

Photic Desynchronization of Two Subgroups of Circadian Oscillators in a Network Model of the Suprachiasmatic Nucleus with Dispersed Coupling Strengths

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Abstract

The suprachiasmatic nucleus (SCN) is the master circadian clock in mammals and is composed of thousands of neuronal oscillators expressing different intrinsic periods. These oscillators form a coupled network with a free-running period around 24 h in constant darkness and entrainable to the external light-dark cycle (T cycle). Coupling plays an important role in setting the period of the network and its range of entrainment. Experiments in rats have shown that two subgroups of oscillators within the SCN, a ventrolateral (VL) subgroup that receives photic input and a dorsomedial (DM) subgroup that is coupled to VL, can be desynchronized under a short (22-h) T cycle, with VL entrained to the cycle and DM free-running. We use a modified Goodwin model to understand how entrainment of the subgroups to short (22-h) and long (26-h) T cycles is influenced by light intensity, the proportion of neurons that receives photic input, and coupling heterogeneity. We find that the model's critical value for the proportion of photically-sensitive neurons is in accord with actual experimental estimates, while the model's inclusion of dispersed coupling can account for the experimental observation that VL and DM desynchronize more readily under the 22-h than under the 26-h T cycle. Heterogeneous intercellular coupling within the SCN is likely central to the generation of complex behavioral patterns.

Citation: Gu C, Liu Z, Schwartz WJ, Indic P (2012) Photic Desynchronization of Two Subgroups of Circadian Oscillators in a Network Model of the Suprachiasmatic Nucleus with Dispersed Coupling Strengths. PLoS ONE 7(5): e36900. doi:10.1371/journal.pone.0036900

Editor: Shin Yamazaki, Vanderbilt University, United States of America

Received: November 2, 2011; **Accepted:** April 9, 2012; **Published:** May 16, 2012

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Funding: This work was supported by the National Natural Science Foundation (NNSF) of China under grant No.10975053 (<http://www.nsf.gov.cn/Portal0/default106.htm>) and by the PHD program scholarship fund of East China Normal University under grant No. 2010027 (<http://www.ecnu.edu.cn/english/>). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

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Introduction

Circadian (~24 h) rhythms in physiological and behavioral measures are universal in living things, reflecting the period of the earth's rotation. In mammals, circadian rhythms are regulated by a master clock in the suprachiasmatic nucleus (SCN) of the hypothalamus, composed of approximately 20,000 neuronal oscillators; SCN neurons are nonidentical, express different intrinsic periods, and are coupled together to form a network with a coherent output [1]. The period of the network's output signal is adaptable. Under constant darkness, the rhythm has a free-running period close to 24 h; whereas under an external light-dark cycle (T cycle), it is precisely entrained to a period identical to the external cycle.

The SCN network is heterogeneous [2,3,4]. It can be divided into distinct functional subgroups, including a ventrolateral part (VL), which receives photic input from the retina, and a dorsomedial part (DM), which is coupled to VL; both VL and DM contribute to the generation of overt circadian rhythms in physiological and behavioral measures. Peptide neurotransmitters differ between the VL and DM subdivisions, with neurons that express vasoactive intestinal polypeptide (VIP) in the VL and arginine vasopressin in the DM. Periods may vary in different

regions of the SCN, with DM running faster than VL in tissue slices [5]. Gamma aminobutyric acid (GABA) neurons are present throughout the SCN and may play a role in coupling the two subdivisions [6]. It has been shown that the circadian oscillation between VL and DM can desynchronize with exposure to short T cycles [7] or after a phase shift of the light-dark cycle [6,8,9]; the VL appears to set the final phase of the SCN after the phase shift [6,8,9].

Much experimental [10,11,12] and theoretical [13,14,15] work has been motivated by a desire to understand how this heterogeneous SCN network is reliably entrained and able to generate a coherent output signal, and neuropeptidergic mechanisms appear to be necessary elements [16,17,18]. Modeling studies suggest that the circadian clock's free-running period is proportional to the average intercellular coupling strength [13] and that coupling governs the clock's range of entrainment to T cycles [15]. However, coupling strength between cells in the SCN network is unlikely to be uniform. The effects of heterogeneous coupling on network synchronization have been studied previously in multi-oscillator models. Daido considered the dispersion of coupling strengths in the Kuramoto model and studied the synchronization property of the network [19,20]; coupling

strength between two oscillators was chosen from a normal distribution. Hong and Strogatz considered a heterogeneous network with excitatory (positive) and inhibitory (negative) coupling in the Kuramoto model to understand the relative contributions of excitatory and inhibitory properties on network synchronization [21]. Our recent work (C.G. and Z.L.) has demonstrated that the dispersion of coupling strengths between SCN cellular oscillators can influence the emergent free running period of the network [22]. To our knowledge, however, there has been no work on the relationship between coupling dispersion and network entrainment.

We examine this issue in the present work, inspired by an interesting experiment performed by de la Iglesia et al. [7] in which rats were exposed to an artificially short 22-h T cycle (11 h light alternating with 11 h darkness). Individual animals expressed two separate circadian motor activity rhythms, with one rhythm entrained by the light and oscillating with a period equal to the external cycle, while the other was not entrained and expressed a period greater than 24 h. Analyses of SCN gene expression suggested that these two motor activity rhythms reflected the stable forced desynchronization of VL and DM subdivisions, respectively. Here we model how entrainment of the subdivisions is influenced by coupling dispersion, as well as by the proportion of cellular oscillators that receive photic input (i.e., that are within VL) and the light intensity.

