

Differences in the Population Structure of Invasive Streptococcus suis Strains Isolated from Pigs and from Humans in the Netherlands

Constance Schultsz^{1,2,3}*, Ewout Jansen³, Wendy Keijzers⁴, Anja Rothkamp^{5¤}, Birgitta Duim⁶, Jaap A. Wagenaar⁶, Arie van der Ende^{3,4}

1 Academic Medical Center, Department of Global Health, University of Amsterdam, Amsterdam, The Netherlands, 2 Oxford University Clinical Research Unit, Hospital for Tropical Diseases, Ho Chi Minh City, Vietnam, 3 Center for Infection and Immunity Amsterdam (CINIMA), Department of Medical Microbiology, Academic Medical Center, Amsterdam, The Netherlands, 4 The Netherlands Reference Center for Bacterial Meningitis, Academic Medical Center, Amsterdam, The Netherlands, 5 GD - Animal Health Service, Deventer, The Netherlands, 6 Department of Infectious Diseases and Immunology, Faculty of Veterinary Medicine, Utrecht University, Utrecht, The Netherlands

Abstract

Streptococcus suis serotype 2 is the main cause of zoonotic *S. suis* infection despite the fact that other serotypes are frequently isolated from diseased pigs. Studies comparing concurrent invasive human and pig isolates from a single geographical location are lacking. We compared the population structures of invasive *S. suis* strains isolated between 1986 and 2008 from human patients (N = 24) and from pigs with invasive disease (N = 124) in the Netherlands by serotyping and multi locus sequence typing (MLST). Fifty-six percent of pig isolates were of serotype 9 belonging to 15 clonal complexes (CCs) or singleton sequence types (ST). In contrast, all human isolates were of serotype 2 and belonged to two non-overlapping clonal complexes CC1 (58%) and CC20 (42%). The proportion of serotype 2 isolates among *S. suis* strains isolated from humans was significantly higher than among strains isolated from pigs (24/24 vs. 29/124; P<0.0001). This difference remained significant when only strains within CC1 and CC20 were considered (24/24 vs. 27/37,P = 0.004). The Simpson diversity index of the *S. suis* population isolated from humans (0.598) was smaller than of the population isolated from pigs (0.765, P = 0.05) indicating that the *S. suis* population isolated from infected pigs was more diverse than the *S. suis* population isolated from human patients. *S. suis* serotype 2 strains of CC20 were all negative in a PCR for detection of genes encoding extracellular protein factor (EF) variants. These data indicate that the polysaccharide capsule is an important correlate of human *S. suis* infection, irrespective of the ST and EF encoding gene type of *S. suis* strains.

Citation: Schultsz C, Jansen E, Keijzers W, Rothkamp A, Duim B, et al. (2012) Differences in the Population Structure of Invasive Streptococcus suis Strains Isolated from Pigs and from Humans in the Netherlands. PLoS ONE 7(5): e33854. doi:10.1371/journal.pone.0033854

Editor: Tara C. Smith, University of Iowa, United States of America

Received November 4, 2011; Accepted February 22, 2012; Published May 1, 2012

Copyright: © 2012 Schultsz et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: The work published in this article has received partial funding from the European Commission Seventh Framework Programme under ANTIGONE with project number 278976. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: Anja Rothkamp was employed by GD-Animal Health Service, Deventer, the Netherlands and is currently employed by vaxxinova GmbH diagnostics, Münster, Germany. This does not alter the authors' adherence to all the PLoS ONE policies on sharing data and materials.

1

- * E-mail: schultsz@gmail.com
- ¤ Current address: vaxxinova GmbH diagnostics, Münster, Germany

Introduction

Streptococcus suis is an emerging human pathogen. The number of human *S. suis* infections reported worldwide has increased significantly in the past years, with most cases originating in Southeast Asia [1]. Human *S. suis* infection is acquired through exposure to contaminated pigs or pig derived products. Meningitis and sepsis are the most common clinical manifestations of *S. suis* infections; hearing loss is a frequent complication [1].

