Ascorbate peroxidase-related (APx-R) is not a duplicable gene

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Phylogenetic, genomic and functional analyses have allowed the identification of a new class of putative heme peroxidases, so called APx-R (Apx-Related). This new class, mainly present in the green lineage (including green algae and land plants), can also be detected in other unicellular chloroplastic organisms. Except for recent *Apx-R* extra-copies lost after chromosomal or segmental duplications. In a similar way, most *Apx-R* co-expressed genes in Arabidopsis genome do not have conserved extra-copies after chromosomal duplications and are predicted to be localized in organelles, as are the Apx-R. The member of this gene network can be considered a unique gene, well conserved through the evolution due to a strong negative selection pressure and a low evolution rate.

Ascorbate peroxidases (APx) belong to the class I peroxidase. They have been detected in all chloroplastic containing organisms in which they form a small multigenic family in green lineage.¹ They were subjected to some species specific duplications which produced punctual variation in a number of isoforms from 3 to 10. These duplications are probably associated with subfunctionalization. Indeed three major subclasses are defined based on their cellular localizations: cytoplasmic, peroxisomal and chloroplastic/mitochondrial.² Recently, an additional group of sequences closely related to APx has been characterized and named ascorbate peroxidase-related (APx-R). Noteworthy, this new class does not seem to be subjected to functional duplication.³

Exhaustive datamining of multiple sequence resources have been performed with available genome and EST libraries to confirm the previous observations. No functional APx-R gene duplication has been detected. Duplicated APx-R are only observed in polyploid organisms: Triticum aestivum, an allohexapolyploid, possesses 3 APx-R,⁴ and Brassica napus, an allotetraploid, contains at least two independent expressed APx-R with no evidence of conservation of all expected paralogs sequences. Glycine max, an ancient polyploid (palaeopolyploid, tetraploid) ⁵ possesses a single APx-R sequence and one pseudogene while most of APx genes have been detected in duplicated forms. Exhaustive data mining shows that APx-R genes are present in green algae (Chlorophyceae such as Chlamydomonas reinhardtii and Charaphyceae such as *Klebsormidium flaccidum*) and streptophytes, even though two marginal presences have been detected in chloroplastic diatoms (Table 1). APx-R sequence can be considered as good functional molecular marker because APx-R phylogenetic tree

and taxonomic tree are congruent (Fig. 1). More genomic data are needed to determine if all APx-R sequences share the same ancestral sequence or if APx-R from diatoms resulted from a convergent evolution.

The search performed in EST libraries demonstrated that APx-R are poorly or not expressed in all analyzed organisms with an expression average of 0.003%. Expression analysis in *Arabidopsis thaliana* with Genevestigator⁶ confirmed the low level of expression.

In addition to the absence of conserved duplication, high level of sequence conservation is detected (minimum of 50% identity between green algae and streptophyte, and 40% between chloroplastic diatoms and streptophytes). High variability intron positions and number is observed in diatoms and green algae (Fig. 1). However, intron positions and number are highly conserved in higher plants. Only low conservation of the gene structure is observed in the 5'end of the sequences which coincides with the variability of the coding sequence.

Detailed analysis of Arabidopsis thaliana APx-R co-expression network demonstrated that among the 42 genes listed, 31 encode proteins that are predicted to be localized in organelles, in most cases chloroplasts. These proteins display a great variety of biological functions, but a considerable number of them are implicated in chloroplasts protection against photooxidative damage, which suggests that APx-R could play a role in this protective mechanism as well. Interestingly, more than half of those genes are present as single-copy or as lowcopy number in Arabidopsis thaliana (24 among the 42 genes, Table 2), but also in Oryza sativa, Populus trichocarpa and Vitis vinifera genomes. This data confirms that plant proteins predicted to be targeted to organelles are more likely single-copy

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Table 1. Ascorbate peroxidase-related (APx-R)-encoding genes identified in different plant species

