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***Staphylococcus aureus*: an introduction**

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Shortly after finishing his undergraduate studies at the University of Aberdeen in Scotland and embarking on his career as a surgeon, Alexander Ogston presented in 1880 at the Ninth Surgical Congress in Berlin his work establishing the causative role of bacteria in wound infection and subsequent septicemia. Building on the teachings of his senior contemporaries, Louis Pasteur and Joseph Lister, Ogston had observed pus from 88 human abscesses under his microscope and noted Gram-positive spherical “micrococci” [1]. Taking from the Greek word for “bunches of grapes,” he named the organism *Staphylococcus*. After injecting the isolated bacteria into healthy guinea pigs and mice and recreating the abscesses from which the isolates were derived, he had conclusively introduced the world to the infectious agent, now known as *Staphylococcus aureus* due to its golden color in culture, that continues to burden human health today [1].

For decades, treatment options were limited to the topical application of carbolic acid (phenol), the same antiseptic technique introduced by Lister and promoted by Ogston [2]. The excitement that accompanied the introduction of penicillin in the 1940s turned into concern only 2 years later when resistance to the once-heralded miracle drug first appeared in penicillinase-producing strains of *S. aureus*, and then into surrender by 1950 when the frequency of resistance mandated alternative therapy [3–7]. Strains of *S. aureus* with an altered penicillin-binding protein soon countered the semi-synthetic beta-lactam antibiotics that had replaced penicillin, with methicillin-resistant *Staphylococcus aureus* (MRSA) first described in 1961 [5, 8]. After decades of being a predominantly hospital-associated infection, in the twenty first century, MRSA has spread beyond hospitals to become a significant “community-associated” public health burden, as outlined by Fowler and colleagues in this special issue, that threatens to once again outpace antibiotic development.

Beyond antibiotic resistance, the success of *S. aureus* as a human pathogen stems from the arsenal of virulence factors it has evolved to combat host defense mechanisms. The battle over metal ions is an often-overlooked case in point, detailed here by Skaar and colleagues. For example, the body sequesters essential iron to limit bacterial growth [9–11]. However, staphylococcal siderophores overcome host attempts to starve it of iron from both heme [12] and transferrin [13]. *S. aureus* also uses the pore-forming toxin alpha-hemolysin to lyse red blood cells and access hemoproteins [14, 15]. Similarly, *S. aureus* has evolved incompletely examined mechanisms to circumvent host sequestration of other essential nutrients such as manganese, zinc, and copper.

The more conventional battle at the front lines of innate immunity places neutrophils at the center of the host response. *S. aureus* infection promotes neutrophil recruitment through induction of a variety of cytokines, chemokines, and chemokine receptors, such as IL-17 [16], CXCL8 [17], and CXCR2 [18]. Once engaged, neutrophils can phagocytose opsonized

bacteria and trigger oxidative killing and other antimicrobial mechanisms [19], reviewed here by DeLeo and colleagues. The importance of neutrophils in the defense against staphylococcal infections is apparent in the increased susceptibility of patients with abnormalities in neutrophil numbers, such as congenital or chemotherapy-induced neutropenia [20–22]. Other susceptible patients include those with defects in functional oxidative burst (chronic granulomatous disease [20], myeloperoxidase deficiency [22], specific granule deficiency [23]), chemotaxis (leukocyte adhesion deficiency type I [22]), or both (diabetes mellitus [24], end-stage renal failure [25]). To counter neutrophil attack, *S. aureus* produces factors to limit chemotaxis [26], extravasation [27], and superoxide function [28]. It can induce neutrophil apoptosis with its Pantone–Valentine leukocidin [29, 30], whose contribution to the virulence of epidemic community-associated clones has generated recent controversy that is addressed in several articles in this issue. Inside neutrophils, *S. aureus* can survive for days before lysing the cell and escaping back into the surrounding tissue [29]. Even the golden color seen in culture that inspired the “aureus” namesake has revealed itself to be an antioxidant virulence factor that neutralizes oxidative killing by neutrophils [31].

