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# Human longevity and common variations in the *LMNA* gene: a meta-analysis

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# Summary

A mutation in the *LMNA* gene is responsible for the most dramatic form of premature aging, Hutchinson-Gilford progeria syndrome (HGPS). Several recent studies have suggested that protein products of this gene might have a role in normal physiological cellular senescence. To explore further LMNA's possible role in normal aging, we genotyped 16 SNPs over a span of 75.4 kb of the LMNA gene on a sample of long-lived individuals (US Caucasians with age 95 years, N=873) and genetically matched younger controls (N=443). We tested all common non-redundant haplotypes (frequency 0.05) based on subgroups of these 16 SNPs for association with longevity. The most significant haplotype, based on 4 SNPs, remained significant after adjustment for multiple testing (OR = 1.56,  $P=2.5\times10^{-5}$ , multiple-testing-adjusted P=0.0045). To attempt to replicate these results, we genotyped 3448 subjects from four independent samples of long-lived individuals and control subjects from 1) the New England Centenarian Study (NECS) (N=738), 2) the Southern Italian Centenarian Study (SICS) (N=905), 3) France (N=1103), and 4) the Einstein Ashkenazi Longevity Study (N=702). We replicated the association with the most significant haplotype from our initial analysis in the NECS sample (OR = 1.60, P=0.0023), but not in the other three samples (P>.15). In a meta-analysis combining all five samples, the best haplotype remained significantly associated with longevity after adjustment for multiple testing in the initial

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and follow-up samples (OR = 1.18,  $P=7.5\times10^{-4}$ , multiple-testing-adjusted P=0.037). These results suggest that *LMNA* variants may play a role in human lifespan.

#### Keywords

longevity gene; human; longevity; genetics

# Introduction

Single gene mutations have been shown to extend the lifespan of worms, yeast, and mice (Kenyon 2005). Similarly, twin studies have shown that 25–30% of the variance in human lifespan is due to genetic differences (Herskind et al. 1996; Ljungquist et al. 1998; Skytthe et al. 2003). With the completion of the International HapMap Project, power to detect common variants for common and complex diseases has increased (Manolio et al. 2008), and the search for genetic contributors to human aging and lifespan has been accelerated. The results thus far have been somewhat discouraging: only APOE and FOXO3A have been consistently associated with longevity across populations in candidate gene studies (Christensen et al. 2006; Novelli et al. 2008). Even strong candidates such as the WRN gene associated with the premature aging syndrome, Werner syndrome (Castro et al. 2000), have displayed inconsistent results (Christensen et al. 2006). Similarly, all longevity-associated SNPs reaching genome-wide significance in two recently published genome-wide association studies were associated with the APOE gene (Deelen et al. 2011; Nebel et al. 2011). However, because variants with subtler associations with longevity will not be detectable at genome-wide significance levels with conventional sample sizes, candidate gene studies remain a useful approach.

One major candidate gene for longevity that has not yet been explored is *LMNA*. Defects in the *LMNA* gene, which encodes the nuclear envelope proteins lamin A and lamin C, have been associated with at least 13 diseases, including Hutchinson-Gilford progeria syndrome (HGPS) (Gruenbaum *et al.* 2005; Capell & Collins 2006). HGPS is a rare but devastating condition in which symptoms resembling premature aging appear at a very young age, typically within a year of birth. Due to accelerated cardiovascular disease leading to eventual heart attack or stroke, individuals with HGPS have an average life expectancy of 13 years (Capell *et al.* 2007; Merideth *et al.* 2008).

Scientists have long questioned whether HGPS is truly a model of normal human aging. Although displaying many features typically associated with normal aging (alopecia, skin wrinkling, atherosclerosis), HGPS apparently lacks other common features such as tumors or dementia. However, recent evidence obtained since the identification of mutations in LMNA as the cause of HGPS suggests that there may in fact be a molecular relationship. For example, there is evidence that a very small amount of the mutant lamin A protein produced in HGPS, termed "progerin," is produced in normal cells (Scaffidi & Misteli 2006; Cao et al. 2007). Levels of the progerin transcript increase with age in normal cells (Rodriguez et al. 2009), and work using a monoclonal antibody specific for progerin has demonstrated that progerin accumulates in human skin in an age-dependent manner (McClintock et al. 2007). Further work in normal human fibroblasts has demonstrated that progerin production is activated by progressive telomere damage during cellular senescence (Cao et al. 2011). Finally, it has been established that numerous other abnormalities present in HGPS are also common in cells from aged individuals. These include nuclear blebbing, particular epigenetic changes, and increased levels of DNA damage. Most remarkably, suppressing the production of the mutant progerin protein through the use of an antisense morpholino reversed all of these abnormalities in cell culture (Scaffidi & Misteli 2006).

