

Genome Sequence of the Model Hyperthermophilic Archaeon *Thermococcus litoralis* NS-C

Andrew F. Gardner, Sanjay Kumar, and Francine B. Perler

New England Biolabs, Inc., Ipswich, Massachusetts, USA

The hyperthermophilic archaeon *Thermococcus litoralis* strain NS-C, first isolated in 1985, has been a foundational organism for archaeal research in biocatalysis, DNA replication, metabolism, and the discovery of inteins. Here, we present the genome sequence of *T. litoralis* with a focus on the replication machinery and inteins.

Thermococcus litoralis strain NS-C was isolated from a shallow submarine hot spring at Lucrino Beach near Naples, Italy (1), and successfully grown in culture (14). Since then, *T. litoralis* has been the focus of studies on biocatalysis (10), archaeal metabolism (2, 3, 6, 7, 9, 11, 13, 17, 21), DNA replication (4, 5, 8, 12, 20), and protein splicing (15).

T. litoralis was grown at the New England BioLabs fermentation facility as described previously (1). Genomic DNA was purified, and DNA libraries were constructed for paired-end Illumina sequencing-by-synthesis at Cofactor Genomics, Inc. (St. Louis, MO). Sequencing reads were assembled using the Cofactor Genomics short oligonucleotide analysis package (SOAP)-based *de novo* assembly pipeline and annotated using the NCBI Prokaryotic Genomes Automatic Annotation Pipeline (PGAAP).

The *T. litoralis* chromosome was assembled into 77 contigs with a total assembled genome length of 2,309,438 bp, similar to other sequenced *Thermococcus* genomes. The chromosome contains 2,724 predicted open reading frames, 47 tRNA genes, 1 16S and 1 23S rRNA gene, and 2 5S rRNA genes. The *T. litoralis* genome G+C content is 43%, which is similar to the 38% calculated from the melting point (T_m) determination reported by Belkin and Jannasch in 1985 (1).

The majority of *T. litoralis* replication proteins had the highest similarity to homologs from *Thermococcus sibiricus* MM 739, although the DNA polymerase D large subunit was most similar to its homolog in *Pyrococcus yayanosii* CH1, DNA polymerase B (commercialized as Vent DNA polymerase) to its homolog in *Thermococcus* species strain AM4, and replication factor C (RFC) small subunit to its homolog in *Methanocaldococcus jannaschii*. A lesion bypass DNA polymerase gene similar to *dpo4* or *dinB* was not found.

T. litoralis played a pivotal role in our understanding of inteins. Inteins are mobile genetic elements that can be horizontally transferred after cleavage of the intein insertion site in empty host protein genes (termed exteins) by the associated homing endonuclease domain present in many inteins using a simple double-strand break repair mechanism (16). This insertion is apparently neutral, because the intein protein can splice itself out of the precursor protein to yield a fully functional mature extein protein. The inteins found in *T. litoralis* DNA polymerase B (Vent DNA polymerase) were the first to have Ser or Thr as nucleophiles instead of Cys at both splice junctions (15), and the Tli pol-2 intein was the first to have different N- and C-terminal nucleophiles (Ser and Thr, respectively). This allowed positioning of the splice site prior to the intein N-terminal Ser or Cys nucleophile, which was

later confirmed by N-terminal sequencing of excised inteins. Thirteen inteins are present in the *T. litoralis* genome in 8 different genes. These inteins all contain dodecapeptide (DOD) family homing endonuclease domains except for the Tli MCM-1 intein. All are standard, class 1 inteins except for the Tli KlbA intein, which splices using the class 2 mechanism (18, 19). The Tli MCM-2 intein insertion site (VLVLADMGIA/CIDEIDKMSD, CDC21-e) has not been previously noted in InBase, the on-line intein database (16).

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [AHVB0000000](https://doi.org/10.1093/nar/36/11/AHVB0000000). The version described in this paper is the first version, AHVB01000000.

ACKNOWLEDGMENTS

We thank Dean Wilbur and Tim Bowen at New England BioLabs for technical assistance, Tom Evans, Harriet Strimpel, and Bill Jack for helpful discussions, and Don Comb for fostering a supportive research environment.

This study was funded by New England BioLabs, Inc.