The recruitment and activation of neutrophils during staphylococcal infection is triggered by the activation of pattern recognition receptors, such as toll-like receptors (TLR), on keratinocytes and other cells involved in the innate immune response [32]. Upon activation, these cells also secrete directly bactericidal products such as human beta defensins [33], cathelicidin (LL-37) [34], and RNase 7 [35]. Atopic dermatitis is associated with both decreased levels of these antimicrobial peptides as well as increased rates of *S. aureus* colonization and infection, highlighting the potential clinical importance of these compounds in combating *S. aureus* [36]. Susceptibility to staphylococcal infections in patients with deficiencies in the IL-1 receptor/TLR-associated signaling molecules IRAK4 [37] or MyD88 [38] further demonstrates the contribution of this critical innate immunity pathway to control of this pathogen in humans.

While there is a vast literature on measuring the adaptive immune responses to *S. aureus*, few, if any, have found evidence that such responses are protective against subsequent infections [39]. Clinical phenotypes seen in Hyper IgE syndrome (STAT3 [40, 41] or DOCK8 [42, 43] deficiency), mucocutaneous candidiasis [44], APECED [45], HIV [46], and atopic dermatitis [47] suggest that altered T cell responses predispose to staphylococcal infections, in particular implicating a protective role for IL-17-secreting CD4 helper T cells (Th17 cells). In contrast, patients suffering from psoriasis have higher levels of IL-17 in their skin and are relatively protected from staphylococcal infections [47]. Further supporting a role for T cells in the response to *S. aureus* are the bacterial mechanisms that have evolved to counter adaptive cellular immunity. Enterotoxins shift the T cell response away from Th1/Treg and towards Th2 [48]. Superantigens such as toxic shock syndrome toxin function to induce non-specific and thus non-targeted T cell excitation [49], and protein A causes similarly non-targeted expansion of B cells [50]. Protein A, among other factors, also helps negate protective effects mediated by complement [51], TNF receptor [52], and the epidermal growth factor receptor [53].

Staphylococcal infection presents most commonly in the skin and soft tissues. These infections cause over ten million outpatient visits and nearly a half-million hospital admissions per year in the USA [54]. Interestingly, nearly a third of people are colonized with *S. aureus* [55]. Although this is a risk factor for subsequent infection [56], it is notable that so many people have a seemingly peaceful co-existence with this potentially lethal organism. Furthermore, the ubiquitous colonization of people with less virulent *Staphylococcus* species, such as *Staphylococcus epidermidis* and other coagulase-negative staphylococci, suggests that we can gain insights into *S. aureus* virulence by studying the

colonization and virulence strategies of these closely related organisms, reviewed here by Otto. Miller and colleagues examine recent discoveries that elucidate the role of innate and adaptive immune mechanisms in cutaneous host defense against *S. aureus*.

Breach of skin and mucosal defenses predisposes toward potentially life-threatening invasive staphylococcal infection. Most commonly presenting as pneumonia and blood-stream infections, these invasive infections are becoming more prevalent [57–59], primarily as an iatrogenic consequence of increasingly complex medical procedures and therapies. Furthermore, an increased incidence of pneumonia has been seen during the ongoing epidemic of community-associated MRSA infection [59]. The pathogenesis of staphylococcal pulmonary infection is detailed in the chapter by Prince and colleagues, highlighting the role of the previously mentioned immunomodulatory virulence mechanisms [52, 60–62] as well as factors that aid in adhesion such as collagen-binding protein [63] and clumping factors A and B [64, 65]

In recent years, new reservoirs for drug-resistant *S. aureus* have been recognized, with a large proportion of both US meat and poultry samples [66] as well as Brazilian food products [67] testing positive for MRSA. The threats posed by this consequence of widespread agricultural and medical antibiotic use have ironically led to efforts to expand the antibiotic armamentarium against *S. aureus*, as reviewed here by Fowler and colleagues. Topical treatments aimed at colonization, such as mupirocin [68] and chlorhexidine [69], have also gained interest, revisiting mechanisms to kill *S. aureus* that are practically identical to that of carbolic acid used in the 1900s [70, 71]. In this issue, Liu and colleagues consider targeting staphylococcal virulence factors and host defense mechanisms as alternative treatment strategies to supplement traditional antibiotics. Schneewind and colleagues take a similar approach to consider novel vaccine targets. Spellberg and Daum reflect on the multiple past failures and future prospects for a successful vaccine, which still remains the holy grail for controlling infectious pathogens, especially one that has proven adept at outmaneuvering antibiotics and host defenses. This special issue comprises articles from a collection of experts that aim to capture our current knowledge of staphylococcal virulence and host defense as we work to understand this human commensal and formidable pathogen.

