleeson F, Hall GA, McIntyre A, Williams A, Kraft SL, Clark S, Gooch K, et al. Determination of lesion volume by MRI and stereology in a macaque model of tuberculosis. Tuberculosis (Edinb.). 2009; 89:405–416. [PubMed: 19879805]
- 33. Langermans JA, Andersen P, van Soolingen D, Vervenne RA, Frost PA, van der Laan T, van Pinxteren LA, van den Hombergh J, Kroon S, Peekel I, et al. Divergent effect of bacillus Calmette-Guérin (BCG) vaccination on *Mycobacterium tuberculosis* infection in highly related macaque species: implications for primate models in tuberculosis vaccine research. Proc. Natl. Acad. Sci. USA. 2001; 98:11497–11502. [PubMed: 11562492]
- 34. Lin PL, Rodgers M, Smith L, Bigbee M, Myers A, Bigbee C, Chiosea I, Capuano SV, Fuhrman C, Klein E, Flynn JL. Quantitative comparison of active and latent tuberculosis in the cynomolgus macaque model. Infect. Immun. 2009; 77:4631–4642. [PubMed: 19620341]
- Lin PL, Pawar S, Myers A, Pegu A, Fuhrman C, Reinhart TA, Capuano SV, Klein E, Flynn JL. Early events in *Mycobacterium tuberculosis* infection in cynomolgus macaques. Infect. Immun. 2006; 74:3790–3803. [PubMed: 16790751]
- Doherty TM, Demissie A, Olobo J, Wolday D, Britton S, Eguale T, Ravn P, Andersen P. Immune responses to the *Mycobacterium tuberculosis*-specific antigen ESAT-6 signal subclinical infection among contacts of tuberculosis patients. J. Clin. Microbiol. 2002; 40:704–706. [PubMed: 11826002]
- Flynn JL, Chan J. Immunology of tuberculosis. Annu. Rev. Immunol. 2001; 19:93–129. [PubMed: 11244032]
- Lawn SD, Myer L, Edwards D, Bekker LG, Wood R. Short-term and long-term risk of tuberculosis associated with CD4 cell recovery during antiretroviral therapy in South Africa. AIDS. 2009; 23:1717–1725. [PubMed: 19461502]
- Diedrich CR, Mattila JT, Klein E, Janssen C, Phuah J, Sturgeon TJ, Montelaro RC, Lin PL, Flynn JL. Reactivation of latent tuberculosis in cynomolgus macaques infected with SIV is associated with early peripheral T cell depletion and not virus load. PLoS ONE. 2010; 5:e9611. [PubMed: 20224771]
- Jones BE, Young SM, Antoniskis D, Davidson PT, Kramer F, Barnes PF. Relationship of the manifestations of tuberculosis to CD4 cell counts in patients with human immunodeficiency virus infection. Am. Rev. Respir. Dis. 1993; 148:1292–1297. [PubMed: 7902049]
- 41. Lazarevic V, Flynn J. CD8+ T cells in tuberculosis. Am. J. Respir. Crit. Care Med. 2002; 166:1116–1121. [PubMed: 12379557]
- van Pinxteren LAH, Cassidy JP, Smedegaard BHC, Agger EM, Andersen P. Control of latent *Mycobacterium tuberculosis* infection is dependent on CD8 T cells. Eur. J. Immunol. 2000; 30:3689–3698. [PubMed: 11169412]

- Serbina NV, Flynn JL. Early emergence of CD8(+) T cells primed for production of type 1 cytokines in the lungs of *Mycobacterium tuberculosis*-infected mice. Infect. Immun. 1999; 67:3980–3988. [PubMed: 10417164]
- Woodworth JS, Wu Y, Behar SM. *Mycobacterium tuberculosis*-specific CD8+ T cells require perforin to kill target cells and provide protection in vivo. J. Immunol. 2008; 181:8595–8603. [PubMed: 19050279]