We use the Goodwin model, a network model of coupled oscillators that has been widely used to describe the mammalian circadian clock [13,22,23,24,25,26] (defined in Methods). An individual cellular oscillator of the Goodwin model has three variables: a clock gene mRNA, a clock protein, and a transcriptional inhibitor, all of which form a transcription-translation negative feedback loop. It is assumed that light induces the clock gene mRNA, that a neurotransmitter is increased by the clock gene mRNA, and that neurotransmitters from different neurons form a mean field that couples the neurons together. We consider that pN neurons receive photic input, where N is the total number of neurons in the SCN network and p is the ratio of the number of VL neurons to the total number of SCN neurons. We take T cycles of 22 h and 26 h as examples, i.e., symmetrically distant from 24 h. We chose mean field coupling for all of the neurons in the Goodwin model. The coupling strength g_i of all the N neurons satisfies a normal distribution with mean value $\langle g \rangle$ and deviation η .

Results

T-cycle Entrainment of an SCN Network without Dispersion of Coupling Strengths

To determine the effects of p and light intensity, K_f , on the entrainment of VL and DM to T cycles, we have numerically simulated the Goodwin model with no dispersion of coupling strengths, i.e., $\eta=0$. Figure 1 shows the mean field time series of VL and DM oscillations in the 22-h light-dark cycle. Similar to previous observations [13], we find that the time series show quasi-periodic behavior with low light intensity. In (A), the behavior of VL follows the 22-h cycle and sustains a stable phase relationship to it, while the behavior of DM loses its phase relationship to the cycle and runs with a period close to the intrinsic period of the network. This dissociation mimics the forced desynchronization of motor activity rhythms in rats under such a T cycle, as noted previously [7,27]. When p is increased, both VL and DM can be entrained, as in (B). Here the peak of the mean field time series of VL appears around the onset of darkness, whereas that of the DM is phase delayed. This change also can be implemented by K_f . If

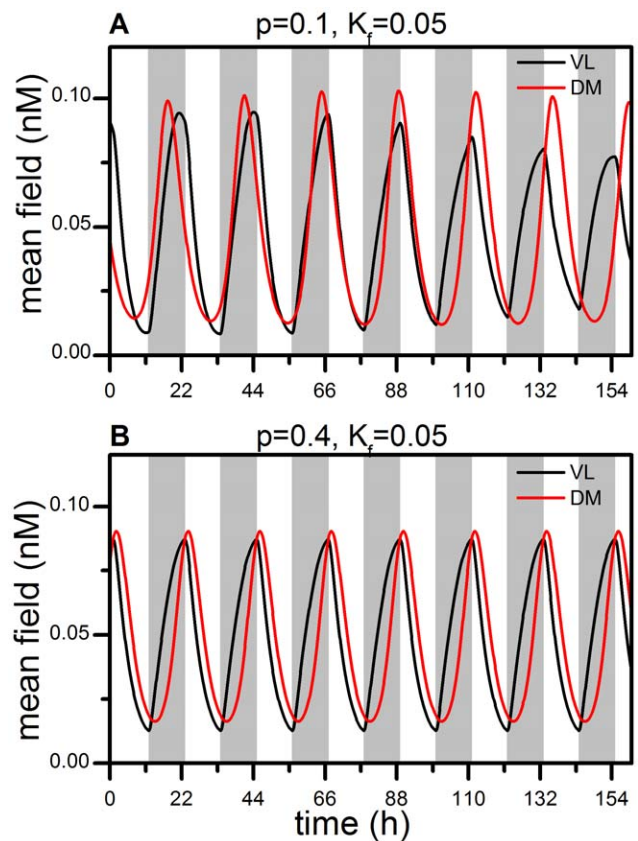


Figure 1. Mean field oscillations of VL and DM during a 22-h T cycle. (A) VL follows the T cycle, whereas DM free runs for the parameters $p=0.1$ and $K_f=0.05$. (B) Both VL and DM follow the T cycle for the parameters $p=0.4$ and $K_f=0.05$. The dispersion of the coupling strengths, η , is set to zero in both (A) and (B). The grey bar indicates the dark phase, and the white bar the light phase, of the T cycle. doi:10.1371/journal.pone.0036900.g001

K_f is reduced, neither VL nor DM entrain to the 22-h cycle. If K_f is increased, both VL and DM can be entrained, as in (B). In sum, both the number of neurons receiving light and the light intensity are important factors for entrainment of the entire SCN network to the T-cycle.

To understand the influence of the parameters p and K_f on entrainment, we have calculated the phase diagram of the period of the mean fields of VL and DM in the p - K_f plane under short and long T cycles, i.e., of 22 h and 26 h, with $\eta=0$ (Figure 2). (A) and (C) show that the behavior of VL follows the T cycle for all values of p , provided that the light intensity is greater than a critical value, such as $K_f > 0.02$. When $K_f < 0.02$, the period of VL may not be entrained to the T cycle, depending on p ; for example, in Fig. 2A, the period of VL can be 23 h or 24 h. For DM to follow the cycle, however, p also must be larger than some threshold; that is, there must be a sufficient number of light-receiving neurons in VL in order to drive the neurons in DM. For a given level of K_f , increasing p may allow the entire network to entrain to the driving T cycle. Surprisingly, when p is decreased under the 26-h T cycle (D), there is a threshold of p at which the period of DM suddenly jumps to a value of 20.8 h to form a locking ratio of 4:5 with the 26-h T cycle; with further decreased p , the period monotonically increases to reach a value of 24 h at $p=0$.