REFERENCES

1. Belkin S, Jannasch HW. 1985. A new extremely thermophilic, sulfur-reducing heterotrophic, marine bacterium. *Arch. Microbiol.* **141**:181–186.
2. De Stefano L, et al. 2008. Enzymes and proteins from extremophiles as hyperstable probes in nanotechnology: the use of D-trehalose/D-maltose-binding protein from the hyperthermophilic archaeon *Thermococcus litoralis* for sugars monitoring. *Extremophiles* **12**:69–73.
3. Diez J, et al. 2001. The crystal structure of a liganded trehalose/maltose-binding protein from the hyperthermophilic archaeon *Thermococcus litoralis* at 1.85 Å. *J. Mol. Biol.* **305**:905–915.
4. Gardner A, Jack W. 1999. Determinants of nucleotide sugar recognition in an archaeon DNA polymerase. *Nucleic Acids Res.*
5. Gardner AF, Joyce CM, Jack WE. 2004. Comparative kinetics of nucleotide analog incorporation by vent DNA polymerase. *J. Biol. Chem.* **279**:11834–11842.
6. Greller G, Horlacher R, DiRuggiero J, Boos W. 1999. Molecular and biochemical analysis of MalK, the ATP-hydrolyzing subunit of the trehalose/maltose transport system of the hyperthermophilic archaeon *Thermococcus litoralis*. *J. Biol. Chem.* **274**:20259–20264.
7. Imamura H, et al. 2003. Crystal structures of 4- α -glucanotransferase

Received 6 February 2012 Accepted 13 February 2012

Address correspondence to Andrew F. Gardner, gardner@neb.com.

Copyright © 2012, American Society for Microbiology. All Rights Reserved.

doi:10.1128/JB.00123-12

- from *Thermococcus litoralis* and its complex with an inhibitor. *J. Biol. Chem.* **278**:19378–19386.
8. Kong H, Kucera RB, Jack WE. 1993. Characterization of a DNA polymerase from the hyperthermophile archaea *Thermococcus litoralis*. Vent DNA polymerase, steady state kinetics, thermal stability, processivity, strand displacement, and exonuclease activities. *J. Biol. Chem.* **268**:1965–1975.
 9. Lee S-J, et al. 2003. TrmB, a sugar-specific transcriptional regulator of the trehalose/maltose ABC transporter from the hyperthermophilic archaeon *Thermococcus litoralis*. *J. Biol. Chem.* **278**:983–990.
 10. Littlechild JA. 2011. Thermophilic archaeal enzymes and applications in biocatalysis. *Biochem. Soc. Trans.* **39**:155–158.
 11. Ma K, Robb FT, Adams MW. 1994. Purification and characterization of NADP-specific alcohol dehydrogenase and glutamate dehydrogenase from the hyperthermophilic archaeon *Thermococcus litoralis*. *Appl. Environ. Microbiol.* **60**:562–568.
 12. Mattila P, Korpela J, Tenkanen T, Pitkänen K. 1991. Fidelity of DNA synthesis by the *Thermococcus litoralis* DNA polymerase—an extremely heat stable enzyme with proofreading activity. *Nucleic Acids Res.* **19**:4967–4973.
 13. Mukund S, Adams MW. 1993. Characterization of a novel tungsten-containing formaldehyde ferredoxin oxidoreductase from the hyperthermophilic archaeon, *Thermococcus litoralis*. A role for tungsten in peptide catabolism. *J. Biol. Chem.* **268**:13592–13600.
 14. Neuner A, Jannasch H, Belkin S, Stetter K. 1990. *Thermococcus litoralis* sp. nov.: a new species of extremely thermophilic marine archaeobacteria. *Arch. Microbiol.* **153**:205–207.
 15. Perler FB, et al. 1992. Intervening sequences in an archaea DNA polymerase gene. *Proc. Natl. Acad. Sci. U. S. A.* **89**:5577–5581.
 16. Perler FB. 2002. InBase: the Intein Database. *Nucleic Acids Res.* **30**:383–384.
 17. Rinker KD, Kelly RM. 1996. Growth physiology of the hyperthermophilic archaeon *Thermococcus litoralis*: development of a sulfur-free defined medium, characterization of an exopolysaccharide, and evidence of biofilm formation. *Appl. Environ. Microbiol.* **62**:4478–4485.
 18. Southworth MW, Benner J, Perler FB. 2000. An alternative protein splicing mechanism for inteins lacking an N-terminal nucleophile. *EMBO J.* **19**:5019–5026.
 19. Tori K, et al. 2010. Splicing of the mycobacteriophage Bethlehem DnaB intein: identification of a new mechanistic class of inteins that contain an obligate block F nucleophile. *J. Biol. Chem.* **285**:2515–2526.
 20. Tubeleviciute A, Skirgaila R. 2010. Compartmentalized self-replication (CSR) selection of *Thermococcus litoralis* Sh1B DNA polymerase for diminished uracil binding. *Protein Eng. Des. Sel.* **23**:589–597.
 21. Xavier KB, Peist R, Kossmann M, Boos W, Santos H. 1999. Maltose metabolism in the hyperthermophilic archaeon *Thermococcus litoralis*: purification and characterization of key enzymes. *J. Bacteriol.* **181**:3358–3367.