The corollary to this observation is the possibility that variants of *LMNA* that produce less progerin might be protective and thus promote a longer lifespan. To investigate further the potential links of HGPS, *LMNA*, and progerin to the normal aging process, we sought to determine whether common variants in the *LMNA* gene might be associated with extreme human longevity. Towards this end, we tested variants of the *LMNA* gene for association with extreme longevity in five independent samples of long-lived individuals (LLI) and control subjects.

# Results

Characteristics of the five study samples are shown in Table 1. In the Stage 1 sample (US Caucasians with age 95 years, N=873) and genetically matched younger controls (N=443), we genotyped 16 SNPs covering two coherent linkage disequilibrium (LD) blocks, corresponding to the LMNA gene and promoter region (Figure 1). 2414 non-redundant haplotypes (i.e., haplotypes not in perfect linkage disequilibrium with another haplotype) involving two to sixteen SNPs were present with estimated frequency 0.05 in our Stage 1 analysis. We tested all SNPs and all 2414 haplotypes for association with longevity. SNP 6 showed the strongest single-SNP association (P=0.0137) in Stage 1, but was not significant upon correction for multiple testing via permutation tests. However, an excess of significant haplotypes over what is expected under no association was evident. We examined the proportion of haplotypes that were significant at a range of significance thresholds ( $\alpha =$ 0.00005, 0.0001, 0.0005, 0.001, 0.005, 0.01, and 0.05) and in all cases observed significantly more than expected under no association. P-values for association tests of each of these haplotypes are plotted against their quantiles in Figure S1. The most significant haplotype, involving SNPs 6, 7, 13, and 15 (GTCT), remained significant after adjustment for multiple testing via permutation tests ( $P = 2.5 \times 10^{-5}$ , multiple-testing-adjusted P =0.0045). Haplotypes involving SNPs 4 and 5 were also significant, and the most significant 6-SNP haplotype involved SNPs 4, 5, 6, 7, 13, and 15 (GAGTCT) ( $P = 4.0 \times 10^{-5}$ ). 140 nonredundant haplotypes involving different combinations of two to six of these six SNPs were present with estimated frequency>0.05.

To follow up on these 140 haplotypes, we selected SNPs 4, 5, 6, 7, 13, and 15 for genotyping in our four Stage 2 samples (Table 1). One of the six SNPs was not successfully genotyped in the French sample, but for purposes of haplotype inference we were able to impute genotypes for this SNP using MaCH 1.0 (Li *et al.* 2010) in conjunction with phased haplotype information from the HapMap CEU sample (estimated  $r^2$  between true underlying genotype and imputed genotype = 0.97). Additional *LMNA* SNPs beyond SNPs 4, 5, 6, 7, 13, and 15 were also genotyped based on convenience and were included in the SNP association analysis in Table 2 but not the haplotype analysis. SNP association results for all five samples are shown in Table 2. When we combined results across samples, SNP 15 showed the strongest single-SNP association in the meta-analysis (OR=1.17, *P*=0.0056) with odds ratios ranging from 1.09–1.22 in the four samples tested, but was not significant after correction for multiple testing via permutation tests (multiple-testing-adjusted *P*=0.105).

In replication analyses of the 140 haplotypes of SNPs 4, 5, 6, 7, 13, and 15 in the four Stage 2 samples, an excess of significant haplotypes was observed in the NECS sample (Figure S2; P < 0.01) for all significance thresholds examined ( $\alpha = 0.00005$ , 0.0001, 0.0005, 0.001, 0.005, 0.001, and 0.05) but not in the other three Stage 2 samples. When the results for all 140 followed-up haplotypes were combined across all five samples in a meta-analysis (Table 3 and Figure 2), the original GTCT haplotype of SNPs 6, 7, 13, and 15 was again the most significant ( $P=7.5\times10^{-4}$ ) and remained significant after permutation-based adjustment for multiple testing in both stages of the analysis (multiple-testing-adjusted P=0.037).