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References

1. Ogston A. On Abscesses. *Rev Infect Dis*. 1984; 6:122–128. [PubMed: 6369479]
2. Report on the Germacidal Action of Chloroform on Pathogenic Bacteria In Vitro and In Vivo. Chicago: 1921.
3. Barber M. Sensitization of penicillin-resistant staphylococci. *Lancet*. 1948; 1:730. [PubMed: 18857152]
4. Barber M, Rozwadowska-Dowzenko M. Infection by penicillin-resistant staphylococci. *Lancet*. 1948; 2:641–644. [PubMed: 18890505]
5. Chambers HF, Deleo FR. Waves of resistance: *Staphylococcus aureus* in the antibiotic era. *Nat Rev Microbiol*. 2009; 7:629–641. [PubMed: 19680247]
6. Lowy FD. Antimicrobial resistance: the example of *Staphylococcus aureus*. *J Clin Invest*. 2003; 111:1265–1273. [PubMed: 12727914]
7. Otto M. Basis of virulence in community-associated methicillin-resistant *Staphylococcus aureus*. *Annu Rev Microbiol*. 2010; 64:143–162. [PubMed: 20825344]

8. Jevons MP, Coe AW, Parker MT. Methicillin resistance in staphylococci. *Lancet*. 1963; 1:904–907.
9. Beasley FC, et al. Characterization of staphyloferrin A biosynthetic and transport mutants in *Staphylococcus aureus*. *Mol Microbiol*. 2009; 72:947–963. [PubMed: 19400778]
10. Friedman DB, et al. *Staphylococcus aureus* redirects central metabolism to increase iron availability. *PLoS Pathog*. 2006; 2:e87. [PubMed: 16933993]
11. Horsburgh MJ, Ingham E, Foster SJ. In *Staphylococcus aureus*, fur is an interactive regulator with PerR, contributes to virulence, and is necessary for oxidative stress resistance through positive regulation of catalase and iron homeostasis. *J Bacteriol*. 2001; 183:468–475. [PubMed: 11133939]
12. Skaar EP, Humayun M, Bae T, DeBord KL, Schneewind O. Iron-source preference of *Staphylococcus aureus* infections. *Science*. 2004; 305:1626–1628. [PubMed: 15361626]
13. Meiwes J, et al. Isolation and characterization of staphyloferrin A, a compound with siderophore activity from *Staphylococcus hyicus* DSM 20459. *FEMS Microbiol Lett*. 1990; 55:201–205. [PubMed: 2139423]
14. Bubeck. Poring over pores: a-hemolysin and Panton– Valentine leukocidin in *Staphylococcus aureus* pneumonia. *Nat Med*. 2007; 13:1405–1406. [PubMed: 18064027]
15. Kennedy AD, et al. Targeting of alpha-hemolysin by active or passive immunization decreases severity of USA300 skin infection in a mouse model. *J Infect Dis*. 2010; 202:1050–1058. [PubMed: 20726702]
