



Published in final edited form as:

J Phys Chem B. 2012 April 5; 116(13): 3918–3927. doi:10.1021/jp211904j.

Structure and Elasticity of Lipid Membranes with Genistein and Daidzein Bioflavonoids Using X-ray Scattering and MD Simulations

Mohit Raghunathan^{1,a}, Yuriy Zubovski^{1,b}, Richard M. Venable², Richard W. Pastor², John F Nagle¹, and Stephanie Tristram-Nagle^{1,*}

¹Biological Physics Group, Physics Dept., Carnegie Mellon University, 5000 Forbes Avenue, Pittsburgh, PA 15213

²Laboratory of Computational Biology, National Heart, Lung and Blood Institute, 5635 Fishers Lane T-900 Suite, Rockville, MD 20852

Abstract

This work reports the effects of the bioflavonoids genistein and daidzein on lipid bilayers as determined by volume measurements, X-ray scattering and molecular dynamics simulations. The experimental and simulated total molecular volumes were found to be in outstanding agreement with each other before the addition of genistein and daidzein and also after their addition. Both bioflavonoids inserted into the hydrocarbon region of both DOPC and DiphytanoylPC near the carbonyls of the lipids and both decreased the bilayer thicknesses. The long axes of both bioflavonoids were oriented nearly parallel to the plane of the bilayer with their carbonyl groups preferentially pointed towards the proximal surface. A difference is that daidzein had a solubility limit ~0.14 mole fraction in DOPC (~0.12 mole fraction in DiphytanoylPC) whereas genistein was soluble at least to 0.20 mole fraction in both lipid membranes. Measurements of bending modulus K_C and simulation results for area compressibility modulus K_A indicate that both bioflavonoids soften bilayers.

Keywords

hydrophobic mismatch; bending modulus; area compressibility modulus; gramicidin A; electron density profiles

Introduction

Ion-channel modifiers are generally thought to regulate protein phosphorylation and dephosphorylation. One class of modifiers, the bioflavonoids, includes the well-studied genistein^{1–2}. For the cystic fibrosis transmembrane conductance regulator (CFTR) channel, genistein affects the wild-type channel, and activates a mutant channel due to a change in its specific binding site³. As a tyrosine kinase inhibitor, genistein could serve as an antitumor agent since kinase activity is strongly correlated with the ability of retroviruses to transform

*Corresponding Author: Tel.: 412-268-3174; Fax: 412-681-0648, stn@cmu.edu.

^aCurrent address: Dept. of Physics and Astronomy, Rice University, 6100 Main St., Houston, TX 77005

^bCurrent address: Capital IQ, 55 Water St., NY, NY 10041

Supporting information description: The supporting information (SI) section contains five figures that have been described in the main paper and calculations for the concentration of bioflavonoids in lipid membranes. This information is available free of charge via the Internet at <http://pubs.acs.org>.

cells⁴⁻⁶. Another specific effect of genistein is to mimic the action of estrogen⁷⁻⁸. Most estrogen effects are mediated by estrogen receptors, which are transcriptional regulators, but some are mediated by membrane receptors linked to calcium metabolism⁹⁻¹⁰. The roles of estrogen, genistein and another bioflavonoid, daidzein, on murine osteoclasts were investigated; estrogen > genistein > daidzein all reduced osteoclast differentiation which may protect bone¹¹.

In addition to these specific effects that require binding to proteins, bioflavonoids have also been reported to modulate ion-channel activity in a non-specific way, i.e., by altering the properties of the lipid membrane surrounding the channel¹²⁻¹³. By changing the length of a gramicidin A (gA) channel¹² and by using lipid membranes of varying thickness¹³, the importance of hydrophobic mismatch in limiting both the onset and the lifetime of gA channel formation was demonstrated. It was suggested that genistein shifts the equilibrium from non-conducting monomers to conducting gA dimers by compensating for hydrophobic mismatch. This conclusion was reached since the magnitude of the effect of genistein increased with increasing hydrophobic mismatch between the channel length and the membrane thickness¹³. It was further hypothesized that genistein affects protein-lipid coupling by changing the elastic properties of the membrane, which involve the lipid area compressibility modulus K_A and the bending modulus K_C . An interesting contrast was that, in DPhyPC/n-decane black lipid membranes, daidzein did not increase gA channel lifetime¹³, although later it was determined that daidzein did increase gA channel lifetime, but only half as much as genistein¹⁴. Another reported difference between these two bioflavonoids is that daidzein, but not genistein, was reported to aggregate liposomes¹⁵.

In the present investigation, we use x-ray diffuse scattering to measure K_C and to provide basic structural data for the effect of genistein and daidzein in DOPC and DPhyPC membranes. These data are used to validate molecular dynamics (MD) simulations which then provide the area/unit cell of DOPC with bioflavonoids at 20 and 14 mole%, and the bioflavonoid positions in the DOPC membrane. In addition, we measure the molecular volume and use a volume-conserving fit of our diffuse x-ray scattering data to a bilayer model for the component groups¹⁶ as an alternative way to determine the position of both bioflavonoids in membranes. These structural and elasticity results provide insight into the effects of bioflavonoids on lipid membranes.

Materials and Methods

Samples

Dioleoylphosphatidylcholine (DOPC) (Lot 18:1-228) and diphytanoyl-phosphatidylcholine (DPhyPC) (Lots 4Me160-118, 4Me160-121) were purchased from Avanti Polar Lipids in the lyophilized form. Bioflavonoids, genistein and daidzein, were purchased from Sigma/Aldrich (St. Louis, MO); their chemical structures are shown in Fig. 1. Lipid/bioflavonoid mixtures in the mole fractions 0.05, 0.10, 0.15 and 0.20 were prepared by mixing with lipids in organic solvent. Bioflavonoids are insoluble in most common organic solvents at room temperature, including chloroform, methanol, acetone, hexafluoroisopropanol and trifluoroethanol, but soluble in hot methanol and DMSO. Four mg DOPC/bioflavonoid mixtures (in duplicate) were dissolved in 200 μ l HPLC hot methanol and plated onto 30mm \times 15mm \times 1mm silicon wafers in an oven at 50°C using the rock and roll technique¹⁷. For DPhyPC/bioflavonoid mixtures, evaporation from chloroform in the fume hood was more successful than hot methanol at producing well-oriented samples. After drying overnight in the hood, samples were trimmed to a 5 mm \times 30 mm strip in the center of the wafer. Hydration of oriented samples from water vapor was then carried out in a thick-walled hydration chamber¹⁸. Unoriented multilamellar vesicles (MLV) in excess water were prepared by weighing 1–2 mg of dry lipid mixture with 40 μ l milli-Q water and thoroughly

mixing in small nalgene vials, then thermally cycling three times with vortexing between $-20\text{ }^{\circ}\text{C}$ and $50\text{ }^{\circ}\text{C}$ before loading into 1mm diameter glass capillaries (Charles Supper, Cambridge, MA). Thin layer chromatography using chloroform:methanol:7N NH_4OH (46:18:3, v/v) and a molybdic acid stain indicated negligible lysolipid before and 0.1–0.5 % lysolipid after X-ray exposure, with genistein samples showing slightly more degradation than daidzein samples.

Volume determination

Volumes in fully hydrated MLV were determined at $30\pm 0.01\text{ }^{\circ}\text{C}$ using an Anton-Paar USA DMA4500 (Ashland, VA) vibrating tube densimeter. Volume of the 'unit cell', V_1 , is defined to be the volume V_L of one lipid plus the proportional bioflavinoid volume $V_{BF}f/(1-f)$ where the mole fraction f is the number of moles of bioflavinoid divided by the number of moles of lipid and bioflavinoid and V_{BF} is the volume of one bioflavinoid. V_1 was calculated from

$$V_1 = \frac{MW_l}{0.6022 \rho_s} \left[1 + \frac{m_w}{m_M} \left(1 - \frac{\rho_s}{\rho_w} \right) \right], \quad (1)$$

where ρ_s and ρ_w are the measured densities of the samples and pure water, respectively, m_w is the mass of water and m_M is the sum of the masses of the lipid and the bioflavinoid in the mixture. In accordance with the definition of V_1 , MW_l is the molecular weight of one lipid plus $f/(1-f)$ times the molecular weight of the bioflavinoid.

X-ray scattering

X-ray data from oriented fluid phase lipid mixtures at $30\text{ }^{\circ}\text{C}$ were obtained on two trips to the Cornell High Energy Synchrotron Source (CHESS) using the G1 station managed by Dr. Arthur Woll. The wavelength was set with a WB_4/C multilayer monochromator to 1.1803 \AA on trip 1 and to 1.1825 \AA on trip 2, with a total beam intensity of 5×10^{11} photons/sec/mm². Beam width was 0.26 mm and the beam height was 0.9–1.2 mm. The samples were ~10 micron thick along the normal to the ~2000 bilayers. The angle of the flat samples was cycled uniformly from -3 to 7 and back to -3 degrees relative to the beam once a second during the 30–60 s exposures. Data were collected using a Flicam CCD (Finger Lakes Instrumentation, Lima, NY) with a 1024×1024 pixel array with average pixel size $71\text{ }\mu\text{m}/\text{pixel}$. The sample-to-CCD distance was 371 or 354 mm, calibrated using a silver behenate standard with D-spacing 58.4 \AA . Temperature was controlled with a Neslab Controller (Portsmouth, NH) and monitored using a Cole-Parmer thermistor (Vernon Hills, IL). To obtain fully hydrated D-spacings, MLV samples were X-rayed at CMU at $30\text{ }^{\circ}\text{C}$ using a Rigaku RUH3R microfocuss rotating anode (Woodlands, TX) equipped with Xenocs FOX2D (Sassenage, France) focusing collimation optics.

The analysis of diffuse data from oriented stacks of fluctuating fluid bilayers has been previously described^{18–21} and will only briefly be summarized here. The scattering intensity for a stack of oriented bilayers is the product: $I(\mathbf{q}) = S(\mathbf{q})|F(q_z)|^2/q_z$, where $\mathbf{q} = (q_r, q_z)$, $S(\mathbf{q})$ is the structure interference factor, $F(q_z)$ is the bilayer form factor and q_z^{-1} is the usual low angle x-ray scattering (LAXS) approximation to the Lorentz factor for which all the sample remains in the beam for all relevant q . The first step of the analysis obtains the bilayer bending modulus K_C and the compression modulus B by fitting to the q_r dependence of the diffuse X-ray scattering. $|F(q_z)|^2/q_z$ is then determined by dividing $I(q)$ by the $S(q)$ derived from validated liquid crystal theory. A geometric undulation correction²² is used to multiply the q_z axis of $F(q_z)$ by a factor slightly larger than 1, for example, 1.02.