S. suis infections are causing huge losses in pig production worldwide. Although multiple putative virulence factors of S. suis have been identified [2,3,4,5], including the polysaccharide capsule [6,7], none of these have been shown to be essential for infection. The eff gene encoding Extracellular Protein Factor (EF), is a virulence-associated gene and is used as a virulence marker although an eff mutant was not severely attenuated in virulence in experimental infection [8,9]. Repeats-containing variants of this gene (eff*, encoding EF*) were found in non-pathogenic or weakly

pathogenic strains [10]. In addition, Muramidase Released Protein (MRP) and the hemolysin (suilysin), have been used as virulence markers [8,11].

Healthy pigs can carry multiple serotypes of *S. suis* in their nasal cavities, tonsils, and upper respiratory, genital and alimentary tracts. Of the 33 serotypes, serotypes 1–9 and 14 are most commonly associated with clinical infections in pigs [12,13]. Across the infectious serotypes, differences in virulence can be observed under experimental conditions. Thus, serotype 2 is considered to be more pathogenic for pigs than serotype 9 [14]. However, the latter serotype is endemic in the pig population causing clinical illness under conditions such as stress and suboptimal farm management, in certain geographical areas [12]. *S. suis* serotype 2 is the most common cause of *S. suis* infection in humans, in contrast to the situation in pigs. Serotypes 1, 4, 14 and 16 have been reported in a limited number of patients only [1]. It is unknown why isolates with serotypes which are highly prevalent amongst diseased pigs, such as serotypes 7 and 9,

have not been isolated from human cases of S. suis infections. One possible explanation for this observation is the lack of human exposure to strains with serotypes 7 and 9 due to a much higher prevalence of serotype 2 strains in pigs in geographical areas where humans are highly exposed. However, another plausible explanation is that humans are more susceptible to infection with serotype 2 strains than to infection with strains of serotypes 7 or 9.

Multi Locus Sequence Typing (MLST) is a recognized tool for characterization of bacterial population structures. A MLST scheme was designed for S. suis [15], and the currently available comprehensive data set indicates that strains belonging to clonal complex 1 (CC1), which includes sequence type 1 (ST1), are associated with invasive disease in both pigs and humans. Other CCs including invasive strains isolated from both humans and pigs include CC25, CC28, and CC104 [16,17].

S. suis infection is a well recognized problem within the porkrearing industry in the Netherlands [18]. In addition, human cases of S. suis infection occur regularly in the Netherlands albeit at a very low frequency [19,20,21]. Surveillance of S. suis infections and storage of bacterial isolates for both animals and humans are centralized. The combined sets of stored invasive isolates provide a unique opportunity to study and compare the population structures of S. suis strains isolated from humans and pigs with invasive disease in a single geographical area in order to identify differences across S. suis strains infecting humans and pigs.

Materials and Methods

Bacterial Strains

All isolates cultured from cerebrospinal fluid or blood from patients with bacterial meningitis in the Netherlands are submitted to the National Reference Laboratory of Bacterial Meningitis (NRLBM) at the Academic Medical Centre, University of Amsterdam, for further typing and storage, as part of the continuous surveillance of bacterial meningitis in the Netherlands. For this study the isolates were anonymized. Additional institutional review board approval is not required for studying anonymized submitted strains without patients data.

All S. suis strains, isolated from cerebrospinal fluid or blood of human patients between 1986 and 2007, which were available at the NRLBM, were included in the study.

S. suis isolates from pigs were obtained from samples sent to the Animal Health Service, Deventer, or the Veterinary Microbiological Diagnostic Center of the Faculty of Veterinary Medicine, Utrecht in the Netherlands. These samples were submitted by veterinary practitioners for diagnosis of S. suis infection. Anonymized non-duplicate invasive S. suis isolates isolated in the period 1996-2008, were used in this study. A representative sample of each year from a total number of 2773 strains was obtained by choosing every 20th isolate irrespective of source or serotype. S. suis isolates obtained from pigs were defined as invasive if they were cultured from brain tissue, cerebrospinal fluid, blood, or joints from pigs with clinical disease compatible with S. suis infection.

Strains were identified using Gram-stain, catalase test and API20Strep (bioMerieux). Serotyping was performed using PCR for detection of serotypes 1 (and 14), 1/2 and 2, 7, and 9 as described by Wisselink and colleagues [22], including positive and negative control strains, and by slide agglutination using serotype specific antibodies for strains which were untypeable by PCR (H. Smith, Central Veterinary Institute, Lelystad, The Netherlands).

