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Females of an African Cichlid Fish Display Male-Typical Social Dominance Behavior and Elevated Androgens in the Absence of Males

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Abstract

Social environment can affect the expression of sex-typical behavior in both males and females. Males of the African cichlid species *Astatotilapia burtoni* have long served as a model system to study the neural, endocrine, and molecular basis of socially plastic dominance behavior. Here we show that in all-female communities of *A. burtoni*, some individuals acquire a male-typical dominance phenotype, including aggressive territorial defense, distinctive color patterns, and courtship behavior. Furthermore, dominant females have higher levels of circulating androgens than either subordinate females or females in mixed-sex communities. These male-typical traits do not involve sex change, nor do the social phenotypes in all-female communities differ in relative ovarian size, suggesting that factors other than gonadal physiology underlie much of the observed variation. In contrast to the well-studied situation in males, dominant and subordinate females do not differ in the rate of somatic growth. Dominant females are not any more likely than subordinates to spawn with an introduced male, although they do so sooner. These results extend the well known extraordinary behavioral plasticity of *A. burtoni* to the females of this species and provide a foundation for uncovering the neural and molecular basis of social dominance behavior while controlling for factors such as sex, gonadal state and growth.

Keywords

Aggression; social dominance; sex steroid hormones; reproduction; growth

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Authors Contributions

H.A.H, S.C.P.R. conceived the experiment. S.C.P.R. and E.J.F. conducted the behavioral observations. B.C.T. conducted the hormone assays. E.J.F., S.C.P.R, N.A.-H. and H.A.H. wrote the paper. All authors have read and approved the manuscript.

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Introduction

Aggressiveness varies among species, populations, environments, life stages, sexes, and individuals. Understanding the proximate causes of variation in aggressiveness is necessary in order to understand the evolution of social behavior and to explain mechanisms causing the observed trade-offs between aggressiveness and other fitness-linked traits (Knapp et al., 1999; McGlothlin and Ketterson, 2008). However, important physiological or experimental limitations often confound mechanistic studies of aggressiveness. It is often impossible to know to what extent molecular and mechanistic differences between individuals displaying varying levels of aggressiveness are directly related to the observed behavioral phenotype. While not independent of the behavior, these associated physiological changes complicate efforts to identify the mechanisms underlying variation in aggressiveness. Studying aggressiveness in alternate contexts might allow us to control some of these potentially confounding factors.

Social fish species have emerged as important model systems for the study of aggressive behavior and its underlying physiological mechanisms (Arnott and Elwood, 2009). The African cichlid fish *Astatotilapia burtoni* has become a particularly powerful model system in social neuroscience (for review: (Fernald, 2004; Robinson et al., 2008; Wong and Hofmann, 2010), while at the same time its ecology and behavior have been well characterised in nature (Fernald and Hirata, 1977a, b). We aim to use social hierarchies within experimentally manipulated female communities to experimentally dissociate the physiological and molecular mechanisms underlying social dominance from some of the potentially confounding variables, such as sex, gonadal state, and growth rate.

At any given time, males of *A. burtoni* assume one of two social states (Hofmann, 2003). Dominant males are colourful and display a characteristic melanocyte-based lachrymal stripe across their face (so-called eye-bar). They aggressively maintain territories from which they court females. In addition to actual physical encounters, dominant males exhibit ritualized displays of aggression directed at subordinate males and other dominant males. The dominant males have elevated levels of testosterone and 11-KT, and they are reproductively active, producing sperm and spawning with females to fertilize eggs. Subordinate males do not hold a territory, look and act like females, have low androgen levels, and are reproductively inactive with regressed gonads. Individuals shift between dominant and subordinate states throughout life, in part due to differential growth rates (Hofmann, 2000; Hofmann et al., 1999; Parikh et al., 2006). Importantly, these transitions can be experimentally induced in order to gain insights into the hormonal and molecular mechanisms underlying these endpoints (Renn et al., 2008) and the transition process (Burmeister, 2005; Maruska and Fernald, 2010).

Female *A. burtoni* behave much like subordinate males under normal conditions in the field and laboratory, i.e., they school and feed with other females, juveniles, and subordinate males (Fernald, 1977a; Fernald and Hirata, 1977a, b) (White and Fernald, 1993). Once gravid (full with mature eggs), a female will spawn with dominant males, after which she incubates the fertilized eggs in her buccal cavity for several weeks. Upon release of the fry a female may defend a territory and continue to exhibit maternal care for a short period (Renn et al., 2009; Fernald and Hirata, 1977a). Aggressiveness in females provides an alternate paradigm in which to address the mechanistic basis of such behavior.