Structural Analysis

The x-ray $|F(q_z)|$ data were fit using a recently devised modeling procedure abbreviated SDP for Scattering Density Profile¹⁶. This procedure satisfies the important principle of volume conservation which requires that the volume probabilities of all the components sum to one at all z along the bilayer normal. The SDP procedure guarantees an important relation between the area A_1 and the zeroth order x-ray form factor $F(0)$ ²³:

$$A_1 F(0) = 2(n_1 - \rho_w V_1), \quad (2)$$

where V_1 is the measured volume of the lipid/bioflavinoid mixture in Eq. (1), $n_1 = n_L + n_{BF}f/(1-f)$ is the number of electrons, n_L is 434 for DOPC and 470 for DPhyPC, and n_{BF} is 140 for genistein and 132 for daidzein, $\rho_w = 0.333e/\text{\AA}^3$ is the electron density of water at 30 °C, and A_1 is the area of the unit cell, similar to the above definition of V_1 . The original SDP application parsed the lipid molecule into components, called the SDP model, that were especially appropriate for simultaneous analysis of x-ray and neutron scattering data¹⁶. For x-ray only data in this paper we have parsed the system differently, combining some of the features of the older H2²⁴ and HB models²⁵. We represented the phosphatidylcholine part of the headgroup with a PC Gaussian, the carbonyl/glycerol part of the headgroup with a CG Gaussian, and the hydrocarbon chains with a combination of error functions and a Gaussian for the terminal methyls as previously described¹⁶. The bioflavinoid was represented by an additional Gaussian. The fitting procedure allows the use of outside information to impose constraints on the model parameters. Informed by our volume measurements, we constrained the ratio of bioflavinoid volume to lipid volume, and the chain terminal methyl to the chain methylene volume ratio to 1.96. Consistent with the MD simulations, we constrained the widths of the lipid headgroup peaks to 2.5 Å, the bioflavinoid width to 3.0 Å, the methyl trough width to 3.0 Å and the width of the hydrocarbon interface to 2.4 Å. We also constrained D_{H1} , the difference in distance between the maximum in the electron density profile and the Gibbs dividing surface for the hydrocarbon region, to 4.95 Å, and ΔD_H , the difference in distance between the PC and CG Gaussians to 4.6 Å. Otherwise, the locations of the components remained free to provide estimates for D_{HH} and the position of the bioflavinoid in the membrane.

MD Simulations

Release version c35b4 of the CHARMM program²⁶ was used for all molecular dynamics simulations, model building, and either analysis of the simulation data, or extraction of data for further analysis with other programs. Distributed CHARMM force field parameters were utilized for DOPC²⁷ and water²⁸; CHARMM compatible parameters for genistein were obtained from Burendahl et al.²⁹, and parameters for daidzein were derived by analogy from them. A force switched Lennard-Jones method was used for the van der Waals term of the potential, with a cutoff of 12 Å, and the switching function starting at 10 Å. Electrostatics were computed with the particle-mesh Ewald method using a 12 Å real space cutoff, $\kappa=0.32$, a 5th order spline interpolation for the complementary error function, and a grid density of about 1 Å. All MD simulations used the Verlet leapfrog integrator and the extended pressure system implemented in CHARMM.

Pure DOPC systems were either an extension to 105 ns of an NPT ensemble simulation from Klauda et al.²⁷, or derived from that system and simulated via the NPAT ensemble (fixed area/lipid) for 35 ns. Starting with the NPT system ($\langle \text{area/lipid} \rangle = 69.0 \text{ \AA}^2$), the number density z profiles for each uniquely labeled atom of a molecule were first extracted from the simulation trajectories, and then combined and formatted with a Fortran program for subsequent use as input to the SIMtoEXP program³⁰. The SIMtoEXP program compares x-ray structure factors calculated from scattering data with those calculated from the

simulation number density profiles. In this case, the comparison indicated the area cross section ($69 \text{ \AA}^2/\text{lipid}$) was too small, and that simulations at larger area/lipid were required to provide atomic level insight to the experimental results. Two additional DOPC simulations were generated in the NPAT ensemble, with fixed area/lipid values of 72.4 and 75 \AA^2 . The last 25 ns of these two simulations were used for data analysis, while the last 90 ns of the NPT simulation were included for the calculation of K_A . For further model building, a library of individual DOPC conformations was derived from the latter part of the NPT simulation coordinate trajectory; random selection from this library was used to pack the mixtures described below.

Systems containing 20% bioflavonoids were constructed *de novo*, by placing an oriented lipid or bioflavonoid at grid points in a hexagonal lattice via random selection of the molecule type. Using a spacing derived from initial area estimates from the scattering experiments, a 7×7 grid was used for each leaflet, placing 36 lipids and 9 bioflavonoids, with 4 unoccupied lattice points, giving a total of 72 lipids and 18 bioflavonoids. After successful packing, the systems were hydrated with TIP3P water, energy minimized, adjusted to give a total water count of 2400 molecules (based on estimates of hydration for the experiments), and more extensively minimized. Replicate models for each bioflavonoid were built in this fashion, and three were chosen for NPT ensemble simulations. After heating from 203 K, the Hoover thermostat at 303 K was employed for NPT simulations for a total run time of 45 ns. Analogous to the case for DOPC, comparison of the experimental x-ray structure factor curves with those from the last 35 ns of the NPT simulations indicated that the simulation area cross sections ($73\text{--}75 \text{ \AA}^2/\text{lipid}$) were too small to match the experiments, and that NPAT simulations at larger areas were required. Systems containing 14% daidzein were obtained by removing 3 bioflavonoids from each leaflet, energy minimizing, adjusting the area cross section via NP γ T simulations, and running for an additional 35 ns.

A genistein coordinate set from the 45 ns point of one of the NPT simulations was chosen as a starting point for further simulations at fixed areas of 80, 83, and $86 \text{ \AA}^2/\text{lipid}$. In order to expand the area cross section, short NP γ T simulations were used, with $\gamma = 15 \text{ dyn/cm/leaflet}$ sufficient to attain the target areas. Three matching daidzein conformations were created by modification of the genistein molecules (converting $-\text{OH}$ to $-\text{H}$). Six NPAT simulations were started, with each bioflavonoid simulated at each of the three areas given above. Based on a preliminary comparison to experimental x-ray structure factors, additional daidzein simulations were done at $A=78 \text{ \AA}^2/\text{lipid}$ for mole fraction $f=0.20$ and at $A=79$ and $82 \text{ \AA}^2/\text{lipid}$ for $f=0.14$, as well as a genistein simulation with $A=84.5 \text{ \AA}^2/\text{lipid}$ at $f=0.20$. These 10 NPAT simulations were run for 35 ns, and the final 25 ns of each used for all data analysis.

Results

Volumes

Table 1 shows our measured unit cell volume $V_1(\text{exp})$, consisting of one lipid plus the fraction $f/(1-f)$ of a bioflavonoid. Addition of mole fraction $f=0.2$ bioflavonoids increased $V_1(\text{exp})$ compared to the molecular volumes of pure DOPC and DPhyPC measured previously^{31–33}. Table I also shows that our MD simulations for DOPC give values $V_1(\text{sim})$ that agree very well with $V_1(\text{exp})$. If we assume that the volume of the lipid does not change upon addition of bioflavonoids, then we obtain the volumes $V_{\text{BF}}(\text{calc})$ of genistein and daidzein listed in Table I. Another way to estimate the volumes V_{BF} employs a volumetric analysis of components in simulations³⁴. This gives the larger volumes $V_{\text{BF}}(\text{sim})$ for the bioflavonoids in DOPC shown in Table I, and requires a corresponding condensation of the lipid volumes (not shown).

Structural x-ray Data

The lobes of diffuse scattering from oriented samples are shown numbered in Fig. 2. The diffuse scattering for DOPC at 30 °C extends beyond 0.8 \AA^{-1} with three strong lobes (1–3) and two lobes of weaker scattering not visible in Fig. 2A. The two weaker lobes (4 and 5) are barely visible for DPhyPC in Fig. 2D. It is important to emphasize that accurate measurement of weak or zero intensity increases the spatial resolution of structural data. Our diffuse data contain information equivalent to about 8 orders of Bragg diffraction from rather dry samples and more than the usual 2–4 orders obtained in many investigations of more hydrated samples. There are also sharp peaks corresponding to orders $h=1$ and 2 from which the lamellar D-spacing was obtained; these peaks are barely visible in Fig. 2 because their intensities were reduced by a factor of ~ 1700 by a 0.1 mm thick molybdenum attenuator. As the relative humidity in the sample chamber was increased, the measured D-spacing increased towards the fully hydrated D-spacing determined from MLV samples in excess water in capillaries. These D-spacings did not change significantly with addition of bioflavonoids; their values are shown in Fig. S1. The diffuse x-ray data were collected within 5 \AA of the fully hydrated D-spacing. At the highest concentration ($f=0.20$) of daidzein, Fig. 2C and Fig. 2F show several sharp reflections, presumably due to the presence of daidzein crystals. The measured D-spacing of the strongest sharp reflection was 8.4 \AA , comparable to the length of a tilted daidzein molecule. These reflections were removed for the structural analysis, but this crystallization limited the concentration of bioflavonoid in this study. The ratio of the intensity of the sharp reflections to the intensity of the diffuse scattering was larger for our nominal $f=0.2$ daidzein samples than for our nominal $f=0.15$ daidzein samples. By extrapolating to zero intensity of the sharp rings (data not shown), we estimate that the solubility limit of daidzein in DOPC is $f=0.14$ and in DPhyPC it is $f=0.12$ in these oriented samples. Crystal rings did not appear in any of the genistein samples.

Structural Results

Figure 3 compares the X-ray form factors obtained from MD simulations of 20 mole% genistein in DOPC at three different unit cell areas A_1 . As A_1 increases and the thickness of the bilayer decreases, the simulated lobes of $|F(q_z)|$ move to larger q_z . Comparison to the experimental $|F(q_z)|$ obtained from the data in Fig. 2B indicates that the best simulated value of A_1 is closer to 83 \AA^2 than to 80 \AA^2 or 86 \AA^2 . Similar comparisons of simulations and experiment were performed for DOPC and 20% (14%) daidzein in DOPC to obtain the simulated values of A_1 shown in Table I. Figure 3 also shows $|F(q_z)|$ obtained from modeling. Of course, with a sufficiently complex model with many unconstrained parameters, it is possible to fit the experimental $|F(q_z)|$ quite well, but this yields component distributions in real space that are not realistic. It is therefore important that the model be constrained to be realistic and the simulations are valuable for that, as we show next.