Multi Locus Sequence Typing

Multi Locus Sequence Typing (MLST) was performed as described by King et al. [15]. The nucleotide sequences of 7 housekeeping genes were generated by PCR and sequencing, using primers as described in the S. suis MLST scheme, and the sequence type (ST) was determined on the basis of the S. suis MLST database available at www.mlst.net. To identify clonal complexes, i.e groups of related genotypes (STs), isolates were grouped with all isolates present in the S. suis database using the eBURST algorithm (http://eburst.mlst.net) with the software provided by the MLST website. Clonal complexes consisted of sequence types that shared 6 of 7 alleles with at least 1 other sequence type in the complex and named after the putative founder (i.e. the ST that has the greatest number of single-locus variants) of the group or after the most frequent ST of the group. Sequence types that did not group with other sequence types in the database were defined as singletons. For calculation of the genetic diversity amongst isolates cultured from humans or from pigs, the

Simpson's index of diversity was used,
$$D = f\left(1 - \frac{\sum_{i=1}^{p} n_i(n_i - 1)}{N(N - 1)}\right)$$

where n_i is the number of strains belonging to ith type and N is the total number of strains in the sample population [23].

Phylogenetic Analyses

A distance matrix in Nexus format was generated from the set of allelic profiles using SplitsTree at http://pubmlst.org/analysis/. This file was then used for phylogenetic analyses in SplitsTree 4.0, by generating a Neighbor-Joining tree [24].

epf gene Typing

The presence of the epf gene, and its larger size variants designated epf* encoding EF or EF*, was determined for all strains isolated from human patients using PCR, as described by Wisselink et al. [22].

Results

Human Isolates

During the period 1986–2007, 24 S. suis isolates obtained from human patients with meningitis were submitted to the NRLBM. Submitted isolates were equally distributed over time and originated from multiple provinces with a high pig density, throughout the country. All patients were male and were aged between 27 and 66 years. Exposure to pigs or pork was confirmed for 15 (63%) of patients, 14 of whom were keeping pigs or were working in slaughterhouses.

All 24 human strains were identified as serotype 2 strains using PCR, with a PCR product of the expected size for serotype 2 and a negative PCR for detection of serotype 1 (Table 1 and supplementary Table S1). MLST identified two distinct populations. One group of 12 strains (50%) was assigned to ST1 (CC1) whilst the other group of 10 strains (42%) to ST20 (CC20). Two strains were assigned new sequence types (ST134, ST146) on the basis of sequence variation within a single locus when compared with allele sequences of ST1 (single locus variants). Strains of ST1 and ST20 were equally distributed over time without clustering.

All strains with ST1 or single locus variants of ST1 were positive in the PCR for detection of the epf gene. Ten strains carried the epf gene and 4 strains the epf* gene. All strains with ST20 were negative in this PCR.

Table 1. Distribution of *Streptococcus suis* isolates among patients and pigs in the Netherlands, according to serotype and genotype.

host	serotype	Genotype (Clonal Complex)							
		1	13	16	20	25	27	Singleton	total
human	2	14			10				24
pig	1	3	4						7
	1/2	1					1		2
	2	24			3	1			28
	3			2					2
	4				3				3
	7			1		5		3	9
	8			2					2
	9	2		62	1			5	70
	NT							1	1
All		44	4	67	17	6	1	9	148

doi:10.1371/journal.pone.0033854.t001

Pig Isolates

A total of 130 pig isolates were obtained from the collection. Six strains did not yield the expected PCR products in the MLST assays and were excluded from further analysis. *S. suis* strains isolated from pigs had a much more diverse serotype distribution than strains isolated from humans (Table 1 and Table S1). Serotype 9 was the predominant serotype. Amongst 124 pig isolates, 70 (56%) had serotype 9 and 29 (23%) serotype 2 (p<0.001, χ 2 test; human vs pig strains for serotype 2).

