

Neuroprotection of retinal ganglion cells with GDNF –Loaded biodegradable microspheres in experimental glaucoma

Jian-He Xiao, Mao-Nian Zhang

Department of Ophthalmology, the Chinese PLA General Hospital, Beijing 100853, China

Correspondence to: Mao-Nian Zhang. Department of Ophthalmology, the Chinese PLA General Hospital, Beijing 100853, China. zhangmaonian@msn.com

Received:2010-06-28 Accepted:2010-08-23

Abstract

• Glaucoma is the second leading cause of blindness worldwide, and also the most common optic neuropathy. The ultimate cause of vision loss in glaucoma is thought to be retinal ganglion cell (RGC) death. Neuroprotection of RGC is therefore an important goal of glaucoma therapy. Currently, glaucoma treatment relies on pharmacologic or surgical reduction of intraocular pressure (IOP). It is critical to develop treatment approaches that actively prevent the death of RGCs at risk in glaucoma. Neurotrophic factors have the ability to promote the survival and influence the growth of neurons. Neurotrophic factor deprivation has been proposed as one mechanism leading to RGC death in glaucoma. Effective neuroprotection in glaucoma likely requires the consistent availability of the active agent for prolonged periods of time. Biodegradable microspheres are especially attractive as drug delivery vehicles for a number of reasons. Sustained GDNF delivery by biodegradable microspheres offers significant neuroprotection to injured RGC in experimental glaucoma. PLGA microsphere-delivered GDNF represents an important neuroprotective strategy in the treatment of glaucomatous optic neuropathy and provides direction for further investigations of this hypothesis.

• **KEYWORDS:** glaucoma; neuroprotection; biodegradable microspheres; GDNF

DOI:10.3980/j.issn.2222-3959.2010.03.01

Xiao JH, Zhang MN. Neuroprotection of retinal ganglion cells with GDNF–Loaded biodegradable microspheres in experimental glaucoma. *Int J Ophthalmol* 2010;3(3):189–191

INTRODUCTION

Glaucoma is the second leading cause of blindness worldwide, and also the most common optic

neuropathy, affecting about 60 million people worldwide in its most common forms. This figure is expected to rise to 80 million by 2020 [1-3]. In glaucoma, progressive death of the retinal ganglion cells (RGCs) leads to optic nerve degeneration and vision loss. The ultimate cause of vision loss in glaucoma is thought to be retinal ganglion cell (RGC) death. Neuroprotection of RGC is therefore an important goal of glaucoma therapy. Therefore, a major therapeutic aim is to facilitate the survival of RGCs [4]. Currently, glaucoma treatment relies on pharmacologic or surgical reduction of intraocular pressure (IOP). However, many patients suffer progressive visual field loss despite what appears to be adequate control of IOP, as it does in normal tension glaucoma. For these reasons, it is critical to develop treatment approaches that actively prevent the death of RGCs at risk in glaucoma [5-12]. Recently, Jiang *et al* [13] show in a rat model of glaucoma that Intravitreal injections of GDNF-loaded biodegradable microspheres significantly increased the survival of RGCs and their axons, suggesting that GDNF delivered by PLGA microspheres may be useful as a neuroprotective tool in the treatment of glaucomatous optic neuropathy.

Neurotrophic factors have the ability to promote the survival and influence the growth of neurons. Neurotrophic factor deprivation has been proposed as one mechanism leading to RGC death in glaucoma [14-18]. Retrograde axonal transport of neurotrophic factors synthesized in target structures has been specifically associated with RGC survival. In addition, the growing recognition that glaucoma is a form of optic neuropathy suggests that neuroprotection, *i.e.*, therapy directed at preventing neuronal loss, may represent an efficacious adjunctive therapy in this setting [6]. The neurotrophin (NT) hypothesis proposes that the obstruction of retrograde transport at the optic nerve head results in the deprivation of neurotrophic support to retinal ganglion cells (RGC) leading to apoptotic cell death in glaucoma. An important corollary to this concept is the implication that appropriate enhancement of neurotrophic support will

prolong the survival of injured RGC indefinitely. This hypothesis is, perhaps, the most widely recognized theory to explain RGC loss resulting from exposure of the eye to elevated intraocular pressure (IOP)^[19].

Glial-cell-line-derived neurotrophic factor (GDNF) is a 20kDa glycosylated homodimer belonging to the TGF- β super family that was first recognized for its ability to increase the survival of dopaminergic neurons in animal models of Parkinson's disease^[20]. Recent work has established that GDNF signals directly through the cell surface receptor GFR- α and indirectly through the transmembrane Ret receptor tyrosine kinase^[21]. Both receptors have been identified on embryonic chick RGCs as well as on amacrine and horizontal cells^[22]. Exogenous GDNF also increased RGC survival in axotomized rats and in mice following liquid injection and adenoviral transmission^[23-26]. Intravitreal microsphere-delivered GDNF significantly increased long-term RGC survival in the DBA/2J mouse glaucoma model^[27].