Figure 4 compares the real space electron density distributions of the DOPC lipid components and 20% genistein as obtained from the simulation at $A_1=83 \text{ \AA}^2$ with those obtained from modeling of the experimental $|F(q_z)|$. Although the widths of the model distributions have been constrained, it is important to emphasize that the unconstrained positions z_{BF} of the modeled genistein and the headgroups are nearly the same as those of the simulations. This strongly supports the validity of the simulations. Table I shows the good agreement between the values of the simulated and experimental head-head spacing D_{HH} in the electron density profiles and the average position of both bioflavonoids.

Figure 5 shows the X-ray form factors $|F(q_z)|$ from oriented samples of DOPC and bioflavonoids, and Fig. 6 shows the X-ray form factors from oriented samples of DPhyPC and bioflavonoids that came from the CCD images shown in Fig. 2. The solid curves in Figures 5 and 6 result from the model fits. The vertical dotted lines help compare the effects

of the bioflavinoids on the positions of the zero between lobes 1 and 2. The data for 20% genistein in both DOPC and DPhyPC shifts to higher q_z which is an indication of a thinning of the bilayers.

Figures 7 and 8 show electron density profiles obtained from modeling experimental data. The most robustly determined quantity is the head-to-head thickness, D_{HH} , which is defined as the distance between the maxima in the total electron density profiles. The vertical dotted lines help compare the effects of the bioflavinoids on D_{HH} of DOPC. Results for the highest concentrations of bioflavinoid are given in Table I. Genistein decreased the thickness of DOPC somewhat more than daidzein as shown in Figs. 7 and 8 and quantified in Table 1. With increasing concentration, genistein moved slightly towards the bilayer center, while daidzein moved slightly outwards from the bilayer center. Similar but smaller thinnings were observed when these bioflavinoids were added to DPhyPC as shown in Figs.S2, S3 in SI and quantified in Table 1. Figs S2 and S3 also note slight movements in the bioflavinoids in DPhyPC with increasing concentration.

Elasticity

Results for the bending modulus K_C are shown in Figure 9. The K_C values were averaged from data at different D spacings from the same sample as well as from different samples. For any one sample, K_C did not vary systematically with D -spacing, consistent with it being a property of single bilayers. As shown in Fig. 9, the effect of either bioflavinoid on K_C was relatively small, especially in the case of genistein.

The values of the area compressibility modulus, $K_A = 2A_1(\partial\gamma/\partial A_1)$, given in Table I were obtained from the simulated results for the leaflet surface tension γ in Fig. 10. K_A was clearly smaller with either bioflavinoid than for the DOPC control. As determined from the slopes in Fig. 10, K_A for $f=0.14$ daidzein (190 mN/m) was closer to the control value (240 mN/m) than the K_A for $f=0.20$ genistein (160 mN/m).

Figure 11 shows two snapshots from the final 30 nsec point of the NPAT simulations. Electron density profiles resulting from the simulations at three fixed areas are shown in Fig. S4 in SI.

Discussion

This study emphasizes the synergy between experiments and MD simulations. While the x-ray method provides firm structural data to the highest resolution possible in fully fluid lipid bilayer systems, the extraction of meaningful structural results becomes difficult upon the incorporation of additives that require more modeling parameters. On the other hand, simulations involve uncertainties in the force fields of the additives, and there are concerns that simulation times may not be long enough for additives to equilibrate to their experimental locations. The main test of the simulations in this paper is that the Fourier transforms of the real space electron density profiles agree with the x-ray $|F(q_z)|$ data. Best agreement was obtained by adjusting the unit cell areas A_1 in NPAT simulations and this then provides estimates of A_1 , the head-head thickness D_{HH} , and the positions z_{BF} of the bioflavinoids within the bilayer. The thickness D_{HH} was obtained from the experimental data with a minimal amount of modeling. The good agreement of the simulations for D_{HH} is an important test of our procedure for obtaining A_1 from the simulations. Importantly, for direct modeling of the experimental data, the simulations provide the widths of the distributions of the bioflavinoids and the lipid component groups; this means that the modeling does not require so many totally free parameters. Without imposing simulation results for A_1 or z_{BP} , the modeling of the experimental $|F(q_z)|$ data gives good agreement with the simulations for those quantities.

We find both genistein and daidzein located closer to the headgroup than to the center of the bilayer as qualitatively shown in Fig. 11. For quantitative evaluation of differences, we use our values of $D_{HH}/2$ as a measure for the surface of the bilayer. Then, the depth of insertion $D_{HH}/2 - z_{BF}$ from Table I is only ~ 0.1 Å greater for genistein than for daidzein in DOPC although it is ~ 1.1 Å greater in DPhyPC. These small differences in depth of penetration are consistent with genistein being more hydrophobic than daidzein. At first, this seems contrary to genistein having an additional OH group. However, that extra OH group can participate in intra-molecular hydrogen bonding with the adjacent carbonyl (Fig. 1), thereby decreasing the hydrophilicity of the carbonyl on genistein relative to that of daidzein. Our simulation indicates this intramolecular hydrogen bond is formed 95% of the time. Intramolecular hydrogen bonding in genistein is supported by ^1H NMR linewidth experiments of these bioflavonoids in SDS micelles³⁵ and the relative hydrophobicities are supported by the partitioning of genistein into octanol ($\log P = 3.04$) being greater than for daidzein ($\log P = 2.51$)³⁶. Consistently, the C=O was associated with a water hydrogen 50% of the time for daidzein and only 25% of the time for genistein. Histograms of the bioflavonoid center of mass $|z|$ coordinate (Fig. S5 in SI) also suggest genistein is slightly more hydrophobic; the distribution outliers, shown by enlarging the baseline in the inset, are inward for genistein and outward for daidzein.

It is also especially encouraging that the difference between the volumes $V_{BF}(\text{calc})$ of the two bioflavonoids shown in Table I are the same in DOPC and DPhyPC and that these differences are reasonable given the replacement of COH in genistein by CH in daidzein. However, it seems unlikely that the volume of either bioflavonoid is so much different, as given in Table I, in DPhyPC compared to DOPC. More likely is that either bioflavonoid alters the volume of different lipids by different amounts. For example, if genistein condensed DOPC to 1294 \AA^3 and did not condense DPhyPC at all, then its volume would be 363 \AA^3 in both lipids. We attempted to obtain some perspective on this from our simulations. The fact that the simulations agreed so well with the experimental 'unit cell' volumes in Table I is another important test of the simulations. This success suggested trying to obtain the bioflavonoid volume using a well established component volume method. However, as noted in the original paper³⁴, high accuracy cannot be expected from this method when there are closely overlapping distributions; indeed the results for $V_{BF}(\text{sim})$ listed in Table I yield the unlikely result that genistein is much larger than daidzein. Nevertheless, these results are consistent with the bioflavonoids condensing rather than expanding DOPC. We propose that the values for $V_{BF}(\text{calc})$ shown in Table I for DPhyPC are likely to be good estimates when bioflavonoids are added to most lipid bilayers.

Experimental validation of the simulation allows us to extract additional structural information that cannot be obtained from our experimental data. The narrow distributions of the bioflavonoids along the z direction (Fig. S5) are consistent with their long axes being preferentially parallel to the bilayer surface (see Figure 1 for definition of axes). This is further confirmed in the simulation by the two hydroxyl oxygens at opposite ends of the bioflavonoids having nearly equal average distances from the center of the bilayer and by directly obtaining the angle β of the long axes relative to the bilayer normal, shown in Fig. 12B. Hydrogen bonding of these two hydroxyls to water or phosphate in the interfacial region is the likely interaction responsible for this orientation, as well as for maintaining the location of the bioflavonoids close to the bilayer surface. The simulation also provides the angle α of the short axes of the bioflavonoids around their long axes. The distribution of α , shown in Fig. 12A, is peaked about $\alpha=0$ which we define to be the value of α at which the plane of the rings is perpendicular to the bilayer surface and the C=O points towards the proximal bilayer surface.

Study of the elastic properties also benefits from the synergy of experiment and simulation in that our experiments provide the bending modulus K_C and our simulations provide the area compressibility modulus K_A . These two moduli are often related by

$$K_C = K_A (2D_C)^2 / N \quad (3)$$

where $2D_C$ is the hydrocarbon thickness. The value $N=24$ from the polymer brush theory fits experimental data for pure lipid bilayers quite well³⁷, although the relation breaks down as cholesterol is added³⁸. We observed that, within uncertainty, both bioflavonoids decreased K_A relative to the control DOPC by about the same 40% when either bioflavonoid concentration was 20% and proportionately less with 14% daidzein. In contrast, K_C remained nearly constant for genistein and decreased only about 20% for daidzein, so satisfying Eq. (3) would require D_C to increase considerably. However, if we define D_C to include only the hydrocarbons of the lipid, then our simulated D_C decreases from 13.0 Å for DOPC alone to 11.8 Å with 20% genistein and to 12.6 Å with 14% daidzein. Even if we define D_C to include the largely hydrophobic bioflavonoids, D_C remains at 13.0 Å with 20% genistein and increases to 13.6 Å with 20% daidzein, so we conclude that Eq. (3) breaks down. Interestingly, Eq. (3) breaks down in the opposite way as for addition of cholesterol³⁸. For DOPC, cholesterol increases K_A and $2D_C$ while keeping K_C the same, so the right hand side of Eq. (3) becomes larger than the left hand side for cholesterol, rather than smaller as we find for bioflavonoids. This contrasting elastic behavior appears to be correlated with the structural difference that the long axis of cholesterol is preferentially oriented parallel to the bilayer normal whereas the long axis of the bioflavonoids is preferentially oriented parallel to the bilayer surface.

The largest difference that we observe between the effects of genistein and daidzein on the DOPC bilayer, both in modeling of experimental data and in our simulations, is in the area of the unit cell A_1 . The average increase of $A_1(\text{exp})$ and $A_1(\text{sim})$ for 20 mole% genistein is ~12% increase and ~8% for daidzein at its solubility limit of 14 mole%. In DPhyPC the increase of $A_1(\text{exp})$ is also ~12% increase for 20 mole% genistein and ~8% for 12% daidzein at its solubility limit. Therefore, the area increase is proportional to the concentration of either bioflavonoid in either lipid. These increases may be compared to the 6% increase reported for 20 mole% of another bioflavonoid, curcumin, added to DOPC³⁹.