- 45. Stenger S, Hanson DA, Teitelbaum R, Dewan P, Niazi KR, Froelich CJ, Ganz T, Thoma-Uszynski S, Melián A, Bogdan C, et al. An antimicrobial activity of cytolytic T cells mediated by granulysin. Science. 1998; 282:121–125. [PubMed: 9756476]
- 46. Grotzke JE, Lewinsohn DM. Role of CD8+ T lymphocytes in control of *Mycobacterium tuberculosis* infection. Microbes Infect. 2005; 7:776–788. [PubMed: 15823514]
- 47. Chen CY, Huang D, Wang RC, Shen L, Zeng G, Yao S, Shen Y, Halliday L, Fortman J, McAllister M, et al. A critical role for CD8 T cells in a nonhuman primate model of tuberculosis. PLoS Pathog. 2009; 5:e1000392. [PubMed: 19381260]
- Chan J, Tanaka K, Carroll D, Flynn J, Bloom BR. Effects of nitric oxide synthase inhibitors on murine infection with *Mycobacterium tuberculosis*. Infect. Immun. 1995; 63:736–740. [PubMed: 7529749]
- MacMicking JD, North RJ, LaCourse R, Mudgett JS, Shah SK, Nathan CF. Identification of nitric oxide synthase as a protective locus against tuberculosis. Proc. Natl. Acad. Sci. USA. 1997; 94:5243–5248. [PubMed: 9144222]
- 50. Flynn JL, Scanga CA, Tanaka KE, Chan J. Effects of amino-guanidine on latent murine tuberculosis. J. Immunol. 1998; 160:1796–1803. [PubMed: 9469439]
- Choi HS, Rai PR, Chu HW, Cool C, Chan ED. Analysis of nitric oxide synthase and nitrotyrosine expression in human pulmonary tuberculosis. Am. J. Respir. Crit. Care Med. 2002; 166:178–186. [PubMed: 12119230]
- Nathan C. Role of iNOS in human host defense. Science. 2006; 312:1874–1875. author reply 1874–1875. [PubMed: 16809512]
- Cooper AM, Magram J, Ferrante J, Orme IM. Interleukin 12 (IL-12) is crucial to the development of protective immunity in mice intravenously infected with *mycobacterium tuberculosis*. J. Exp. Med. 1997; 186:39–45. [PubMed: 9206995]
- Ottenhoff TH, Verreck FA, Lichtenauer-Kaligis EG, Hoeve MA, Sanal O, van Dissel JT. Genetics, cytokines and human infectious disease: lessons from weakly pathogenic mycobacteria and salmonellae. Nat. Genet. 2002; 32:97–105. [PubMed: 12205477]
- 55. Sada-Ovalle I, Chiba A, Gonzales A, Brenner MB, Behar SM. Innate invariant NKT cells recognize *Mycobacterium tuberculosis*-infected macrophages, produce interferon-gamma, kill intracellular bacteria. PLoS Pathog. 2008; 4:e1000239. [PubMed: 19079582]
- 56. Singh SB, Davis AS, Taylor GA, Deretic V. Human IRGM induces autophagy to eliminate intracellular mycobacteria. Science. 2006; 313:1438–1441. [PubMed: 16888103]
- Casanova JL, Abel L. Genetic dissection of immunity to mycobacteria: the human model. Annu. Rev. Immunol. 2002; 20:581–620. [PubMed: 11861613]
- 58. Flynn JL, Goldstein MM, Chan J, Triebold KJ, Pfeffer K, Lowenstein CJ, Schreiber R, Mak TW, Bloom BR. Tumor necrosis factor-alpha is required in the protective immune response against *Mycobacterium tuberculosis* in mice. Immunity. 1995; 2:561–572. [PubMed: 7540941]
- Mohan VP, Scanga CA, Yu K, Scott HM, Tanaka KE, Tsang E, Tsai MM, Flynn JL, Chan J. Effects of tumor necrosis factor alpha on host immune response in chronic persistent tuberculosis: possible role for limiting pathology. Infect. Immun. 2001; 69:1847–1855. [PubMed: 11179363]
- Algood HM, Lin PL, Flynn JL. Tumor necrosis factor and chemokine interactions in the formation and maintenance of granulomas in tuberculosis. Clin. Infect. Dis. 2005; 41 Suppl 3:S189–S193. [PubMed: 15983898]