Name	Taxonomic group	Organism	Sequence Status	Expression (EST count)	Intron number
PtrAPx-R	Bacillariophyta (diatoms)	Phaeodactylum tricornutum	complete	4/133887	0
TpsAPx-R	Bacillariophyta (diatoms)	Thalassiosira pseudonana	complete	0/61913	0
CreAPx-R	Chlorophyta (green algae)	Chlamydomonas reinhardtii	complete	18/204076	6
CvarAPx-R	Chlorophyta (green algae)	Chlorella variabilis	complete	0/413	5
MpuAPx-R	Chlorophyta (green algae)	Micromonas pusilla	complete	no	0
OIAPx-R	Chlorophyta (green algae)	Ostreococcus lucimarinus	complete	0/17592	1
OtAPx-R	Chlorophyta (green algae)	Ostreococcus tauri	complete	no	1
VcaAPx-R	Chlorophyta (green algae)	Volvox carteri	partial	/132038	
KfIAPx-R	Other Streptophyta	Klebsormidium flaccidum	complete	*	na
AcvAPx-R	Cryptogam	Adiantum capillus-veneris	partial	1/30540	na
MpAPx-R	Cryptogam	Marchantia polymorpha	partial	1/33692	na
PpaAPx-R	Cryptogam	Physcomitrella patens	complete	7/362131	10
SmAPx-Ra_0	Cryptogam	Selaginella moellendorffii	complete	4/93811	10
SmAPx-Rb_8	Cryptogam	Selaginella moellendorffii	complete	0/93811	10
PgAPx-R	Gymnospermae	Picea glauca (white spruce)	partial	3/313110	na
PsiAPx-R	Gymnospermae	Picea sitchensis (Sitka spruce)	partial	1/186637	na
AmaAPx-R	Eudicotyledons	Antirrhinum majus (snapdragon)	partial	1/25310	na
AfpAPx-R	Eudicotyledons	Aquilegia formosa x Aquilegia pubescens	complete	4/85039	na
AlyAPx-R	Eudicotyledons	Arabidopsis lyrata	complete	no	9
AtAPx-R	Eudicotyledons	Arabidopsis thaliana	complete	17/1529700	9
BnAPx-R-1	Eudicotyledons	Brassica napus (oilseed rape)	complete	3/643937	na
BnAPx-R-2	Eudicotyledons	Brassica napus (oilseed rape)	partial	1/643937	na
BoAPx-R-1	Eudicotyledons	Brassica oleracea (Cauliflower)	complete	5/179150	na
BrAPx-R-1	Eudicotyledons	Brassica rapa	complete	0/194305	9
CcIAPx-R	Eudicotyledons	Citrus clementina	complete	0/118365	9
CsAPx-R	Eudicotyledons	Citrus sinensis	complete	1/213830	9
CsaAPx-R	Eudicotyledons	Cucumis sativus	partial	0/8128	9
EgraAPx-R	Eudicotyledons	Eucalyptus grandis	complete	0/1910	9
EeAPx-R	Eudicotyledons	Euphorbia esula	partial	1/47543	na
GmAPx-R	Eudicotyledons	Glycine max (soybean)	complete	13/1461624	9
GmAPx-R[P]	Eudicotyledons	<i>Glycine max</i> (soybean)	pseudogene	no	nd
GhAPx-R	Eudicotyledons	Gossypium hirsutum (cotton)	complete	8/273779	na
GrAPx-R	Eudicotyledons	Gossypium raimondii	complete	3/63577	na
HarAPx-R	Eudicotyledons	Helianthus argophyllus	partial	1/35720	na
LjAPx-R	Eudicotyledons	Lotus japonicus	partial	10/242432	na
LeAPx-R	Eudicotyledons	Lycopersicon esculentum (Tomato)	complete	6/298289	9
MdAPx-R	Eudicotyledons	Malus domestica (apple tree)	complete	2/324565	9
MeAPx-R	Eudicotyledons	Manihot esculenta (cassava)	partial	1/80681	9
MtAPx-R	Eudicotyledons	Medicago truncatula (barrel medic)	complete	4/269238	9
MguAPx-R	Eudicotyledons	Mimulus guttatus	complete	20/261907	8
NtAPx-R	Eudicotyledons	Nicotiana tabacum	partial	1/332667	na
PtAPx-R	Eudicotyledons	Populus trichocarpa (poplar)	complete	1/89943	9
PpeAPx-R	Eudicotyledons	Prunus persica (peach)	complete	0/79584	9