Sequence typing showed more diversity for pig isolates than for human isolates. Of 124 strains typed by MLST, 53 (43%) had ST16 and 28 (23%) had ST1. ST20 was found in only 3 strains isolated from pigs (p<0.001, Fisher's exact test; human vs pig strains for ST20). A new ST was defined for each of 19 pig isolates (Table S1). The higher number of genotypes (STs) and CCs among S. suis strains isolated from pigs indicated that the S. suis population isolated from infected pigs was more diverse than the S. suis population isolated from human patients. Comparison of diversity of the S. suis populations isolated from pigs and humans using the Simpson's diversity index, showed a significant difference for both STs (0.775 and 0.598, respectively, P=0.05; T-test) and CCs (0.648 and 0.507, respectively; P=0.05; T-test).

Association between Genotype and Serotype

To assess the clonal complex distribution among the Dutch isolates, isolates were grouped with all isolates (823 isolates on August 2, 2011) present in the MLST database at http://ssuis. mlst.net/, using ST profiles in an eBURST analysis (Figure 1). Among 148 isolates, comprising 30 unique STs, 6 clonal complexes were identified. CC1, CC13, CC16, CC20, CC25 and CC27 comprised 44, 4, 67, 17, 6 and 1 isolates, respectively (Table 1, Table S1). The remaining 9 isolates were singletons. Of 124 pig isolates, 67 (54%) were of CC16. Human isolates were only seen in CC1 and CC20. Among human isolates the proportion of isolates belonging to CC1 (58%) or to CC20 (42%) was significantly higher than among pig isolates, (24%, p<0.002, χ 2 test; and 6% (p<0.0001, Fisher exact test; respectively).

The distribution of clonal complexes across the different serotypes is shown in Table 1 and supplementary Table S1. Of 44 CC1 isolates, 38 (86%) were of serotype 2, 3 of serotype 1, 1 of serotype 1/2 and 2 of serotype 9. Of 17 CC20 isolates, 13 (76%) were of serotype 2, 3 of serotype 4, and one of serotype 9. The vast majority of CC16 isolates (all from pigs) were of serotype 9 (62/67; 93%), 2 were serotype 3, one serotype 7 and two serotype 8.

Thus the two major serotypes 2 and 9 were distributed over multiple generally non-overlapping clonal complexes. Serotype 2 strains were found in CC1 (38/53; 72%), CC20 (13/53; 25%), CC27 (1/53; 2%) and CC29 (1/53; 2%). Serotype 9 comprised isolates of CC1 (2/70; 3%), CC16 (62/70; 89%), and CC20 (1/70; 1%), as well as 5 (7%) singletons.

The proportion of serotype 2 strains among *S. suis* strains isolated from humans was significantly higher than among strains from pigs (24/24 vs 29/124; P<0.001). This difference remained significant when only strains belonging to CC1 and CC20 were considered. Amongst CC1 and CC20 strains, all 24 strains (100%) isolated from humans were of serotype 2, compared with 27 of 37 (73%) pig strains (P=0.004; Fisher's exact).

Phylogenetic Analysis

A Neighbor-Joining cluster analysis of allelic profiles using SplitsTree4 showed groups corresponding with clonal complexes identified with eBURST (Figure 2). Again, clusters comprised multiple STs as well as serotypes. Moreover, some STs were associated with multiple serotypes.

Discussion

We observed significant differences between the *S. suis* populations isolated from human patients and from diseased pigs in the Netherlands. Whilst serotype 9 was predominant amongst invasive pig isolates, serotype 2 was responsible for all human infections, during the same time period. In case series and other reports on *S. suis* infections in humans, serotype 2 strains were responsible for more than 95% of infections [1]. Limited data are available on the prevalence of serotype 2 and other serotypes in the pig populations in areas where most human infections occur, such as in Vietnam and Thailand. A high prevalence of invasive