- Clay H, Volkman HE, Ramakrishnan L. Tumor necrosis factor signaling mediates resistance to mycobacteria by inhibiting bacterial growth and macrophage death. Immunity. 2008; 29:283–294. [PubMed: 18691913]
- 62. Lin PL, Myers A, Smith L, Bigbee C, Bigbee M, Fuhrman C, Grieser H, Chiosea I, Voitenek NN, Capuano SV, et al. Tumor necrosis factor neutralization results in disseminated disease in acute

and latent *Mycobacterium tuberculosis* infection with normal granuloma structure in a cynomolgus macaque model. Arthritis Rheum. 2010; 62:340–350. [PubMed: 20112395]

- 63. Möller M, Flachsbart F, Till A, Thye T, Horstmann RD, Meyer CG, Osei I, van Helden PD, Hoal EG, Schreiber S, et al. A functional haplotype in the 3' untranslated region of TNFRSF1B is associated with tuberculosis in two African populations. Am. J. Respir. Crit. Care Med. 2010; 181:388–393. [PubMed: 20007930]
- Keane J. Tumor necrosis factor blockers and reactivation of latent tuberculosis. Clin. Infect. Dis. 2004; 39:300–302. [PubMed: 15306994]
- Windish HP, Lin PL, Mattila JT, Green AM, Onuoha EO, Kane LP, Flynn JL. Aberrant TGF-beta signaling reduces T regulatory cells in ICAM-1-deficient mice, increasing the inflammatory response to *Mycobacterium tuberculosis*. J. Leukoc. Biol. 2009; 86:713–725. [PubMed: 19454651]
- 66. Scott HM, Flynn JL. *Mycobacterium tuberculosis* in chemokine receptor 2-deficient mice: influence of dose on disease progression. Infect. Immun. 2002; 70:5946–5954. [PubMed: 12379669]
- 67. Algood HM, Lin PL, Yankura D, Jones A, Chan J, Flynn JL. TNF influences chemokine expression of macrophages in vitro and that of CD11b+ cells in vivo during *Mycobacterium tuberculosis* infection. J. Immunol. 2004; 172:6846–6857. [PubMed: 15153503]
- Peters W, Scott HM, Chambers HF, Flynn JL, Charo IF, Ernst JD. Chemokine receptor 2 serves an early and essential role in resistance to *Mycobacterium tuberculosis*. Proc. Natl. Acad. Sci. USA. 2001; 98:7958–7963. [PubMed: 11438742]
- Keane J, Balcewicz-Sablinska MK, Remold HG, Chupp GL, Meek BB, Fenton MJ, Kornfeld H. Infection by *Mycobacterium tuberculosis* promotes human alveolar macrophage apoptosis. Infect. Immun. 1997; 65:298–304. [PubMed: 8975927]
- Hinchey J, Lee S, Jeon BY, Basaraba RJ, Venkataswamy MM, Chen B, Chan J, Braunstein M, Orme IM, Derrick SC, et al. Enhanced priming of adaptive immunity by a proapoptotic mutant of *Mycobacterium tuberculosis*. J. Clin. Invest. 2007; 117:2279–2288. [PubMed: 17671656]
- Latorre I, De Souza-Galvão M, Ruiz-Manzano J, Lacoma A, Prat C, Fuenzalida L, Altet N, Ausina V, Domínguez J. Quantitative evaluation of T-cell response after specific antigen stimulation in active and latent tuberculosis infection in adults and children. Diagn. Microbiol. Infect. Dis. 2009; 65:236–246. [PubMed: 19822269]
- Chee CB, Barkham TM, Khinmar KW, Gan SH, Wang YT. Quantitative T-cell interferon-gamma responses to *Mycobacterium tuberculosis*-specific antigens in active and latent tuberculosis. Eur. J. Clin. Microbiol. Infect. Dis. 2009; 28:667–670. [PubMed: 19020909]