In the present study, we show that when all males are removed from the social group, some females acquire morphological and behavioral traits typical of the dominant territorial male phenotype, exhibiting a change in their sex-role without undergoing sex change. These aggressive females acquire dark eye-bars, defend territories, with vigorous chasing, threat

displays and border threats, and even court and spawn with other females. At the physiological level, these behavioral changes are accompanied by the increase in circulating levels of androgens. Our results establish a powerful experimental system for uncovering the molecular basis of specific behavioral patterns. While not thought to mimic ecologically relevant conditions, the experimental manipulations, behavioral observations and physiological measures presented here establish a foundation for the analysis of female behavior in this important model of social behavior. Furthermore, the ability to study dominance behavior in this alternate context (i.e. in females) presents the opportunity to remove, or reduce the influence of, confounding factors (e.g. gonadal state, somatic growth) that have complicated efforts to specifically relate gene expression to behavior in past mechanistic studies of male behavior (e.g. Renn et al., 2008).

Methods

Housing of Fish

Fish derived from a laboratory stock were kept in 110 L aquaria at 28°C and pH 8.5 under full spectrum 12hr light/12 hr dark with 10 minute dim light periods to simulate dawn and dusk, mimicking the conditions found in the natural habitat in the East African Lake Tanganyika. Fish were fed daily and provided with gravel and terracotta flowerpots to simulate natural shelters positioned in each corner and one near the center. All experiments were conducted in accordance with the animal care and use guidelines of Harvard University (IACUC protocol number 22–22).

Behavioral measures for female *A. burtoni*

In order to quantify female behavior, we adapted the systematic description previously developed by Fernald (1977a) for dominant and subordinate *A. burtoni* males. Aggressive behavior patterns included chasing or biting, threatening, and engaging in border threats. Submissive behavior was measured by counting fleeing events triggered by an attacker. Mating behaviors consisted of digging, courting, and spawning. Courting was scored only for male-typical courtship behavior that involves lateral display, tail quiver, and leading another female toward the spawning territory. These behavioral measures were used to calculate a dominance index as the sum of all aggressive behaviors (biting, threatening and border threats) minus the number of fleeing events performed by an individual (Renn et al., 2008; White et al., 2002).

Although spawning was never directly observed during any of the focal observation periods, fish did spawn between observations and these events were recorded based on the visual observation of eggs in the buccal cavity. In addition to behavioral observations we recorded body coloration as either blue, yellow, or grey according to subjective report, and we recorded reproductive status as brooding, non-gravid or gravid by visual inspection of buccal or body distension. Schooling time and time in territory were estimated as the proportion of time the focal female was within 2 body lengths of either the majority of the fish in the tank or the terracotta flowerpot that she normally defended. The proportion of time the characteristic eye-bar was displayed was also estimated as a proportion of the observation period.

Observation Protocol

Three all-female experimental communities were established in 110-liter aquaria, with 9–10 females and 4–5 available pot shard territories each. While mean mass and length varied between communities (mean mass: 4.90g±1.6 s.d. – 7.48g±0.58 s.d.; mean standard length 5.59cm±0.41 s.d. – 6.36cm±0.18 s.d.), the females within a community were roughly size matched. These aquaria were maintained for >90 days and are referred to as the “long-term”

observations. Five additional all-female experimental communities were maintained for behavioral measures and neural tissue samples (for gene expressions studies reported elsewhere). Here, each 110-liter aquarium was divided in two by a clear divider and each side contained 5 females and 3 available potshard territories. Here, females were roughly size matched across all five communities (mean of mass: $3.54\text{g}\pm 0.66$ s.d.; mean standard length: $5.03\text{cm}\pm 0.29$ s.d.). These communities are referred to as “short-term” as they were maintained for only 45 days. Three mixed-sex control communities comprised of 5–6 males and 5–6 females with 4 pot shard territories were maintained in 110-liter aquaria. Here, the females were roughly size matched across all three communities (mean female mass: $4.19\text{g}\pm 0.70$ s.d.; mean standard length: $5.12\text{cm}\pm 0.13$ s.d.). Males were age matched to females, and therefore were larger (not measured). The females in these mixed-sex communities are referred to as control females. Two of these mixed sex communities were observed in parallel with the short-term observations and one in parallel with the long-term observation experiment. In all cases, females of unspecified reproductive cycle were taken from standard stock tanks and individually marked with a colored bead held through the dorsal muscle by a plastic tag (Avery Dennison, Pasadena CA).

In all communities, three-minute real-time focal observations were performed 2–3 times per week between 08:30 and 10:00 hours (0.5–2 hours after light onset) following a 5–10 min. accommodation period during which the observer sat in front of each aquarium. All females in a single tank were observed sequentially on the same day, and the order was varied each observation day. All female fish were surveyed daily for brooding status. Behavioral data from females in both the long-term and short-term observation communities are reported only for those fish that exhibited a consistent behavioral phenotype for the four weeks preceding euthanasia (DOM: $n=15$; SUB: $n=21$, Control: $n=14$) and included four to ten observations per fish (mean = 7.94 observations).