Glaucoma is a chronic disease. Effective neuroprotection in glaucoma likely requires the consistent availability of the active agent, such as GDNF, for prolonged periods of time. Neurotrophic factors present in the vitreous humor are rapidly degraded by free extracellular proteases, including any released as a consequence of RGC degeneration. In addition, neurotrophic factors may be taken up and degraded in the retina by resident microglia. Repeated injections of unprotected neurotrophic factors over the life of the patient might not be sufficient to consistently confer a significant visual advantage and could be expected to result in an unacceptable rate of serious complications such as retinal detachment and endophthalmitis.

Biodegradable microspheres are especially attractive as drug delivery vehicles for a number of reasons. First of all, they are relatively inert in the vitreous cavity, inciting only a minimal host immune response. Furthermore, they can be formulated in ways so as to alter the duration and magnitude of drug release. In addition, they can be reproduced with high consistency and at low cost^[27-29].

Recently, Jiang et al. investigated the potential survival enhancing role for glial cell line-derived neurotrophic factor (GDNF) using the hypertonic saline model^[13,30]. After showing that biodegradable microspheres persist in the vitreous for at least 6 weeks, they injected GDNF and control microspheres at 1 week after an initial hypertonic saline injection. At 8 weeks following a second hypertonic saline injection, retinas and optic nerves were collected and analyzed by immunohistochemistry and histology. Jiang

et al^[13] showed that the pressure levels rose gradually, stabilizing at 3 weeks at about twice normal values in all experimental groups. Immunolabeling for GDNF and its receptors showed that these proteins were localized, in part, to RGC. In retinas treated with GDNF spheres, the authors reported decreased nerve head cupping, increased nerve fiber layer thickness, significantly increased inner plexiform layer thickness, more importantly, significantly increased RGCs and axonal survival. In addition, retinal glial activation, which has been proposed as an important factor contributing to RGC death in glaucoma^[31,32], significantly reduced^[13]. These results provide the best evidence so far that sustained GDNF delivery by biodegradable microspheres offers significant neuroprotection to injured RGC in experimental glaucoma. Although these results provide hope for the glaucomatous optic neuropathy, they also have implications for treatment strategies in other retinal degenerative diseases. In summary, these results show that PLGA microsphere-delivered GDNF represents an important neuroprotective strategy in the treatment of glaucomatous optic neuropathy and provides direction for further investigations of this hypothesis.

REFERENCES

- 1 Quigley HA. Number of people with glaucoma worldwide. *Br J Ophthalmol* 1996;80(5):389-393
- 2 Quigley HA, Broman AT. The number of people with glaucoma worldwide in 2010 and 2020. *Br J Ophthalmol* 2006;90(3):262-267
- 3 Dahlmann-Noor AH, Vijay S, Limb GA, Khaw PT. Strategies for optic nerve rescue and regeneration in glaucoma and other optic neuropathies. *Drug Discov Today* 2010;15(7-8):287-299
- 4 Kuehn MH, Fingert JH, Kwon YH. Retinal ganglion cell death in glaucoma: mechanisms and neuroprotective strategies. *Ophthalmol Clin North Am* 2005;18(4):383-395
- 5 Levin LA. Retinal ganglion cells and neuroprotection for glaucoma. *Surv Ophthalmol* 2003;48(Suppl 1):S21-S24
- 6 Levin LA. Neuroprotection and regeneration in glaucoma. *Ophthalmol Clin North Am* 2005;18(4):585-596
- 7 Ritch R. Complementary therapy for the treatment of glaucoma: a perspective. *Ophthalmol Clin North Am* 2005;18(4): 597-609
- 8 Ritch R. Neuroprotection: is it already applicable to glaucoma therapy? *Curr Opin Ophthalmol* 2000;11(2):78-84
- 9 Osborne NN, Chidlow G, Layton CJ, Wood JPM, Casson RJ, Melena J. Optic nerve and neuroprotection strategies. *Eye* 2004;18(11):1075-1084
- 10 Thanos C, Emerich D. Delivery of neurotrophic factors and therapeutic proteins for retinal diseases. *Expert Opin Biol Ther* 2005;5(11):1443-1453
- 11 Quigley HA. New paradigms in the mechanisms and management of glaucoma. *Eye* 2005;19(12):1241-1248
- 12 Veinreb RN, Levin LA. Is neuroprotection a viable therapy for glaucoma? *Arch Ophthalmol* 1999;117(11):1540-1544
- 13 Jiang C, Moore M, Zhang X, Klassen H, Langer R, Young M. Intravitreal injections of GDNF-loaded biodegradable microspheres are neuroprotective in a rat model of glaucoma. *Mol Vis* 2007;13:1783-1792
- 14 Pearson HE, Stoffer DJ. Retinal ganglion cell degeneration following loss of