- Janssens JP, Roux-Lombard P, Perneger T, Metzger M, Vivien R, Rochat T. Quantitative scoring of an interferon-gamma assay for differentiating active from latent tuberculosis. Eur. Respir. J. 2007; 30:722–728. [PubMed: 17537773]
- 74. Arend SM, Thijsen SF, Leyten EM, Bouwman JJ, Franken WP, Koster BF, Cobelens FG, van Houte AJ, Bossink AW. Comparison of two interferon-gamma assays and tuberculin skin test for tracing tuberculosis contacts. Am. J. Respir. Crit. Care Med. 2007; 175:618–627. [PubMed: 17170386]
- 75. Goletti D, Vincenti D, Carrara S, Butera O, Bizzoni F, Bernardini G, Amicosante M, Girardi E. Selected RD1 peptides for active tuberculosis diagnosis: comparison of a gamma interferon wholeblood enzyme-linked immunosorbent assay and an enzyme-linked immunospot assay. Clin. Diagn. Lab. Immunol. 2005; 12:1311–1316. [PubMed: 16275946]
- Davidow AL. Interferon-gamma release assay test characteristics depend upon the prevalence of active tuberculosis. Int. J. Tuberc. Lung Dis. 2009; 13:1411–1415. [PubMed: 19861015]
- 77. del Corral H, París SC, Marín ND, Marín DM, López L, Henao HM, Martínez T, Villa L, Barrera LF, Ortiz BL, et al. IFNgamma response to *Mycobacterium tuberculosis*, risk of infection and disease in household contacts of tuberculosis patients in Colombia. PLoS ONE. 2009; 4:e8257. [PubMed: 20011589]
- 78. Abebe M, Doherty TM, Wassie L, Aseffa A, Bobosha K, Demissie A, Zewdie M, Engers H, Andersen P, Kim L, et al. Expression of apoptosis-related genes in an Ethiopian cohort study

correlates with tuberculosis clinical status. Eur. J. Immunol. 2010; 40:291–301. [PubMed: 19877018]

- 79. Tsao TC, Hong J, Li LF, Hsieh MJ, Liao SK, Chang KS. Imbalances between tumor necrosis factor-alpha and its soluble receptor forms, interleukin-1beta and interleukin-1 receptor antagonist in BAL fluid of cavitary pulmonary tuberculosis. Chest. 2000; 117:103–109. [PubMed: 10631206]
- Boherty TM, Wallis RS, Zumla A. Biomarkers of disease activity, cure, and relapse in tuberculosis. Clin. Chest Med. 2009; 30:783–796. [PubMed: 19925967]
- Scott-Browne JP, Shafiani S, Tucker-Heard G, Ishida-Tsubota K, Fontenot JD, Rudensky AY, Bevan MJ, Urdahl KB. Expansion and function of Foxp3-expressing T regulatory cells during tuberculosis. J. Exp. Med. 2007; 204:2159–2169. [PubMed: 17709423]
- 82. Chen X, Zhou B, Li M, Deng Q, Wu X, Le X, Wu C, Larmonier N, Zhang W, Zhang H, et al. CD4(+)CD25(+)FoxP3(+) regulatory T cells suppress *Mycobacterium tuberculosis* immunity in patients with active disease. Clin. Immunol. 2007; 123:50–59. [PubMed: 17234458]
- Guyot-Revol V, Innes JA, Hackforth S, Hinks T, Lalvani A. Regulatory T cells are expanded in blood and disease sites in patients with tuberculosis. Am. J. Respir. Crit. Care Med. 2006; 173:803–810. [PubMed: 16339919]
- 84. Burl S, Hill PC, Jeffries DJ, Holland MJ, Fox A, Lugos MD, Adegbola RA, Rook GA, Zumla A, McAdam KP, Brookes RH. FOXP3 gene expression in a tuberculosis case contact study. Clin. Exp. Immunol. 2007; 149:117–122. [PubMed: 17465993]
- Green A, Mattila JT, Bigbee CL, Bongers KS, Lin PL, Flynn JL. CD4 regulatory T cells in a cynomolgus macaque model of Mycobacterium tuberculosis infection. J. Infect. Dis. 2010 In press.