Exhaustive data mining was performed with all available resources (JGI, NCBI, Phytozome...). When available, EST count and intron number were determined and included in the 5th and 6th columns. no: no EST was found; nd: gene structure cannot be determined; na: no genomic sequence available; *sequence kindly provided by R.Timme.

Table 1. Ascorbate peroxidase-related (APx-R)-encoding genes identified in different plant species (continued)

Table 1. Ascolutate perovidase-related (Ar x-h)-encoding genes identified in different plant species (continued)								
RcAPx-R	Eudicotyledons	Ricinus communis	complete	1/62582	9			
StAPx-R	Eudicotyledons	Solanum tuberosum (Potato)	partial	6/249614	na			
ToAPx-R	Eudicotyledons	Taraxacum officinale (dandelion)	partial	2/41296	na			
VvAPx-R	Eudicotyledons	Vitis vinifera (Grape)	complete	7/362674	9			
AGcAPx-R	Monocotyledons	Agrostis capillaris	partial	1/7743	na			
AsAPx-R	Monocotyledons	Avena sativa (Oat)	partial	1/25344	na			
BdiAPx-R	Monocotyledons	Brachypodium distachyon	complete	23/128092	10			
FarAPx-R	Monocotyledons	Festuca arundinacea	complete	4/63758	na			
HvAPx-R	Monocotyledons	Hordeum vulgare (barley)	complete	19/525781	na			
OmAPx-R	Monocotyledons	Oryza minuta	partial	1/5760	na			
OsiAPx-R	Monocotyledons	<i>Oryza sativa</i> (indica)	complete	?/203447	10			
OsAPx-R	Monocotyledons	<i>Oryza sativa</i> (japonica)	complete	?/987318	10			
ShyAPx-R	Monocotyledons	Saccharum hybrid cultivar (sugarcane)	partial	3/282809	na			
SiAPx-R	Monocotyledons	Setaria italica	complete	0/2741	10			
SbAPx-R	Monocotyledons	Sorghum bicolor	complete	6/209828	10			
TaAPx-Ra	Monocotyledons	Triticum aestivum (bread wheat)	complete	4/1071453	na			
TaAPx-Rb	Monocotyledons	Triticum aestivum (bread wheat)	partial	2/1071453	na			
TaAPx-Rd	Monocotyledons	Triticum aestivum (bread wheat)	partial	2/1071453	na			
ZmAPx-R	Monocotyledons	Zea mays	complete	16/2019105	10			
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Exhaustive data mining was performed with all available resources (JGI, NCBI, Phytozome...). When available, EST count and intron number were determined and included in the 5th and 6th columns. no: no EST was found; nd: gene structure cannot be determined; na: no genomic sequence available; *sequence kindly provided by R.Timme.