One motivation for this work was to test the attractive hypothesis that the increased gA ion-channel lifetimes induced by bioflavonoids could be due to genistein reducing the energy of hydrophobic mismatch of the gA channel to the lipid bilayer by reducing the appropriate elastic moduli, thereby making the bilayer softer and more easily deformable. By consideration of the details of deformations, the appropriate elasticity moduli have been proposed to be $K_C/40$ and both K_C and $K_A/41$. A simpler expression has been given^{14, 42-44} as

$$\Delta G_{\text{bilayer}} = H_B (L - d_0)^2, \quad (4)$$

where L is hydrophobic channel length, d_0 is bilayer hydrocarbon thickness and H_B is a phenomenological spring constant to mimic the effects of both K_C and K_A . Decreasing K_C and/or K_A would decrease H_B and $\Delta G_{\text{bilayer}}$, which, according to the hypothesis, would then increase channel lifetimes, as was observed. The hypothesis is therefore broadly consistent, for either bioflavonoid, with our results for K_A and K_C . One might also consider that bioflavonoids could induce monolayer intrinsic curvature, but Lundbaek et al. reported that monolayer intrinsic curvature does not play a major role¹².

Another important hypothesis that explains how bioflavonoids could increase gA channel lifetimes follows from our result for D_{HH} in Table 1 that bioflavonoids thin lipid bilayers. The hydrocarbon thickness d_0 has been given as $2D_C=27.1 \text{ \AA}$ for DOPC22 and 27.2 \AA for DPhyPC⁴⁵ and the hydrophobic channel length L of wild-type gA has been given as 22 \AA ¹³, so bioflavonoids would reduce the hydrophobic mismatch free energy in Eq. (4) by reducing $L-d_0$. Of course, the average thinning will be smaller for the lower concentrations used in gA experiments¹⁴, but local thinning near a bioflavonoid could be comparable to the results in Table 1. Such thin local regions would then provide an effectively attractive interaction between the bioflavonoids and gA dimers. Once formed, gA dimers with neighboring bioflavonoids would be relatively more stable with respect to breaking into gA monomers and this would increase channel lifetimes.

However, neither of the preceding hypotheses is consistent with the reported experimental result that genistein is twice as effective as daidzein for increasing gA channel lifetimes¹⁴. Regarding the first hypothesis, our results are that the decrease in K_A is nearly the same for both bioflavonoids up to the solubility limit of daidzein and that there is an even larger decrease in K_C for daidzein on a per mole basis, so daidzein would be expected to be even more effective than genistein for increasing channel lifetimes. Regarding the second hypothesis, we find that thinning is nearly the same for daidzein and genistein on a per mole basis. As the concentration of daidzein in the gA lifetime experiments was about four times smaller than the limiting mole fraction $f=0.14$ (calculation given in supplementary material), the solubility limit does not account for the difference in lifetime results. Instead, the reason that genistein has a greater effect than daidzein on channel lifetimes is much simpler. The bioflavonoid concentrations reported in the gA channel lifetime experiments were based on the total amount of gA added to the system consisting of a black lipid membrane with its associated decane annulus and approximately 1000 times as much water by volume. Because genistein is more hydrophobic than daidzein, more of it partitioned into the bilayer when the overall reported concentrations were equal. From reported values of the partition coefficient, the $\log P$ of 3.04 for genistein and 2.51 for daidzein³⁶ would give three times as much genistein as daidzein in the bilayer for equal aqueous concentrations. However, the finite ratio of aqueous to bilayer volume in the gA experiments¹⁴ alters the relative concentrations in the bilayer from a factor of 3 to a factor of 2 when equal overall amounts of bioflavonoid are added (see supporting material for the calculation). This alone accounts for the differences in the effect of the bioflavonoids genistein and daidzein on gA channel lifetimes.

Summary and Conclusions

This work combines X-ray diffuse scattering, volume measurements, and MD simulations to determine structural and elastic properties of DOPC and DPhyPC bilayers with the bioflavonoids genistein and daidzein. The unit cell volume, V_1 , obtained by direct measurements of lipid/bioflavonoid mixtures is in excellent agreement with V_1 obtained from the MD simulations. There is good agreement between the electron density profiles and bioflavonoid positions obtained by MD simulations and model fits to the experimental form factor data. Both genistein and daidzein are located at the surface of the hydrocarbon region near the glycerol/carbonyl with the long axis parallel to the bilayer and the C=O preferentially pointing towards the bilayer surface. Genistein thins DOPC by $\sim 2.3 \text{ \AA}$ at 20 mole %, while daidzein thins DOPC by $\sim 1.5 \text{ \AA}$ at 14 mole %. Consistent with a theory for the effect of bioflavonoids on gramicidin lifetimes, both bioflavonoids generally softened bilayers. However, since the bending modulus K_C decreased more with daidzein than with genistein and K_A decreased the same for both on a per mole basis, softening of the membrane is not the reason that genistein increases gA channel lifetimes twice as much as daidzein. Similarly, our result that both bioflavonoids thin the bilayer is consistent with the

other hypothesis that bioflavonoids alleviate the hydrophobic free energy penalty for forming gA channels, but our result that the thinning is the same on a per mole basis is inconsistent with the larger reported effect of genistein. That factor of two is quantitatively explained by the relative water/hydrocarbon partition coefficients³⁶. Our simulation results and our experimental solubility limit results for daidzein are consistent with the partition coefficient result that genistein is more hydrophobic than daidzein.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This research was supported in part by Grant Number GM 44976 from NIGMS/NIH (STN,JN), the Howard Hughes Medical Institute (MR, YZ) and the Charles E. Kaufman Foundation (STN). It was supported in part (RV, RP) by the Intramural Research Program of the NIH, National Heart, Lung and Blood Institute, and utilized the high-performance computational capabilities at the National Institutes of Health, Bethesda, MD (NHLBI LoBoS cluster). X-ray scattering data were taken at the Cornell High Energy Synchrotron Source (CHESS), which is supported by the National Science Foundation and the National Institutes of Health/National Institute of General Medical Sciences under National Science Foundation award DMR-0225180. We especially thank Dr. Arthur Woll for obtaining our beam and for general support during our data collection at the G1 station. We thank Dr. Jianjun Pan for help with sample preparation and x-ray data collection at CHESS, Prof. Olaf Andersen for helpful discussions, and especially Dr. Gilman Toombes for very insightful criticism of a first draft.

Bibliography

1. Dixon RA, Ferreira D. *Phytochemistry*. 2002; 60:205–211. [PubMed: 12031439]
2. Birt DF, Hendrich S, Wang WQ. *Pharmacol Therapeut*. 2001; 90:157–177.
3. Derand R, Bulteau-Pignoux L, Becq F. *J Biol Chem*. 2002; 277:35999–36004. [PubMed: 12124395]
4. Akiyama T, Ishida J, Nakagawa S, Ogawara H, Watanabe S, Itoh N, Shibuya M, Fukami Y. *J Biol Chem*. 1987; 262:5592–5595. [PubMed: 3106339]
5. Bishop JM. *Annu Rev Biochem*. 1983; 52:301–354. [PubMed: 6351725]
6. Coward L, Barnes NC, Setchell KDR, Barnes S. *J Agr Food Chem*. 1993; 41:1961–1967.
7. Kuiper GGJM, Carlsson B, Grandien K, Enmark E, Haggblad J, Nilsson S, Gustafsson JA. *Endocrinology*. 1997; 138:863–870. [PubMed: 9048584]
8. Kuiper GGJM, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, van der Burg P, Gustafsson JA. *Endocrinology*. 1998; 139:4252–4263. [PubMed: 9751507]
9. Rickard DJ, Subramaniam M, Spelsberg TC. *J Cell Biochem*. 1999:123–132. [PubMed: 10629111]
10. Fanti P, Monier-Faugere MC, Geng Z, Schmidt J, Morris PE, Cohen D, Malluche HH. *Osteoporosis Int*. 1998; 8:274–281.
11. Palacios VG, Robinson LJ, Borysenko CW, Lehmann T, Kalla SE, Blair HC. *J Biol Chem*. 2005; 280:13720–13727. [PubMed: 15644335]
12. Lundbaek JA, Birn P, Tape SE, Toombes GES, Sogaard R, Koeppe RE, Gruner SM, Hansen AJ, Andersen OS. *Mol Pharmacol*. 2005; 68:680–689. [PubMed: 15967874]
13. Hwang TC, Koeppe RE, Andersen OS. *Biochemistry-US*. 2003; 42:13646–13658.
14. Lundbaek JA, Koeppe RE, Andersen OSP. *Natl Acad Sci USA*. 2010; 107:15427–15430.
15. Lehtonen JYA, Adlercreutz H, Kinnunen PKJ. *Biochimica Et Biophysica Acta-Biomembranes*. 1996; 1285:91–100.
16. Kučerka N, Nagle JF, Sachs JN, Feller SE, Pencer J, Jackson A, Katsaras J. *Biophys J*. 2008; 95:2356–2367. [PubMed: 18502796]
17. Tristram-Nagle SA. *Methods Mol Biol*. 2007; 400:63–75. [PubMed: 17951727]
18. Kučerka N, Liu YF, Chu NJ, Petrache HI, Tristram-Nagle ST, Nagle JF. *Biophys J*. 2005; 88:2626–2637. [PubMed: 15665131]