- 86. Jurado JO, Alvarez IB, Pasquinelli V, Martínez GJ, Quiroga MF, Abbate E, Musella RM, Chuluyan HE, García VE. Programmed death (PD)-1:PD-ligand 1/PD-ligand 2 pathway inhibits T cell effector functions during human tuberculosis. J. Immunol. 2008; 181:116–125. [PubMed: 18566376]
- Einarsdottir T, Lockhart E, Flynn JL. Cytotoxicity and secretion of gamma interferon are carried out by distinct CD8 T cells during *Mycobacterium tuberculosis* infection. Infect. Immun. 2009; 77:4621–4630. [PubMed: 19667047]
- Caccamo N, Guggino G, Meraviglia S, Gelsomino G, Di Carlo P, Titone L, Bocchino M, Galati D, Matarese A, Nouta J, et al. Analysis of *Mycobacterium tuberculosis*-specific CD8 T-cells in patients with active tuberculosis and in individuals with latent infection. PLoS ONE. 2009; 4:e5528. [PubMed: 19436760]
- Darrah PA, Patel DT, De Luca PM, Lindsay RW, Davey DF, Flynn BJ, Hoff ST, Andersen P, Reed SG, Morris SL, et al. Multifunctional TH1 cells define a correlate of vaccine-mediated protection against *Leishmania major*. Nat. Med. 2007; 13:843–850. [PubMed: 17558415]
- 90. Sutherland JS, Adetifa IM, Hill PC, Adegbola RA, Ota MO. Pattern and diversity of cytokine production differentiates between *Mycobacterium tuberculosis* infection and disease. Eur. J. Immunol. 2009; 39:723–729. [PubMed: 19224636]
- Mueller H, Detjen AK, Schuck SD, Gutschmidt A, Wahn U, Magdorf K, Kaufmann SH, Jacobsen M. *Mycobacterium tuberculosis*-specific CD4+, IFNgamma+, and TNFalpha+ multifunctional memory T cells coexpress GM-CSF. Cytokine. 2008; 43:143–148. [PubMed: 18603443]
- 92. Khader SA, Bell GK, Pearl JE, Fountain JJ, Rangel-Moreno J, Cilley GE, Shen F, Eaton SM, Gaffen SL, Swain SL, et al. IL-23 and IL-17 in the establishment of protective pulmonary CD4+ T cell responses after vaccination and during *Mycobacterium tuberculosis* challenge. Nat. Immunol. 2007; 8:369–377. [PubMed: 17351619]
- Lockhart E, Green AM, Flynn JL. IL-17 production is dominated by gammadelta T cells rather than CD4 T cells during *Mycobacterium tuberculosis* infection. J. Immunol. 2006; 177:4662– 4669. [PubMed: 16982905]
- 94. Chen X, Zhang M, Liao M, Graner MW, Wu C, Yang Q, Liu H, Zhou B. Reduced Th17 response in patients with tuberculosis correlates with IL-6R expression on CD4+ T Cells. Am. J. Respir. Crit. Care Med. 2010; 181:734–742. [PubMed: 20019339]
- 95. Meintjes G, Lawn SD, Scano F, Maartens G, French MA, Worodria W, Elliott JH, Murdoch D, Wilkinson RJ, Seyler C, et al. International Network for the Study of HIV-associated IRIS.

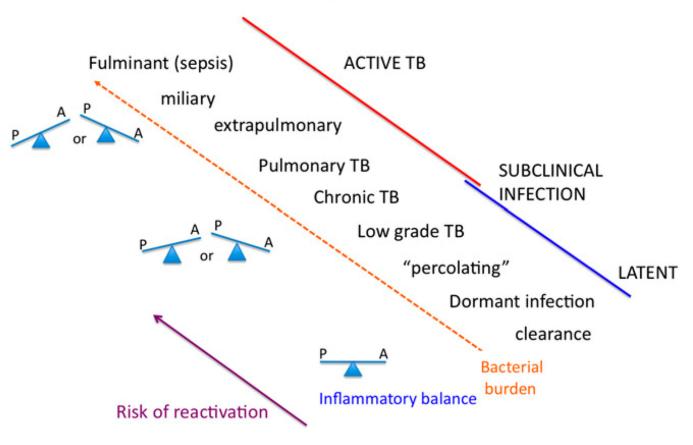
Tuberculosis-associated immune reconstitution inflammatory syndrome: case definitions for use in resource-limited settings. Lancet Infect. Dis. 2008; 8:516–523. [PubMed: 18652998]

- 96. Lawn SD, Wilkinson RJ, Lipman MC, Wood R. Immune reconstitution and "unmasking" of tuberculosis during antiretroviral therapy. Am. J. Respir. Crit. Care Med. 2008; 177:680–685. [PubMed: 18202347]
- 97. Gagneux S, Small PM. Global phylogeography of *Mycobacterium tuberculosis* and implications for tuberculosis product development. Lancet Infect. Dis. 2007; 7:328–337. [PubMed: 17448936]
- Parwati I, van Crevel R, van Soolingen D. Possible underlying mechanisms for successful emergence of the *Mycobacterium tuberculosis* Beijing genotype strains. Lancet Infect. Dis. 2010; 10:103–111. [PubMed: 20113979]