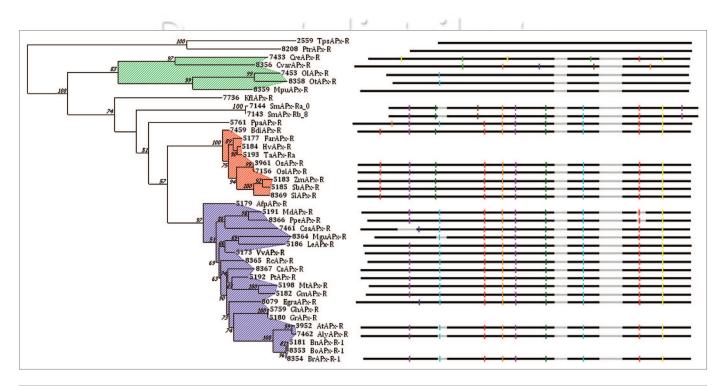


Figure 1. Phylogenetic analyses of APx-R were conducted using complete protein sequences from 39 organisms (Gene accession numbers and name are given as PeroxiBase, peroxibase.toulouse.inra.fr/⁹). Alignment was performed with MAFFT¹⁰ and gap columns were discarded. Phylogenetic analysis was conducted with PhyML, using JTT substitution matrix (4), and among-site rate variation was modeled using a Gamma distribution plus a percent of invariant sites and observed frequencies, as advised by ProtTest (JTT + I + G + F). Branch support was calculated by approximate Likelihood Ratio Test (aLTR), and displayed when higher than 50%. Intron positions relatively to the protein alignment are shown, as obtained with CIWOG¹¹ and were included in front of each branch. The hatched areas in green, red and purple stand for green algae, monocots and dicots respectively.

Gene	Annotation	Subcellular Localization			Single-copy	Low-copy
		Target P	Psort	TAIR	gene*	gene**
At1g05385	LOW PSII ACCUMULATION 19 (LPA19)	Chlo	Chlo	chloroplast, chloroplast thy- lakoid lumen	Yes	-
At1g08550	NON-PHOTOCHEMICAL QUENCHING 1 (NPQ1); ARABIDOPSIS VIOLAXANTHIN DE-EPOXIDASE 1 (AVDE1)	Other	Cyto	chloroplast photosystem II, chloroplast thylakoid lumen	No	Yes
At1g10830	15-CIS-ZETA-CAROTENE ISOMERASE (Z-ISO)	Chlo	Chlo	chloroplast	Yes	-
At1g27385	Unknown protein	Chlo	Chlo	chloroplast	No	Yes
At1g33290	Sporulation protein-related	Chlo	Chlo	n/d	No	No
At1g33330	Peptide chain release factor	Mito	Chlo	chloroplast	Yes	-
At1g54520	Unknown protein	Chlo	Chlo	chloroplast	Yes	-
At1g64430	Unknown protein	Chlo	Chlo	n/d	No	Yes
At1g67840	CHLOROPLAST SENSOR KINASE (CSK)	Chlo	Chlo	chloroplast, chloroplast stroma	No	Yes
At1g76730	5-formyltetrahydrofolate cyclo-ligase family pro- tein	Chlo	Chlo	chloroplast	No	Yes
At1g78140	Methyltransferase-related protein	Mito	Chlo	chloroplast, plastoglobule	No	No
At1g78995	Unknown protein	Chlo	Chlo	n/d	No	Yes
At2g01620	MATERNAL EFFECT EMBRYO ARREST 11 (MEE11)	Other	Chlo	n/d	No	No
At2g03390	uvrB/uvrC motif-containing protein	Chlo	Chlo	chloroplast	No	No
At2g20860	LIPOIC ACID SYNTHASE 1 (LIP1)	Mito	Chlo	mitochondrial matrix, mito- chondrion	No	No
At2g30170	Unknown protein	Chlo	Chlo	chloroplast	No	No
At2g37920	EMBRYO DEFECTIVE 1513 (emb1513)	Chlo	Chlo	n/d	No	Yes
At2g38270	CAX-INTERACTING PROTEIN 2 (CXIP2); GLUTAREDOXIN (ATGRX2)	Chlo	Chlo	chloroplast, chloroplast stroma	Yes	-
At3g10970	Haloacid dehalogenase-like hydrolase family pro- tein	Chlo	Chlo	chloroplast	Yes	-
At3g48560	CHLORSULFURON/IMIDAZOLINONE RESISTANT 1 (CSR1); ACETOLACTATE SYNTHASE (ALS); ACETOHYDROXY ACID SYNTHASE (AHAS); TRIAZOLOPYRIMIDINE RESISTANT 5 (TZP5); IMIDAZOLE RESISTANT 1 (IMR1)	Chlo	Chlo	chloroplast	No	No
At3g53920	RNA POLYMERASE SIGMA-SUBUNIT C (SIGC); SIGMA FACTOR 3 (SIG3)	Chlo	Chlo	chloroplast	No	No
At3g55630	A. THALIANA DHFS-FPGS HOMOLOG D (ATDFD)	Other	Cyto	cytosol	No	No
At4g02260	RELA-SPOT HOMOLOG 1 (RSH1); RELA-SPOT HOMOLOG 1 (AT-RSH1); RELA/SPOT HOMOLOG 1 (ATRSH1)	Chlo	Plast	chloroplast	No	No
At4g10000	Electron carrier protein; disulfide oxidoreductase	Chlo	Chlo	chloroplast	Yes	-
At4g25650	ACD1-LIKE (ACD1-LIKE); PROTOCHLOROPHYLLIDE- DEPENDENT TRANSLOCON COMPONENT 52 KDA (PTC52)	Chlo	Plast	chloroplast, chloroplast envelope	No	No
At4g27600	NECESSARY FOR THE ACHIEVEMENT OF RUBISCO ACCUMULATION 5 (NARA5)	Chlo	Chlo	chloroplast	Yes	-
At4g30310	Ribitol kinase protein	Other	Chlo	chloroplast	No	No
At4g32320	ASCORBATE PEROXIDASE-RELATED (APX-R)	Chlo	Chlo	cytosol	Yes	-

