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Thieno[2,3-*d*]pyrimidinedione derivatives as antibacterial agents

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Abstract

Several thieno[2,3-*d*]pyrimidinediones have been synthesized and examined for antibacterial activity against a range of Gram-positive and Gram-negative pathogens. Two compounds displayed potent activity (2–16 mg/L) against multi-drug resistant Gram-positive organisms, including methicillin, vancomycin-intermediate, and vancomycin-resistant *Staphylococcus aureus* (MRSA, VISA, VRSA) and vancomycin-resistant enterococci (VRE). Only one of these agents possessed moderate activity (16–32 mg/L) against Gram-negative strains. An examination of the cytotoxicity of these agents revealed that they displayed low toxicity (40–50 mg/L) against mammalian cell and very low hemolytic activity (2–7%). Taken together, these studies suggest that thieno[2,3-*d*]pyrimidinediones are interesting scaffolds for the development of novel Gram-positive antibacterial agents.

1. Introduction

The increasing prevalence of pathogenic bacteria that are resistant to currently available antibiotics represents an alarming threat to public health. The most commonly encountered antibiotic-resistant bacteria, methicillin-resistant *Staphylococcus aureus* (MRSA), has had a major impact on infections in both the hospital and community setting.[1, 2] While vancomycin continues to be the standard treatment option for antibiotic-resistant infections, the isolation of vancomycin-resistant *Staphylococcus* (VRSA) and *Enterococci* (VRE) foreshadows a day in which the utilization of vancomycin may become limited.[3] Unfortunately, as antibiotic-resistant organisms have become more commonplace, the pipeline for the discovery of new antimicrobial agents has decreased.[4] Thus, there is a pressing need for new antimicrobial agents that are capable of treating resistant bacterial strains.

Thienopyrimidines are interesting heterocyclic compounds and a number of derivatives of these compounds display therapeutic activity as antimicrobial [5–7], antiviral [8, 9], anti-inflammatory [10] antidiabetic [11] and anti cancer [12, 13] agents.[14–16] Despite the breadth of biological activities displayed by these agents, the antibacterial activity of this

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class of compounds has been underexplored. El-Sherbeny and colleagues examined the antimicrobial and antiviral activity of cyclopenteno and cyclohexeno [b]thieno[2,3-d]-3,4-dihydropyrimidine-4-one derivatives (Figure 1a).[8] These agents displayed reasonable activity (MIC values 6.25–25 mg/L) against both Gram-positive and Gram-negative bacteria; however, these agents were significantly more potent against herpes simplex virus. [8] Furanyl-thieno[2,3-d]pyrimidin-4-ones (Figure 1b) were examined by Bahekar et. al. for their antibacterial activity. [7] These agents displayed MIC values in the range of 4–100 mg/L against a collection of Gram-positive and Gram-negative microbes. Interestingly, these compounds also displayed antimycobacterial activity.[7] The antibacterial activity of thieno[2,3-d]pyrimidindiones (Figure 1c) has not been reported in the literature; however, these compounds have been examined for antiviral activity.[15]

Recently, during a study on thieno [2,3-d]pyrimidinones, we discovered a set of compounds that possessed antibacterial activity (Figure 1d). These agents are structurally unrelated to any clinically used antibiotic and display discreet structural overlap with thieno [2,3-d]pyrimidines that have been reported in the literature. In this report, we discuss the synthesis of thieno[2,3-d]pyrimidinediones and their antibacterial and cytotoxic activities.

2. Results and Discussion

2.1. Chemistry

For a project on the development of antiviral therapeutics, we required the synthesis of two constrained (**1** and **3**) and unconstrained (**2** and **4**) thieno[2,3-*d*]pyrimidine-2,4-dione derivatives (Figure 1d), neither of which had been described in the literature. A retrosynthetic analysis of these agents suggested that an amino thiophene ester ring would be prepared first using the standard Gewald reaction. [17, 18] Once the thiophene was in hand, the pyrimidine ring could be prepared by converting the amine into a urea followed by cyclizing with the ester under basic conditions.

The synthesis of compounds **1–4** is shown in schemes 1–3. The synthesis of the constrained derivatives **1** and **3** starts from the commercially available ethyl-2-cyanoacetate and racemic benzylated isopropyl cyclohexanone (**8**, prepared from **7**). These were reacted with sulfur to obtain the amino thiophene ester, **9**, in decent yields. Activation of **9** with *p*-nitrophenyl chloroformate generated the unstable intermediate **10**, which upon reaction with amines **5** or **6** produced urido compounds **11a** and **11b**. [19] Formation of the pyrimidinedione ring was accomplished with refluxing sodium methoxide to provide the final compounds **1** and **3** (Scheme 1). [20] The unconstrained compounds **2** and **4** were prepared using similar methodology starting from the ethyl 3-oxo-5-phenylpentanoate, **13** (Scheme 2).

The amines, **5** and **6**, were prepared, as shown in the Scheme 3, from commercially available compounds (**17** and **21**) using procedures published for related compounds. [21–27] Amine **5** was prepared from *p*-hydroxycinnamic acid (**17**) by first protecting the phenol as the silyl ether followed by conversion of the acid into the azide (**20**). Selective reduction of the azide to **5** was accomplished using catalytic hydrogenation in the presence of Lindlar's catalyst (Pd/CaCO₃). Amine **6** was synthesized from *p*-iodoanisole (**21**) using a Heck reaction with acrylonitrile to generate *E*-cinnamonitrile (**22**). Reduction of the nitrile to the amine was accomplished with LiAlH₄.

2.2. In vitro Antibacterial activity

The initial examination of compounds **1–4** failed to detect antiviral activity. As part of a further investigation of the biological activities of these compounds, we examined the antibacterial activity of compounds **1–4** and intermediates **11a**, **11b**, **16a** and **16b** against a

panel of five Gram-positive and four Gram-negative bacteria (Table 1). Compounds **1** and **2** demonstrated significant antibacterial activity with MIC values in the 2–16 mg/L range against Gram-positive bacteria such as MRSA, VRSA, VISA, VRE and *S. pneumoniae*. The Gram-negative activity of **1** and **2** was weak with MIC values in the range of 16 to over 32 mg/L. Compounds **3** and **4** displayed almost no antibacterial activity with the exceptions being the moderate activity (8 mg/L) of **3** displayed against *E. aerogenes*. Compounds **11a**, **16a** and **16b** showed no activity against any bacterial strain; compound **11b** was insoluble in media, water and DMSO and thus could not be tested. Taken together, the data support the conclusion that **1** and **2** possess Gram-positive antibacterial activity, while compounds **3**, **4**, **11a**, **16a** and **16b** are essentially inactive.

2.3. In vitro cytotoxicity

We examined the cytotoxic activity of compounds **1–4** against mammalian cells (NIH-3T3) using the MTS assay. As shown in Figure 2, all compounds displayed moderate levels of toxicity at 100 mg/L (< 50% of cell viability). The LD₅₀ of each compound was determined from a dose-response curve of cytotoxicity versus concentration (Table 1). As shown in the table, the LD₅₀ (~52 mg/L) of the most potent compound, **2**, is 25 times higher than its MIC value indicating that **2** is selectively toxic to bacteria. In contrast, **1** displayed a therapeutic index of only 5 suggesting that the toxicity of the compound may have also played a role in its antibacterial activity.

2.4. In vitro Hemolytic activity

Another common measure of the toxicity of an agent is lysis of red blood cells. We examined the ability of compounds **1–2** to lyse Sheep erythrocytes. As shown in Figure 3, compound **1** displayed approximately 7% hemolysis at a concentration 25 times its MIC value. In contrast, **2**, which is the most selective antibacterial agent described here, showed only 2% hemolysis at 50 times its MIC value. These results indicate that the antibacterial activity observed for compounds **1** and **2** is not due to non-specific membrane damage. This would suggest that there are specific antibacterial targets for these agents in bacteria.

2.5. Discussion

In this report we have explored thieno[2,3-d]pyrimidinedione derivatives as antibacterial agents. While thieno[2,3-d]pyrimidines have been extensively explored for their varied biological activity, the compounds reported here are unique and have not been prepared in the literature. Among the compounds synthesized, compound **2** showed selective antibacterial activity against Gram-positive antibiotic-resistant bacterial strains such as MRSA, VRSA, VISA and VRE. Unfortunately, these compounds were inactive against Gram-negative pathogens.

A complete structure-activity relationship cannot be determined from the limited set of compounds prepared here. However, inactivity of urido compounds (**11a**, **16a** and **16b**) clearly demonstrates the importance of the pyrimidine ring for the activity and our data also suggests that the presence of a phenol is critical for activity (compare compound **2** to compound **4**). Furthermore, the presence of a methoxy group increases the cytotoxicity by 2-fold when compared to the phenol. A well-known method for enhancing potency of compounds is to lock flexible portions of the molecule into a fixed conformation. Tethering the side chains located on the thiophene results in the racemic compound **1**. While **1** displayed antibacterial activity against Gram-positive pathogens, the MIC values were 2–8-fold higher than for the constrained analog **2**. Thus, the flexibility seen in compound **2** appear to be necessary for the compound to adopt a biologically activity conformation.

The serendipitous discovery of antibacterial activity for these agents unfortunately means that the mechanism of action is unknown. While there have been reports of antibacterial activity for structurally unrelated thieno-pyrimidinones (see Introduction), the mechanism of action of those agents has never reported. Thieno-pyrimidines and thieno-pyrimidinediones have been reported as inhibitors of proteases[28], kinases[29–31] and folate utilizing enzymes[8, 9, 32, 33] suggesting that the mechanism of action of our agent could be due to inhibition of a specific protein. The inhibition of folate utilizing enzymes (i.e. dihydrofolate reductase, etc.) is especially interesting given the fact that antibacterial agents specifically targeting bacterial folate enzymes are well known. Of course, numerous other pathways could be responsible for the antibacterial activity of the compounds, including non-specific membrane effects. We are currently investigating the mechanism of action of these thieno[2,3-d]pyrimidinediones and hope to report on their mechanism in due course.