19. Lyatskaya Y, Liu YF, Tristram-Nagle S, Katsaras J, Nagle JF. *Phys Rev E*. 2001; 63:0119071–0119079.
20. Liu YF, Nagle JF. *Phys Rev E*. 2004; 69:040901–040904(R).
21. Kučerka N, Tristram-Nagle S, Nagle JF. *Biophys J*. 2006; 90:L83–L85. [PubMed: 16617085]
22. Nagle JF, Tristram-Nagle S. *Biochim Biophys Acta-Reviews on Biomembranes*. 2000; 1469:159–195.
23. Nagle JF, Wiener MC. *Biophys J*. 1989; 55:309–313. [PubMed: 2713444]
24. Klauda JB, Kučerka N, Brooks BR, Pastor RW, Nagle JF. *Biophys J*. 2006; 90:2796–2807. [PubMed: 16443652]
25. Wiener MC, Suter RM, Nagle JF. *Biophys J*. 1989; 55:315–325. [PubMed: 2713445]
26. Brooks BR, Brooks CL 3rd, Mackerell AD Jr, Nilsson L, Petrella RJ, Roux B, Won Y, Archontis G, Bartels C, Boresch S, et al. *J Comput Chem*. 2009; 30:1545–1614. [PubMed: 19444816]
27. Klauda JB, Venable RM, Freites JA, O'Connor JW, Tobias DJ, Mondragon-Ramirez C, Vorobyov I, MacKerell AD Jr, Pastor RW. *J Phys Chem B*. 2010; 114:7830–7843. [PubMed: 20496934]
28. Jorgensen WL, Chandrasekhar J, Madura JD, Impey RW, Klein ML. *J Chem Phys*. 1983; 79:926–935.
29. Burendahl S, Danciulescu C, Nilsson L. *Proteins*. 2009; 77:842–856. [PubMed: 19626711]
30. Kučerka N, Katsaras J, Nagle JF. *J Membr Biol*. 2010; 235:43–50. [PubMed: 20407764]
31. Greenwood AI, Tristram-Nagle S, Nagle JF. *Chem Phys Lipids*. 2006; 143:1–10. [PubMed: 16737691]
32. Tristram-Nagle S, Petrache HI, Nagle JF. *Biophys J*. 1998; 75:917–925. [PubMed: 9675192]
33. Tristram-Nagle S, Kim DJ, Akhuzada N, Kucerka N, Mathai JC, Katsaras J, Zeidel M, Nagle JF. *Chem Phys Lipids*. 2010; 163:630–637. [PubMed: 20447383]
34. Petrache HI, Feller SE, Nagle JF. *Biophys J*. 1997; 72:2237–2242. [PubMed: 9129826]
35. Whaley WL, Rummel JD, Kastrapeli N. *Langmuir*. 2006; 22:7175–7184. [PubMed: 16893213]
36. Rothwell JA, Day AJ, Morgan MRA. *J Agr Food Chem*. 2005; 53:4355–4360. [PubMed: 15913295]
37. Rawicz W, Olbrich KC, McIntosh T, Needham D, Evans E. *Biophys J*. 2000; 79:328–339. [PubMed: 10866959]
38. Pan JJ, Tristram-Nagle S, Nagle JF. *Phys Rev E*. 2009; 80:021931 (021931–021912).
39. Sun Y, Lee CC, Hung WC, Chen FY, Lee MT, Huang HW. *Biophys J*. 2008; 95:2318–2324. [PubMed: 18515370]
40. Huang HW. *Biophys J*. 1986; 50:1061–1070. [PubMed: 2432948]
41. Harroun TA, Heller WT, Weiss TM, Yang L, Huang HW. *Biophys J*. 1999; 76:3176–3185. [PubMed: 10354442]
42. Lundbaek JA. *J Gen Physiol*. 2008; 131:421–429. [PubMed: 18411332]
43. Lundbaek JA. *J Phys-Condens Mat*. 2006; 18:S1305–S1344.
44. Lundbaek JA, Collingwood SA, Ingolfsson HI, Kapoor R, Andersen OS. *J R Soc Interface*. 2010; 7:373–395. [PubMed: 19940001]
45. Tristram-Nagle S, Kim DJ, Akhuzada N, Kucerka N, Mathai JC, Katsaras J, Zeidel M, Nagle JF. *Chem Phys Lipids*. 2010; 163:630–637. [PubMed: 20447383]

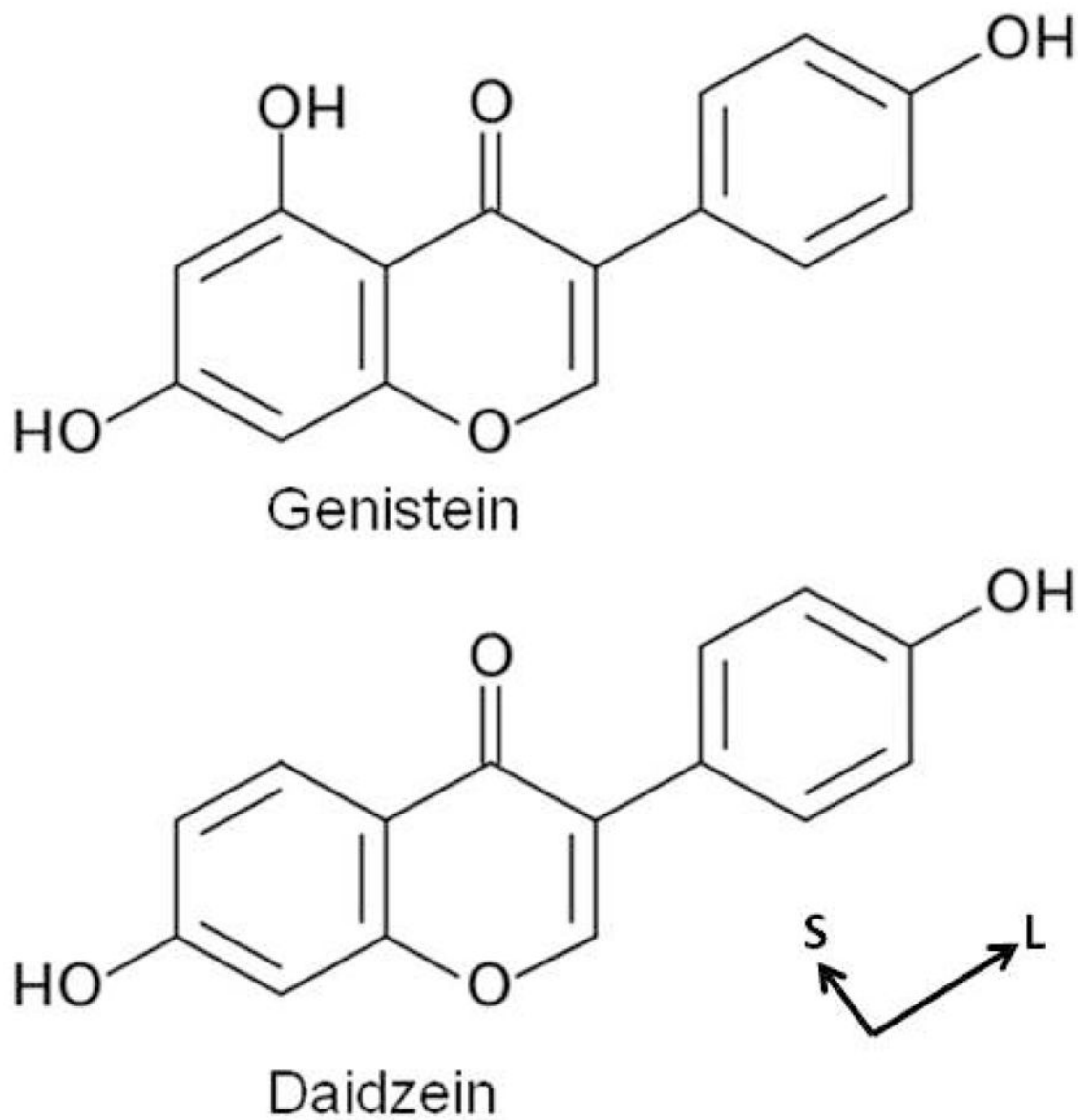


Figure 1. Chemical structures of the bioflavonoids, genistein and daidzein. The long (L) and short (S) axes of the bioflavonoids are shown.

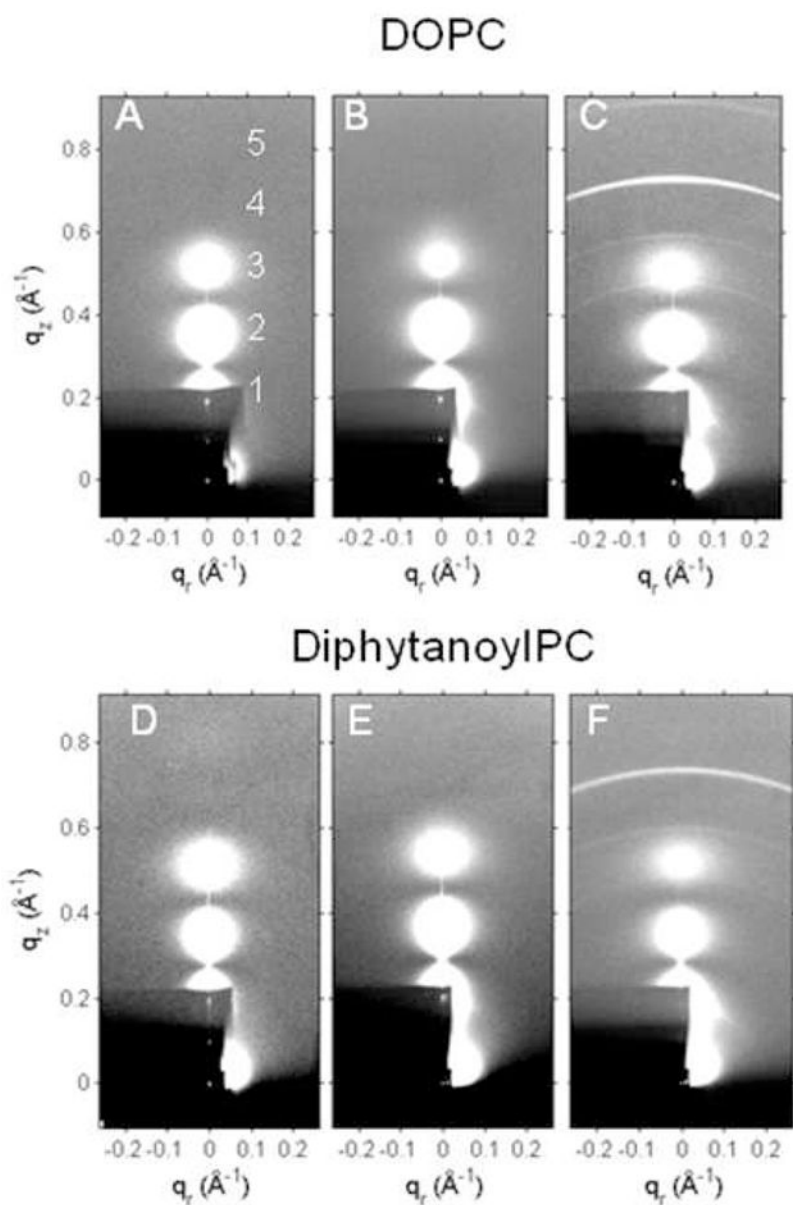


Figure 2. 2D CCD images of LAXS diffuse scattering, white is highest intensity. A. DOPC, B. DOPC/20% genistein, C. DOPC/20% daidzein, D. DPhyPC, E. DPhyPC/20% genistein, F. DPhyPC/20% daidzein. The dark shadows are caused by attenuators through which the beam and orders 1 and 2 can be seen.

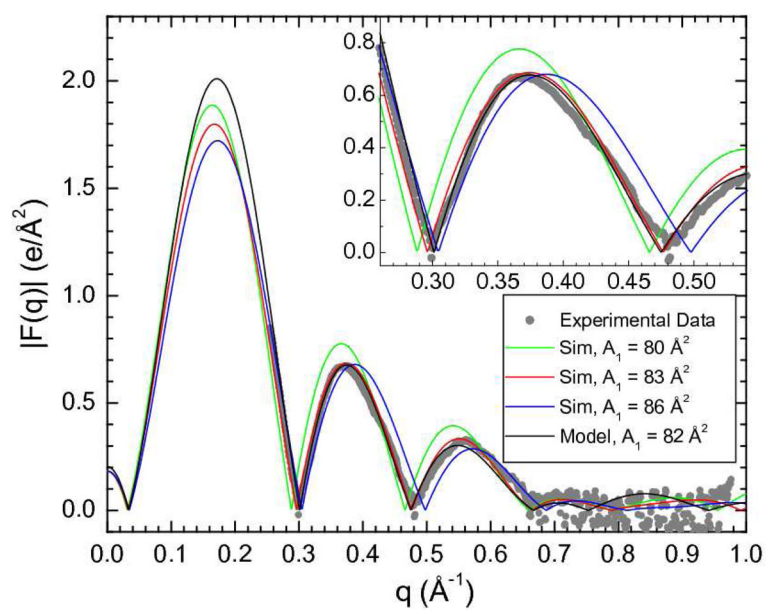


Figure 3.