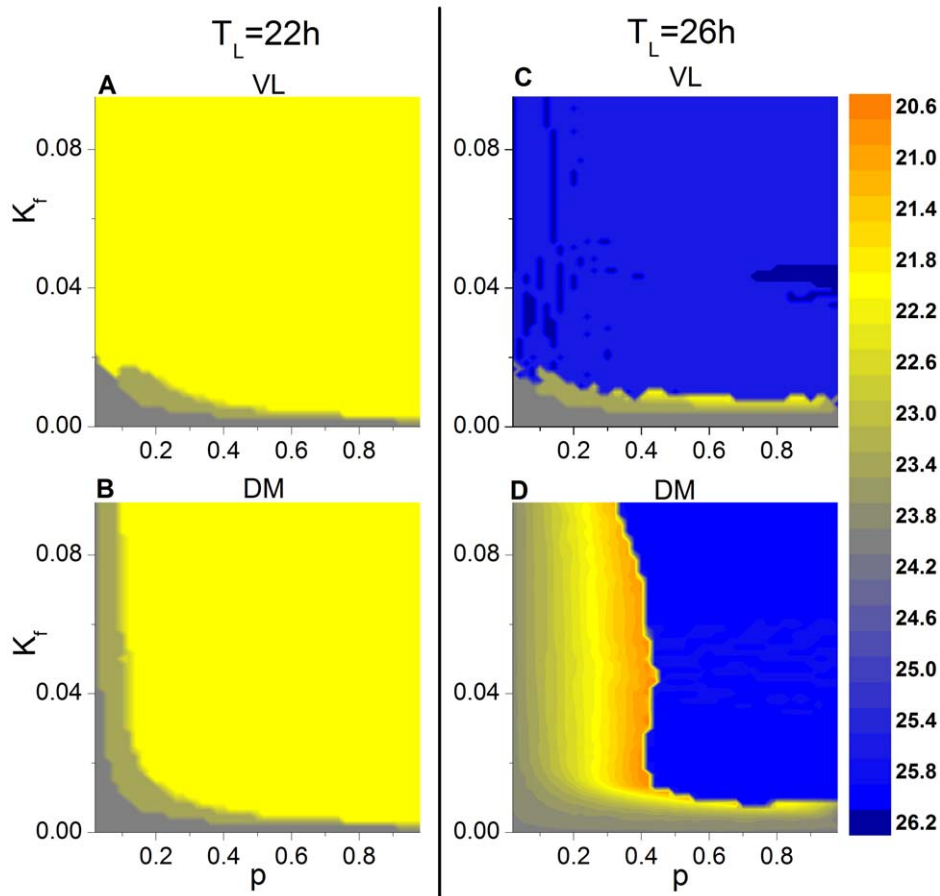


Figure 2. Period of the mean fields of VL and DM in the p - K_f plane. The case for the 22-h T cycle is shown for VL (A) and DM (B), and the case for the 26-h T cycle is shown for VL (C) and DM (D). The coupling strengths are identical for all the oscillators (i.e., $\eta = 0$). Entrainment of the sub-network to the 22-h cycle is represented by the yellow region, and entrainment of the sub-network to the 26-h cycle is represented by the blue region.

doi:10.1371/journal.pone.0036900.g002

A comparison of (B) and (D) shows that the threshold of p for entrainment of DM to the 26 h T cycle is greater than that for the 22 h cycle, suggesting that desynchronization between VL and DM might be more likely under long than under short T cycles. Experimentally, however, this appears not to be the case [28,29], prompting us to consider the influence of heterogeneous coupling strengths on the behavior of the SCN network.

T-cycle Entrainment of an SCN Network with Dispersion of Coupling Strengths

Figure 3 shows the phase diagram of the period of the mean fields of VL and DM in the p - K_f plane using two values for η . Although qualitatively similar to the diagrams in Figure 2, there are quantitative differences when network coupling strengths are dispersed. In the case of the 22-h T cycle, entrainment is only modestly affected; in contrast, in the case of the 26-h T cycle, increased coupling dispersion significantly reduces the critical value of p for DM entrainment, suggesting that the network can be entrained to the long T cycle with a relatively lower K_f .

For weak K_f and different values of η , we find that the critical p (p_c) for the 22-h and 26-h T cycles reaches approximately the values of 0.28 and 0.20, respectively. Figure 4 represents the variation in p_c for different values of K_f and η . In the case of the 22-h T cycle (A), there is little variation in p_c for different K_f , e.g., p_c is between 0.14 and 0.24 for a $K_f = 0.04$. However, p_c does

change significantly in the case of the 26-h T-cycle (B). Thus, dispersion of coupling strengths affects entrainment in an asymmetric way, with an influence that is larger for the long than for the short T cycle.

Instead of randomly assigning coupling values to the network, we also studied the network by selectively assigning coupling values. In two separate trials, we assigned the strongest coupling values to either VL or DM. The phase diagram of the period of the mean fields of VL and DM in the p - K_f plane was similar to that previously reported.