- postsynaptic target neurons in the dorsal lateral geniculate nucleus of the adult cat. *Exp Neurol* 1992;116(2):163-171
- 15 Schulz M, Raju T, Ralston G. A retinal ganglion cell neurotrophic factor purified from the superior colliculus. *J Neurochem* 1990;55(3):832-841
- 16 Quigley HA, Mckinnon SJ, Zack DJ. Retrograde axonal transport of BDNF in retinal ganglion cells is blocked by acute IOP elevation in rats. *Invest Ophthalmol Vis Sci* 2000;41(11):3460-3466
- 17 Pease ME, Mckinnon SJ, Quigley HA. Obstructed axonal transport of BDNF and its receptor TrkB in experimental glaucoma. *Invest Ophthalmol Vis Sci* 2000;41(3):764-774
- 18 Ko ML, Hu DN, Ritch R. Patterns of retinal ganglion cell survival after brain-derived neurotrophic factor administration in hypertensive eyes of rats. *Neurosci Lett* 2001;305(2):139-142
- 19 Johnson EC, Guo Y, Cepurna WO, Morrison JC. Neurotrophin roles in retinal ganglion cell survival: lessons from rat glaucoma models. *Exp Eye Res* 2009;88(4):808-815
- 20 Lin LF, Doherty DH, Lile JD, Bektesh S, Collins F. GDNF: A glial cell line-derived neurotrophic factor for midbrain dopaminergic neurons. *Science* 1993;260(5111):1130-1132
- 21 Sariola H, Saarna M. Novel functions and signaling pathway for GDNF. *J Cell Sci* 2003;116(Pt 19):3855-3862
- 22 Karlsson M, Lindqvist N, Mayordomo R, Hallbook F. Overlapping and specific patterns of GDNF, c-ret and GFR alpha mRNA expression in the developing chicken retina. *Mech Dev* 2002;114(1-2):161-165
- 23 Yan Q, Wang J, Matheson CR, Ulrich JL. Glial cell line-derived neurotrophic factor (GDNF) promotes the survival of axotomized retinal ganglion cells in adult rats: comparison to and combination with brain-derived neurotrophic factor (BDNF). *J Neurobiol* 1999;38(3):382-390
- 24 Koeberle PD, Ball AK. Effects of GDNF on retinal ganglion cell survival following axotomy. *Vision Res* 1998;38(10):382-390
- 25 Straten, G, Schmeer C, Kretz A, Gerhardt E, Kugler S, Schulz JB, Gravel C, Bahr M, Isenmann S. Potential synergistic protection of retinal ganglion cells from axotomy-induced apoptosis by adenoviral administration of glial cell line-derived neurotrophic factor and X-chromosome-linked inhibitor of apoptosis. *Neurobiol Dis* 2002;11(1):123-133
- 26 Schmeer C, Straten G, Kugler S, Gravel C, Bahr M, Isenmann S. Dose-dependent rescue of axotomized rat retinal ganglion cells by adenovirus-mediated expression of glial cell line-derived neurotrophic factor *in vivo*. *Eur J Neurosci* 2002;15(4):637-643
- 27 Ward MS, Khoobehi A, Lavik EB, Langer R, Young MJ. Neuroprotection of retina ganglion cells in DBA/2J mice with GDNF-loaded biodegradable microspheres. *J Pharm Sci* 2007;96(3):558-568
- 28 Giordano GG, Chevez-Barrios P, Refojo MF, Garcia CA. Biodegradation and tissue reaction to intravitreal biodegradable poly (D,L-lactic-co-glycolic) acid microspheres. *Curr Eye Res* 1995;14(9):761-768
- 29 Moritera T, Ogura Y, Honda Y, Wada R, Hyon SH, Ikada Y. Microspheres of biodegradable polymers as a drug-delivery system in the vitreous. *Invest Ophthalmol Vis Sci* 1991;32(6):1785-1790
- 30 Morrison JC, Moore CG, Deppmeier LMH, Gold BG, Meshul C, Johnson EC. A rat model of chronic pressure-induced optic nerve damage. *Exp Eye Res* 1997;64(1):85-96
- 31 Wang X, Tay SS, Ng YK. An immunohistochemical study of neuronal and glial cell reactions in retina of rats with experimental glaucoma. *Exp Brain Res* 2000;132(4):476-484
- 32 Woldemussie E, Wijono M, Ruiz G. Muller cell response to laser-induced increase in intraocular pressure in rats. *Glia* 2004;47(2):109-119