Experimental form factor $|F(q_z)|$ data for 20 mole% genistein in DOPC (grey circles) compared to the form factors resulting from three MD NPAT simulations fixed to areas A_1 of 80\AA^2 (green), 83\AA^2 (red) and 86\AA^2 (blue). Also shown is the model fit (black) that yielded $A_1 = 82 \text{\AA}^2$. The inset shows more detail in the second lobe.

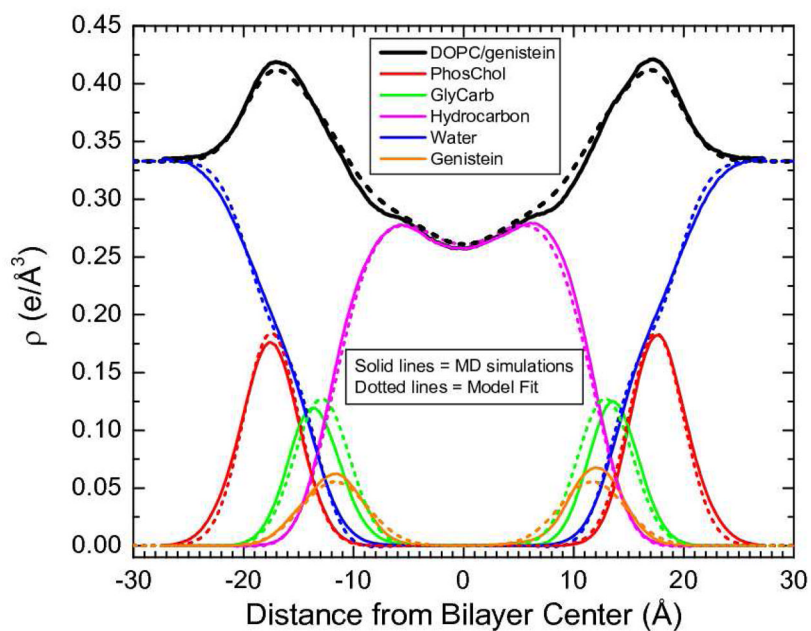


Figure 4. Electron density profiles for DOPC with 20% genistein resulting from MD simulation at $A_1=83 \text{ \AA}^2$ (solid lines) and model fitting to experimental data (dashed lines). Component groups are identified by colors shown in the legend.

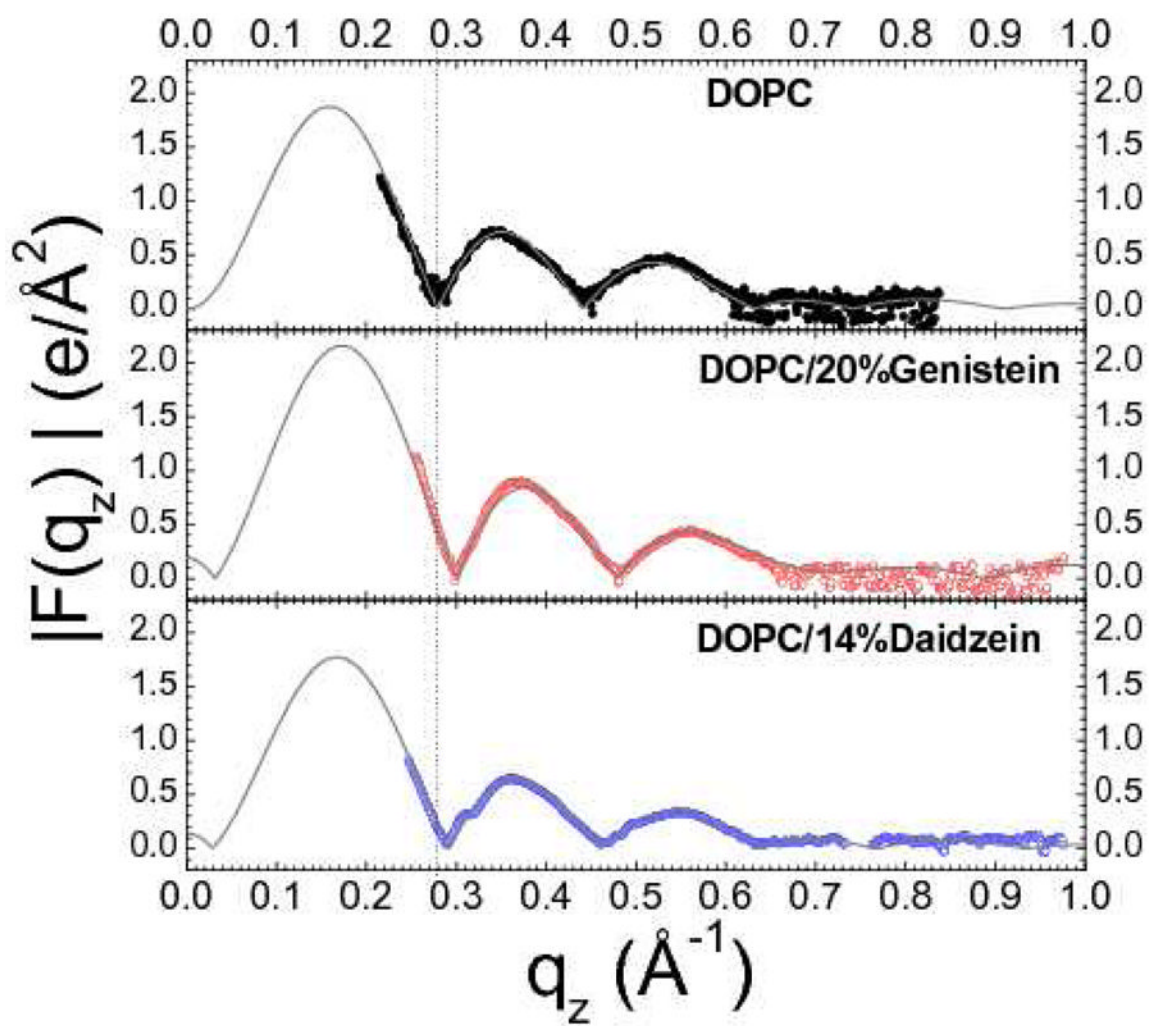


Figure 5. (left) $|F(q_z)|$ data for DOPC (black), DOPC/20% genistein (red) and DOPC/14% daidzein (blue). Lobes are numbered for DOPC.

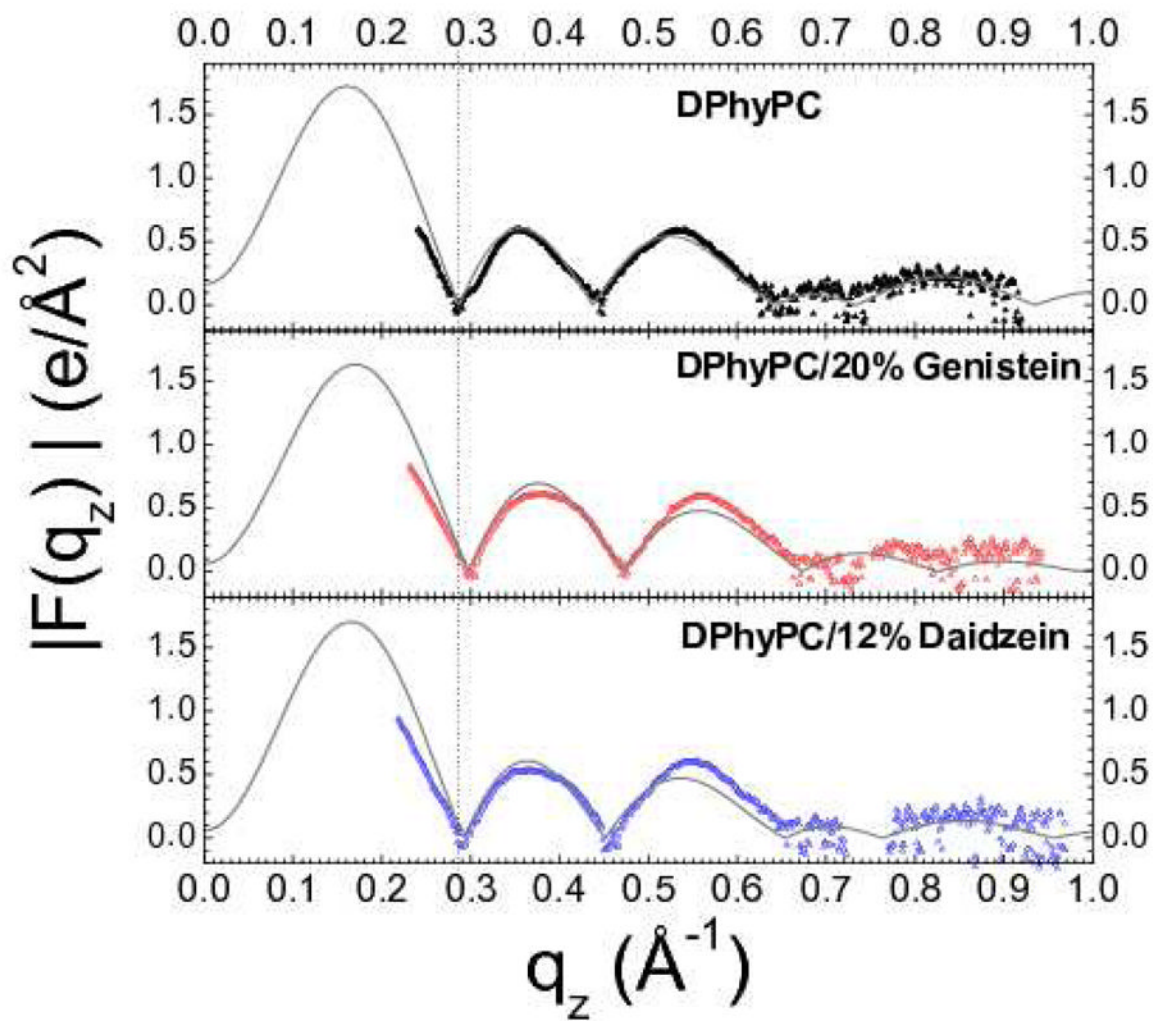


Figure 6.
(right) $|F(q_z)|$ data for DPhyPC (black), DPhyPC/20% genistein (red) and DPhyPC/12% daidzein (blue).