We also simulated the network with dispersed oscillator periods, rather than dispersed coupling strengths, by selecting different values for the standard deviation of period (σ) for each individual oscillator, such that the period distribution has a mean of 24 h with variability. Without dispersed coupling, we do not observe DM entrainment to the long T cycle at any p until σ is increased to a value greater than 5 h. Since such a large non-identical intrinsic period is not realistic, the dispersion of coupling strengths is likely a crucial factor affecting the entrainment of the network to different T cycles.

Importantly, dispersion of coupling influences the mean field amplitude (Figure 5). The amplitude of VL as a function of p changes modestly as the dispersion η is increased in the 22-h as well as the 26-h T cycle (A and C). On the other hand, for DM in the 22-h T cycle (B), amplitude decreases as η increases. In the 26-

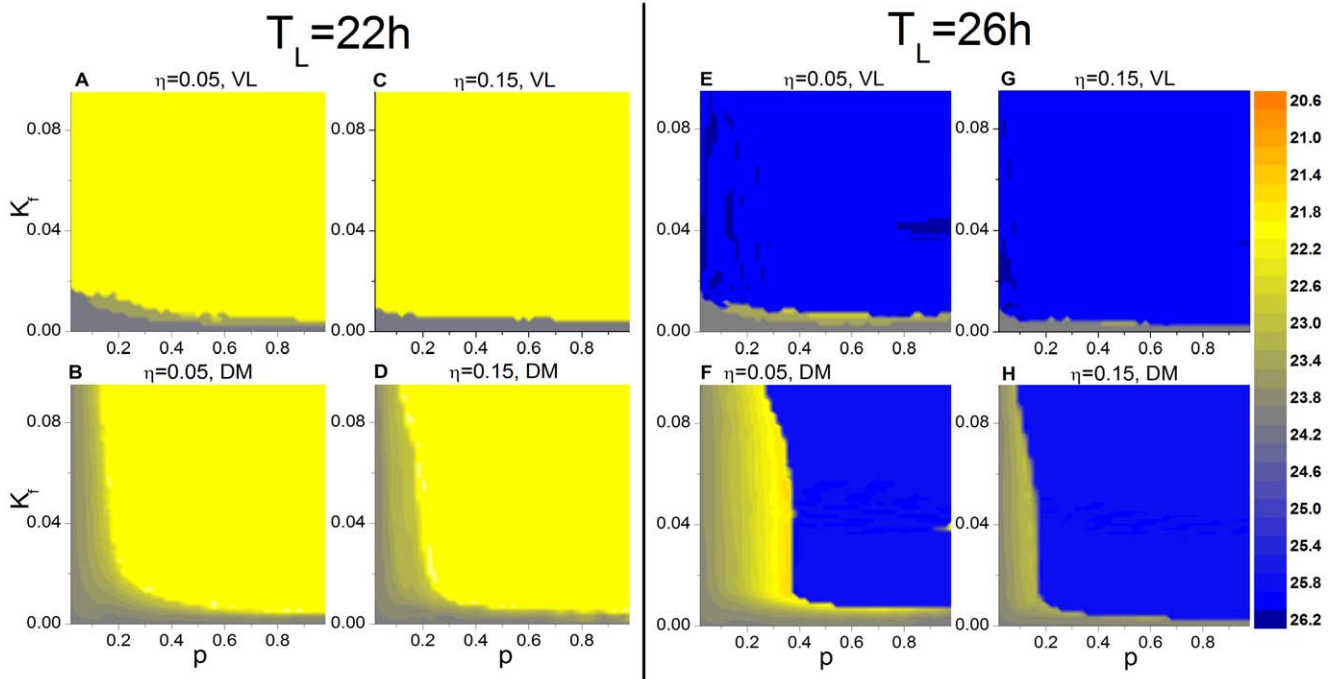


Figure 3. Effect of coupling dispersion on the period of the mean fields of VL and DM in the $p-K_f$ plane. The case for the 22-h T cycle is shown for VL (A) and DM (B) with $\eta = 0.05$ and for VL (C) and DM (D) with $\eta = 0.15$. The corresponding case for the 26-h T cycle is represented in (E) - (H).
doi:10.1371/journal.pone.0036900.g003

h T cycle (D), DM amplitude as a function of p changes dramatically, with relatively diminished amplitude as p is increased; dispersion η counteracts this effect. The enhancement of DM amplitude by increased η in the 26-h cycle could be due to enhanced phase synchronization of the oscillators in the network, increased amplitude of the individual oscillators, or both. To begin

to distinguish among these possibilities, we studied the effect of dispersed coupling on the order parameter, a measure that represents phase synchronization of the network.

Effect of Coupling Dispersion on the Order Parameter of the Network

Order parameter characterizes the synchronization property of a network [30,31], and it is defined here by estimating the phases of the oscillators in VL and DM (see Methods). The order parameter will be unity if all oscillators in the network are perfectly synchronized and zero if they are completely uncorrelated. When their behavior is between these two extremes, the order parameter will be in $(0, 1)$, i.e., representing a phase difference between VL and DM or desynchronization of individual oscillators within VL and/or DM.

