

A new vector and RNase H method for the subtractive hybridization

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Subtractive hybridization is an attractive method to clone cDNAs but requires a large amount of mRNA. In addition, non-hybridized cDNA is not always completely separated from hybridized cDNAs by a hydroxyapatite column. To overcome these limitations, we explored a new vector (T7 λ gt10-1) and procedure (RNase H method) for preparing, subtracted cRNA probes of a given cDNA library. A synthetic 57mer fragment, 5'-AATTGTAATACGACTCACTATAGGGTCTAGACCCCGAATTCGCGGCCGCGAGCGGCCA-3', was inserted into the *Eco* RI site of λ gt10 to make a vector that has the T7 promoter containing *Sfi* I, *Not* I, *Eco* RI and *Xba* I sites (Fig.1). cDNA inserts were synthesized according to Gubler-Hoffman (1) using GCGGCCG(T)₁₂ as a primer. After ligation with *Eco* RI linker, cDNAs were digested with

Fig.1



Fig.2

Probes DNA on filters	Subtracted		Mixture	
	A	B	A	B
1 ng				
0.1 ng				
0.01 ng				

Eco RI and *Not* I, and inserted into *Eco* RI-*Not* I site of T7 λ gt10-1. From this cDNA library, we could produce transcripts equivalent to mRNA by T7 RNA polymerase without any plasmid sequence. To separate non-hybridized RNA from the hybrid, we introduced the RNase H digestion method. A model experiment is described below. We have prepared two RNA probes by transcribing human IgC γ and V H par gene plasmids (templates A and B respectively) with T7 polymerase(2). A mixture of probes A and B (50ng each) was hybridized with 5 μ g of denatured template DNA B at 65 °C for 20 hrs in 200 μ l of 0.4 M NaCl-1mM EDTA-40mM PIPES(pH6.7). Hybridized RNA was digested with 65 units of RNase H(Takara) in 300 μ l of 40mM Tris-Cl(pH7.5)-4mM MgCl₂-1mM DTT at 30°C for 40 min. After removal of digested oligonucleotides by ethanol precipitation, probes and a template were denatured and the same

process was repeated once again. The template DNA in the subtracted probe was digested with RNase-free DNase(Promega). This subtracted probe and the original mixture of probes for A and B were hybridized with various amounts of template DNAs on nitrocellulose filters. The subtracted probe hybridized to template B 1/100 as strongly as the original mixture probe whereas both probes hybridized to template A with the same intensity (Fig.2). The result indicates that the subtracted RNA probes prepared as above were enriched at least 100 folds.

Reference

- (1) Gubler, U. and Hoffman, B.J. (1983) *Gene*:25, 263.
- (2) Melton, D.A. *et al.* (1984) *Nucleic Acids Res.*:12, 7035.