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If the genetic load is greater for older individuals, restricting the definition of LLI to include only centenarians should lead to a stronger genetic effect (Tan *et al.* 2008). For the four samples where age and sex information was available (Stage 1, NECS, SICS, and the Ashkenazi sample), we re-performed the association test for the GTCT haplotype within each sample after restricting the definition of LLI to include individuals with age 100. However, the odds ratio for GTCT did not change substantially in any of the four samples. We also performed a sex-stratified version of the haplotype analysis. Similar results were observed in the sex-stratified analysis for each sample, and results were similar for males and females within each sample.

The Stage 1 sample had previously been tested for the presence of population stratification between LLI and control subjects (see Supporting Information) (Geesaman et al. 2003). However, to ensure that the association with the GTCT haplotype was not due to subtle population stratification, we re-fit the model including the principal components of genotypes from 73 ancestry-informative markers (AIMs) from a published panel of European substructure AIMs (Tian et al. 2008). Inclusion of the first four principal components from the AIM genotypes as covariates in the analyses did not substantially diminish the strength of the association. For the 1242 individuals for whom AIMs were available, a similar association between the GTCT haplotype and longevity was observed before (OR=1.47, P=0.00039) and after (OR=1.46, P=0.00060) inclusion of the principal components. Although a Tracy-Widom test (Patterson et al. 2006) suggested that four principal components was sufficient to account for potential population stratification, the observed result was robust to the number of principal components included in the model (P ranged from 0.00057 to 0.00062 when one to ten principal components were included). Similarly, the top two principal components from genome-wide data were available for 671 of the NECS individuals; inclusion of these components as covariates in the analysis slightly strengthened the result. In both cases, the inclusion of principal components in the analyses affected the results only minimally, suggesting that the observed haplotype association in the Stage 1 and NECS samples is not driven by population stratification. Similarly, analysis of the first two principal components did not reveal evidence of population stratification in a genome-wide study of SICS cases and controls (Malovini et al. 2011). The French sample had been previously tested for population stratification on 57 randomly selected microsatellite markers. As described in Supporting Information,  $\chi^2$  1.00 in all cases, indicating that the French LLI and controls were well-matched.

## Discussion

We observed evidence for association with longevity involving a 4-SNP haplotype, and an excess of significant associations among the same set of correlated haplotypes, in an initial sample of centenarians and younger controls. This result was replicated in one of four follow-up samples, and the association was sufficiently strong in those two samples to remain significant when combined in a meta-analysis of all five.

Had our initial haplotype association been unreplicated in all four follow-up samples, it would have been difficult to argue against the possibility of a false positive. However, the probability of observing a second false positive of similar strength is low (<0.005): in the same 10,000 permutations used to adjust the meta-analysis *P*-value for multiple testing, the best haplotype observed in the Stage 1 sample was followed up with an association in the same direction with |Z| > 3.05 in one or more of the four follow-up samples only 46 times.

Other possible explanations for incomplete replication include undetected population structure, use of under-powered replication samples, and heterogeneity between samples. However, adjustment for potential population stratification via principal components only

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modestly diminished the associations reported here. Similarly, differential power based on sample size differences does not appear to explain the incomplete replication since the NECS sample was arguably the least well-powered due to small sample size and skewed case-control ratio. Heterogeneity between samples can occur for a variety of reasons. For example, the GTCT haplotype could tag a functional variant in some samples but not others due to differential LD patterns. Alternatively, a functional variant tagged by GTCT could correlate with the longevity phenotype only in the NECS and Stage 1 samples; this can result from differential ascertainment of cases or from between-sample differences in environmental exposures (McCarthy et al. 2008). One potential source of heterogeneity is differential ascertainment of LLI, since the minimum age for inclusion varied by study (Table 1). Notably, the association with GTCT was observed in two of the three samples that had ascertained older sets of LLI (mean age > 100); Tan et al. (2008) has argued that the power of association studies investigating exceptional longevity markedly increases with the use of centenarian subjects compared to nonagenarians. However, in the three samples where individual ages were available, we observed similar results when the definition for cases was narrowed to include only centenarians. Another potential source of heterogeneity is environmental differences across samples due to region of origin. The association between GTCT and longevity was observed in two of the three US samples, but not in the European samples. One region-specific environmental exposure which could interact with agingrelated genes to affect longevity is diet. Dietary differences between Europe and the US have been proposed as explanations for regional differences in rates of coronary mortality (Renaud & de Lorgeril 1992; de Lorgeril et al. 1994), and caloric restriction and several dietary antioxidants, including resveratrol, found in red wine, can alter expression of agingrelated genes in the hearts and brains of mice (Park et al. 2009).