We have studied the influence of η on the order parameter. Figure 6 shows the dependence of order parameter R on the parameters p and K_f in the $p-K_f$ plane. To reveal the effect of coupling dispersion, we have considered two cases, one with $\eta = 0.15$ and the other with $\eta = 0.0$. Under the 22-h T cycle, coupling dispersion reduces R for larger p and K_f values; whereas under the 26-h T cycle, coupling dispersion enhances R . As p and K_f increase from $(0,0)$, the relationship between VL and DM changes; comparison of Figure 6 with Figures 2 and 3 visualizes the regions where $R < 1$, i.e., either when VL and DM express different periods or when VL and DM express the same period but with a large phase difference between them. Thus, for the 22-h cycle, although higher p and K_f values enhance both VL and DM entrainment to the cycle, the reduction of R with dispersed coupling suggests that individual oscillators are not fully synchronized within the network, with a greater vulnerability to perturbations of light intensity. For the 26-h cycle, coupling dispersion synchronizes network oscillation for $p > 0.6$; the gradual

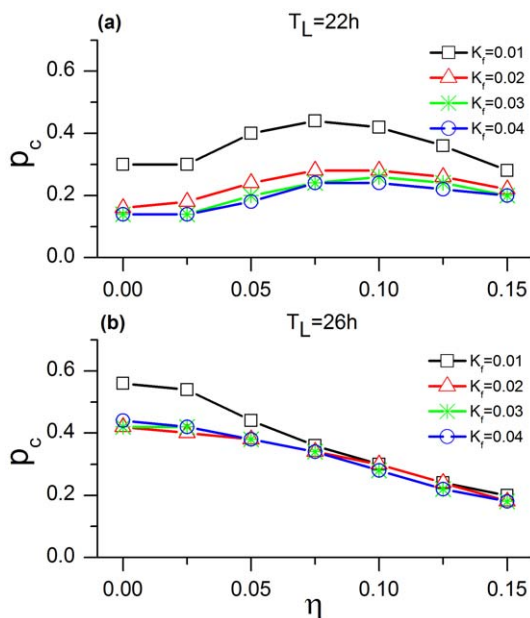


Figure 4. Effect of coupling dispersion on the critical p . Shown are the cases for the 22-h (A) and 26-h (B) T cycles.
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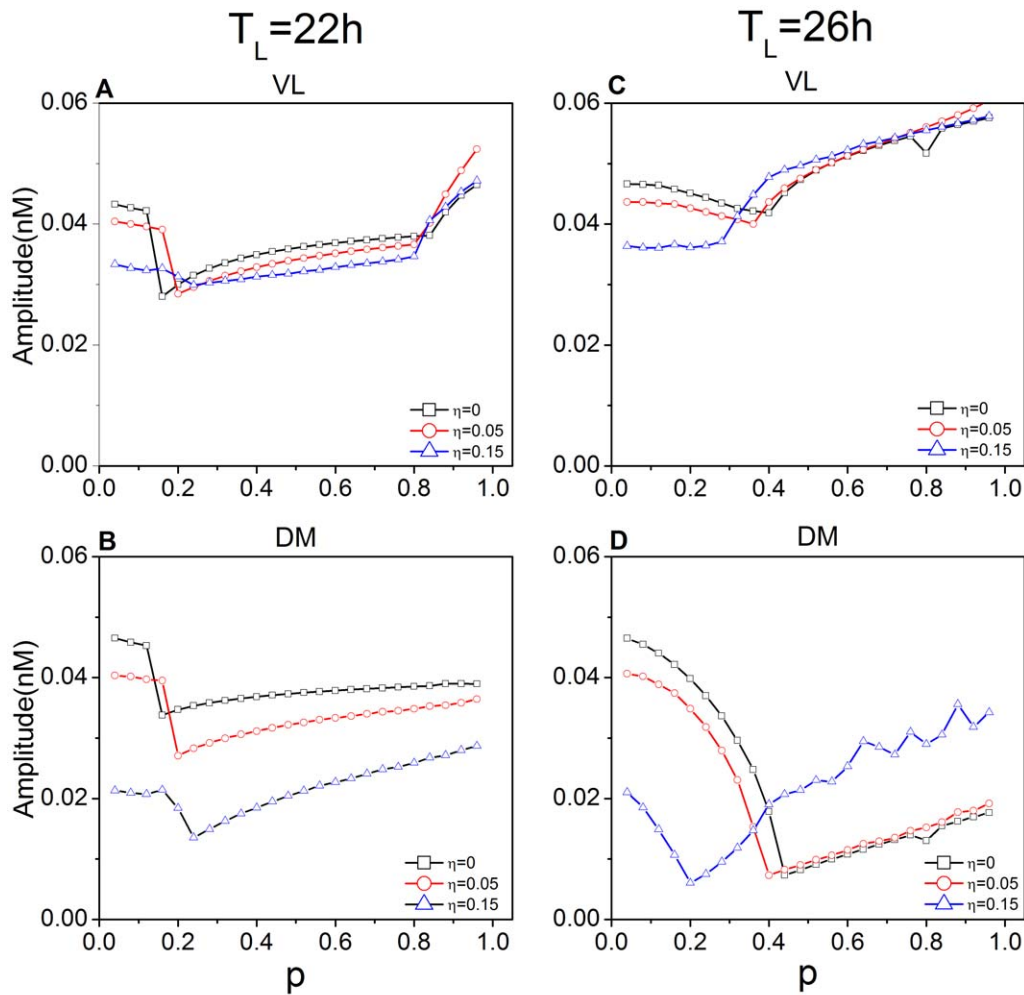


Figure 5. Effect of coupling dispersion on the amplitude of the mean fields of VL and DM. The case for the 22-h T cycle is shown for VL (A) and DM (B), and the case for the 26-h T cycle is shown for VL (C) and DM (D).
doi:10.1371/journal.pone.0036900.g005

increase of DM mean field amplitude as p increases further (Figure 5D) is thus attributable to increased individual oscillator amplitude.