3. Conclusion

We have synthesized constrained and unconstrained thieno[2,3-d]pyrimidinedione derivatives and examined their antibacterial, cytotoxicity and hemolytic activity. Only one compound, **2**, displayed potent antibacterial activity against a wide range of antibiotic-resistant bacteria including MRSA, VRSA, VISA and VRE. This compound was minimally cytotoxic against mammalian cells and possessed essentially no hemolytic activity. While the mechanism of action of this compound is unknown, efforts to determine the reason for its antibacterial activity are ongoing and will be reported in due course.

4. Experimental

Materials and Instruments

All chemicals were purchased from Acros, Sigma-Aldrich, Matrix Scientific, EMD or Frinton labs and used without further purification. ^1H NMR and ^{13}C NMR spectra were recorded on Varian DRX400. The mass spectra of respective final compounds were recorded on Waters-Micromass ZQ quadupole located in the central instrumentation facility at Wayne State University, Detroit, MI. For the MTS and hemolysis assays, absorbance was collected using Synergy 2 multi-mode plate reader from Biotek Instruments Inc. Winooski, VT.

2-benzyl-4-isopropylcyclohexanone (**8**)

To a cooled ($-20\text{ }^\circ\text{C}$) solution of diisopropylamine (0.160 g, 1.57 mmols) in dry toluene, n-butyllithium (1.6 M in hexanes, 0.95 mL, 1.5 mmols) was added and stirred for 15 min. To the solution, isopropyl cyclohexanone (**7**) (0.200 g, 1.43 mmols), dissolved in 5 mL of dry toluene, was added drop wise. The reaction was stirred at $-20\text{ }^\circ\text{C}$ for 30 min and then cooled to $-78\text{ }^\circ\text{C}$. To the chilled solution, benzylbromide (0.270 g, 1.57 mmols), dissolved in 5 mL dry toluene, was added drop wise and the reaction mixture was warmed to $-45\text{ }^\circ\text{C}$ and stirred at this temperature for 18h. The reaction was quenched with 0.5N HCl (10mL) and extracted with ether (150 mL). The combined organic layers were washed with brine, H_2O and then dried over anhydrous Na_2SO_4 before being concentrated *in vacuo*. The crude product was purified by flash silica column chromatography (1:9 ethyl acetate:hexanes) to yield pure **10** (0.26, 80%). ^1H NMR (400 MHz, CD_2Cl_2) δ 0.85, (d, $J=2.8\text{Hz}$, 3H, CH_3), 0.86, (d, $J=2.4\text{Hz}$, 3H, CH_3), 1.44–1.59, (m, 4H, CH_2 , CH_2), 2.31–2.37 (m, 4H, CH_2 , CH_2), 2.38–2.65, (m, 2H, CH, CH), 3.2 (dd, $J_a=14\text{Hz}$, $J_b=5.2\text{Hz}$, 1H, CH), 7.15–7.28, (m, 5H, Ph); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 19.47, 20.00, 30.61, 32.22, 35.64, 37.13, 41.71, 43.16, 51.50, 125.96, 128.34, 129.29, 141.02, 212.38.

Ethyl 2-amino-4-benzyl-6-isopropyl-4,5,6,7-tetrahydrobenzo[b]thiophene-3-carboxylate (**9**)

To a 50 mL round bottom flask, **8** (0.341 g, 1.48 mmols), sulfur (0.048 g, 1.48 mmols), ethylcyanoacetate (0.167 g, 1.48 mmols) and morpholine (0.129 g, 1.48 mmols) were added along with 15 mL of absolute ethanol. The resulting solution was refluxed for 48h, cooled and then evaporated to dryness *in vacuo*. The resulting crude product was purified by flash silica column chromatography (1:9 ethyl acetate: hexanes) to generate 0.21 g (39%) of **9**. ¹H NMR (400 MHz, CD₂Cl₂) δ 0.85, (d, *J*=7.2Hz, 3H, CH₃), 0.89, (d, *J*=6.8Hz, 3H, CH₃), 1.35, (t, *J*=5.2Hz, 3H, CH₃), 1.78–1.89 (m, 2H, CH, CH), 2.45, (t, *J*=2.8Hz, 2H, CH₂), 2.56–2.62, (m, 2H, CH₂, CH₂), 3.15 (dd, *J*_a=13.6Hz, *J*_b=2.8Hz, 1H, CH), 4.41, (q, *J*=8.4Hz, 2H, CH₂), 6.03, (s, 2H, NH₂), 7.20–7.32, (m, 5H, Ph); ¹³C NMR (100 MHz, CD₂Cl₂) δ 14.93, 19.56, 20.10, 30.64, 32.27, 40.54, 41.75, 43.19, 51.55, 59.74, 118.50, 126.03, 128.41, 129.35, 136.61, 141.05, 141.94, 162.80, 165.63. HRMS (ES+) *m/z* (M+H) calcd for C₂₁H₂₇NO₂S 358.1839, found 358.1837 (M+H).

Ethyl 4-benzyl-6-isopropyl-2-((4-nitrophenoxy)carbonylamino)-4,5,6,7-tetrahydrobenzo[b]thiophene-3-carboxylate (**10**)

A solution of dry pyridine (0.16 g, 2.0 mmols) and **9** (0.358 g, 1.0 mmols) in 20 mL of dry CH₂Cl₂ was stirred at room temperature for 30 min. To the reaction, a solution of *p*-nitrophenyl chloroformate (0.303g, 1.5 mmols), in 5 mL of dry CH₂Cl₂, was added drop wise over a period of 15 min. The reaction was stirred at room temperature for 12h and then evaporated *in vacuo* to yield the crude product which was purified by flash silica column chromatography (3:7 ethyl acetate : hexanes) to give 0.41 g (77%) of the desired product. ¹H NMR (400 MHz, CD₂Cl₂) δ 0.86, (d, *J*=6.8Hz, 3H, CH₃), 0.91, (d, *J*=6.8Hz, 3H, CH₃), 1.46, (t, *J*=7.6Hz, 3H, CH₃), 1.83–1.88 (m, 2H, CH, CH), 2.50–2.82, (m, 4H, CH₂, CH₂), 3.14–3.17, (m, 2H, CH₂), 3.59–3.62 (m, 1H, CH), 4.47, (q, *J*=7.2Hz, 2H, CH₂), 7.16–7.27, (m, 5H), 7.48, (d, *J*=8.0Hz, 2H, Ph), 8.32, (d, *J*=8.0Hz, 2H, Ph), 11.09, (s, 1H, NH); ¹³C NMR (100 MHz, CD₂Cl₂) δ 14.67, 19.62, 19.99, 28.32, 28.54, 32.36, 35.19, 40.85, 61.25, 111.58, 122.21, 125.37, 126.12, 127.81, 128.35, 129.30, 135.87, 141.30, 145.48, 148.67, 150.12, 155.50, 166.09. HRMS could not be obtained because of decomposition in the mass spectrometer.

(E)-ethyl 4-benzyl-2-(3-(3-(4-(tert-butylidimethylsilyl)phenyl)allyl)ureido)-6-isopropyl-4,5,-6,7-tetrahydrobenzo[b]thiophene-3-carboxylate (**11a**)

Compound **5** (0.250 g, 0.95 mmols) was dissolved in 20 mL of dry THF followed by the addition of pyridine (0.090 g, 1.14mmols) and DMAP (0.017 g, 0.14 mmols). The resulting solution was stirred for 20 min. A solution of **10** (0.496 g, 0.95 mmols) in 15 mL of dry THF was added drop wise to the reaction mixture and the reaction was stirred for an additional 12–24 h. The reaction was then evaporated to dryness *in vacuo* and the residue was purified by flash silica column chromatography (2:8 ethyl acetate: hexanes) to yield 0.49 g (79%) of the expected products. ¹H NMR (400 MHz, CD₂Cl₂) δ 0.86, (d, *J*=6.8Hz, 3H, CH₃), 0.91, (d, *J*=6.8Hz, 3H, CH₃), 1.46, (t, *J*=7.6Hz, 3H, CH₃), 1.83–1.88 (m, 2H, CH, CH), 2.50–2.82, (m, 4H, CH₂, CH₂), 3.14–3.17, (m, 2H, CH₂), 3.59–3.62 (m, 1H, CH), 4.47, (q, *J*=7.2Hz, 2H, CH₂), 7.16–7.27, (m, 5H), 7.48, (d, *J*=8.0Hz, 2H, Ph), 8.32, (d, *J*=8.0Hz, 2H, Ph), 11.09, (s, 1H, NH); ¹³C NMR (100 MHz, CD₂Cl₂) δ 14.67, 19.62, 19.99, 28.32, 28.54, 32.36, 35.19, 40.85, 61.25, 111.58, 122.21, 125.37, 126.12, 127.81, 128.35, 129.30, 135.87, 141.30, 145.48, 148.67, 150.12, 155.50, 166.09. HRMS could not be obtained because of decomposition in the mass spectrometer.