Male Reintroduction

In order to determine whether a female’s mating success was related to her social status we reintroduced males into all-female communities. Three additional 110-liter aquaria were established, each of which housed two communities of 5 females separated by a clear divider. Focal observations were performed twice every week between 08:00 and 10:00 hours for seven weeks in order to identify dominant and subordinate individuals and quantify the total number of days each individual displayed a dominant phenotype. These animals are not included in the detailed behavioral analysis. After seven weeks a single male was introduced to each community. Three times per week, for the following 30 days, females were visually inspected for evidence of eggs in the buccal cavity.

Growth and Condition

At intervals of two to five weeks, fish were weighed and standard length (SL) was recorded. As was already observed by Hofmann et al. (1999), removal of fish for weighing and measuring does not disrupt hierarchies; the behavior patterns displayed before and after each measurement do not differ qualitatively, and dominant females always returned to their original territories. Growth rate (GR) was calculated as the average weekly relative change in SL during the final four weeks of the experiment when the animals’ social status was stable. Condition factor (CF) was calculated based on the residuals from the regression of body mass on standard length at the end of the observation periods for those animals that held a stable social status for the preceding four weeks (i.e. the individuals for which behavior data are also reported). Gonadosomatic index (GSI) was calculated as (gonad mass/body mass) $\times 100$ for the subset of females that had displayed a consistent social phenotype for four weeks and were killed at the end of the short-term observation experiment.

Hormone Assays

Using heparinized butterfly infusion sets (SURFLO 26G, #SV-25BLK), we collected blood from the dorsal aorta from dominant (n=8) and subordinate (n=15) females in the short-term communities as well as from control females (n=6). As for the reported behavioral measures, only individuals that had demonstrated consistent social phenotype for 28 days or longer were used. The plasma was separated from the serum using a tabletop centrifuge at 5000rpm for 10min and stored at -80°C . We measured testosterone with a direct radioimmunoassay (Diagnostic Systems Laboratories, Webster, TX) in plasma samples diluted 1:12.5 (Trainor and Hofmann, 2006). The crossreactivity of the assay for dihydrotestosterone was 5.8% and 2.3% for androstenedione. We measured estradiol in plasma samples diluted 1:10 also using a direct radioimmunoassay (Diagnostic Systems Laboratories, Webster, TX). The sensitivity of both assay systems was 0.1 ng/mL. There was not sufficient sample volume to also measure 11-ketotestosterone. Furthermore, in *A. burtoni* circulating levels of this fish-specific androgen have always been tightly correlated with testosterone, independent of sex, albeit at much lower concentrations (Kidd et al., 2010; Parikh et al., 2006). For each assay system, we followed the manufacturer's protocol. For each hormone assay, assayed concentrations for serial dilutions of an *A. burtoni* plasma pool were compared with standards and computed regression lines did not differ in slope ($p > 0.05$). All samples were assayed in duplicate in each assay. The intra-assay coefficient of variation was 3.3% for testosterone and 8.4% for estradiol.

Statistical Analysis

All statistical analyses were performed in R, the open source software environment for statistical computing and graphics (R development group). Due to non-normal distribution of the behavioral, hormonal and GSI data, p-values were calculated using one-way ANOVA in a randomization test based on 10,000 replications. To identify significant differences in each pair wise comparison of dominant, subordinate and control females, the probability of obtaining a difference at least as great as observed was calculated from the randomization and those p-values were adjusted for multiple comparisons (Benjamini and Hochberg, 1995). Behavioral data, averaged over the final four weeks, were analyzed together for females in the long-term (DOM: n=9; SUB: n=15, Control: n=12) and short-term (DOM: n=6; SUB: n=6, Control: n=4) observation communities because there were no statistically significant differences between the tanks. Linear Least Squares Regression was applied between hormone measures and Spearman rank correlations were computed to assess the relationship between hormone titers and specific behaviors (averaged over just one week prior to blood collection).

Results

Female Appearance and Behavior Can Resemble that of Dominant Males

In both long-term and short-term all-female communities, a subset of the females displayed an eye-bar approximately 90% of the time, while this characteristic display of social dominance was observed only 10% of the time in the remainder of the females in the all female communities and only 5% of the time in the control females. These females displaying the eye-bar, like dominant males, exhibited either yellow or blue coloration; however, it was not as vivid as is seen in males (Figure 1). The females that displayed the eye-bar 10% of the time or less did not exhibit any body color. The red humeral patch and the distinctive egg spots on the anal fin, which are two other color patterns typical of dominant males, were never seen in any of the females.

