

Analysis of subcellular localization of auxin carriers PIN, AUX/LAX and PGP in *Sorghum bicolor*

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Auxin transport at least correlates to the three gene families: efflux carriers PIN-formed (PIN), p-glycoprotein (PGP), and influx carrier auxin resistant 1/like aux1 (AUX/LAX) in *Arabidopsis thaliana*. In monocotyledon *Sorghum bicolor*, the biological function of these genes retains unclear. Our previous study reported that the member analysis, organ-specific expression and expression profiles of the auxin transporter PIN, PGP and AUX/LAX gene families in *Sorghum bicolor* under IAA, brassinosteroid, polar auxin transport inhibitors and abiotic stresses. Here we further supply the prediction of subcellular localization of SbPIN, SbLAX and SbPGP proteins and discuss the potential relationship between the subcellular localization and stress response. The predicted results showed that the most of SbPIN, SbLAX and SbPGP proteins are localized to the plasma membrane, except few localized to vacuolar membrane and endoplasmic reticulum. This data set provides novel information for investigation of auxin transporters in *Sorghum bicolor*.

asymmetric subcellular localization of PIN in each transporting cell. Since, uncovering the mechanisms controlling the subcellular dynamics of auxin transport machinery is important for plant response to all developmental signals from embryogenesis to organogenesis, vascular tissue differentiation and tropisms.⁷ In the other hand, the report of auxin transporter related to abiotic stresses was gradually revealed recently. Auxin influx transport AUX1 is essential for the lateral root proliferation component of the salt stress-induced morphogenic response;⁸ the promoters of the *SbPIN*, *SbLAX* and *SbPGP* genes contain numerous DNA elements predicted to respond to abscisic acid, drought and high salt stresses.⁹ To make clear if the subcellular localization of SbPIN, SbLAX and SbPGP proteins also related to abiotic stresses response, here we first predicted their subcellular localization by <http://wolfsort.org/> (Table 1).¹⁰ *SbPIN2*, 5, 6, 7, 8 and 10 were showed the more motifs localized in PM when *SbPIN1*, 3, 4, 9 and 11 were showed the more motifs localized in vacuolar membrane (VM). The five *SbLAXs* all showed the most motifs in PM, even though there are fewer motifs in E.R or VM. The 24 *SbPGPs* members almost were localized in PM besides the both *SbPGP5* and *SbPGP11* also contained the more motifs localized in chloroplast. The above data were similar with the prediction of subcellular localization of PIN in *Arabidopsis* (Table 1). Compare the prediction with the published data of subcellular localization of six *AtPINs*, the prediction analysis of the *AtPIN1*, 3, 4 and 7 were consistent with the published data in reference 2–6. Thus, these analyses

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The intercellular auxin transport depends on the polar subcellular localization of PIN-formed (PIN) auxin efflux carriers at the plasma membrane (PM) and endoplasmic reticulum (ER).¹⁻³ In *Arabidopsis*, PIN1–PIN4 and PIN7 localize to the PM when PIN5, 6 and 8 localize to the ER, which likely mediate the transport of auxin between the ER lumen and cytosol, and then regulate the cellular auxin homeostasis.²⁻⁶ The direction of auxin flow within tissues is mainly determined by the

Table 1. Prediction of subcellular localization of auxin carriers PIN, AUX/LAX and PGP in *Sorghum bicolor*