(E)-ethyl 4-benzyl-6-isopropyl-2-(3-(3-(4-methoxyphenyl)allyl)ureido)-4,5,6,7-tetrahydrobenzo[b]thiophene-3-carboxylate (11b)

Compound **6** (0.097 g, 0.60 mmols) was dissolved in 15 mL of dry THF followed by the addition of pyridine (0.052g, 0.65mmols) and DMAP (0.008g, 0.06 mmols). To the reaction, which was stirred for 20 min, a solution of **10** (0.310g, 0.60 mmols) in 10 mL of dry THF was added drop wise. The reaction was stirred at room temperature for 12–24 h and evaporated to dryness in *vacuo*. The expected product (0.16 g, 69%) was obtained from the residue after purification by flash silica column chromatography (3:7 ethyl acetate: hexanes). ¹H NMR (400 MHz, CD₂Cl₂) δ 0.85, (d, *J*=6.8Hz, 3H, CH₃), 0.90, (d, *J*=6.8Hz, 3H, CH₃), 1.19–1.26 (m, 1H, CH), 1.37, (t, *J*=7.6Hz, 3H, CH₃), 1.41–1.48 (m, 1H, CH), 1.79–1.85 (m, 1H, CH), 2.28 (t, *J*=5.6Hz, 2H, CH₂), 2.55 (d, *J*=6.2Hz, 2H), 3.13, (d, *J*=6.0Hz, 2H, CH₂), 3.76 (s, 3H, CH₃), 4.05 (t, *J*=6.0Hz, 2H, CH₂) 4.41, (q, *J*=6.8Hz, 2H, CH₂), 5.71 (t, *J*=4.8Hz, 1H, NH), 6.08–6.17 (sextet, *J*= 15.6Hz, 1H, CH=CH), 6.53 (d, *J*= 15.6Hz, 1H, CH=CH), 6.84 (d, *J*=8.4Hz, 2H, Ph), 7.19–7.33, (m, 7H, Ph), 10.80, (s, 1H, NH); ¹³C NMR (100 MHz, CD₂Cl₂) δ 14.77, 19.66, 19.81, 28.48, 32.83, 35.23, 36.72, 40.87, 42.96, 55.41, 60.68, 108.39, 114.15, 123.76, 125.72, 126.00, 127.75, 128.40, 129.16, 129.44, 131.52, 134.52, 141.62, 152.25, 153.85, 159.60, 166.76., HRMS (ES+) *m/z* (M+H) calcd for C₃₂H₃₈N₂O₄S 547.2632, found 547.2631 (M+H)

(E)-ethyl 4-benzyl-6-isopropyl-2-(3-(3-(4-methoxyphenyl)allyl)thieno[2,3-d]pyrimidine-2,4-dione (3)

Compound **11b** (0.050 g, 0.09 mmols) was refluxed in sodium methoxide (prepared from NaOCH₃ (0.005 g, 0.10 mmols) and 10 mL of methanol) for 3h. The reaction was cooled and neutralized with Dowex 50 H⁺ resin which resulted in a white precipitate. The precipitate was dissolved in hot methanol and filtered through a sintered glass funnel. The resulting filtrate [NOTE: check on this] was dried and purified using flash silica column chromatography (1:1 ethyl acetate: hexane) to yield 0.035 g (76%) of the final product. ¹H NMR (400 MHz, DMSO) δ 0.86, (d, *J*=6.8Hz, 3H, CH₃), 0.91, (d, *J*=6.4Hz, 3H, CH₃), 1.46–1.58, (m, 1H, CH), 1.81–2.12, (m, 1H, CH), 2.26–2.43 (m, 4H, CH₂, CH₂) 2.73, (dd, *J*_a=11.6Hz, *J*_b=4.8Hz, 1H, CH), 3.18 (d, *J*=12.0Hz, 2H, CH₂), 3.73 (s, 3H, CH₃), 4.61 (d, *J*=8.0Hz, 2H, CH₂), 6.15–6.19 (sextet, *J*= 16Hz, 1H, CH=CH), 6.48 (d, *J*= 16Hz, 1H, CH=CH), 6.85 (d, *J*=8.8Hz, 2H, Ph), 7.18–7.38, (m, 7H, Ph), 12.18, (s, 1H, NH); ¹³C NMR (100 MHz, CD₂Cl₂) δ 20.25, 20.41, 27.31, 28.84, 32.61, 35.65, 37.46, 42.09, 55.75, 112.18, 114.66, 122.72, 126.57, 126.82, 128.19, 128.84, 129.65, 129.78, 131.73, 135.63, 141.86, 150.87, 151.07, 158.99, 159.50; HRMS (ES+) *m/z* (M+H) calcd. for C₃₀H₃₂N₂O₃S 501.2212, found 501.2210 (M+H).

(E)-ethyl 4-benzyl-2-(3-(3-(4-(hydroxy)phenyl)allyl) thieno[2,3-d]pyrimidine-2,4-dione (1)

Compound **11a** (0.100 g, 0.15 mmols) was refluxed in sodium methoxide (prepared from NaOCH₃ (0.010 g, 0.19 mmols) and 15 mL of methanol) for 3h. The reaction was cooled and neutralized with Dowex 50 H⁺ resin which resulted in a white precipitate. The precipitate was dissolved in hot methanol and filtered through a sintered glass funnel. The resulting filtrate [NOTE: check on this] was dried and purified using flash silica column chromatography (1:1 ethyl acetate: hexane) to yield 0.058 g (77%) of the final product. ¹H NMR (400 MHz, DMSO) δ 0.86, (d, *J*=6.8Hz, 3H, CH₃), 0.90, (d, *J*=6.8Hz, 3H, CH₃), 1.44–1.60, (m, 2H, CH, CH), 2.25–2.42 (m, 4H, CH₂, CH₂) 2.70–2.75, (m, 1H, CH), 3.16 (d, *J*=7.8Hz, 2H, CH₂), 4.59 (d, *J*=7.0Hz, 2H, CH₂), 6.04–6.14 (sextet, *J*= 16Hz, 1H, CH=CH), 6.44 (d, *J*= 16Hz, 1H, CH=CH), 6.67 (d, *J*=8.8Hz, 2H, Ph), 7.18–7.38, (m, 7H, Ph), 9.47 (s, 1H, OH), 12.18, (s, 1H, NH); ¹³C NMR (100 MHz, DMSO) δ 20.26, 20.42, 27.31, 28.85, 32.62, 35.66, 37.47, 42.15, 60.44, 112.21, 116.04, 121.49, 126.58, 126.86,

128.04, 128.25, 128.86, 129.79, 132.29, 135.65, 141.86, 150.82, 150.90, 157.78, 158.97; HRMS (ES+) m/z (M+H) calcd for C₂₉H₃₀N₂O₃S 487.2055, found 487.2046 (M+H).

Ethyl 3-oxo-5-phenylpentanoate (13)

Sodium ethylacetoacetate (**12**) (5 g, 32.87 mmols) was dissolved in 50 mL of dry THF and cooled to 0°C under argon. Once cooled, a solution (1.6 M in hexane) of n-butyl lithium (21.88 mL, 34.51 mmols) was added drop wise and the reaction was stirred for 30 min at 0°C to generate the enolate. Benzyl bromide (5.62 g, 32.87 mmols) was then added and the reaction continued stirring for another 90 min at 0°C. Upon completion, the reaction was poured into a cold solution of saturated potassium hydrogen phosphate and the resulting aqueous solution was extracted with ethyl ether (3 × 100 mL). The combined organic layers were washed with water, dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The resulting material was purified by flash silica column chromatography (1:1:8 CH₂Cl₂: ethyl acetate: hexanes) to obtain 4.75 g (66%) of the final product as eluent. ¹H NMR (400 MHz, CD₂Cl₂) δ 1.25, (t, *J* = 7.2 Hz, 3H, CH₃), 2.87–2.92, (m, 4H, CH₂, CH₂), 3.42, (s, 2H, CH₂) 4.15, (q, *J* = 6.8 Hz, 2H, CH₂), 7.19–7.31, (m, 5H, Ph); ¹³C NMR (100 MHz, CD₂Cl₂) δ 14.1, 29.52, 44.56, 49.58, 61.46, 126.31, 128.5, 128.64, 141.05, 167.26, 202.11.

Diethyl 5-amino-3-phenethylthiophene-2,4-dicarboxylate (14)

Compound **13** (3.04 g, 13.8 mmols), sulfur (0.443 g, 13.8 mmols), ethylcyanoacetate (1.56 g, 13.8 mmols) and morpholine (1.2 g, 13.8 mmols) were refluxed in 60 mL of absolute ethanol for 48h. Upon completion, the solvent was evaporated *in vacuo* and the resulting material was purified by flash silica column chromatography (1:2:7 ethyl acetate: CH₂Cl₂: hexanes) to yield 1.78 g (38%) of the final product. ¹H NMR (400 MHz, CD₂Cl₂) δ 1.31, (t, *J* = 7.2 Hz, 3H, CH₃), 1.37, (t, *J* = 7.2 Hz, 3H, CH₃), 2.82, (t, *J* = 8.4 Hz, 2H, CH₂) 3.52, (t, *J* = 8.4 Hz, 2H, CH₂), 4.24, (q, *J* = 7.2 Hz, 2H, CH₂), 4.34, (q, *J* = 7.2 Hz, 2H, CH₂), 6.58, (s, 2H, br, NH₂), 7.19 (q, *J* = 4.8 Hz, 1H, Ph), 7.29, (d, *J* = 4.4 Hz, 4H, Ph); ¹³C NMR (100 MHz, CD₂Cl₂) δ 14.47, 14.54, 31.25, 36.73, 60.44, 60.73, 107.79, 109.02, 125.96, 128.42, 128.63, 142.64, 151.57, 162.51, 165.91, 166.83. HRMS (ES+) m/z (M+Na) calcd for C₁₈H₂₁NO₄SNa 370.1079, found 370.1083 (M+Na).