These considerations imply that VL and DM desynchronize more readily under the 22-h than under the 26-h T cycle and that dispersion of coupling strengths improves network robustness preferentially under the 26-h cycle.

Effect of Coupling Dispersion on the Network's Phase Response Curve to a Light Pulse

The network's capacity to generate phase advances or delays can be quantified as a phase-response curve (PRC), measured by plotting the phase shifts that occur in the rhythm when discrete light pulses are applied at different phase points across the circadian cycle [24,32,33,34]. Figure 7 represents the family of PRC's obtained to a 1-h light pulse of increasing intensities, showing that the phase response region (i.e., the area under the delay and advance zones) increases in magnitude with increasing K_f . Notably, as the value of η increases, the area under the delay zone increases relatively more than that under the advance zone, as calculated in Table 1, where S represents the ratio of the area under the delay zone to the area under the advance zone.

Advances should correspond to the capacity of the network to follow a T cycle less than 24 h, while delays should correspond to its capacity to follow a T cycle greater than 24 h [35].

Discussion

Here we analyze the photic desynchronization of two subgroups of circadian oscillators in a network model of the suprachiasmatic nucleus. As also demonstrated in experiments with rats exposed to a short T cycle of low light intensity [7,36], a subgroup of oscillators receiving photic input (VL) can entrain to the external cycle while the other, coupled subgroup (DM) expresses an unentrained period greater than 24 h.

Granada et al. [37] have modeled this forced desynchronization of rat activity rhythms as a single oscillator with oscillatory interactions (modulation and superposition) between the external cycle and the internal clock, while Schwartz et al. [38] have modeled entrainment to the T cycle by two coupled oscillators forced by a Zeitgeber. Casiraghi et al. [39] have used a two oscillator model to analyze a chronic jet lag paradigm that leads to forced desynchrony, and they observed an asymmetry in its behavior similar to our findings reported

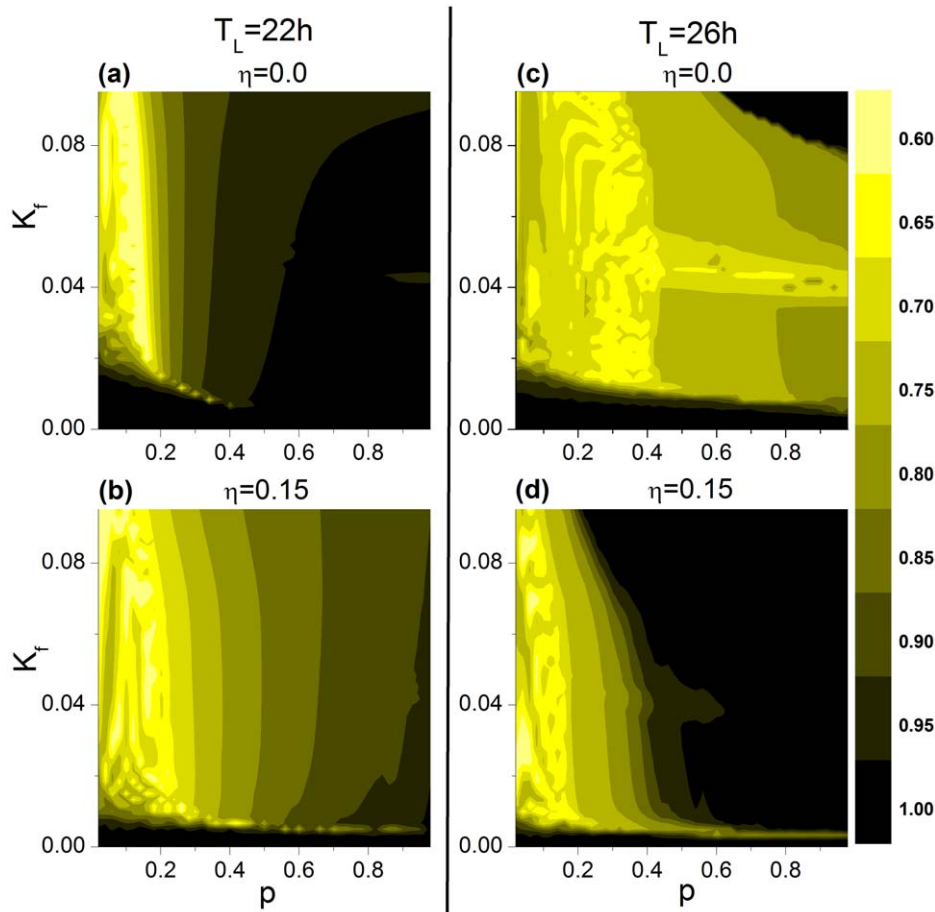


Figure 6. Effect of coupling dispersion on the order parameter of the network. The case for the 22-h T cycle is shown with coupling dispersion $\eta=0.0$ (A) and $\eta=0.15$ (B), and the case for the 26-h T cycle is shown with coupling dispersion $\eta=0.0$ (C) and $\eta=0.15$ (D). doi:10.1371/journal.pone.0036900.g006

here. We have taken the Goodwin model and extended it to include p , the proportion of all SCN cellular oscillators that receive photic input, and η , the dispersion of coupling strengths. We find, first, that network desynchronization (with an entrained VL but an unentrained DM) depends on light intensity and the value of p . Relatively higher light intensities

protect the network from desynchronization, as reported experimentally [36]. Experiments estimate that the value of p for the rodent SCN ranges from 20% to 33%, based on molecular, electrophysiological, and computational studies [40,41]. Comparing these results to our simulations in Figure 4, we find that $\eta=0.15$ is a good parameter value to