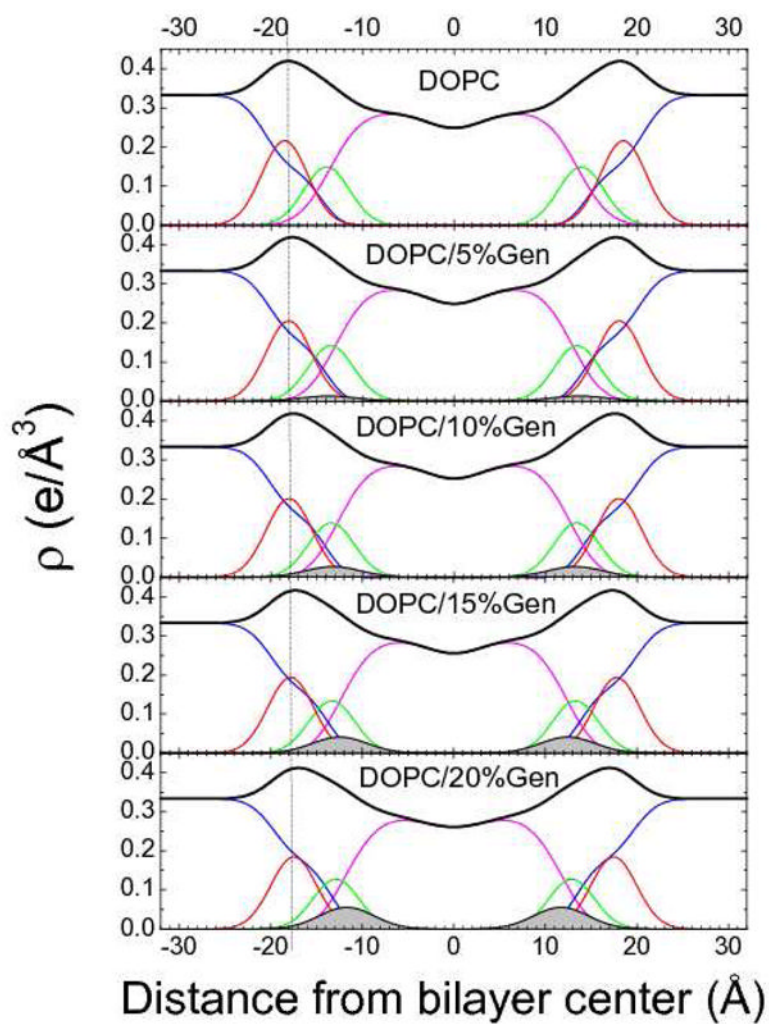


Figure 7. (left). Electron density profiles of DOPC with increasing concentration of genistein obtained using the SDP procedure. Component groups are phosphate (red), carbonyl-glycerol (green), methylenes and terminal methyl group (magenta), water (blue), bioflavinoid (filled grey) and total (black).

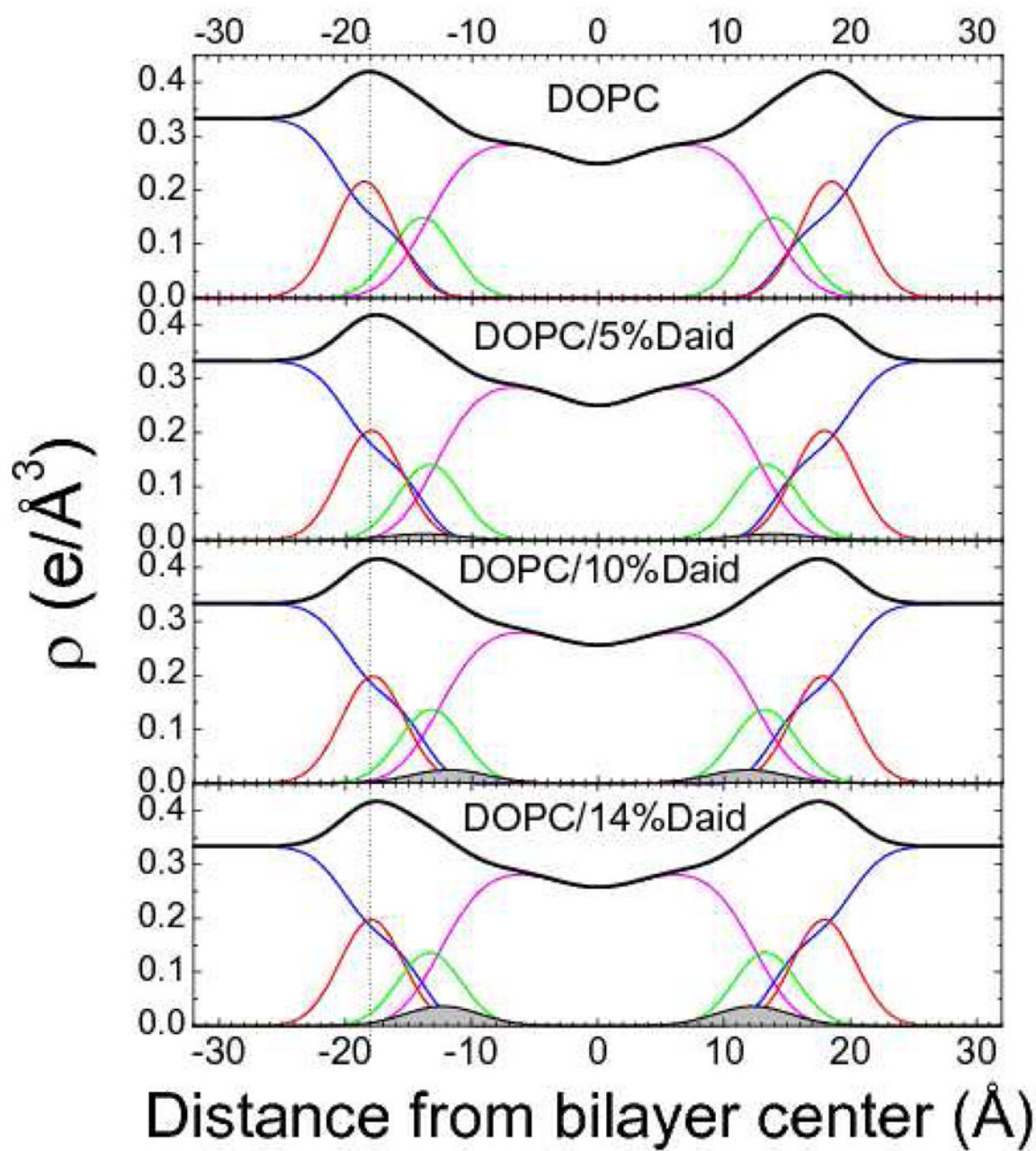


Figure 8. (right). Electron density profiles of DOPC with increasing concentration of daidzein obtained using the SDP procedure. Line colors as for Figure 7.

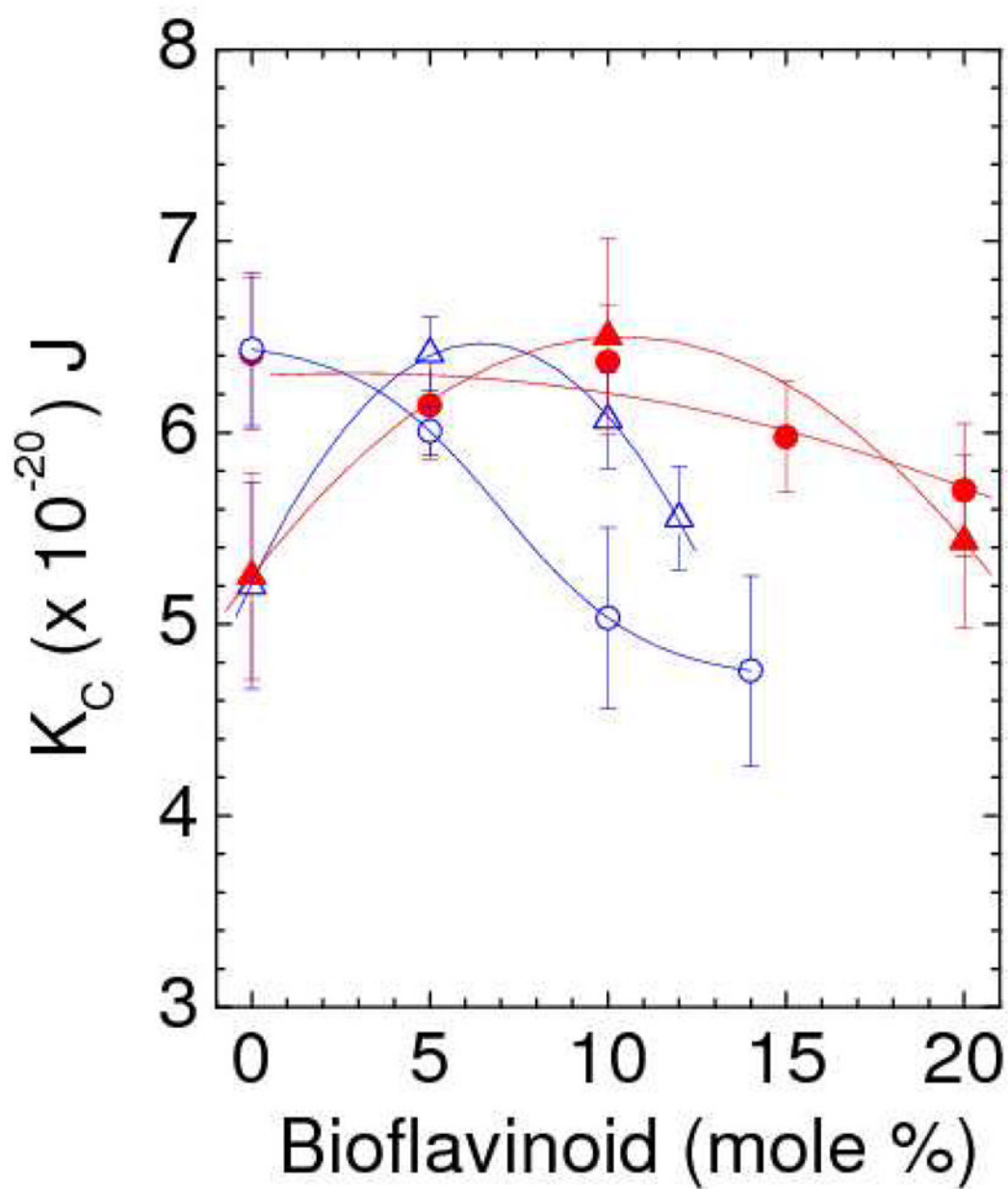


Figure 9. Bending modulus, K_C , vs. concentration of genistein (solid symbols) and daidzein (open symbols) in DOPC (circles) and DPhyPC (triangles).