Diethyl 5-((4-nitrophenoxy)carbonylamino)-3-phenethylthiophene-2,4-dicarboxylate (15)

A mixture of dry pyridine (0.12 g, 1.5 mmols) and **14** (0.300 g, 0.86 mmols) in 20 mL of dry CH₂Cl₂ was placed under argon and stirred for 30 min at room temperature. To the solution, *p*-nitrophenyl chloroformate (0.186 g, 0.93 mmols) dissolved in 10 mL of dry CH₂Cl₂ was added drop wise over a period of 15 min and the resulting mixture was stirred at room temperature for 18h. The solvent was evaporated *in vacuo* and the residue was purified by flash silica column chromatography (2:2:6 ethyl acetate: CH₂Cl₂: hexanes) to give 0.34 g (77%) of the final, pure product. ¹H NMR (400 MHz, CD₂Cl₂) δ 1.35, (t, *J* = 6.8 Hz, 3H, CH₃), 1.43, (t, *J* = 7.2 Hz, 3H, CH₃), 2.86, (t, *J* = 8.4 Hz, 2H, CH₂) 3.61, (t, *J* = 8.4 Hz, 2H, CH₂), 4.3, (q, *J* = 7.2 Hz, 2H, CH₂), 4.42, (q, *J* = 7.2 Hz, 2H, CH₂), 7.19–7.34, (m, 5H, Ph), 7.47 (d, *J* = 9.2 Hz, 2H, Ph), 8.31, (d, *J* = 9.2 Hz, 2H, Ph), 11.36 (s, 1H, NH); ¹³C NMR (100 MHz, CD₂Cl₂) δ 14.33, 30.63, 36.85, 61.2, 61.86, 114.15, 118.61, 122.19, 125.48, 126.1, 128.46, 128.59, 142.17, 145.73, 148.85, 150.24, 153.67, 155.17, 162.25, 166.15. HRMS could not be obtained because of decomposition in the mass spectrometer.

(E)-diethyl 3-phenethyl-5-(3-(3-(4-(trimethylsilyloxy)phenyl)allyl)ureido)thiophene-2,4-dicarboxylate (16a)

Compound **5** (0.170 g, 0.65 mmols) was dissolved in 15 mL of dry THF and to the resulting solution, pyridine (0.061 g, 0.78 mmols) and DMAP (0.012 g, 0.10 mmols) were added. The amine solution was stirred at room temperature for 20 min followed by a drop wise addition

of a solution of **15** (0.331 g, 0.65 mmols) in 15 mL of dry THF. The reaction was stirred for an additional 12 h and then evaporated *in vacuo*. The residue was purified by flash silica column chromatography (2:8 ethyl acetate: hexanes) to yield 0.33 g (80%) of pure **16a**. ¹H NMR (400 MHz, CD₂Cl₂) δ 0.19, (s, 6H, CH₃), 0.98, (s, 9H, *t*Bu) 1.30–1.39, (m, 6H, CH₃), 2.82 (t, *J* = 8.0 Hz, 2H, CH₂), 3.57 (t, *J* = 8.0 Hz, 2H, CH₂), 4.07 (t, *J* = 5.8 Hz, 2H, CH₂), 4.26–4.37, (m, 4H, CH₂, CH₂), 5.49, (t, *J* = 5.2 Hz, 1H, NH), 6.12 (sextet, *J* = 15.6 Hz, 1H, CH=CH), 6.55 (d, *J* = 16.4 Hz, 1H, CH=CH), 6.80 (d, *J* = 8.8 Hz, 2H, Ph), 7.25–7.30, (m, 7H, Ph), 11.04 (s, 1H, NH); ¹³C NMR (100 MHz, CD₂Cl₂) δ -4.49, 14.44, 25.66, 30.69, 31.91, 32.44, 36.87, 53.67, 60.86, 61.35, 120.15, 120.44, 123.54, 126.02, 127.75, 128.45, 128.59, 129.42, 130.56, 131.89, 142.50, 148.59, 153.86, 155.77, 162.88, 166.78. HRMS (ES+) *m/z* (M+H) calcd for C₃₄H₄₄N₂O₆SSi 637.2755, found 637.2715 (M+H).

(E)-diethyl 5-(3-(3-(4-methoxyphenyl)allyl)ureido)-3-phenethylthiophene-2,4-dicarboxylate (16b)

To a solution of **6** (0.173 g, 1.06 mmols), dissolved in 20 mL of dry THF, pyridine (0.092 g, 1.17 mmols) and DMAP (0.013 g, 0.106 mmols) were added and the resulting solution was stirred at room temperature for 20 min. To the amine solution, a solution of **15** (0.543 g, 1.06 mmols) in 15 mL of dry THF was added drop wise and the resulting reaction was allowed to stir for an additional 12 h. Upon completion, the solvent was evaporated *in vacuo* and the resulting mixture was purified by flash silica column chromatography (3:7 ethyl acetate: hexanes) to generate 0.49 g (85%) of **16b**. ¹H NMR (400 MHz, CD₂Cl₂) δ 1.32, (t, *J* = 7.2 Hz, 3H, CH₃), 1.37, (t, *J* = 7.2 Hz, 3H, CH₃), 2.83, (t, *J* = 8.4 Hz, 2H, CH₂) 3.60, (t, *J* = 8.4 Hz, 2H, CH₂), 3.8 (s, 3H, CH₃), 4.10 (t, *J* = 5.6 Hz, 2H, CH₂), 4.28, (q, *J* = 7.2 Hz, 2H, CH₂), 4.37, (q, *J* = 7.6 Hz, 2H, CH₂), 5.57 (t, *J* = 5.2 Hz, 1H, NH), 6.07–6.14 (sextet, *J* = 15.6 Hz, 1H, CH=CH), 6.56 (d, *J* = 15.6 Hz, 1H, CH=CH), 6.84 (d, *J* = 8.4 Hz, 2H, Ph), 6.95, (d, *J* = 8.8 Hz, 2H, Ph), 7.17–7.31 (m, 3H, Ph), 8.13 (d, *J* = 9.2, 2H, Ph), 11.14 (s, 1H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 14.35, 30.58, 36.82, 55.47, 61.26, 61.57, 114.20, 115.94, 126.07, 126.32, 127.79, 128.46, 128.55, 129.31, 142.27, 148.87, 154.18, 156.30, 159.70, 162.61, 163.32, 166.82. HRMS (ES+) *m/z* (M+Na) calcd for C₂₉H₃₂N₂O₆SNa 559.1869, found 559.1872 (M+Na).

(E)-ethyl 3-(3-(4-methoxyphenyl)allyl)-2,4-dioxo-5-phenethyl-1,2,3,4-tetrahydrothieno[2,3-d]pyrimidine-6-carboxylate (4)

Compound **16b** (0.385 g, 0.72 mmols) was refluxed in sodium methoxide (prepared from NaOCH₃ (0.043 g, 0.79 mmols) and 30 mL methanol) for 3h. The reaction was cooled and neutralized with Dowex 50 H⁺ resin which resulted in a white precipitate. The precipitate was dissolved in hot methanol and filtered through a sintered glass funnel. The resulting filtrate [NOTE: check on this] was dried and purified using flash silica column chromatography (3:7 ethyl acetate: hexane) to yield 0.31 g (88%) of the final product. ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.28, (t, *J* = 6.8 Hz, 3H, CH₃), 2.78, (t, *J* = 7.6 Hz, 2H, CH₂) 3.55, (t, *J* = 8.0 Hz, 2H, CH₂), 3.73 (s, 3H, CH₃), 4.25, (q, *J* = 7.2 Hz, 2H, CH₂), 4.60, (d, *J* = 6.0 Hz, 2H, CH₂), 6.11–6.18 (sextet, *J* = 16 Hz, 1H, CH=CH), 6.52 (d, *J* = 16 Hz, 1H, CH=CH), 6.87 (d, *J* = 8.8 Hz, 2H, Ph), 7.19 (t, *J* = 6.8 Hz, 1H, Ph), 7.25–7.32 (m, 4H, Ph), 7.36 (d, *J* = 8.8 Hz, 2H, Ph), 12.56 (s, 1H, NH); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 14.91, 30.51, 36.47, 42.17, 55.76, 61.33, 114.68, 116.51, 123.02, 126.53, 126.88, 128.17, 128.91, 128.97, 129.77, 131.79, 140.27, 142.27, 149.31, 159.48, 160.03, 162.55, 164.69; HRMS (ES+) *m/z* (M+Na) calcd for C₂₇H₂₆N₂O₅SNa 513.1460, found 513.1453 (M+Na).

(E)-ethyl 3-(3-(4-hydroxyphenyl)allyl)-2,4-dioxo-5-phenethyl-1,2,3,4-tetrahydrothieno[2,3-d]pyrimidine-6-carboxylate (2)

Compound **16a** (0.100 g, 0.16 mmols) was refluxed in sodium methoxide (prepared from NaOCH₃ (0.009 g, 0.17 mmols) in 30 mL methanol) for 3h. The reaction was cooled and neutralized with Dowex 50 H⁺ resin which resulted in a white precipitate. The precipitate was dissolved in hot methanol and filtered through a sintered glass funnel. The resulting filtrate was dried and purified using flash silica column chromatography (3:7 ethyl acetate:hexane) to yield 0.083 g (90%) of the final product. ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.28, (t, *J* = 7.2Hz, 3H, CH₃), 2.78, (t, *J* = 8.0Hz, 2H, CH₂) 3.55, (t, *J* = 7.8Hz, 2H, CH₂), 4.26, (q, *J* = 6.8Hz, 2H, CH₂), 4.58, (d, *J* = 6.0Hz, 2H, CH₂), 6.06 (sextet, *J* = 15.6Hz, 1H, CH=CH), 6.47 (d, *J* = 16Hz, 1H, CH=CH), 6.69 (d, *J* = 8.4Hz, 2H, Ph), 7.17–7.32 (m, 7H, Ph), 9.48 (s, 1H, OH), 12.55 (s, 1H, NH); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 14.62, 30.30, 36.33, 42.31, 61.60, 114.01, 115.89, 117.27, 120.99, 126.50, 128.05, 128.20, 128.83, 128.90, 132.69, 141.99, 148.87, 150.47, 154.81, 157.67, 159.20, 162.20. HRMS HRMS (ES+) *m/z* (M+Na) calcd for C₂₆H₂₄N₂NaO₄S 499.1304, found 499.1306 (M+Na)

(E)-3-(4-(tert-butyl dimethylsilyloxy)phenyl)acrylic acid (18)

p-hydroxycinnamic acid (**17**) (5.0 g, 30.46 mmols) and imidazole (5.2 g, 76.15 mmols) were dissolved in 25 mL of dry DMF and the solution was cooled to 0 °C. To the chilled solution, *t*-butyldimethylsilylchloride (11.5 g, 76.15 mmols) was added and the reaction was stirred at room temperature for 3h. The resulting reaction mixture was poured into 125 mL of water. The white, cloudy solution was extracted with ethyl acetate (3 × 100 mL) and the combined organic layers were washed with brine and dried with sodium sulfate. The organic layer was evaporated to dryness and the residue was purified by flash silica column chromatography (3:7 ethyl acetate:hexanes) to yield 8.32 g (98%) of the final product. ¹H NMR (400 MHz, CDCl₃) δ 0.23, (s, 6H, CH₃), 0.99, (s, 9H, *t*Bu) 6.33 (d, *J* = 16Hz, 1H, CH=CH), 6.85 (d, *J* = 8.4Hz, 2H, Ph), 7.46 (d, *J* = 8.4Hz, 2H, Ph), 7.74 (d, *J* = 16Hz, 1H, CH=CH); ¹³C NMR (CDCl₃) δ -4.16, 18.46, 25.83, 114.77, 120.80, 127.59, 130.24, 146.94, 158.49, 171.23.