16. Cho JS, et al. IL-17 is essential for host defense against cutaneous *Staphylococcus aureus* infection in mice. *J Clin Invest*. 2010; 120:1762–1773. [PubMed: 20364087]
17. Standiford TJ, et al. Lipoteichoic acid induces secretion of interleukin-8 from human blood monocytes: a cellular and molecular analysis. *Infect Immun*. 1994; 62:119–125. [PubMed: 8262617]
18. Ley K, Laudanna C, Cybulsky MI, Nourshargh S. Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nat Rev Immunol*. 2007; 7:678–689. [PubMed: 17717539]
19. Segal AW. How neutrophils kill microbes. *Annu Rev Immunol*. 2005; 23:197–223. [PubMed: 15771570]
20. Bouma G, Ancliff PJ, Thrasher AJ, Burns SO. Recent advances in the understanding of genetic defects of neutrophil number and function. *Br J Haematol*. 2010; 151:312–326. [PubMed: 20813010]
21. Lakshman R, Finn A. Neutrophil disorders and their management. *J Clin Pathol*. 2001; 54:7–19. [PubMed: 11271792]
22. Andrews T, Sullivan KE. Infections in patients with inherited defects in phagocytic function. *Clin Microbiol Rev*. 2003; 16:597–621. [PubMed: 14557288]
23. Gombart AF, Koefler HP. Neutrophil specific granule deficiency and mutations in the gene encoding transcription factor C/EBP(epsilon). *Curr Opin Hematol*. 2002; 9:36–42. [PubMed: 11753076]
24. Alba-Loureiro TC, et al. Neutrophil function and metabolism in individuals with diabetes mellitus. *Braz J Med Biol Res*. 2007; 40:1037–1044. [PubMed: 17665039]
25. Chonchol M. Neutrophil dysfunction and infection risk in end-stage renal disease. *Semin Dial*. 2006; 19:291–296. [PubMed: 16893406]
26. Postma B, et al. Chemotaxis inhibitory protein of *Staphylococcus aureus* binds specifically to the C5a and formylated peptide receptor. *J Immunol*. 2004; 172:6994–7001. [PubMed: 15153520]
27. Athanasopoulos AN, et al. The extracellular adherence protein (Eap) of *Staphylococcus aureus* inhibits wound healing by interfering with host defense and repair mechanisms. *Blood*. 2006; 107:2720–2727. [PubMed: 16317095]
28. Karavolos MH, Horsburgh MJ, Ingham E, Foster SJ. Role and regulation of the superoxide dismutases of *Staphylococcus aureus*. *Microbiology*. 2003; 149:2749–2758. [PubMed: 14523108]
29. Kubica M, et al. A potential new pathway for *Staphylococcus aureus* dissemination: the silent survival of *S. aureus* phagocytosed by human monocyte-derived macrophages. *PLoS One*. 2008; 3:e1409. [PubMed: 18183290]