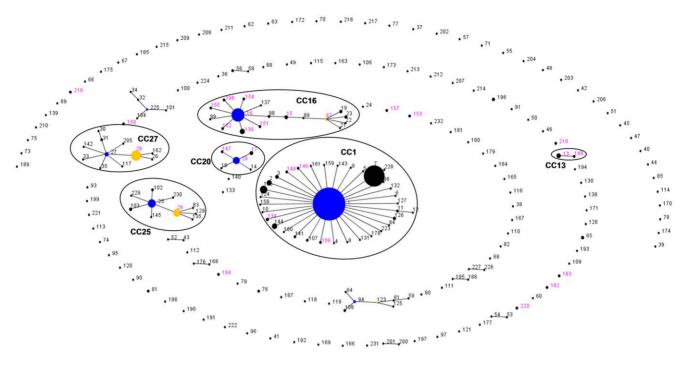


Figure 1. Combined eBURST analysis of the entire *S. suis* MLST database (accessed on 2 August 2011) and the complete collection of strains isolated from pigs and humans with invasive *S. suis* infection described in this study. Clonal complexes and the predicted founder STs are indicated by blue dots. Secondary founders are indicated by yellow dots. The size of the dots is relative to the number of isolates with the respective ST present in the combined databases. Numbers in magenta correspond to the STs of the Dutch isolates in this study. Clonal complexes relevant to this study are circled and labeled. Isolates cultured from human patients in this study are of CC1 (ST1, ST134 and ST148); and of CC20 (ST20).

doi:10.1371/journal.pone.0033854.g001

serotype 2 strains in the pig population may explain a predominance of these strains as a cause of zoonotic infections. Indeed, in a study on the characteristics of invasive strains isolated from diseased pigs in China, the most prevalent serotype was serotype 2, followed by serotype 3 [25], supporting this view. In addition, serotype 2 was the most prevalent serotype carried in the tonsils of healthy slaughterhouse pigs in Vietnam [26]. In contrast, serotype 9 was the most prevalent serotype in diseased pigs in our study performed in the Netherlands, but this serotype was not isolated from human patients. Data on the prevalence of serotype 2 and serotype 9 strains in healthy pigs in the Netherlands are not available. However, since serotype 2 is considered more virulent than serotype 9 [14], one would expect this serotype to be isolated more frequently from diseased pigs than serotype 9 strains if serotype 2 was more prevalent than serotype 9 in the healthy pig population.

The number of human strains studied is relatively small compared with the number of pig strains. In addition, invasive pig strains were isolated in a period only partly spanning the period in which human strains were isolated (1996 to 2007 vs 1986 to 2007). We analyzed the distribution of genotypes of the human isolates in time and found similar distributions in the two periods 1986 to 1995 (6 strains each CC1 and CC 20) and 1996 to 2007 (8 strains CC1, 4 strains CC20, Fisher's exact test not significant). The population of invasive pig *S. suis* strains was more diverse than that of S. suis isolated from human patients as indicated by the estimated Simpson's diversity indices, suggesting selection in the transmission from pigs to human. Human patients in the Netherlands appear to be infected by only two, distinct, clonal complexes of S. suis. Whilst each of these clonal complexes is associated with multiple serotypes, human patients were infected only by S. suis serotype 2. The structure of the serotype 2 capsular polysaccharide of S. suis was recently determined. The backbone sequence was found to be identical to that of Streptococcus agalactiae type VIII and Streptococcus pneumoniae type 23F, whilst the repeating unit contained a terminal 2,6-linked sialic acid molecule. Bacterial capsular sialic acid has been shown to contribute to immune evasion in a number of pathogens [27,28]. Although reports on the possible role of sialic acid in porcine disease pathogenesis during S. suis infection were inconclusive [29,30], it is tempting to speculate on the role of capsular sialic acid in human S. suis infection. Strikingly, the only other serotype of S. suis increasingly found to infect humans, albeit at low numbers, is serotype 14 [31,32,33], which was shown to also contain genes involved in sialic acid synthesis, in contrast to serotype 7 and 9 strains [34]. Serotype 14 strains were not observed in our study. Although the capsule of serotype 1 also contains sialic acid [34], human infection with S. suis of serotype 1 has only been reported once [35]. In our study, 7/124 (6%) of the porcine isolates were of serotype 1 making transfer from pigs to human less likely than that of serotype 2 isolates (29/124 [23%]).

EF, encoded by the epf gene, is associated with but not essential for virulence. Similarly, other proteins, such as Muramidase Released Protein (MRP) and the hemolysin (sly) have been associated with virulence and these proteins are now often used as markers to differentiate virulent and less-virulent strains. Amongst the serotype 2 strains which were isolated from human patients, those with ST20 (CC20) were negative in a PCR for detection of the epf gene or its high molecular weight variants, as opposed to the strain belonging to CC1. This indicates that EF is not required for S. suis serotype 2 invasive disease caused by ST20 strains in human patients and suggests that the presence of these genes is associated with the genotype.