(E)-3-(4-(tert-butyl dimethylsilyloxy)phenyl)prop-2-en-1-ol (19)

Compound **18** (5.0 g, 17.98 mmols) was dissolved in 25 mL of dry CH₂Cl₂ and the solution was cooled to -78 °C. To the chilled solution, DIBAL (20% solution in toluene, 61 mL, 71.91 mmol) was added drop wise and the reaction was stirred at -78 °C for 8h. Upon completion, 75 mL of ether was added to the reaction mixture and the mixture was slowly warmed to 0 °C. Once the solution was equilibrated to 0 °C, 2.5 mL of H₂O was added drop wise, followed by 5 mL of 1N NaOH and an additional 5 mL of H₂O. The solution was then warmed to room temperature and stirred for an additional 15 min and anhydrous MgSO₄ was added slowly with stirring another 15 min until no hydrated MgSO₄ aggregates formed. The salts were filtered and filtrate evaporated to obtain the pure product in 2.43 g (59%) yield. ¹H NMR (400 MHz, CD₂Cl₂) δ 0.22 (s, 6H, CH₃), 1.01 (s, 9H, *t*Bu) 2.13 (s, br, 1H, OH), 4.25 (t, *J* = 4.8Hz, 2H, CH₂), 6.24 (quintet, *J* = 16Hz, 1H, CH=CH), 6.54 (d, *J* = 16Hz, 1H, CH=CH), 6.81 (d, *J* = 8.8Hz, 2H, Ph), 7.28 (d, *J* = 8.4Hz, 2H, Ph); ¹³C NMR (100 MHz, CD₂Cl₂) δ -4.48, 18.34, 25.67, 63.74, 120.43, 127.14, 127.77, 130.44, 130.51, 155.64.

(E)-(4-(3-azidoprop-1-enyl)phenoxy)(tert-butyl)dimethylsilane (20)

Compound **19** (0.300 g, 1.13 mmols) was dissolved in 10 mL of dry THF followed by the addition of triethylamine (0.172 g, 1.7 mmol). Methanesulfonyl chloride (0.195 g, 1.7 mmols) was added drop wise to the reaction and the reaction was stirred at room temperature for 3 h. The reaction was then quenched by the addition of 20 mL H₂O and the resulting solution was extracted with ethyl acetate (3 × 25mL). The combined organic layers

were then washed with brine and dried over anhydrous Na_2SO_4 . The organic layer was evaporated *in vacuo* to obtain the crude product which was used immediately in the next reaction.

The crude mesylated compound was dissolved in dry DMF and sodium azide (0.067 g, 1.64 mmol) was added. The reaction was stirred at room temperature for 5h and then poured into 50 mL H_2O to generate a white, cloudy solution. The solution was ethyl acetate (3 \times 50mL) and the combined organic layers were washed with brine and dried over anhydrous Na_2SO_4 . The organic layer was evaporated *in vacuo* to dryness and the crude product was purified by flash silica column chromatography (1:9 ethyl acetate:hexanes) to give 0.25 g (82%) of the final product. ^1H NMR (400 MHz, CDCl_3) δ 0.22 (s, 6H, CH_3), 1.01 (s, 9H, *t*Bu) 3.93 (d, $J = 6.8\text{Hz}$, 2H, CH_2), 6.12 (quintet, $J = 16\text{Hz}$, 1H, $\text{CH}=\text{CH}$), 6.60 (d, $J = 16\text{Hz}$, 1H, $\text{CH}=\text{CH}$), 6.83 (d, $J = 8.4\text{Hz}$, 2H, Ph), 7.30 (d, $J = 8.4\text{Hz}$, 2H, Ph); ^{13}C NMR (100 MHz, CDCl_3) δ -4.17, 18.47, 53.45, 120.40, 120.52, 128.08, 129.53, 134.58, 156.12.

(E)-3-(4-(tert-butylidimethylsilyloxy)phenyl)prop-2-en-1-amine (5)

Compound **20** (0.200 g, 0.69 mmols) was dissolved in 8 mL of ethanol and Lindlar's catalyst (Pd/CaCO_3 , 0.043 g, 0.21 mmol) was added to the solution. The reaction was stirred under 1 atm of H_2 for 5 h, filtered through celite and evaporated to dryness in *vacuo*. The resulting product (0.17 g, 93%) was used without any additional purification. ^1H NMR (400 MHz, CD_2Cl_2) δ 0.19 (s, 6H, CH_3), 0.98 (s, 9H, *t*Bu), 1.19 (br, 2H, NH_2), 3.40 (d, $J = 5.6\text{Hz}$, 2H, CH_2), 6.19 (quintet, $J = 15.6\text{Hz}$, 1H, $\text{CH}=\text{CH}$), 6.42 (d, $J = 16\text{Hz}$, 1H, $\text{CH}=\text{CH}$), 6.78 (d, $J = 8.4\text{Hz}$, 2H, Ph), 7.25 (d, $J = 8.4\text{Hz}$, 2H, Ph); ^{13}C NMR (100 MHz, CDCl_3) δ -4.54, 18.29, 25.63, 44.50, 120.35, 127.41, 128.68, 130.12, 130.99, 155.27.

((E)-3-(4-methoxyphenyl)acrylonitrile (22)

A mixture of 4-iodo anisole (**21**) (20 g, 85.46 mmols), acrylonitrile (18.14 g, 342 mmols), $\text{Pd}(\text{OAc})_2$ (1.92 g, 8.55 mmols), Bu_4NBr (5.51 g, 17.1 mmols) and NaHCO_3 (14.36 g, 171 mmols) were vigorously stirred in 80 mL of distilled water at 80 °C for 10 hrs under argon. The reaction mixture was cooled to room temperature and extracted with diethyl ether (3 \times 150 mL). The combined organic layer was washed with distilled water, brine and then dried over anhydrous Na_2SO_4 . The organic layer was evaporated *in vacuo* and the resulting material purified by silica column chromatography (7:3 CH_2Cl_2 : hexane) yield 4.89 g (48%) of the product as a white solid. ^1H NMR (400 MHz, CD_2Cl_2) δ 3.83, (s, 3H, CH_3), 5.75, (d, $J = 16.8\text{Hz}$, 1H, $\text{CH}=\text{CH}$), 6.92, (d, $J = 8.8\text{Hz}$, 2H, Ph), 7.35, (d, $J = 16.4\text{Hz}$, 1H, $\text{CH}=\text{CH}$), 7.42, (d, $J = 8.4\text{ Hz}$, 2H, Ph); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 30.24, 93.91, 114.97, 119.2, 126.93, 129.61, 150.43, 162.64.

(E)-3-(4-methoxyphenyl)prop-2-en-1-amine (6)

Lithium aluminum hydride (0.751 g, 19.79 mmols) was added to 25 mL of anhydrous ethyl ether and the resulting solution was stirred at room temperature for 20 min. To a solution of the reductant, **22** (3.15g, 19.79 mmols), dissolved in 15 mL of anhydrous ethyl ether, was added drop wise over a 10 min period. The reaction was stirred at room temperature for an additional 30 min and then was quenched by the addition of (2.5 mL) of 15% NaOH . The reaction was extracted with ethyl ether (3 \times 100mL) and the combined organic layers were washed with brine before being dried with anhydrous Na_2SO_4 . The organic layer was evaporated *in vacuo* and the resulting crude residue was purified by flash silica column chromatography (1:10:89 NH_4OH : CH_3OH : CH_2Cl_2) to generate the desired pure product (1.95 g, 61%) in good yield. ^1H NMR (400 MHz, CDCl_3) δ 1.40, (s, 2H, br, NH_2), 3.47, (s, 2H, br, CH_2), 3.81, (s, 3H, CH_3) 6.17–6.22, (quintet, $J = 20\text{ Hz}$, 1H, $\text{CH}=\text{CH}$), 6.45, (d, $J = 16\text{ Hz}$, 1H, $\text{CH}=\text{CH}$), 6.85, (d, $J = 8.4\text{ Hz}$, 2H, Ph), 7.31, (d, $J = 8.4\text{ Hz}$, 2H, Ph); ^{13}C NMR (100 MHz, CDCl_3) δ 29.93, 44.64, 55.51, 114.2, 127.58, 129.16, 130.2, 159.22.