Females that developed these male-typical morphological characteristics also displayed male-typical dominance behavior. Behavioral measures, dominance index and time in

territory see below), not color, were used to designate “dominant” females at the end of each observation period. Here we analyze and present the behavior of only those individuals that consistently expressed the same social phenotype for the last four weeks of continuous observation (DOM: n=15; SUB: n=21; Controls: n=14). We found significant differences according to phenotype for several male-typical aggressive displays (Figure 2; Table 1). We observed significantly more chasing events in dominant females compared to either subordinate ($p < 0.0001$) or control females ($p < 0.0001$) (mean number per 3 minute observation \pm SE: DOM: 11.45 ± 0.86 SUB: 0.83 ± 0.16 CON: 1.50 ± 0.39). Dominant females also displayed significantly more threat displays compared to either subordinate ($p < 0.0001$) or control females ($p < 0.0001$) (DOM: 1.33 ± 0.25 SUB: 0.08 ± 0.04 Control: 0.03 ± 0.02), and stereotypical border threats were observed almost exclusively in dominant females (DOM: 0.48 ± 0.08 SUB: 0.004 ± 0.005 Control: 0.0 ± 0.0). Dominant females showed a tendency to escalate aggressive interactions to a “carousel” display, a male-typical behavior; however, these events were very rare (data not shown). Furthermore, dominant females were more likely than either subordinate ($p = 0.0006$) or control females ($p < 0.0001$) to perform digging behavior (DOM: 0.69 ± 0.17 SUB: 0.19 ± 0.04 Control: 0.02 ± 0.01), which dominant males display to prepare the territorial bower for spawning. Nine of the 15 dominant females were also observed at least once performing male-typical courtship displays, whereas subordinate females never initiated courtship and only once was a similar behavior observed from a control female in the mixed-sex communities. Much of the aggressive behavior observed in dominant females was directed toward the subordinate females. Dominant females fled significantly less often than either subordinate ($p < 0.0001$) or control females ($p = 0.00015$) (DOM: 0.38 ± 0.09 ; SUB: 7.37 ± 0.55 ; Control: 5.43 ± 0.47) and spent a significantly smaller fraction of their time schooling (DOM: $13.06 \pm 3.57\%$; SUB: $88.49 \pm 4.71\%$; Control: 93.93 ± 1.85). Subordinate females closely resembled control females that were housed with males such that no statistically significant behavioral differences were observed between subordinate and control fish, even the difference in the number of observed fleeing events was not significant ($p = 0.097$). For males, aggressive and subordinate behaviors have been used to calculate a dominance index (Renn et al., 2008; White et al., 2002); see methods). This measure also effectively differentiates the female phenotypes (DOM: 12.75 ± 0.83 ; SUB: -6.5 ± 0.54 ; Control: -3.72 ± 0.83 ; $F_{2,48}$; $p < 0.0001$; Figure 3). The index for the dominant females is significantly greater than for subordinate or control females ($p < 0.0001$), but was not statistically different for subordinate compared with control females ($p = 0.3606$).

Dominant females displayed many of the morphological and behavioral characteristics normally associated with dominant males, yet, even in the long-term (up to 140 days) all-female communities, no female changed gonadal sex. This result, observed by gross inspection of gonadal tissue and failure to observe successful fertilizations in the all-female communities, is in agreement with past work on this species (J. Rhodes and R. Fernald personal communication). Brooding was observed for both dominant and subordinate females. Of the 14 females that showed dominance behavior in the long-term observation communities, 11 (78%) had spawned at least once, whereas of the 25 females that displayed subordinate status, 13 (52%) had spawned. This difference is not significantly different (Fisher Exact test; $p = 0.171$). All of the control females spawned at least once.

Territorial Tenure of Dominant Females

Among the three separate long-term communities, a total of 14 out of 29 individuals exhibited the male-typical dominant phenotype for at least some portion of the experiment, underscoring the dynamic social life of this species. Eleven of these 14 females lost their status at least once (three lost it twice; one lost it three times), and all but two of those later regained their status. In the majority of cases (69%) territory loss occurred shortly after spawning. Median duration of tenure as a dominant female was 3.6 weeks, for subordinates

the median was 3.4 weeks. When we disregard the short periods (several days) of subordinate status immediately after brooding, median time spent as dominant was 3.9 weeks and median time as subordinate was 4.9 weeks.

Circulating Sex Steroid Hormones

Blood samples for assaying hormone concentrations were obtained from eight dominant, fifteen subordinate, and six control females from the long-term communities. All of these fish had maintained social status at least four weeks prior to sampling. Circulating testosterone levels varied significantly among phenotypes ($F_{2,26}=5.85$, $p=0.0067$) (DOM: 10.79 ± 1.97 , SUB: 5.19 ± 0.88 , Control: 6.64 ± 0.97 ng/mL). T levels were significantly higher in dominant compared to subordinate females ($p=0.0087$), but not compared to control females ($p=0.0829$; Figure 4A). When we measured circulating estradiol levels, the variation across the experimental groups was significant ($F_{2,26}=3.16$, $p=0.047$). However, while pair wise comparisons were not significant after false discovery rate correction, estradiol levels showed the same pattern as testosterone in that dominant females exhibited higher titers than either subordinate ($p=0.0612$) or control females ($p=0.1205$) (DOM: 6.39 ± 2.32 , SUB: 2.25 ± 0.63 , Control: 2.88 ± 1.19 ng/ml) (Figure 4B).