Replicating genetic association results across diverse populations has generally been difficult in the longevity literature (Christensen *et al.* 2006). For example, a strong association between longevity and a *FOXO3A* SNP (Willcox *et al.* 2008) replicated in some samples (Flachsbart *et al.* 2009; Li *et al.* 2009), including males from the SICS (Anselmi *et al.* 2009), but was not present in others (Kuningas *et al.* 2007) (Pawlikowska *et al.* 2009; Soerensen *et al.* 2010), including females from the SICS (Anselmi *et al.* 2009), although two of these studies reported associations with other *FOXO3A* SNPs Reasons proposed for the lack of replication in some samples included those discussed above, as well as lack of gender stratification in some analyses (Anselmi *et al.* 2009), although as described in Results, this does not appear to be an issue in the present study.

The meta-analysis results presented here complement a growing body of experimental evidence (Scaffidi & Misteli 2006; McClintock *et al.* 2007; Cao *et al.* 2011) in suggesting a potential role for *LMNA* and progerin in aspects of normal aging. However, given the incomplete replication we observed, further work is needed to validate and interpret the results reported here. One of the four SNPs in our reported haplotype, rs4641, is a coding SNP for which allele-specific expression has been demonstrated (Rodriguez & Eriksson, 2011), with 2.3-fold greater expression for transcripts with the C allele, which is part of the GTCT haplotype associated with greater longevity in our meta-analysis. Another SNP in our reported haplotype, rs915179, associated with longevity in a recent genome-wide association study (Sebastiani et al. 2012), though this study included most of our NECS cases and controls along with other subjects. Additional association studies of *LMNA* variants in independent samples and additional experiments focusing on functional consequences of the associated variants would be particularly valuable in establishing and elucidating the role of *LMNA* in the normal aging process.

#### **Experimental Procedures**

#### Samples

We analyzed one initial (Stage 1) and four follow-up (Stage 2) samples of LLI and younger controls. Samples collected for the Stage 1 study by Elixir Pharmaceuticals were Caucasians primarily of Northern European ancestry. Samples were also obtained from the New England (NECS), French, Southern Italian (SICS), and Ashkenazi Jewish Centenarian Studies. Details of sample collection, as well as steps taken to ensure that the study samples did not overlap, can be found in Supporting Information.

#### **SNP** selection

We downloaded all SNP genotype data for the CEU population from -40 kb to +10 kb of *LMNA* from the website of the International HapMap (International HapMap Consortium 2005): http://www.hapmap.org. These genotype data in raw HapMap format were uploaded to Tagger (http://www.broad.mit.edu/mpg/tagger/), and our multiplex assay design (see Supporting Information for details) was created from the list of tag SNPs to capture all variants of interest in our *LMNA* region of interest. We used  $r^2 > 0.8$  as the threshold for accepting tag SNPs. 10 SNPs were selected in HapMap Phase I, release #16 along with the synonymous coding SNP, rs4641, which was not in HapMap Phase I. Upon release of HapMap Phase II, release #19 we selected SNPs in the *LMNA* region again and added 5 additional SNPs not tagged ( $r^2 < 0.8$ ) by the previously selected SNPs. Based on Phase II of the HapMap, these 16 SNPs offered comprehensive coverage and representation of common variants in the *LMNA* region and were genotyped in Stage 1.

For Stage 2 samples, we genotyped the 6 SNPs from the most significant 6-SNP haplotype in Stage 1. Additional SNPs from the original 16 were genotyped in some Stage 2 samples based on convenience; these SNPs were tested for association (Table 2) but were not included in the haplotype analysis. Details on SNP genotyping are provided in Supporting Information.

#### Statistical analysis

We used the MaCH 1.0 haplotype inference and imputation software (Li *et al.* 2010) to infer 16-SNP haplotypes for all individuals in Stage 1, and 6-SNP haplotypes for all individuals in Stage 2. For individuals with missing genotypes, the most likely genotype was inferred based on haplotype frequencies within the sample. We imputed genotypes for SNP rs1468772 that was not genotyped in the French sample, using MaCH (Li *et al.* 2010) in conjunction with phased haplotype information from the HapMap CEU sample.