MTS assay

The MTS assay was performed using the Cell Titer 96 aqueous cell proliferation assay (Promega). Fibroblast cells derived from a NIH Swiss mouse embryo (NIH-3T3) were obtained from ATCC and maintained in Dulbecco's Modified Eagles Medium (DMEM) supplemented with 10% fetal bovine serum (FBS). The cells were grown to 3 passages and approximately 10,000 cells were seeded into each well of a 96-well plate and allowed to incubate overnight to allow cells to attach to the substrate. Individual wells were treated with various concentrations of compounds **1–4** (10, 25, 50 and 100 mg/L in DMSO) in 100 μ L of DMEM/FBS media and incubated for 24 h. To analyze the viability of the cells, the media was removed and replaced with 100 μ L of fresh DMEM media and 20 μ L of MTS reagent solution. The resulting assay mixture was incubated for 1 h at 37 °C in a CO₂ incubator and the absorbance of each well was measured at 490 nm. The assay was conducted in four replicants using DMEM/FBS media as negative control and 5% DMSO as positive control. The results are expressed as the mean percentage of cell viability relative to untreated cells. LD₅₀ values were determined by Prism software using non-linear regression.

Hemolysis assay

Sheep whole blood (Lampire Biological Laboratories, 1 mL) was centrifuged at 3000 rpm for 10 min and the separated plasma and white blood cells were discarded. The remaining red blood cells were washed twice with 2–3 mL of TBS buffer (10 mM Tris, 150 mM NaCl, pH 7.0) and diluted to 1% in TBS. The hemolysis assay was performed, in duplicate, in a 96-well microplate plate using DMSO as the negative control and 1% Triton-X as the positive control. In each well, 120 μ L of the 1% red blood cell stock was treated with increasing concentrations of **1** or **2**, dissolved in DMSO and the final volume was adjusted to 150 μ L by the addition of TBS. The plate was incubated at 37 °C for 1 h. After incubation, the plate was centrifuged at 3800 rpm for 5 min to collect the red blood cells and the supernatant was analyzed by diluting 20 μ L of the supernatant with TBS buffer until the final volume reached 120 μ L. The absorbance at 414 nm was measured to determine the release of hemoglobin. The absorbance of negative control solution was set to 0% hemolysis while the absorbance of the 1% Triton X was set to 100% hemolysis.

Antibacterial activity

Eight bacterial strains (five Gram-positive and 3 Gram-negative pathogens) were selected from the Anti-Infective Research Laboratory collection. Tested isolates included 3 *Staphylococcus aureus* (VISA Mu50, MRSA 494, and VRSA MI) strains, *Streptococcus pneumoniae* (ATCC 49619), *Enterococcus faecium* (VRE 7303), *Escherichia coli* (ATCC 25922), *Pseudomonas aeruginosa* (ATCC 27853), *Enterobacter aerogenes* (CEF 3978) and *Klebsiella pneumoniae* (CEF 3978). Antibacterial activity of the thieno[2,3-d]pyrimidinedione compounds **1–4**, **11a**, **16a**, and **16b** was evaluated by determining minimum inhibitory concentration (MIC) values using the broth microdilution method [34]. Briefly, strains were grown overnight at 35°C on Tryptic soy agar plates (TSA, Difco, Detroit, MI). Bacterial inocula (0.5 Mac Farland, i.e. 10⁸ CFU/mL) were prepared by suspending few colonies grown onto a TSA plate in sterile normal saline solution. Resulted bacterial suspensions were diluted in Mueller-Hinton broth (MHB, Difco, Detroit, MI) in order to reach an inoculum of 1–2 \times 10⁶ CFU/mL. A volume of 100 μ L of this inoculum (final titer 5 \times 10⁵ CFU/mL) was added to 100 μ L of twofold serial dilutions of compounds **1–4** (final concentrations of 0.06 to 32 mg/L) in Mueller-Hinton broth. Inoculated plates were incubated at 35°C for 18–24h, after which the MIC values were visually read as the lowest concentration of compound that prevented bacterial growth.

- Four unique thieno[2,3-d]pyrimidinediones were synthesized by a unique route

- Two compounds displayed potent activity against antibiotic-resistant Gram-positive bacteria
- One compound was selectively cytotoxic to bacteria and displayed no hemolytic activity

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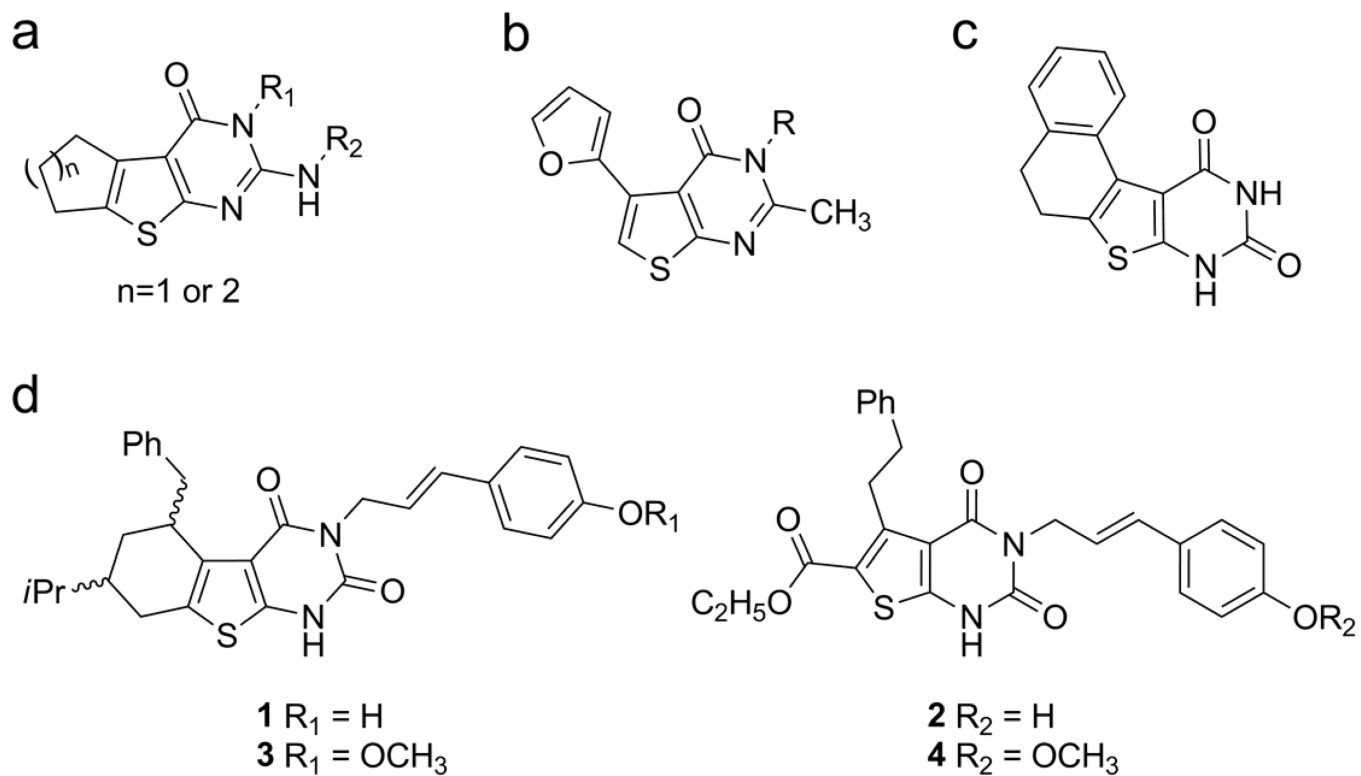


Figure 1.
Thieno[2,3-d]pyrimidineone derivatives

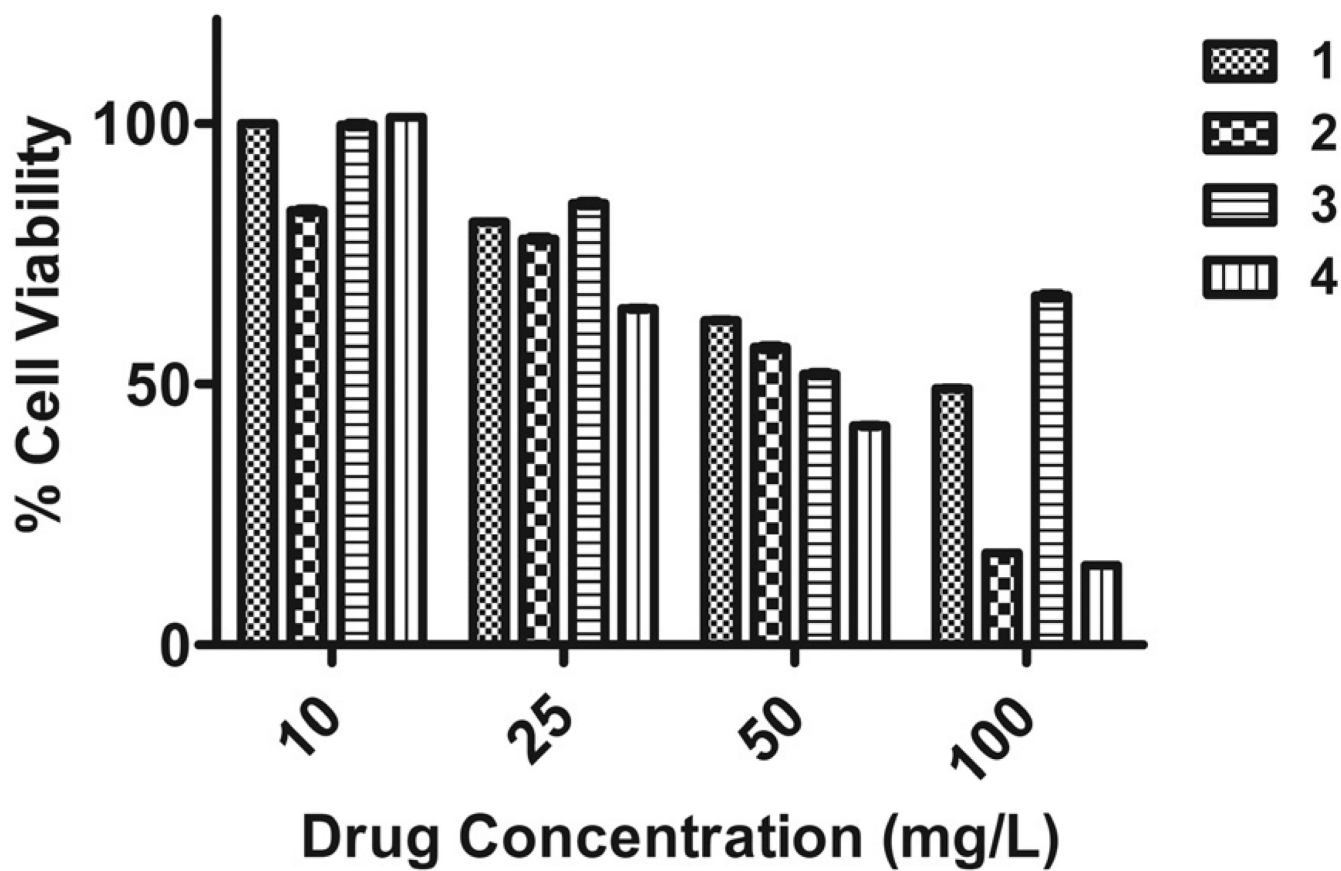


Figure 2. Cytotoxicity of compounds 1–4 obtained by MTS assay against mammalian cells (NIH-3T3), 5% DMSO is used as control.