Gene name ^a	Locus identifier ^b	Accession number ^c	plasma membrane	endoplasmic reticulum	vacuolar membrane	chloroplast
<i>AtPIN1</i>	At01g73590	Q9C6B8	7	2	2	1
<i>AtPIN2</i>	At05g57090	Q9LU77	5		6	
<i>AtPIN3</i>	At01g70940	Q9S7Z8	7			3
<i>AtPIN4</i>	At02g01420	Q8RWZ6	9	2	2	
<i>AtPIN5</i>	At05g16530	Q9FFD0	1		12	
<i>AtPIN6</i>	At01g77110	Q9SQH6	6		4	2
<i>AtPIN7</i>	At01g23080	Q940Y5	8	2		2
<i>AtPIN8</i>	At05g15100	Q9LFP6	5	3	2	3
<i>SbPIN1</i>	Sb02g029210	C5X4P5	4		8	
<i>SbPIN2</i>	Sb03g029320	C5XF44	8		1	2
<i>SbPIN3</i>	Sb03g032850	C5XIA5	1		10	
<i>SbPIN4</i>	Sb03g037350	C5XMI2	3		6	1.5
<i>SbPIN5</i>	Sb03g043960	C5XG98	9	2	1	
<i>SbPIN6</i>	Sb04g028170	ND d	11			1
<i>SbPIN7</i>	Sb05g002150	C5Y431	8	2		3
<i>SbPIN8</i>	Sb07g026370	C5YI36	5		5	1
<i>SbPIN9</i>	Sb10g004430	C5Z4U5	5		9	
<i>SbPIN10</i>	Sb10g008290	C5Z7E9	6	2	2	1
<i>SbPIN11</i>	Sb10g026300	C5Z7A0	6		8	
<i>SbLAX1</i>	Sb01g026240	C5WP27	11	2		
<i>SbLAX2</i>	Sb01g041270	C5WR01	11	3		
<i>SbLAX3</i>	Sb03g040320	C5XQG2	10		3	
<i>SbLAX4</i>	Sb05g004250	C5Y5L4	10	3		
<i>SbLAX5</i>	Sb09g021990	C5YYU5	9	3.5		
<i>SbPGP1</i>	Sb01g039110	C5WPA9	8	3	3	
<i>SbPGP2</i>	Sb02g019540	C5X8A6	13			
<i>SbPGP3</i>	Sb03g011860	C5XI10	10		1	
<i>SbPGP4</i>	Sb03g023740	C5XMA7	12			
<i>SbPGP5</i>	Sb03g031990	C5XHH9	5	2		6
<i>SbPGP6</i>	Sb03g032000	C5XHI0	13			
<i>SbPGP7</i>	Sb03g032030	C5XHI4	11	2		
<i>SbPGP8</i>	Sb03g033290	C5XIE9	12	2		
<i>SbPGP9</i>	Sb03g047490	C5XJF5	8	2	1	1
<i>SbPGP10</i>	Sb04g006087	C5XX25	12		1	
<i>SbPGP11</i>	Sb04g006090	C5XX26	5		2	6
<i>SbPGP12</i>	Sb04g006100	C5XX27	3	2		3
<i>SbPGP13</i>	Sb04g022480	C5XU71	5	2		4
<i>SbPGP14</i>	Sb04g031170	C5Y0R2	11	1		
<i>SbPGP15</i>	Sb06g001440	C5YC52	9	2		
<i>SbPGP16</i>	Sb06g018860	C5Y9T7	12	2		
<i>SbPGP17</i>	Sb06g020350	C5YAT5	5	3		2
<i>SbPGP18</i>	Sb06g030350	C5Y8Z4	11			1
<i>SbPGP19</i>	Sb07g003510	C5YGW7	12			1
<i>SbPGP20</i>	Sb07g003520	C5YGW8	13			
<i>SbPGP21</i>	Sb07g023730	C5YMS8	5	2		
<i>SbPGP22</i>	Sb09g002940	C5YZK3	11			

Table 1. Prediction of subcellular localization of auxin carriers PIN, AUX/LAX and PGP in *Sorghum bicolor* (continued)

Gene name ^a	Locus identifier ^b	Accession number ^c	plasma membrane	endoplasmic reticulum	vacuolar membrane	chloroplast
SbPGP23	Sb09g027320	C5YUY3	14			
SbPGP24	Sb09g027330	C5YUY4	10		1	

suggested that the prediction of subcellular localization of auxin carriers SbPIN, SbLAX and SbPGP was also useful for their experimental research. By previous report, the regulation of PIN protein polarity is needed to quickly respond and adapt plant development to internal and external stimuli.¹¹ At the cellular level, various signals are translated into specific changes in the polar subcellular localization of PIN family thereby guiding the intercellular fluxes of auxin;¹¹ dark treatment affects the subcellular localization of PIN1 and auxin maxima;¹² various environmental and endogenous signals can modulate trafficking and polarity of PIN proteins and then change auxin distribution.¹³ Namely, the subcellular localization of PIN protein may relate to abiotic stresses responses. Future experiments of subcellular localization and abiotic stress responses need to be performed to confirm the bioinformatics prediction.

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