There was a strong positive relationship between testosterone and estradiol levels within dominant and subordinate phenotypes, but not in control females, as revealed by linear regression analysis (DOM: $F_{1,6}=15.79$, $R^2=0.725$, $p=0.007$; SUB: $F_{1,13}=15.47$, $R^2=0.508$, $p=0.002$; Control: $F_{1,4}=1.17$, $R^2=0.226$, $p=0.341$) (Figure 4C). While circulating estradiol did not correlate with any behavioral displays, we found that testosterone levels did tend to correlate with behavior patterns, (Table 2). Both chasing and threat behavior were positively correlated, while fleeing and digging behavior showed a negative correlation.

Gonadosomatic Index

We calculated the gonadosomatic index (GSI) as a measure of reproductive status for a subset of the females from each phenotype in the short-term communities only (DOM: 7.5 ± 2 , $n=4$; SUB: 5.2 ± 1.4 , $n=6$; Control: 0.9 ± 0.2 , $n=6$). While there was significant variation among phenotypes ($F_{2,13}=6.748$, $p=0.0034$), largely due to the low GSI of the control females relative to dominant ($p=0.0081$). Dominant and subordinate females did not differ in this measure ($p=0.1419$). This is not unexpected because the control females were brooding and therefore their ovaries would not be expected to contain any developed oocytes. There were no significant correlations between GSI and behavioral measures when only the dominant and subordinate females were considered.

Somatic Growth and Condition

In the long-term communities, weekly growth rates averaged 0.88% length across the subset of females monitored, with dominant females trending towards faster growth (DOM: $1.6\pm 0.1\%$, $n=14$; SUB: $0.99\pm 0.04\%$, $n=20$; Control: $0.70\pm 0.02\%$, $n=9$). However, this variation in growth rate with respect to social status was not significant ($F_{2,40}=2.965$, $p=0.0589$). Similarly, condition factor (CF) was calculated for a subset of the individuals included in the behavior analysis above (DOM: 3.40 ± 0.109 , $n=14$; SUB: -0.074 ± 0.150 , $n=20$; Control: -0.364 ± 0.299 , $n=9$). While, CF differed among social phenotypes ($F_{2,40}=3.404$, $p=0.043$), pair wise comparisons were not significant after false discovery rate correction (data not shown).

Male Reintroduction

We reasoned that for females a dominant phenotype may convey an advantage by increasing her mating success when a male does become available. To test this hypothesis, we

introduced a single dominant male to each of six all-female communities after the female social hierarchy had been stable for at least four weeks. Of the 13 females that had experienced dominance, only four spawned within the 30-day observation period following male introduction. Importantly, the duration of territorial tenure did not predict whether a dominant female spawned or not (Spearman correlation; $\rho=0.151$, $p=0.44$). The non-spawning dominant females maintained an eye-bar, continued aggressive behavior toward other females, and were vigorously attacked by the introduced males (data not shown). Among the 15 females that never experienced dominance, eight spawned within the 30-day observation period, which is not different from the dominant females (χ^2 test; $p=0.2049$). However, of the individuals that did spawn, the dominant females ($n=4$) mated with the reintroduced male within 5.75 ± 3.09 days, whereas subordinate females ($n=8$) needed 16.12 ± 2.57 days, a significant difference (t-test; $p=0.036$).

Discussion

In the present study we have shown that, for all-female communities of the highly social cichlid species *A. burtoni*, individuals can acquire a social dominance phenotype that closely resembles the well-known male-typical phenotype associated with territory maintenance and access to potential mates. In all-female situations, dominant females show behavioral, morphological, and hormonal changes typical of dominant males of this species. However, dominant social status in females did not correlate with growth rate or reproductive status, which eliminates some of the confounding physiological variables associated with the study of social status in males of this species. This paradigm thus provides a great opportunity to decipher the mechanisms that specifically underlie socially mediated behavioral plasticity.

In females, the dominant phenotype created by removal of males, included direct aggressive behavior such as chasing as well as stereotypical displays of aggression such as lateral displays and border threats. Interestingly, these females also showed male typical behaviors of digging (usually associated with bower construction) and courtship directed toward other females. Contrary to aggression in males, among females, most of the aggressive behavior observed was directed toward the subordinate females rather than other dominant females, reflecting the strong social hierarchy within the community. With regard to behavior, subordinate females provide a suitable comparison to subordinate males. Importantly, subordinate females, unlike subordinate males, do not show decreased GSI nor increased growth rate. Thus, the similar behavioral phenotypes of subordinate females and subordinate males can be compared in the absence of these confounding variables as is also the case for the dominant phenotypes of the two sexes.