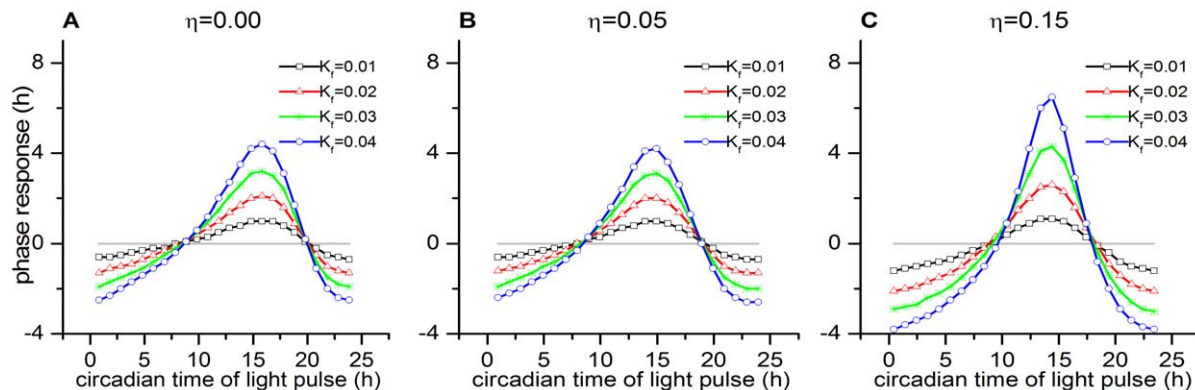


Figure 7. Effect of coupling dispersion on the phase response curve (PRC) of the network. Shown are PRCs with coupling dispersions $\eta=0.0$ (A), $\eta=0.05$ (B), and $\eta=0.15$ (C). Although the network shows relatively larger phase advances and delays with increased coupling dispersion, the area under the phase delay zones is greater than that under the advance zones. The PRCs were similar for all the values of $p \geq p_c$. doi:10.1371/journal.pone.0036900.g007

Table 1. Effect of coupling dispersion on the ratio of the area of the delay zone to the advance zone.

ηK_f	0.01	0.02	0.03	0.04
0.00	0.69	0.70	0.71	0.73
0.05	0.82	0.85	0.89	0.92
0.15	1.80	1.40	1.40	1.40

The ratio increases with increasing coupling dispersion.
doi:10.1371/journal.pone.0036900.t001

fit the experiments. At this value, there is a critical value of p for network entrainment to short (22 h) and long (26 h) T cycles of 0.28 and 0.20, respectively, and the critical value appears fairly insensitive to K_f . We predict that the rat SCN is likely to have this very large heterogeneity in coupling strengths, given that the critical p best matches the experimental estimate for the larger values of η .

Second, we find that the inclusion of dispersed coupling strengths affects network entrainment in a preferential manner, such that increased η significantly reduces the critical value of p for DM entrainment to the 26-h T cycle. A consequence of this η influence is that network robustness is superior under the 26-h cycle while desynchronization is favored under the 22-h cycle. In fact, such an asymmetry has been observed experimentally, with no obvious desynchronization of rat motor activity rhythms during exposure to long T cycles [28].

The basis for this asymmetry is unclear. Coupling dispersion appears to generally increase the effect of light on the system, and since the delay zone of the PRC is greater than the advance zone, a preferential effect on entrainment to the 26-h T cycle might be expected. But such an explanation does not account for the u-shaped, rather than monotonic, p_c function for the 22-h T cycle. Moreover, differences in entrainment to short and long T cycles may not be a general feature of the SCN; it may not be true for other species [42], and the possible effects of locomotor activity itself on SCN network behavior (e.g., in or out of a running wheel, diurnal or nocturnal activity pattern) needs further investigation.

Our findings may provide new elements to the theory of coupled oscillators, especially with regard to chimera states in which one group of the system is synchronized and the other is desynchronized [43,44,45,46,47]; in these studies, the oscillators in the two groups are identical and the chimera states are generally induced by the initial conditions. However, in our case, the discovery that the subgroup VL may be entrained to the T cycle while the group DM remains free-running is similar to the chimera state, but this phenomenon does not depend on the initial conditions. Thus, a novel oscillator theory is needed to explain this robustness to initial conditions and should be a topic for further studies.

Heterogeneous intercellular coupling within the SCN is likely central to the generation of complex behavioral patterns. Non-uniform SCN network architecture also has been implicated in the phase-“splitting” of locomotor activity cycles seen in hamsters maintained in constant environmental light [48]. In the future, we hope to consider how topology influences entrainment, in contrast to the mean field used here.

Methods

We represent each mammalian cell of the network as a Goodwin oscillator. The Goodwin model is a widely used mathematical model to represent the behavior of the gene

regulatory network in single cellular circadian oscillators [13]. The model represents the transcription-translation behavior of the single cell by using three variables that include a clock gene mRNA (x), a clock protein (y), and a transcriptional inhibitor (z).