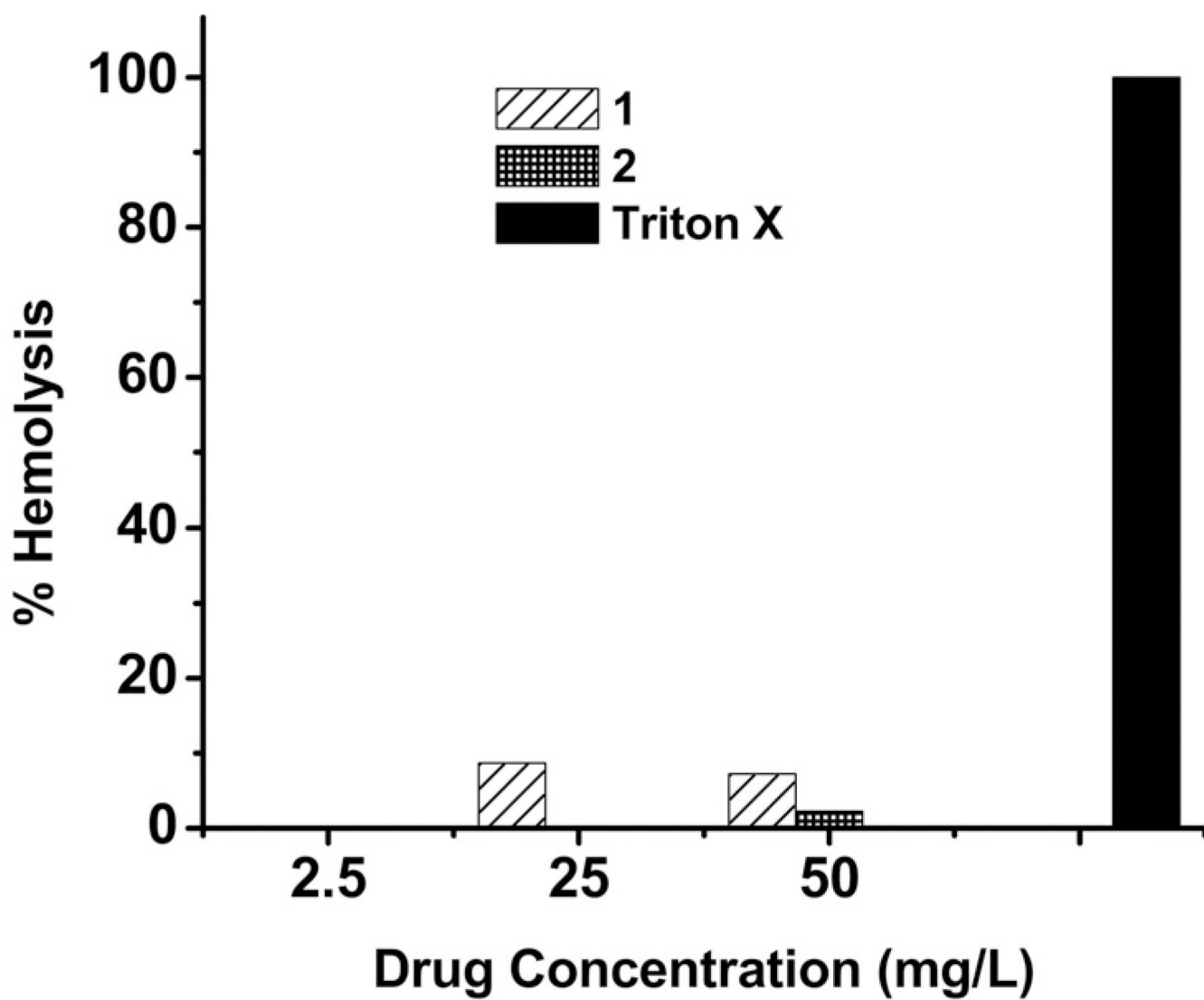
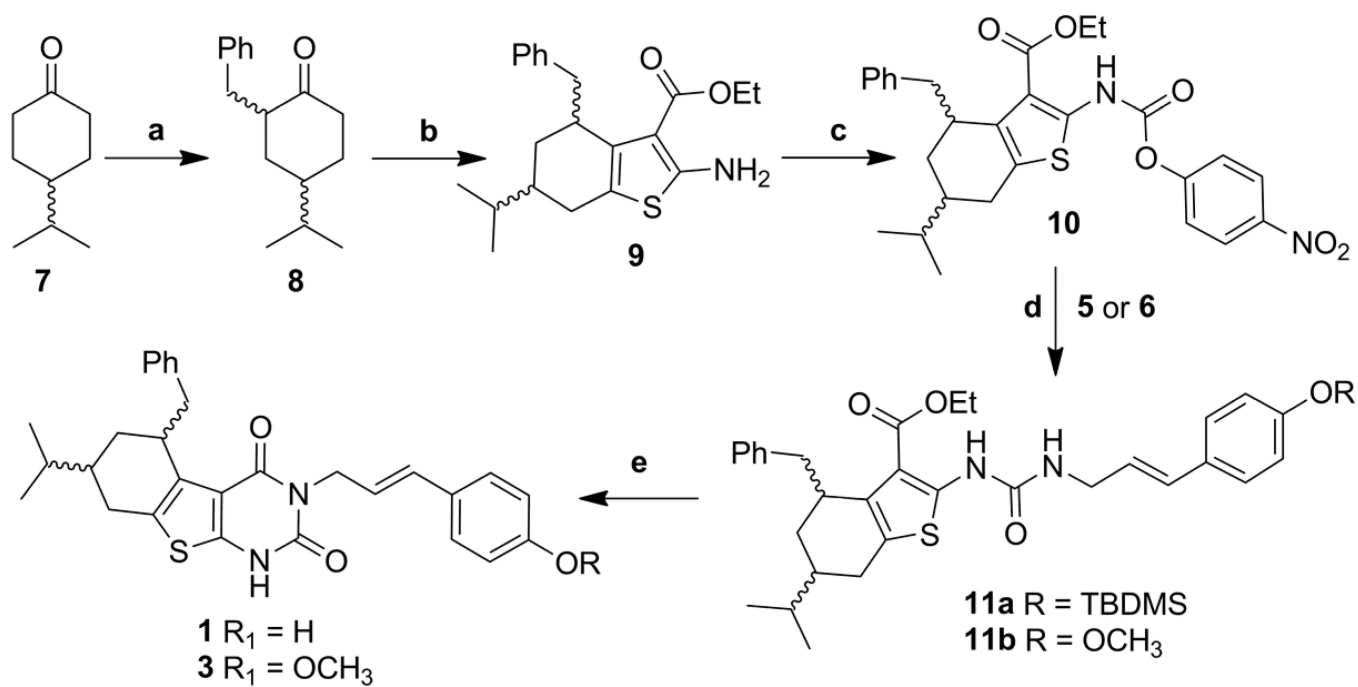
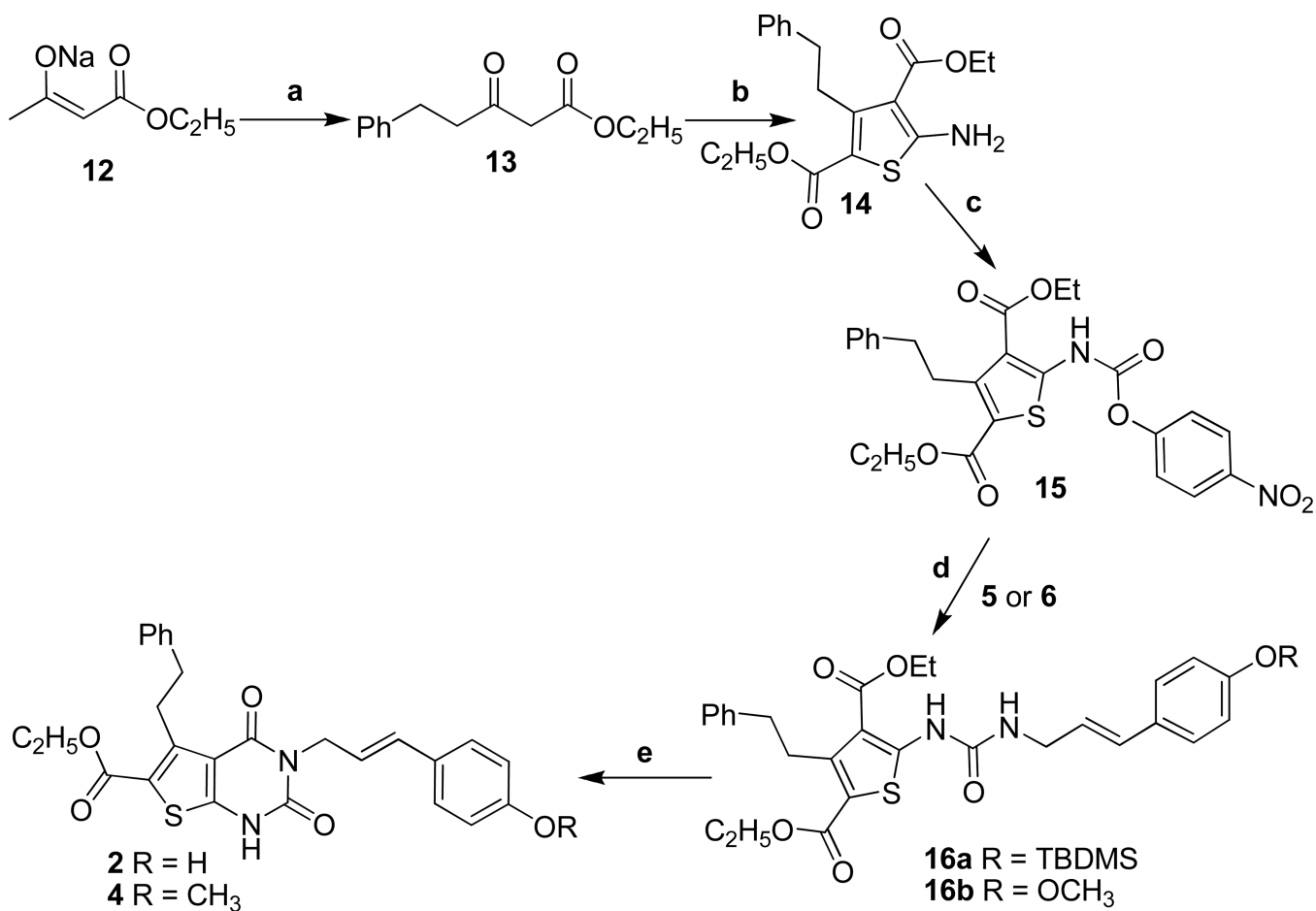