Our behavioral observations demonstrate that female dominance, like male dominance, is a plastic and reversible phenotype. While not every individual attained dominant social status during our observations, there was a high degree of social volatility, similar to the situation in males (Hofmann et al., 1999). Among males, dominance is primarily determined by relative size within the social group (Hofmann et al., 1999). Similarly, in 4 out of the 5 all-female communities, the females that adopted the subordinate phenotype were on average smaller (as measured by mass or by standard length) at the beginning of the experiment than females that eventually established a stable dominant phenotype (data not shown). Because we measured size at time intervals independent of status changes, we cannot conclusively demonstrate this relationship between size and probability to achieve dominance in a given social interaction for the females in this study. However, given the difference in mean size and the similarity in growth rates between dominant and subordinate females, one would expect that females could retain dominant status longer than males. However, the duration of tenure in a particular social status for females is more similar to male tenure under a fluctuating environment (DOM: 3 weeks; SUB: 4 weeks) than under a stable environment

(DOM: 9.5 weeks; SUB: 7 weeks)(Hofmann et al., 1999), suggesting that the social volatility observed in all-female communities may be at least in part caused by their reproductive cycle, including spawning and (abandoned) brooding events.

We found a clear association between social phenotype and circulating testosterone in females as had been seen in males (Parikh et al., 2006). As seen in previous studies, testosterone levels in females were overall considerably lower than those reported for either dominant or subordinate males (e.g., (Greenwood et al., 2008; Trainor and Hofmann, 2006). Nonetheless, the elevated testosterone levels, and correlated increases in estradiol, in dominant females suggest a role for these steroids in female aggression. The lack of strong correlations between circulating levels of steroid hormones and specific behavior patterns suggests that their effects are indirect and/or several other factors play important roles here as well. Either scenario would reduce the amount of variation explained by steroid hormone levels. It could also be that ongoing aggressive interactions – such as border threats – lead to acute fluctuations in androgen levels. We did not test this “challenge hypothesis” (Wingfield et al., 1990) in our all-female communities, however recent reports suggest that such a response does occur during maternal care (Renn et al., 2009) or during aggressive interactions in gravid females (Desjardins personal communication). Finally, it is possible that 11-KT, which we did not measure, is more tightly associated with social behavior than other steroids. However, this is unlikely as numerous studies have clearly shown that in *A. burtoni* (Kidd et al., 2010; Parikh et al., 2006) as well as other Haplochromine cichlids (Dijkstra et al., 2012) this fish-specific androgen is always tightly associated with testosterone at about 10-fold lower concentrations, independent of sex (e.g. Kidd et al., 2010)

The two color phenotypes, yellow (n=9) or blue (n=2), that we observed in dominant females are reminiscent of the color polyphenism typical of dominant males (Korzan et al., 2008), even though several dominant females (n=4) showed no obvious coloration. Some of the females displayed color as early as three days after all-female experimental communities were established. While the intensity of the color was somewhat variable, females do not appear to switch color morphs as readily as males (Korzan et al., 2008). While size clearly plays a role in determining social status, baseline hormone levels and/or color variation (cichlid: Dijkstra et al., 2009; bird: (Pryke, 2007) may be contributing factors. These two traits are known to be correlated with each other and in some cases with aggression levels in a wide range of animals (e.g. cichlid fish: Dijkstra et al., 2012; reptiles: Huyghe et al., 2009; birds: Muller and Eens, 2009; baboons Higham et al., 2008).

Dominant females do not appear to suppress the GSI and reproductive physiology of subordinate females, which is in strong contrast to the well-known situation in males (see, for example, Hofmann and Fernald, 2000). We found that both dominant and subordinate individuals in the all-female communities possessed ovaries of similar relative size (as measured by GSI) and continued to spawn and even incubate (unfertilized) eggs, thus demonstrating that both phenotypes produce mature ova. Similarly, dominant and subordinate females were in similar condition, whereas both GSI and condition were decreased in control females, which refrain from feeding during egg incubation (Renn et al., 2010). The observation that many dominant females transiently lost their social status immediately after spawning suggests that dominance behavior can nevertheless also be regulated by the reproductive cycle in addition to the social environment. The fact that the median tenure as dominant female (3.9 weeks) is almost exactly as long as the female reproductive cycle (Kidd et al., 2012) further suggests a role of reproductive physiology. While in many other teleost species changes in social status are often accompanied by sex change, our results are consistent with previous observations in cichlids of behavioral sex-

role change without subsequent gonadal sex change in cichlids (reviewed by Oldfield, 2005).

It has been suggested that female aggression might be associated with competition for males and/or in response to male aggression at the lek (Karvonen et al., 2000; Desjardins personal communication). We therefore hypothesized that male-typical dominance behavior might enhance a female's future spawning opportunities. However, the duration of territorial tenure did not predict whether a dominant female spawned or not. Subordinate females were just as likely as dominant individuals to spawn with an introduced male, although it took them almost three times as long to obtain a mating. Careful field studies will be needed to determine whether the male-typical dominance behavior we have described here indeed provides females with any adaptive advantage in the context of mate acquisition. Alternatively, the ultimate function of female dominance could derive from other social situations among the females. For example, female aggression and dominance status affect reproductive success in a wide variety of species and contexts (Heinsohn et al., 2005; Kinahan and Pillay, 2008; Sterck et al., 1997). Furthermore, recent evidence from a newly collected wild stock of *A. burtoni* suggests that maternal care after release of the fry from the buccal cavity is much more extensive than previously appreciated (Renn et al., 2009). Maternal care in this species likely involves territorial defense of a temporary nest site, as was first noted in field observations (Fernald and Hirata, 1977a). Clearly, more research is needed in both lab and field settings to further examine these and other adaptive explanations.