Table 2. Co-The list of APx- R co-expressed genes was obtained through the network generated with ATTED-II ver. 6.0 (atted.jp/)

The putative subcellular localization was predicted through TargetP ver. 1.1 (www.cbs.dtu.dk/services/TargetP/) and Psort ver. 3.0 (www.psort.org/ psortb/) and from TAIR databases (www.arabidopsis.org/). The number of copies of each gene was estimated from the data published by Duarte et al., 2010, which listed single and low copy genes in *Oryza sativa, Vitis vinifera, Populus trichocarpa* and *Arabidopsis thaliana* genomes. *Single-copy genes in *Oryza sativa, Vitis vinifera, Populus trichocarpa* and *Arabidopsis thaliana* genomes, according to Duarte et al. 2010.⁷ **Genes present as one or two copies in at least one of the analyzed genomes. ***Not present in *Oryza sativa*. Table 2. Co-The list of APx- R co-expressed genes was obtained through the network generated with ATTED-II ver. 6.0 (atted.jp/) (continued)

Gene	Annotation	Subcellular Localization			Single-copy	Low-copy
Gene		Target P	Psort	TAIR	gene*	gene**
At5g02250	EMBRYO DEFECTIVE 2730 (EMB2730); RIBONUCLEOTIDE REDUCTASE 1 (RNR1); ARABIDOPSIS THALIANA MITOCHONDRIAL RNASE II (ATMTRNASEII)	Chlo	Chlo	chloroplast, mitochondrion	Yes	-
At5g03900	Unknown protein	Chlo	Plast	chloroplast envelope	Yes	-
At5g04360	PULLULANASE 1 (ATPU1); LIMIT DEXTRINASE (ATLDA); PULLULANASE 1 (PU1)	Chlo	Chlo	chloroplast	No	Yes
At5g06340	ARABIDOPSIS THALIANA NUDIX HYDROLASE HOMOLOG 27 (ATNUDX27)	Chlo	Chlo	chloroplast	No	No
At5g08340	Riboflavin biosynthesis protein-related	Other	Chlo	cellular_component unknown	No	No
At5g08410	FERREDOXIN/THIOREDOXIN REDUCTASE SUBUNIT A2 (FTRA2)	Chlo	Chlo	chloroplast	No	Yes
At5g13720	Unknown protein	Chlo	Plast	chloroplast, chloroplast inner membrane, chloroplast envelope	No	No
At5g18140	DNAJ heat shock N-terminal domain-containing protein	Chlo	Nuclear	n/d	No	No
At5g19540	Unknown protein	Chlo	Chlo	chloroplast	No	Yes
At5g26820	MULTIPLE ANTIBIOTIC RESISTANCE 1 (MAR1); IRON REGULATED 3 (IREG3)	Chlo	Plast	chloroplast, chloroplast envelope	No	Yes
At5g38510	Rhomboid family protein	Chlo	Nuclear	integral to membrane	Yes***	-
At5g57040	Lactoylglutathione lyase family protein	Chlo	Chlo	chloroplast	Yes	-
At5g65685	Soluble glycogen synthase-related protein	Chlo	Chlo	chloroplast	No	No