30. Genestier AL, et al. *Staphylococcus aureus* Panton– Valentine leukocidin directly targets mitochondria and induces Bax-independent apoptosis of human neutrophils. *J Clin Invest*. 2005; 115:3117–3127. [PubMed: 16276417]
31. Liu GY, et al. *Staphylococcus aureus* golden pigment impairs neutrophil killing and promotes virulence through its antioxidant activity. *J Exp Med*. 2005; 202:209–215. [PubMed: 16009720]
32. Miller LS. Toll-like receptors in skin. *Adv Dermatol*. 2008; 24:71–87. [PubMed: 19256306]
33. Kisich KO, et al. The constitutive capacity of human keratinocytes to kill *Staphylococcus aureus* is dependent on beta-defensin 3. *J Invest Dermatol*. 2007; 127:2368–2380. [PubMed: 17460726]
34. Braff MH, Zaiou M, Fierer J, Nizet V, Gallo RL. Keratinocyte production of cathelicidin provides direct activity against bacterial skin pathogens. *Infect Immun*. 2005; 73:6771–6781. [PubMed: 16177355]
35. Simanski M, Dressel S, Glaser R, Harder J. RNase 7 protects healthy skin from *Staphylococcus aureus* colonization. *J Invest Dermatol*. 2010; 130:2836–2838. [PubMed: 20668470]
36. Ong PY, et al. Endogenous antimicrobial peptides and skin infections in atopic dermatitis. *N Engl J Med*. 2002; 347:1151–1160. [PubMed: 12374875]
37. Ku CL, et al. Selective predisposition to bacterial infections in IRAK-4-deficient children: IRAK-4-dependent TLRs are otherwise redundant in protective immunity. *J Exp Med*. 2007; 204:2407–2422. [PubMed: 17893200]
38. von Bernuth H, et al. Pyogenic bacterial infections in humans with MyD88 deficiency. *Science*. 2008; 321:691–696. [PubMed: 18669862]
39. Daum RS. Clinical practice. Skin and soft-tissue infections caused by methicillin-resistant *Staphylococcus aureus*. *N Engl J Med*. 2007; 357:380–390. [PubMed: 17652653]
40. Renner ED, et al. Novel signal transducer and activator of transcription 3 (STAT3) mutations, reduced T(H)17 cell numbers, and variably defective STAT3 phosphorylation in hyper-IgE syndrome. *J Allergy Clin Immunol*. 2008; 122:181–187. [PubMed: 18602572]
41. Milner JD, et al. Impaired T(H)17 cell differentiation in subjects with autosomal dominant hyper-IgE syndrome. *Nature*. 2008; 452:773–776. [PubMed: 18337720]
42. Chu EY, Freeman AF, Jing H, Cowen EW, Davis J, Su HC, Holland SM, Turner ML Chanco. Cutaneous manifestations of DOCK8 deficiency syndrome. *Arch Dermatol*. 2012; 148(1):79–84. [PubMed: 21931011]
43. Su HC. Combined immunodeficiency associated with DOCK8 mutations and related immunodeficiencies. *Dis Markers*. 2010; 29:121–122. [PubMed: 21178270]
44. Kisand K, et al. Chronic mucocutaneous candidiasis in APECED or thymoma patients correlates with autoimmunity to Th17-associated cytokines. *J Exp Med*. 2010; 207:299–308. [PubMed: 20123959]
45. Puel A, et al. Autoantibodies against IL-17A, IL-17 F, and IL-22 in patients with chronic mucocutaneous candidiasis and autoimmune polyendocrine syndrome type I. *J Exp Med*. 2010; 207:291–297. [PubMed: 20123958]
46. Prendergast A, et al. HIV-1 infection is characterized by profound depletion of CD161+ Th17 cells and gradual decline in regulatory T cells. *AIDS*. 2010; 24:491–502. [PubMed: 20071976]
47. Guttman-Yassky E, et al. Low expression of the IL-23/Th17 pathway in atopic dermatitis compared to psoriasis. *J Immunol*. 2008; 181:7420–7427. [PubMed: 18981165]
48. Patou J, et al. *Staphylococcus aureus* enterotoxin B, protein A, and lipoteichoic acid stimulations in nasal polyps. *J Allergy Clin Immunol*. 2008; 121:110–115. [PubMed: 17980412]
49. Rajagopalan G, et al. Intranasal exposure to staphylococcal enterotoxin B elicits an acute systemic inflammatory response. *Shock*. 2006; 25:647–656. [PubMed: 16721274]
50. Hakoda M, et al. Molecular basis for the interaction between human IgM and staphylococcal protein A. *Clin Immunol Immunopathol*. 1994; 72:394–401. [PubMed: 8062451]
51. Nguyen T, Ghebrehiwet B, Peerschke EI. *Staphylococcus aureus* protein A recognizes platelet gC1qR/p33: a novel mechanism for staphylococcal interactions with platelets. *Infect Immun*. 2000; 68:2061–2068. [PubMed: 10722602]
52. MacEwan DJ. TNF receptor subtype signalling: differences and cellular consequences. *Cell Signal*. 2002; 14:477–492. [PubMed: 11897488]