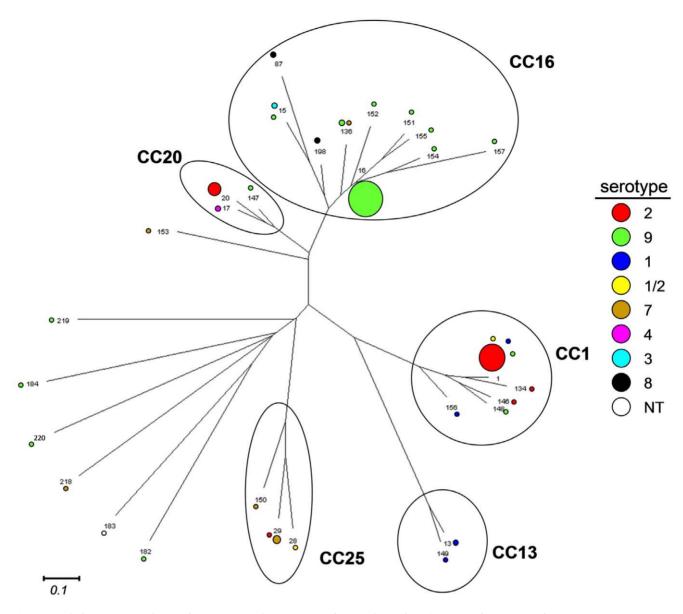


Figure 2. Phylogenetic analyses of concatenated sequences of seven housekeeping gene fragments of *Streptococcus suis* strains isolated from human patients and pigs in the Netherlands. The tree was constructed using Neighbor-Joining algorithm in SplitsTree4 using MLST allelic profiles. Distance matrix was obtained from allelic profiles using the SplitsTree program at http://pubmlst.org/analysis/. ST's comprising the different clonal complexes are circled. Serotypes are indicated by coloured dots with a diameter corresponding to the number of strains. The horizontal line indicates the scale for genetic distance in arbitrary units. doi:10.1371/journal.pone.0033854.g002

Serotype 2 strains which were isolated from human patients were of ST1, ST134 or ST146 (CC1), and of ST20 (CC20). Whilst ST1 is known to contain virulent strains which occur worldwide, invasive isolates with ST20 have not been described before. One isolate of serotype 9 (ST147, CC20) from a diseased pig appeared to be a single locus variant of ST20. These findings, in addition to the observation that multiple serotypes are found in CC16 and CC1, indicate that capsule switch due to horizontal transfer of capsule loci occurs rather frequently in *S. suis*, as was described earlier by King and colleagues [15]. The high prevalence of ST20/CC20 strains amongst the *S. suis* strains isolated from humans in our study, as well as the recently reported strains of clonal complex ST104 isolated from human patients in Thailand, indicate that the population of *S. suis* serotype 2 strains infecting humans is more diverse than previously suspected. Enhanced

surveillance of *S. suis* infections in humans may further increase our knowledge on the population of *S. suis* causing zoonotic infection.

Fifty-four percent of the *S. suis* isolates from pigs belonged to CC16, while the remaining 42% were distributed over five clonal complexes and 9 singletons. Serotypes within CC16 included serotypes 9, 7, and 8. These data indicate that CC16 is another clonal complex representing strains with invasive potential in pigs, in addition to CC1, CC28 and CC25. The high prevalence of ST16 strains in our study, which originated from different farms, regions and were isolated in different years, resulted in ST16 becoming the founder of the CC16 (CC87) complex.

In summary, in this study from the Netherlands, humans were infected by *S. suis* serotype 2 only, while serotype 9 was most prevalent among diseased pigs. Human patients were infected by

two distinct genotypes or clonal complexes (CC1 and CC20). In contrast, strains belonging to CC16 were most frequently isolated from infected pigs. Within CC1 and CC20, serotype 2 was strongly associated with disease in humans. These data indicate that the polysaccharide capsule is an important correlate of human S. suis infection, irrespective of the ST and EF encoding gene type of S. suis strains.