We performed association tests in all five samples and combined the samples in a metaanalysis using R (R Development Core Team 2011). We performed all SNP and haplotype association tests as Cochran-Armitage tests (Cochran 1954; Armitage 1955) for a linear trend between the number of copies of a reference allele (0, 1, or 2) and the likelihood that an individual belongs to the case or control group. We first tested each SNP for association with centenarian status. We computed meta-analysis statistics for all followed-up SNPs and haplotypes as the weighted sum of z-statistics from the trend tests. The weights for sample *i* 

were defined as 
$$\sqrt{\frac{n_i}{n}}$$
, where  $n_i$  = sample size for samples  $i = 1, ..., 5$  and  $n = \sum_{i=1}^{3} n_i$ .

To account for the testing of multiple correlated SNPs and haplotypes in Stage 1, we performed permutation tests by 1) randomly permuting case-control status and 2) recomputing the test statistics for all SNPs and haplotypes. Based on 10,000 permutations, we

then computed adjusted *P*-values that reflected the probability of observing each result by chance alone. To establish whether our most extreme *P*-value was significantly different than what would be expected if no true association existed, we computed an empirical *P*-value as the proportion of permutations in which a more extreme result was observed in any of the SNPs or haplotypes analyzed. To establish whether a significant excess of haplotypes with *P*< $\alpha$  was observed for a given  $\alpha$ -level, we computed empirical *P*-values as the proportion of permutations in which we observed as many haplotypes with *P*< $\alpha$  as in our original data.

To account for multiple testing in the two-stage analysis, we performed permutation tests in R to obtain an empirical *P*-value for the best observed meta-analysis test statistic. Permutation tests for two-stage studies are complicated by the fact that only partial genotyping is performed in the second stage of the study. Our strategy for dealing with this was, for permutation purposes only, to use MACH 1.0, along with inferred haplotypes from the HapMap CEU sample, to impute the full 16-SNP haplotypes for all Stage 2 samples. We were then able to perform a full two-stage analysis in each permutation as follows:

- 1. Randomly re-assign case-control status within each of the five samples.
- **2.** Compute trend test statistics for all haplotypes (or SNPs) in the permuted Stage 1 sample.
- **3.** To mimic the selection of 6 SNPs for genotyping in the Stage 2 samples, as was done in the original study, select the most significant 6-SNP haplotype from the (permuted) Stage 1 analysis for follow-up. Perform trend tests of association for all 6 SNPs and for all haplotypes composed of 2–6 SNP combinations of these 6 SNPs in the four permuted Stage 2 samples.
- **4.** Compute meta-analysis statistics for all haplotypes (or SNPs) tested on all five permuted samples; record the value of the most significant meta-analysis test statistic.

We performed 10,000 permutations, and recorded the best meta-analysis test statistic for each permutation. We computed the empirical *P*-value as the proportion of permutations in which the best statistic from the permutation was greater in magnitude than the best statistic from the original data analysis.

#### **Population stratification**

As described in Supporting Information, the Stage 1 and French samples had previously been tested for the presence of population stratification between LLI and control subjects and the Stage 1 sample had been genetically matched. There was no evidence of population stratification in the French sample (mean  $\chi^2$ =1.00 across 57 random microsatellite markers) or in the genetically matched Stage 1 sample (mean  $\chi^2$ =0.95 across 80 random SNPs) (Geesaman *et al.* 2003).

For the Stage 1 sample, we also genotyped AIMs from a published panel specific to European genetic substructure (Tian *et al.* 2008); genotyping and quality control described in Supporting Information. We used R to compute principal components and to identify the most important principal components via a Tracy-Widom test (Patterson *et al.* 2006). To perform an analysis adjusted for population stratification, we included the top principal-components as covariates in the original analysis.

Principal components based on genotypes from the 370K Illumina SNP chip (collected as part of a separate study – Sebastiani *et al.* 2012) were available for a subset of the NECS sample. We used EIGENSTRAT (Price *et al.* 2006) to compute the principal components

and identified the principal components that were most informative based on visual inspection of a scree plot. To perform an analysis adjusted for population stratification, we included the top principal components as covariates in the original analysis.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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#### Figure 1.

Linkage disequilibrium ( $R^2$  in HapMap CEU sample) between SNPs in *LMNA* region. Labeled SNPs indicate SNPs tested in Table 2. Encircled SNP names indicate SNPs involved in GTCT haplotype in Table 3.