The putative subcellular localization was predicted through TargetP ver. 1.1 (www.cbs.dtu.dk/services/TargetP/) and Psort ver. 3.0 (www.psort.org/ psortb/) and from TAIR databases (www.arabidopsis.org/). The number of copies of each gene was estimated from the data published by Duarte et al., 2010, which listed single and low copy genes in *Oryza sativa, Vitis vinifera, Populus trichocarpa* and *Arabidopsis thaliana* genomes. *Single-copy genes in *Oryza sativa, Vitis vinifera, Populus trichocarpa* and *Arabidopsis thaliana* genomes, according to Duarte et al. 2010.⁷ **Genes present as one or two copies in at least one of the analyzed genomes. ***Not present in *Oryza sativa*.

than expected by chance.⁷ This could happen because these proteins, when present in the organelles, interact with proteins that are encoded by the organellar genome. In this case, the level of nuclear genome encoded proteins has to be very well controlled inside the cell, so the interaction network will not be disturbed. Looking specifically to the network genes that are single-copy in the specified genomes, we noticed that the majority of the extra-copies of these genes were lost after chromosomal duplications, in a situation very similar to APx-R gene. Thus, it is possible to infer that a great number of single and low-copy genes in this co-expression network could reflect a dose-dependent system, where a raise in copy numbers of such genes would not be favorable to the network. In Figure 2, LPA19 (At1g05385), peptide release factor (At1g33330) and 15-cis-zeta-carotene isomerase (At1g10830) genes were used

as examples. The chromosomal segments that contain these genes in Arabidopsis were duplicated during the evolution and genomic analyses showed that the extra copies were lost during this process (red dashed lines).

The hypothesis of conserved unique genes has already been proposed in reference 8. However further analyses are mandatory to precisely evaluate the extension of the proposal of a complex network of unique gene, taking into consideration that many other neighbor genes were also deleted from these genomic regions. The conservation of this unique gene network indicates that they are under a strong negative selection pressure and subjected to low evolution rate.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

References

 Passardi F, Bakalovic N, Teixeira FK, Margis-Pinheiro M, Penel C, Dunand C. Prokaryotic origins of the non-animal peroxidase superfamily and organellemediated transmission to eukaryotes. Genomics 2007; 89:567-79; PMID:17355904; DOI:10.1016/j. ygeno.2007.01.006. Teixeira FK, Menezes-Benavente L, Galvao VC, Margis R, Margis-Pinheiro M. Rice ascorbate peroxidase gene family encodes functionally diverse isoforms localized in different subcellular compartments. Planta 2006; 224:300-14; PMID:16397796; DOI:10.1007/s00425-005-0214-8. Lazzarotto F, Teixeira FK, Rosa SB, Dunand C, Fernandes C, Fontenele AD, et al. Ascorbate peroxidase-related (APx-R) is a new heme-containing protein functionally associated with ascorbate peroxidase but evolutionarily divergent. New Phytol 2011; 191:234-50; PMID:21352234; DOI:10.1111/j.1469-8137.2011.03659.x.