Supporting Information

Table S1 Characteristics of Streptococcus suis strains included in the study. (XLS)

References

- Wertheim HF, Nghia HD, Taylor W, Schultsz C (2009) Streptococcus suis: an emerging human pathogen. Clin Infect Dis 48: 617-625
- Fittipaldi N, Sekizaki T, Takamatsu D, de la Cruz Dominguez-Punaro M, Harel J, et al. (2008) Significant contribution of the pgdA gene to the virulence of Streptococcus suis. Mol Microbiol 70: 1120-1135.
- Fittipaldi N, Sekizaki T, Takamatsu D, Harel J, Dominguez-Punaro Mde L, et al. (2008) D-alanylation of lipoteichoic acid contributes to the virulence of Streptococcus suis. Infect Immun 76: 3587-3594.
- Baums CG, Kaim U, Fulde M, Ramachandran G, Goethe R, et al. (2006) Identification of a novel virulence determinant with serum opacification activity in Streptococcus suis. Infect Immun 74: 6154-6162.
- Li P, Liu J, Zhu L, Qi C, Bei W, et al. (2010) VirA: a virulence-related gene of Streptococcus suis serotype 2. Microb Pathog 49: 305-310.
- Charland N, Harel J, Kobisch M, Lacasse S, Gottschalk M (1998) Streptococcus suis serotype 2 mutants deficient in capsular expression. Microbiology 144 (Pt 2): 325 - 332
- Smith HE, Damman M, van der Velde J, Wagenaar F, Wisselink HJ, et al. (1999) Identification and characterization of the cps locus of Streptococcus suis serotype 2: the capsule protects against phagocytosis and is an important virulence factor. Infect Immun 67: 1750–1756.
- Vecht U, Wisselink HJ, Jellema ML, Smith HE (1991) Identification of two proteins associated with virulence of Streptococcus suis type 2. Infect Immun 59: 3156-3162
- Smith HE, Vecht U, Wisselink HJ, Stockhofe-Zurwieden N, Biermann Y, et al. (1996) Mutants of Streptococcus suis types 1 and 2 impaired in expression of muramidase-released protein and extracellular protein induce disease in newborn germfree pigs. Infect Immun 64: 4409-4412.
- 10. Smith HE, Reek FH, Vecht U, Gielkens AL, Smits MA (1993) Repeats in an extracellular protein of weakly pathogenic strains of Streptococcus suis type 2 are absent in pathogenic strains. Infect Immun 61: 3318-3326.
- 11. Allen AG, Bolitho S, Lindsay H, Khan S, Bryant C, et al. (2001) Generation and characterization of a defined mutant of Streptococcus suis lacking suilysin. Infect Immun 69: 2732-2735
- 12. Wisselink HJ, Smith HE, Stockhofe-Zurwieden N, Peperkamp K, Vecht U (2000) Distribution of capsular types and production of muramidase-released protein (MRP) and extracellular factor (EF) of Streptococcus suis strains isolated from diseased pigs in seven European countries. Vet Microbiol 74: 237-248.
- 13. Fittipaldi N, Fuller TE, Teel JF, Wilson TL, Wolfram TJ, et al. (2009) Serotype distribution and production of muramidase-released protein, extracellular factor and suilysin by field strains of Streptococcus suis isolated in the United States. Vet Microbiol 139: 310-317
- 14. Beineke A, Bennecke K, Neis C, Schroder C, Waldmann KH, et al. (2008) Comparative evaluation of virulence and pathology of Streptococcus suis serotypes 2 and 9 in experimentally infected growers. Vet Microbiol 128:
- 15. King SJ, Leigh JA, Heath PJ, Luque I, Tarradas C, et al. (2002) Development of a multilocus sequence typing scheme for the pig pathogen Streptococcus suis: identification of virulent clones and potential capsular serotype exchange. J Clin
- 16. Takamatsu D, Wongsawan K, Osaki M, Nishino H, Ishiji T, et al. (2008) Streptococcus suis in humans, Thailand. Emerg Infect Dis 14: 181-183.
- Kerdsin A, Dejsirilert S, Puangpatra P, Sripakdee S, Chumla K, et al. (2011) Genotypic profile of Streptococcus suis serotype 2 and clinical features of infection in humans, Thailand. Emerg Infect Dis 17: 835-842.