Conclusions

The novel experimental paradigm introduced here, which utilizes social hierarchies in all-female communities, provides a great opportunity to examine the physiological and molecular mechanisms underlying social dominance in a context that parallels the well-studied situation in males, yet differs in important aspects. We have shown that, in the absence of males, female *A. burtoni* can display male-typical social dominance behavior and associated circulating androgen levels reminiscent of dominant males. In contrast to the situation in males, dominant and subordinate females differ little in reproductive maturity or somatic growth. Thus, the all-female communities examined in the current study in conjunction with the well-established social plasticity in males provide an excellent opportunity to identify common molecular and physiological underpinnings of social dominance independent of sex.

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Highlights

In all-female communities of *A. burtoni* cichlids some individuals exhibit male-typical dominance behavior

Dominant and subordinate females differ in social behavior and circulating sex steroid hormones

Potentially confounding factors such as gonadal state and growth do not vary

This paradigm provides a novel approach to the study of the mechanisms of social behavior.



Figure 1. Representative photographs of females expressing either the dominant (left) or subordinate (right) phenotype typical of male social organization. Note the male-typical eye-bar displayed by the dominant female (arrow).

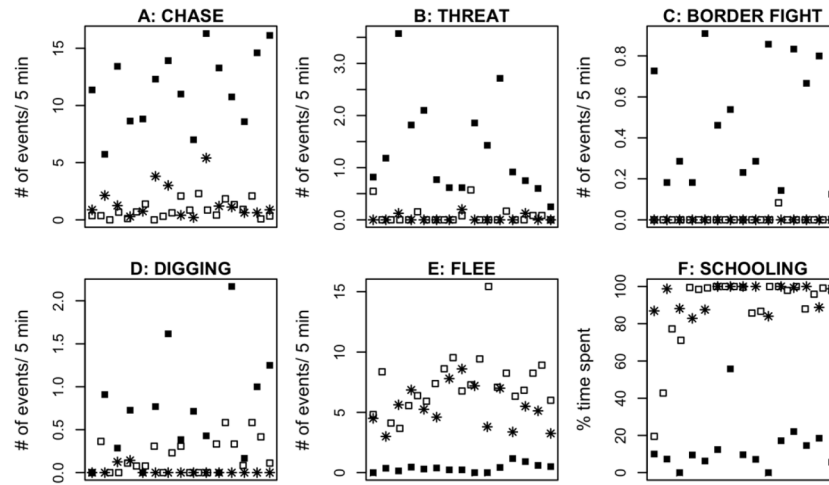


Figure 2. Behavioral female phenotypes. Dominant (solid squares) females exhibit several male-typical aggressive behavior patterns: (A) Chasing (B) Threat Display (C) Border Threats and (D) Digging. Subordinate (open squares), like control (asterisk), females show a high level of (E) Flee and (F) Schooling. Individual fish are arranged along the x-axes in a consistent but not systematic order.