- Palanisamy GS, DuTeau N, Eisenach KD, Cave DM, Theus SA, Kreiswirth BN, Basaraba RJ, Orme IM. Clinical strains of *Mycobacterium tuberculosis* display a wide range of virulence in guinea pigs. Tuberculosis (Edinb.). 2009; 89:203–209. [PubMed: 19251482]
- 100. de Jong BC, Hill PC, Aiken A, Awine T, Antonio M, Adetifa IM, Jackson-Sillah DJ, Fox A, Deriemer K, Gagneux S, et al. Progression to active tuberculosis, but not transmission, varies by *Mycobacterium tuberculosis* lineage in The Gambia. J. Infect. Dis. 2008; 198:1037–1043. [PubMed: 18702608]
- 101. Caws M, Thwaites G, Dunstan S, Hawn TR, Lan NT, Thuong NT, Stepniewska K, Huyen MN, Bang ND, Loc TH, et al. The influence of host and bacterial genotype on the development of disseminated disease with *Mycobacterium tuberculosis*. PLoS Pathog. 2008; 4:e1000034. [PubMed: 18369480]
- 102. Hawn TR, Dunstan SJ, Thwaites GE, Simmons CP, Thuong NT, Lan NT, Quy HT, Chau TT, Hieu NT, Rodrigues S, et al. A polymorphism in Toll-interleukin 1 receptor domain containing adaptor protein is associated with susceptibility to meningeal tuberculosis. J. Infect. Dis. 2006; 194:1127–1134. [PubMed: 16991088]
- 103. Pepper T, Joseph P, Mwenya C, McKee GS, Haushalter A, Carter A, Warkentin J, Haas DW, Sterling TR. Normal chest radiography in pulmonary tuberculosis: implications for obtaining respiratory specimen cultures. Int. J. Tuberc. Lung Dis. 2008; 12:397–403. [PubMed: 18371265]
- 104. Corbett EL, Bandason T, Cheung YB, Munyati S, Godfrey-Faussett P, Hayes R, Churchyard G, Butterworth A, Mason P. Epidemiology of tuberculosis in a high HIV prevalence population provided with enhanced diagnosis of symptomatic disease. PLoS Med. 2007; 4:e22. [PubMed: 17199408]
- 105. Mtei L, Matee M, Herfort O, Bakari M, Horsburgh CR, Waddell R, Cole BF, Vuola JM, Tvaroha S, Kreiswirth B, et al. High rates of clinical and subclinical tuberculosis among HIV-infected ambulatory subjects in Tanzania. Clin. Infect. Dis. 2005; 40:1500–1507. [PubMed: 15844073]
- 106. Amukotuwa S, Choong PF, Smith PJ, Powell GJ, Slavin J, Schlicht SM. Tuberculosis masquerading as malignancy: a multimodality approach to the correct diagnosis - a case report. Int. Semin. Surg. Oncol. 2005; 2:10. [PubMed: 15877821]
- 107. Hofmeyr A, Lau WF, Slavin MA. *Mycobacterium tuberculosis* infection in patients with cancer, the role of 18-fluorodeoxyglucose positron emission tomography for diagnosis and monitoring treatment response. Tuberculosis (Edinb.). 2007; 87:459–463. [PubMed: 17631049]
- 108. American Thoracic Society. Targeted tuberculin testing and treatment of latent tuberculosis infection. This official statement of the American Thoracic Society was adopted by the ATS Board of Directors, July 1999. This is a Joint Statement of the American Thoracic Society (ATS) and the Centers for Disease Control and Prevention (CDC). This statement was endorsed by the Council of the Infectious Diseases Society of America. (IDSA), September 1999, the sections of this statement. Am. J. Respir. Crit. Care Med. 2000; 161:S221–S247. [PubMed: 10764341]
- 109. Opie EAJ. Tubercle bacilli in latent tuberculous lesions and in lung tissue wihtout tuberculous lesions. Arch. Pathol. Lab. Med. 1927; 4:1–21.