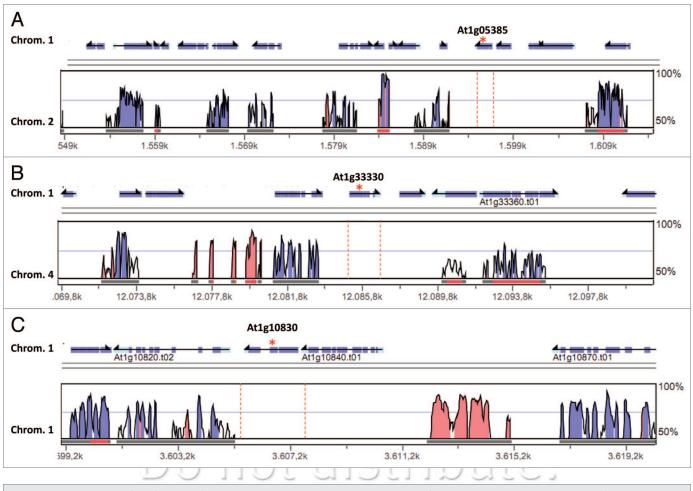


Figure 2. Comparative analyses between the genomic regions containing *APx-R* co-expressed genes *At1g05385* (A), *At1g33330* (B) and *At1g10830* (C) and the DNA segments generated after these regions were duplicated. Peaks correspond to conserved DNA sequences (exons, in most cases) in the Arabidopsis duplicated genomic regions. Horizontal arrows above the graphics represent the annotated genes in Arabidopsis genome (AGI codes are shown). The red dot lines indicate the absence of the *APx-R* co-expressed gene in duplicated region. The VISTA program (www-gsd.lbl.gov/vista/) was used in these analyses.

- Kerby K, Kuspira J. The phylogeny of the polyploid wheats *Triticum aestivum* (bread wheat) and *Triticum turgidum* (macaroni wheat). Genome 1987; 29:722-37; DOI:10.1139/g87-124.
- Schmutz J, Cannon SB, Schlueter J, Ma JX, Mitros T, Nelson W, et al. Genome sequence of the palaeopolyploid soybean. Nature 2010; 463:178-83; PMID:20075913; DOI:10.1038/nature08670.
- Hruz T, Laule O, Szabo G, Wessendorp F, Bleuler S, Oertle L, et al. Genevestigator v3: a reference expression database for the meta-analysis of transcriptomes. Adv Bioinformatics 2008; 2008:420747; PMID:19956698.
- Duarte JM, Wall PK, Edger PP, Landherr LL, Ma H, Pires JC, et al. Identification of shared single copy nuclear genes in Arabidopsis, Populus, Vitis and Oryza and their phylogenetic utility across various taxonomic levels. BMC Evol Biol 2010; 10:61; PMID:20181251; DOI:10.1186/1471-2148-10-61.
- Armisén D, Lecharny A, Aubourg S. Unique genes in plants: specificities and conserved features throughout evolution. BMC Evol Biol 2008; 8:280; PMID:18847470; DOI:10.1186/1471-2148-8-280.
- Koua D, Cerutti L, Falquet L, Sigrist CJA, Theiler G, Hulo N, et al. PeroxiBase: a database with new tools for peroxidase family classification. Nucleic Acids Res 2009; 37:261-6; PMID:18948296; DOI:10.1093/nar/ gkn680.
- Katoh K, Toh H. Recent developments in the MAFFT multiple sequence alignment program. Brief Bioinform 2008; 9:286-98; PMID:18372315; DOI:10.1093/bib/ bbn013.
- Wilkerson MD, Ru YB, Brendel VP. Common introns within orthologous genes: software and application to plants. Brief Bioinform 2009; 10:631-44; PMID:19933210; DOI:10.1093/bib/bbp051.