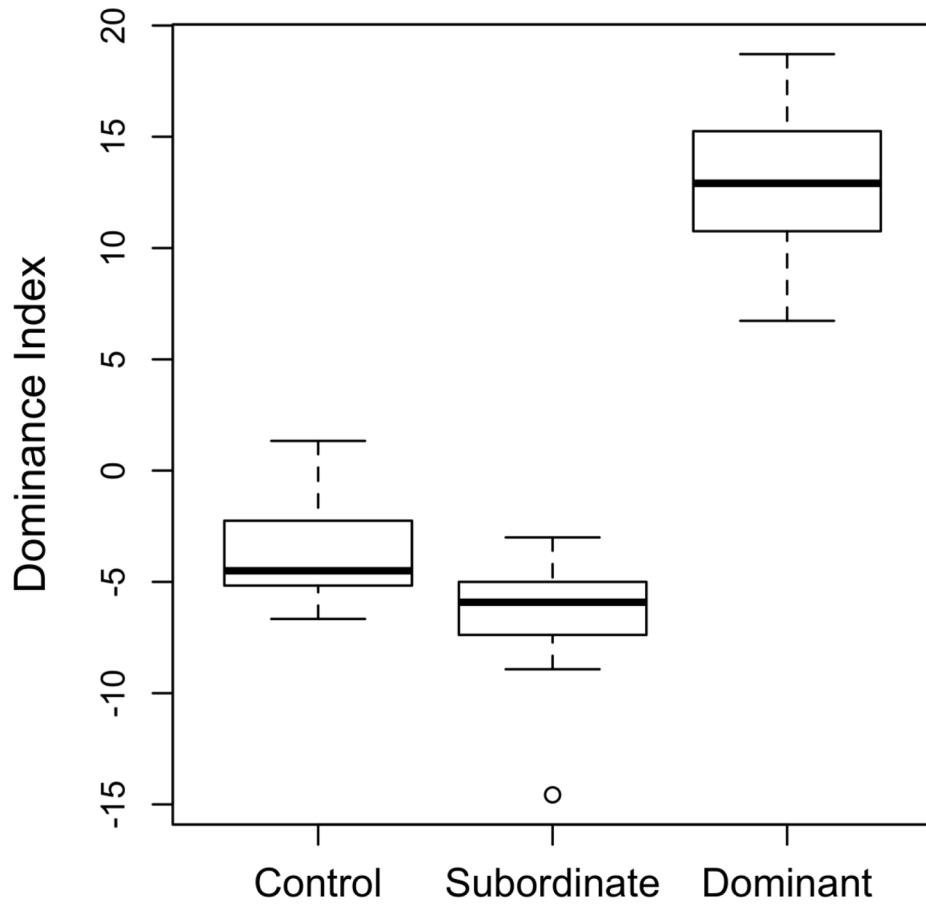


Figure 3. Dominance Index for each female phenotype. Boxplots show the upper and lower quartiles around the median; whiskers indicate the lowest and highest datum still within the 1.5 interquartile range. All pairwise comparisons are significantly different at $\alpha = 0.05$ (see Table 1).

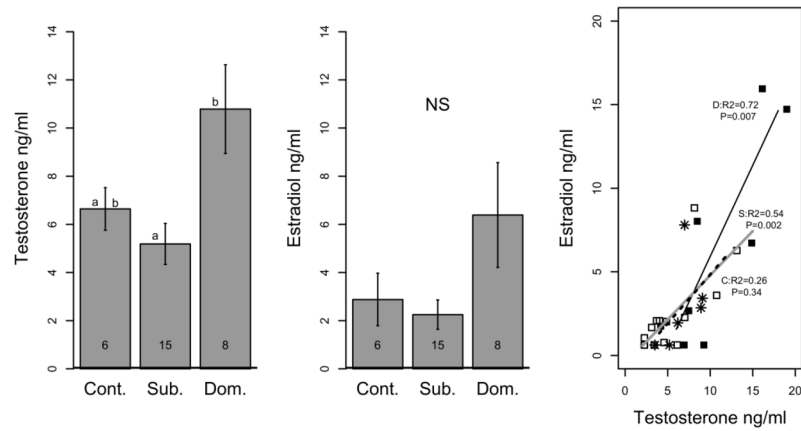


Figure 4. Hormone levels in female phenotypes

Circulating levels (mean \pm SE error) of testosterone (A) and estradiol (B) are shown. Numbers indicate sample sizes, letters indicate statistically significant differences ($p < 0.05$). (C) Correlation between steroid titers for individuals of each phenotype. Regression coefficients and p-values are given for D:dominant (black line), S:subordinate (gray line), and C:control (dashed line) female.

Table 1

Behavioral measures averaged over the last four weeks for female phenotypes in both long term and short term observation tanks. (C = Control; D = Dominant; S = Subordinate)

Behavior	Mean (\pm s.e.m)		One-way ANOVA		P-value (Tukey HSD)			
	Control n=15	Subordinate n=21	Dominant n=15	F _{2,48}	p	C vs. S	C vs. D	S vs. D
Chase	1.50 (0.396)	0.837 (0.162)	11.456 (0.862)	145.99	< 0.0001	0.7027	< 0.0001	< 0.0001
Border Fight	0 (0)	0.004 (0.004)	0.482 (0.079)	48.09	< 0.0001	0.965	< 0.0001	< 0.0001
Threat	0.03 (0.017)	0.08 (0.374)	1.334 (0.246)	33.06	< 0.0001	0.0501	< 0.0001	< 0.0001
Flee	5.43 (0.466)	7.378 (0.551)	0.381 (0.088)	63.63	< 0.0001	0.0965	0.0002	< 0.0001
Digging	0.018 (0.013)	0.187 (0.044)	0.694 (0.168)	14.38	< 0.0001	0.2854	< 0.0001	0.0006
School % time	94.33 (1.846)	88.493 (4.713)	13.058 (3.571)	128.43	< 0.0001	0.6649	< 0.0001	< 0.0001

Table 2

Spearman rho correlation coefficients (n=29) relating circulating steroid hormone levels and behavioral measures (averaged across week before blood collection). Uncorrected p-values are reported in parentheses, with values of $p < 0.05$ highlighted in bold.

Behavior	Hormone	
	[Estradiol]	[Testosterone]
Chase	0.221 (0.248)	0.377 (0.044)
Border Fight	0.114 (0.556)	0.306 (0.105)
Threat	0.235 (0.219)	0.403 (0.029)
Flee	-0.276 (0.146)	-0.375 (0.045)
Digging	-0.114 (0.556)	0.195 (0.309)
School	-0.283 (0.136)	-0.437 (0.018)
Dom. Index	0.204 (0.289)	0.274 (0.015)