#### Figure 2.

Forest plot of odds ratios for meta-analysis of haplotype GTCT on SNPs 6, 7, 13, and 15 across five samples. For each sub-sample, the black box is centered at the estimated odds ratio, and the gray line represents a 95% confidence interval. Area of the black box varies in proportion to the precision of each estimate. For the meta-analysis, the black diamond is centered at the estimated odds ratio and spans the 95% confidence interval for the meta-analysis odds ratio.

#### Table 1

# Samples tested

| Sample                | Ν            | Age range | Mean age |
|-----------------------|--------------|-----------|----------|
| Stage 1 (US)          | 873 LLI      | 95        | 101.5    |
|                       | 443 controls | <50       | 38.6     |
| NECS (US)             | 545 LLI      | 96–113    | 103.2    |
|                       | 193 controls | 55–92     | 76.8     |
| French                | 557 LLI      | 99        | 103.1    |
|                       | 546 controls | 18–70     | 51.2     |
| SICS (Southern Italy) | 455 LLI      | 90-109    | 97.6     |
|                       | 450 controls | 18–46     | 32.9     |
| Ashkenazi Jewish      | 354 LLI      | 95-109    | 97.9     |
|                       | 348 controls | 43–90     | 70.7     |

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| <u> </u>      | live samples         |
|---------------|----------------------|
|               | <b>DINFS ACTOSS</b>  |
| n 1           | <i>P</i> -values lor |
| aindana atau  | meta-analysis        |
| Turned to a f | I renu test and      |

| SNP | RefSNP ID  | Stage 1<br>(N=1316) | NECS<br>(N=738) | French<br>(N=1103) | SICS<br>(N=905) | Ashkenazi<br>Jewish<br>(N=702) | Meta-analysis |
|-----|------------|---------------------|-----------------|--------------------|-----------------|--------------------------------|---------------|
| -   | rs2275075  | 0.54                |                 |                    |                 | 0.73                           | 0.78          |
| 7   | rs3814314  | 0.017               |                 |                    |                 | 0.81                           | 0.039         |
| ю   | rs11578696 | 0.21                |                 | 0.93               |                 |                                | 0.39          |
| 4   | rs4661146  | 0.12                | 0.17            | 0.42               | 0.65            | 0.72                           | 0.97          |
| 5   | rs10737170 | 0.11                | 0.016           | 0.84               | 0.54            | 0.45                           | 0.075         |
| 9   | rs915179   | 0.014               | 0.0086          | 0.96               | 0.66            | 0.59                           | 0.020         |
| ٢   | rs2485662  | 0.054               | 0.11            | 0.81               | 0.82            | 0.35                           | 0.026         |
| ×   | rs547915   | 0.11                |                 | 0.84               |                 | 0.84                           | 0.29          |
| 6   | rs582690   | 0.078               |                 |                    |                 | 0.42                           | 0.35          |
| 10  | rs6686943  | 0.51                |                 |                    |                 |                                |               |
| 11  | rs508641   | 0.014               |                 |                    |                 | 0.95                           | 0.044         |
| 12  | rs12117552 | 0.21                |                 |                    |                 | 0.26                           | 0.71          |
| 13  | rs4641     | 0.50                | 0.14            | 0.61               | 0.30            | 0.45                           | 0.98          |
| 14  | rs12063072 | 0.36                |                 |                    |                 | 0.060                          | 0.065         |
| 15  | rs1468772  | 0.034               | 0.35            |                    | 0.41            | 0.14                           | 0.0056        |
| 16  | rs6669212  | 0.041               |                 | 0.48               |                 | 0.36                           | 0.18          |

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Meta-analysis of haplotype GTCT on SNPs 6, 7, 13, and 15 across five samples

0.0045 (single-stage)

**P-value** 0.000025

4.21

1.56

0.235 0.239

1316

738

0.0023

1.60

0.94

3.05 0.074

0.207

0.208 0.175 0.114

1103

NECS French 0.29 0.16

1.05

1.14

0.157 0.139

905 702

SICS

-1.40

0.80

0.037 (two-stage)

0.00075

3.37

1.18

Full meta-analysis

Ashkenazi

Adjusted P-value

Z statistic

OR

**control** 0.163 0.163

case

z

Stage 1

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|-----------------|
|                 |
|                 |