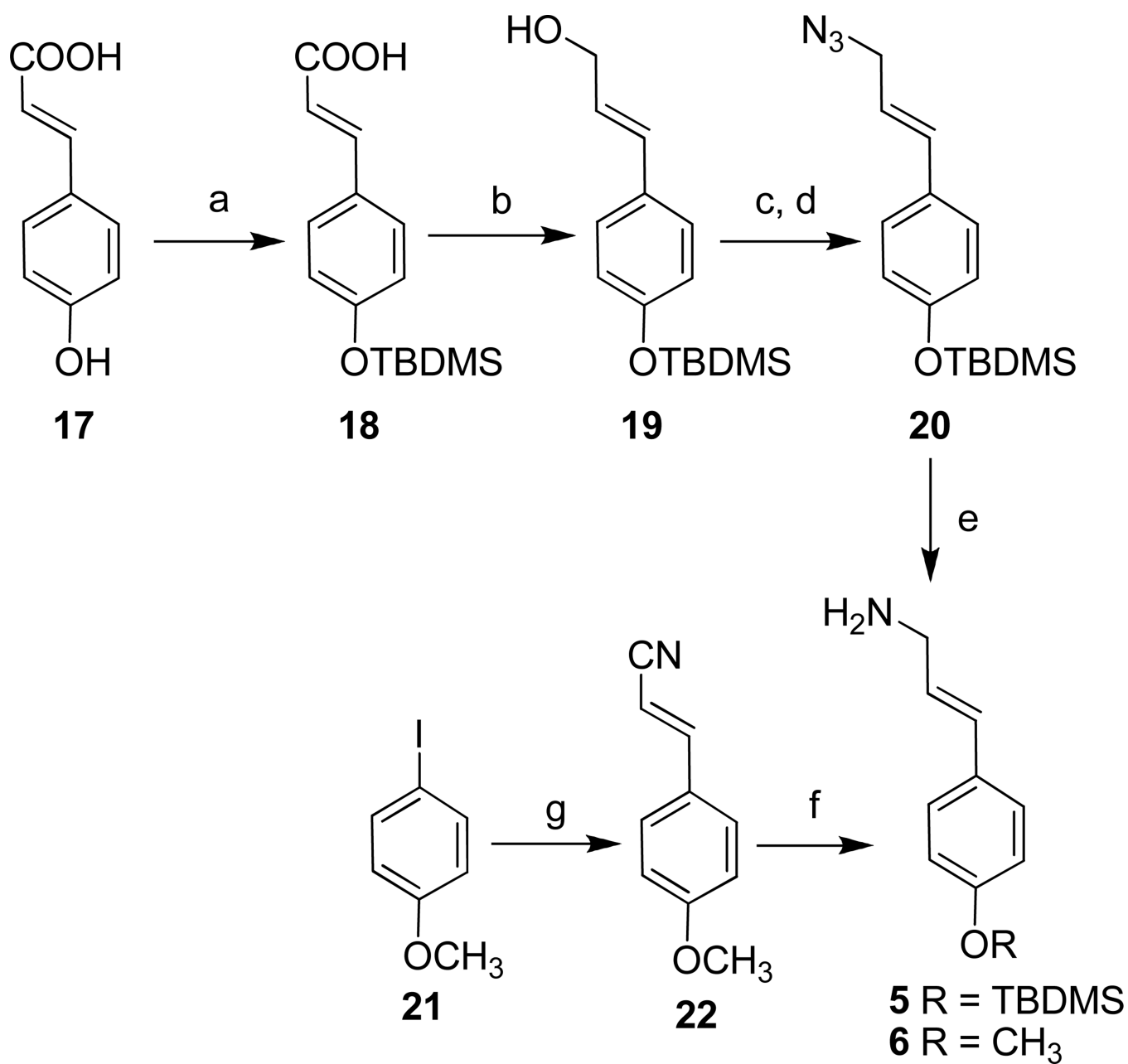
Figure 3. Hemolytic activity of compounds 1 and 2 against Sheep erythrocytes. Hemolysis was calculated using 1% Triton-X as positive control which set to 100% lysis.

**Scheme 1.**

a) $nBuLi$, $BnBr$, THF, $-78\text{ }^\circ\text{C}$ to r.t., 18h, 80% **b)** ethyl 2-cyanoacetate, S_8 , morpholine, EtOH, Δ , 24 – 48h., 39% **c)** p-Nitrophenyl chloroformate, Pyridine, CH_2Cl_2 , r.t. 6 – 12h., 77% **d)** Pyridine, DMAP, THF, r.t. 12–24h., 69–79% **e)** NaOMe, MeOH, reflux, 3h., 76–77%

**Scheme 2.**

a) nBuLi, BnBr, THF, 0–30 °C, 18h, 66% **b)** ethyl 2-cyanoacetate, S₈, morpholine, EtOH, Δ, 24–48h., 38 % **c)** p-Nitrophenyl chloroformate, Pyridine, CH₂Cl₂, r.t. 6–12h, 77% **d)** Pyridine, DMAP, THF, r.t. 12–24h, 80–85% **e)** NaOMe, MeOH, reflux, 3h., 88–90%

**Scheme 3.**

a) TBDMSCl, imidazole/DMF, r.t. 3h., 98% **b)** DIBAL/CH₂Cl₂, -78 °C, 8h., 59% **c)** MsCl, Et₃N/THF **d)** NaN₃/DMF, 82% **e)** Pd / CaCO₃, H₂(g)/EtOH, r.t., 93% **f)** LiAlH₄/Et₂O, r.t. 15min, 61% **g)** Acrylonitrile, Pd(OAc)₂, Bu₄NBr, NaHCO₃/H₂O, 48%

Table 1

MIC and LD₅₀ values for compounds 1–4, 11a, 11b, 16a, and 16b

| | MIC (mg/L) | | | | | | | | | | | LD ₅₀ (mg/L) | Therapeutic Index ^a |
|------------------------|------------------------------------|-----------------------------------|----------------------------------|--------------------------------------|------------------------------------|--------------------------------|---|------------------------------------|-----------------------------------|-----|----|----------------------------|-----------------------------------|
| | <i>S. aureus</i> (VISA Mu50) | <i>S. aureus</i> (MRSA 494) | <i>S. aureus</i> (VRSA MI) | <i>S. pneumoniae</i> (ATCC 49619) | <i>E. faecium</i> (VRE 7303) | <i>E. coli</i> (ATCC 25922) | <i>P. aeruginosa</i> (ATCC 27853) | <i>K. pneumoniae</i> (CEF 2324) | <i>E. aerogenes</i> (CEF 3978) | | | | |
| 1 | 16 | 16 | 8 | 32 | 8 | 32 | 16 | 32 | 32 | 32 | 42 | 5 | |
| 2 | 2 | 2 | 2 | 8 | 4 | >32 | 32 | 32 | 16 | 52 | 26 | | |
| 3 | >32 | >32 | >32 | 32 | >32 | >32 | >32 | >32 | 32 | 120 | 4 | | |
| 4 | >32 | >32 | >32 | >32 | >32 | >32 | >32 | >32 | 8 | 98 | 3 | | |
| 11a | >32 | >32 | >32 | >32 | >32 | >32 | >32 | >32 | >32 | nd | nd | | |
| 11b^b | nd | nd | nd | nd | nd | nd | nd | nd | nd | nd | nd | | |
| 16a | >32 | >32 | >32 | >32 | >32 | >32 | >32 | >32 | >32 | nd | nd | | |
| 16b | >32 | >32 | >32 | >32 | >32 | >32 | >32 | >32 | >32 | nd | nd | | |
| Vancomycin | nd | 0.5 | nd | 0.5 | >32 | nd | >32 | nd | nd | nd | nd | | |

^aTherapeutic index was calculated as the ratio of LD₅₀/MIC for VRSA.^bnot determined due to insolubility of compound in media.