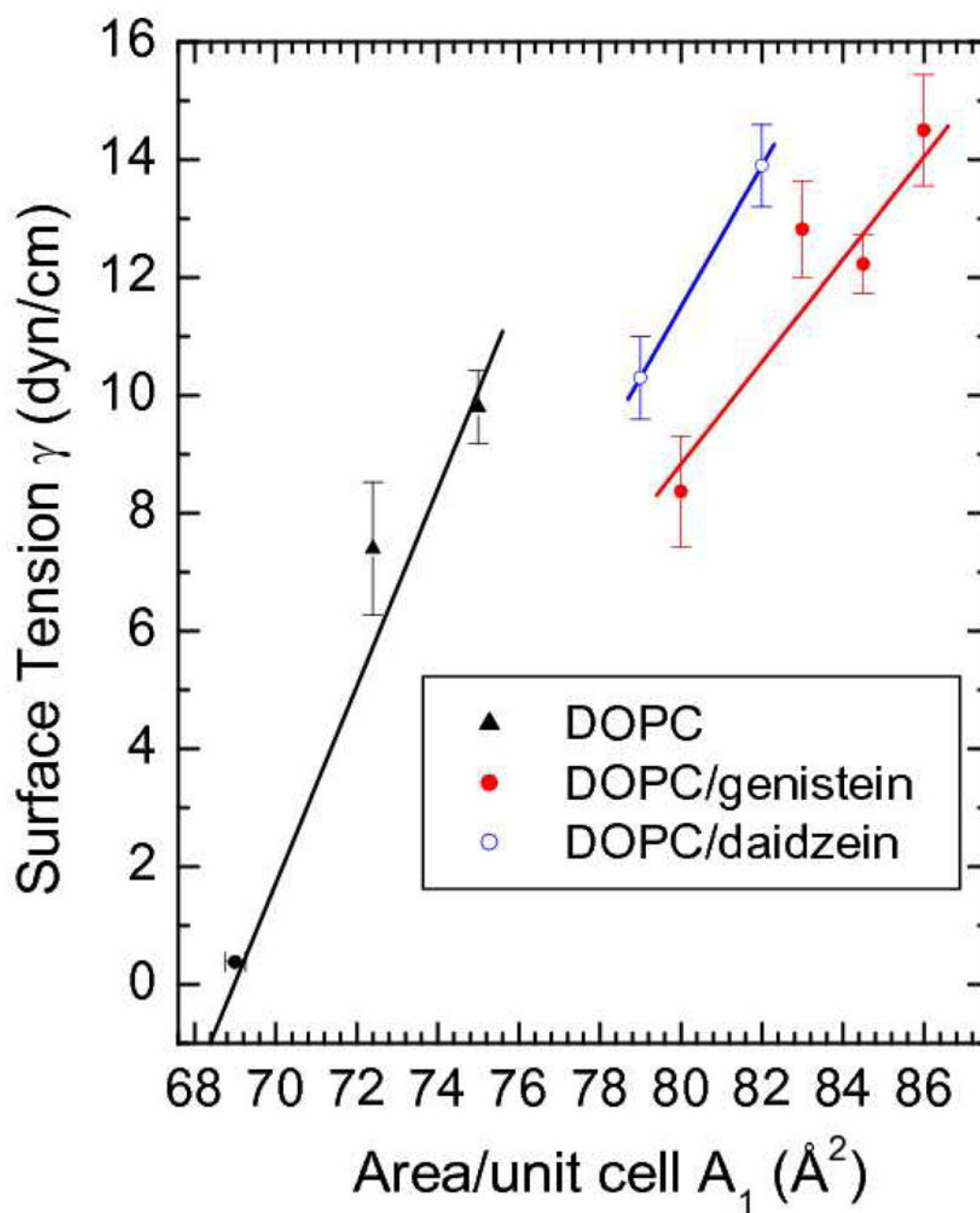


Figure 10. Surface tension, γ , vs. A_1 for DOPC with and without 20 mole% genistein and 14 mole% daidzein from NPAT simulations except the $\gamma=0$ point for pure DOPC which was an NPT simulation.

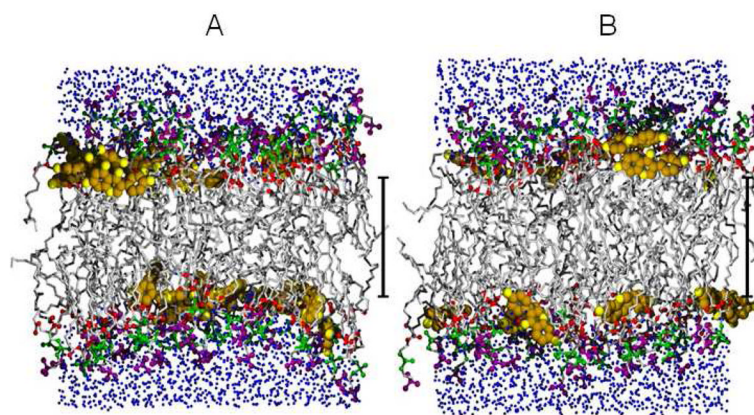


Figure 11. Snapshots of the MD simulations for A) DOPC/20 mole% genistein at $A_1 = 83 \text{ \AA}^2$, and for B) DOPC/14 mole% daidzein at $A_1 = 79 \text{ \AA}^2$. Each snapshot is 1 out of 25,000 coordinate sets that comprise the ensemble; A) collected at 30 nsecs (5 nsecs before the end of the simulation) and B) 35 nsecs. Color coding is: hydrocarbon chains (grey), bioflavonoids (gold and with yellow oxygen atoms), lipid carbonyl oxygens (red), phosphate group (green), choline group (purple) and water (blue). The scale bar marks 20 Å.

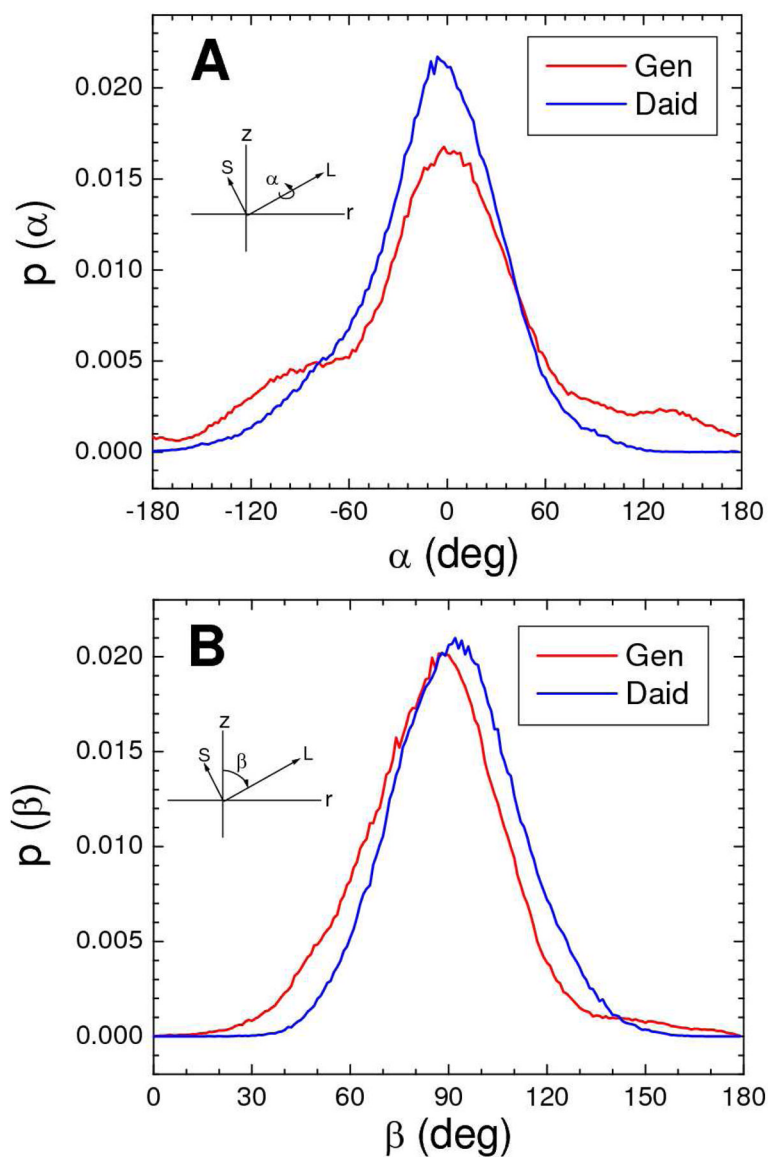


Figure 12. Probability histograms of bioflavonoids in DOPC membranes from the MD simulations for (A) the angle α of the short axis about the long axis and (B) the angle β of the long axis with respect to the bilayer normal. Colors are 20 mole% genistein (red) and 14 mole % daidzein (blue). The insets show α and β with respect to the long (L) and short (S) axes of the bioflavonoids, depicted in Figure 1.

Table 1

Tabulated Results*

lipid	DOPC			DPhyPC		
	none	Gen	Daid	none	Gen	Daid
bioflavonoid						
f	0	0.20	0.14	0	0.20	0.12
$V_1(\text{exp})$	1303	1385	1380 ^a	1427	1518	1513 ^a
$V_1(\text{sim})$	1304	1384	1380 ^a			
$V_{\text{BF}}(\text{calc})$		328	308 ^a		363	343 ^a
$V_{\text{BF}}(\text{sim})$		437	352 ^a			
$A_1(\text{exp})$	72	82	79	83	93	88
$A_1(\text{sim})$	75	83	80			
$D_{\text{HH}}(\text{exp})$	36.1	33.8	34.6	36.4	35.0	35.5
$D_{\text{HH}}(\text{sim})$	36	33.9	34.5			
$Z_{\text{BF}}(\text{exp})$		11.8	12.3		13.0	13.4
$Z_{\text{BF}}(\text{sim})$		11.8	12.3			
$K_A(\text{sim})$	250	160	190			
$K_C(\text{exp})$	6.5	5.7	4.8	5.3	5.4	5.5

* Units are appropriate powers of Å, mN/m for K_A , 10–20 J for K_C , f is mole fraction of bioflavonoid, Gen is genistein and Daid is daidzein, and subscript 1 signifies a unit cell consisting of 1 lipid and f/(1–f) bioflavonoid.

^a indicates results for f = 0.20.