53. Shaykhiev R, Behr J, Bals R. Microbial patterns signaling via Toll-like receptors 2 and 5 contribute to epithelial repair, growth and survival. *PLoS One*. 2008; 3:e1393. [PubMed: 18167552]
54. McCaig LF, McDonald LC, Mandal S, Jernigan DB. *Staphylococcus aureus*-associated skin and soft tissue infections in ambulatory care. *Emerg Infect Dis*. 2006; 12:1715–1723. [PubMed: 17283622]
55. Gorwitz RJ, et al. Changes in the prevalence of nasal colonization with *Staphylococcus aureus* in the United States, 2001–2004. *J Infect Dis*. 2008; 197:1226–1234. [PubMed: 18422434]
56. Hidron AI, et al. Risk factors for colonization with methicillin-resistant *Staphylococcus aureus* (MRSA) in patients admitted to an urban hospital: emergence of community-associated MRSA nasal carriage. *Clin Infect Dis*. 2005; 41:159–166. [PubMed: 15983910]
57. Carrillo-Marquez MA, et al. *Staphylococcus aureus* pneumonia in children in the era of community-acquired methicillin-resistance at Texas Children’s Hospital. *Pediatr Infect Dis J*. 2011; 30:545–550. [PubMed: 21407143]
58. Herold BC, et al. Community-acquired methicillin-resistant *Staphylococcus aureus* in children with no identified predisposing risk. *JAMA*. 1998; 279:593–598. [PubMed: 9486753]
59. Boucher HW, Corey GR. Epidemiology of methicillin-resistant *Staphylococcus aureus*. *Clin Infect Dis*. 2008; 46(Suppl 5):S344–S349. [PubMed: 18462089]
60. Song L, et al. Structure of staphylococcal alpha-hemolysin, a heptameric transmembrane pore. *Science*. 1996; 274:1859–1866. [PubMed: 8943190]
61. Torres VJ, Pishchany G, Humayun M, Schneewind O, Skaar EP. *Staphylococcus aureus* IsdB is a hemoglobin receptor required for heme iron utilization. *J Bacteriol*. 2006; 188:8421–8429. [PubMed: 17041042]
62. Beasley FC, Marolda CL, Cheung J, Buac S, Heinrichs DE. *Staphylococcus aureus* transporters Hts, Sir, and Sst capture iron liberated from human transferrin by Staphyloferrin A, Staphyloferrin B, and catecholamine stress hormones, respectively, and contribute to virulence. *Infect Immun*. 2011; 79:2345–2355. [PubMed: 21402762]
63. Rhem MN, et al. The collagen-binding adhesin is a virulence factor in *Staphylococcus aureus* keratitis. *Infect Immun*. 2000; 68:3776–3779. [PubMed: 10816547]
64. Higgins J, Loughman A, van Kessel KP, van Strijp JA, Foster TJ. Clumping factor A of *Staphylococcus aureus* inhibits phagocytosis by human polymorphonuclear leucocytes. *FEMS Microbiol Lett*. 2006; 258:290–296. [PubMed: 16640587]
65. Palmqvist N, Patti JM, Tarkowski A, Josefsson E. Expression of staphylococcal clumping factor A impedes macrophage phagocytosis. *Microbes Infect*. 2004; 6:188–195. [PubMed: 14998517]
66. Waters AE, et al. Multidrug-resistant *Staphylococcus aureus* in US meat and poultry. *Clin Infect Dis*. 2011; 52:1227–1230. [PubMed: 21498385]
67. Rizek CF, et al. Identification of *Staphylococcus aureus* carrying the *mecA* gene in ready-to-eat food products sold in Brazil. *Foodborne Pathog Dis*. 2011; 8:561–563. [PubMed: 21453120]
68. Hughes J, Mellows G. Inhibition of isoleucyl-transfer ribonucleic acid synthetase in *Escherichia coli* by pseudomonic acid. *Biochem J*. 1978; 176:305–318. [PubMed: 365175]
69. Simor AE, et al. Randomized controlled trial of chlorhexidine gluconate for washing, intranasal mupirocin, and rifampin and doxycycline versus no treatment for the eradication of methicillin-resistant *Staphylococcus aureus* colonization. *Clin Infect Dis*. 2007; 44:178–185. [PubMed: 17173213]
70. Kuyyakanond T, Quesnel LB. The mechanism of action of chlorhexidine. *FEMS Microbiol Lett*. 1992; 79:211–215. [PubMed: 1335944]
71. McDonnell G, Russell AD. Antiseptics and disinfectants: activity, action, and resistance. *Clin Microbiol Rev*. 1999; 12:147–179. [PubMed: 9880479]