Acknowledgments

We thank Hilde Smith for assistance with serotyping and for reviewing the manuscript.

Author Contributions

Conceived and designed the experiments: CS BD JW AvE. Performed the experiments: EJ WK. Analyzed the data: CS EJ WK AvE. Contributed reagents/materials/analysis tools: BD JW AR AvE. Wrote the paper: CS AvF.

- 18. van der Peet-Schwering CMC, Binnendijk GP, Kuijken N, Raymakers R, Lamers J (2008) Beheersing van Streptococcus suis bij gespeende biggen door managementmaatregelen, Rapport 119. Animal Sciences Group. Wageningen UR. 119 119.
- 19. Arends JP, Zanen HC (1988) Meningitis caused by Streptococcus suis in humans. Clin Infect Dis 10: 131-137.
- Halaby T, Hoitsma E, Hupperts R, Spanjaard L, Luirink M, et al. (2000) Streptococcus suis meningitis, a poacher's risk. Eur J Clin Microbiol Infect Dis
- 21. van de Beek D, Spanjaard L, de Gans J (2008) Streptococcus suis meningitis in the Netherlands. J Infect 57: 158-161
- Wisselink HJ, Joosten JJ, Smith HE (2002) Multiplex PCR assays for simultaneous detection of six major serotypes and two virulence-associated phenotypes of Streptococcus suis in tonsillar specimens from pigs. J Clin Microbiol 40: 2922-2929.
- Hunter PR, Gaston MA (1988) Numerical index of the discriminatory ability of typing systems: an application of Simpson's index of diversity. J Clin Microbiol 26: 2465-2466.
- 24. Huson DH, Bryant D (2006) Application of phylogenetic networks in evolutionary studies. Mol Biol Evol 23: 254-267.
- 25. Wei Z, Li R, Zhang A, He H, Hua Y, et al. (2009) Characterization of Streptococcus suis isolates from the diseased pigs in China between 2003 and 2007. Vet Microbiol 137: 196-201.
- Ngo TH, Tran TB, Tran TT, Nguyen VD, Campbell J, et al. (2011) Slaughterhouse pigs are a major reservoir of Streptococcus suis serotype 2 capable of causing human infection in southern Vietnam. PLoS One 6: e17943.
- 27. Maisey HC, Doran KS, Nizet V (2008) Recent advances in understanding the molecular basis of group B Streptococcus virulence. Expert Rev Mol Med 10: e27.
- Vimr E, Lichtensteiger C (2002) To sialylate, or not to sialylate: that is the question. Trends Microbiol 10: 254-257.
- Segura M, Gottschalk M (2002) Streptococcus suis interactions with the murine macrophage cell line J774: adhesion and cytotoxicity. Infect Immun 70: 4312-4322
- 30. Charland N, Kobisch M, Martineau-Doize B, Jacques M, Gottschalk M (1996) Role of capsular sialic acid in virulence and resistance to phagocytosis of Streptococcus suis capsular type 2. FEMS Immunol Med Microbiol 14: 195-203
- 31. Mai NT, Hoa NT, Nga TV, Linh le D, Chau TT, et al. (2008) Streptococcus suis meningitis in adults in Vietnam. Clin Infect Dis 46: 659-667
- 32. Haleis A, Alfa M, Gottschalk M, Bernard K, Ronald A, et al. (2009) Meningitis caused by Streptococcus suis serotype 14, North America. Emerg Infect Dis 15: 350 - 352
- 33. Kerdsin A, Oishi K, Sripakdee S, Boonkerd N, Polwichai P, et al. (2009) Clonal dissemination of human isolates of Streptococcus suis serotype 14 in Thailand. J Med Microbiol 58: 1508-1513.
- Smith HE, de Vries R, van't Slot R, Smits MA (2000) The cps locus of Streptococcus suis serotype 2: genetic determinant for the synthesis of sialic acid. Microb Pathog 29: 127-134.
- 35. Nghia HD, Hoa NT, Linh le D, Campbell J, Diep TS, et al. (2008) Human case of Streptococcus suis serotype 16 infection. Emerg Infect Dis 14: 155-157.