- 110. Lillebaek T, Dirksen A, Baess I, Strunge B, Thomsen VO, Andersen AB. Molecular evidence of endogenous reactivation of *Mycobacterium tuberculosis* after 33 years of latent infection. J. Infect. Dis. 2002; 185:401–404. [PubMed: 11807725]

Lin and Flynn

- 111. Lillebaek T, Dirksen A, Vynnycky E, Baess I, Thomsen VO, Andersen AB. Stability of DNA patterns and evidence of *Mycobacterium tuberculosis* reactivation occurring decades after the initial infection. J. Infect. Dis. 2003; 188:1032–1039. [PubMed: 14513424]
- 112. Lönnroth K, Raviglione M. Global epidemiology of tuberculosis: prospects for control. Semin. Respir. Crit. Care Med. 2008; 29:481–491. [PubMed: 18810682]
- Horsburgh CR Jr. Priorities for the treatment of latent tuberculosis infection in the United States. N. Engl. J. Med. 2004; 350:2060–2067. [PubMed: 15141044]
- 114. Jick SS, Lieberman ES, Rahman MU, Choi HK. Glucocorticoid use, other associated factors, and the risk of tuberculosis. Arthritis Rheum. 2006; 55:19–26. [PubMed: 16463407]
- Wallis RS. Infectious complications of tumor necrosis factor blockade. Curr. Opin. Infect. Dis. 2009; 22:403–409. [PubMed: 19491672]
- 116. Bruns H, Meinken C, Schauenberg P, Härter G, Kern P, Modlin RL, Antoni C, Stenger S. Anti-TNF immunotherapy reduces CD8+ T cell-mediated antimicrobial activity against *Mycobacterium tuberculosis* in humans. J. Clin. Invest. 2009; 119:1167–1177. [PubMed: 19381021]
- 117. Hamdi H, Mariette X, Godot V, Weldingh K, Hamid AM, Prejean MV, Baron G, Lemann M, Puechal X, Breban M, et al. RATIO (Recherche sur Anti-TNF et Infections Opportunistes) Study Group. Inhibition of anti-tuberculosis T-lymphocyte function with tumour necrosis factor antagonists. Arthritis Res. Ther. 2006; 8:R114. [PubMed: 16859506]
- 118. Saliu OY, Sofer C, Stein DS, Schwander SK, Wallis RS. Tumor-necrosis-factor blockers: differential effects on mycobacterial immunity. J. Infect. Dis. 2006; 194:486–492. [PubMed: 16845632]
- Farer LS, Lowell AM, Meador MP. Extrapulmonary tuberculosis in the United States. Am. J. Epidemiol. 1979; 109:205–217. [PubMed: 425959]
- 120. Lawn SD, Bekker LG, Wood R. How effectively does HAART restore immune responses to *Mycobacterium tuberculosis*? Implications for tuberculosis control. AIDS. 2005; 19:1113–1124. [PubMed: 15990564]
- 121. Badri M, Wilson D, Wood R. Effect of highly active antiretroviral therapy on incidence of tuberculosis in South Africa: a cohort study. Lancet. 2002; 359:2059–2064. [PubMed: 12086758]
- 122. Djoba Siawaya JF, Ruhwald M, Eugen-Olsen J, Walzl G. Correlates for disease progression and prognosis during concurrent HIV/TB infection. Int. J. Infect. Dis. 2007; 11:289–299. [PubMed: 17446108]

# SPECTRUM of M. TUBERCULOSIS INFECTION OUTCOMES



#### FIGURE 1.

The spectrum of *M. tuberculosis* infection outcome is depicted. The clinical outcomes of active (red line) and latent (blue line) infection are subdivided to reflect the variability of infection in those categories. Bacterial burden, shown by the dashed orange line, is expected to increase up the spectrum of infection. The seesaws reflect the balance of pro- (P) and anti-inflammatory (A) factors in the granuloma. At the lower end of the latency spectrum, these two factors are well balanced, controlling bacterial growth with minimal pathology. As one advances up the spectrum, the balance can shift with either too much proinflammatory or too much anti-inflammatory activity, which can lead to poor control of bacteria and increased pathology. The purple line reflects the risk of reactivation in the latent spectrum.