As our network model, we consider the mean field-modified Goodwin model proposed by [13] with a global coupling strength. The modified Goodwin model with N oscillators is represented as follows:

$$\begin{aligned}
 \frac{dx_i}{dt} &= \sigma \left(\alpha_1 \frac{k_1^n}{k_1^n + z_i^n} - \alpha_2 \frac{x_i}{k_2 + x_i} + \alpha_c \frac{g_i F}{k_c + g_i F} \right) + L_i \\
 \frac{dy_i}{dt} &= \sigma \left(k_3 x_i - \alpha_4 \frac{y_i}{k_4 + y_i} \right) \\
 \frac{dz_i}{dt} &= \sigma \left(k_5 y_i - \alpha_6 \frac{z_i}{k_6 + z_i} \right) \\
 \frac{dV_i}{dt} &= \sigma \left(k_7 x_i - \alpha_8 \frac{V_i}{k_8 + V_i} \right) \\
 F &= \frac{1}{N} \sum_{i=1}^N V_i \\
 i &= 1, 2, 3, \dots, N
 \end{aligned}
 \tag{1}$$

Where the state variables x_i , y_i , z_i represent the concentrations, respectively, of a clock gene mRNA, a clock protein and a transcriptional inhibitor in each clock cell i . Neurotransmitter V_i is induced by the mRNA x_i . The intercellular coupling is implemented through the neurotransmitter F which acts as a mean field of V_i , the coupling strength g_i represents the sensitivity of the individual oscillator to the neurotransmitter and is required to be a positive value here, and L_i is the light term. We considered the parameters as in [13]:

$$\begin{aligned}
 (\alpha_1 = 0.7nM/h, k_1 = 1.0nM, n = 4.0, \alpha_2 = 0.35nM/h, \\
 k_2 = 1.0nM, k_3 = 0.7/h, \\
 \alpha_4 = 0.35nM/h, k_4 = 1.0nM, k_5 = 0.7/h, \alpha_6 = 0.35nM/h, \\
 k_6 = 1.0nM, k_7 = 0.35/h, \\
 \alpha_8 = 1.0nM/h, k_8 = 1.0nM, \alpha_c = 0.4nM/h, k_c = 1nM).
 \end{aligned}$$

The coupling strength g_i is different for different oscillators and assumes a value from a normal distribution with a mean 0.5 and a standard deviation η . When $\eta \neq 0$, the network is heterogeneous with distribution of coupling.

In order to understand the dissociation behavior observed under a T cycle outside the range of entrainment, we modified the Goodwin model to include the fact that light acts directly on only a portion of the neurons in the network. Furthermore, the light term L_i that is applied to a fraction pN neurons with p being less than one and positive, is considered to be located in the VL subdivision. Mathematically, the effect of light is represented as:

$$L_i = \begin{cases} K_f, & \text{if } i \leq pN \ \& \ \text{mod}(t, T_L) \leq \frac{T_L}{2} \\ 0, & \text{else} \end{cases}$$

Where T_L is the period of the light-dark cycle and K_f is the light intensity.

As pointed out in our previous paper [22], the dispersion of coupling strengths influences the free-running period of the SCN.

In order to compare the influence of different coupling dispersions on entrainment of the SCN network to T cycles, it is necessary to make the free-running period the same for different dispersions. To set the free-running period to 24 h, we multiply a rescaling factor σ to the left hand of equation (1) except for the light term and coupling term, i.e., multiply the same σ to the parameters $\alpha_1, \alpha_2, k_3, \alpha_4, k_5, \alpha_6, k_7, \alpha_8, \alpha_c$ for the deviation η . For example, σ is equal to 1.26 for $\eta=0.0$, 1.22 for $\eta=0.05$, 1.16 for $\eta=0.1$, and 1.13 for $\eta=0.15$.

For simplicity, we refer to the network that is comprised of pN neurons as VL and the network comprised of the remaining $(1-p)N$ neurons as DM. To understand the behavior of the VL and DM subdivisions, we define the mean field of VL and DM respectively as

$$F_{VL} = \frac{1}{pN} \sum_{i=1}^{pN} V_i,$$

$$F_{DM} = \frac{1}{N-pN} \sum_{i=1+pN}^N V_i$$

In addition, to understand the synchronization properties between VL and DM, we have estimated the phase of the individual neurons by using the Hilbert transform [49,50]. From the estimated phase of VL and DM, we introduce an order parameter as:

$$R = \frac{1}{2} \langle |e^{i\theta_{VL}} + e^{i\theta_{DM}}| \rangle$$

where θ is the estimated phase from the mean field output time series of VL or DM and $\langle \rangle$ denotes average over time. The

References

1. Welsh DK, Takahashi JS, Kay SA (2010) Suprachiasmatic nucleus: cell autonomy and network properties. *Annu Rev Physiol* 72: 551–577.
2. Silver R, Schwartz WJ (2005) The suprachiasmatic nucleus is a functionally heterogeneous timekeeping organ. *Methods Enzymol* 393: 451–465.
3. Morin LP (2007) SCN organization reconsidered. *J Biol Rhythms* 22: 3–13.
4. Noguchi T, Watanabe K (2008) Regional differences in circadian period within the suprachiasmatic nucleus. *Brain Res* 1239: 119–126.
5. Noguchi T, Watanabe K, Ogura A, Yamaoka S (2004) The clock in the dorsal suprachiasmatic nucleus runs faster than that in the ventral. *Eur J Neurosci* 20: 3199–3202.
6. Albus H, Vansteensel MJ, Michel S, Block GD, Meijer JH (2005) A GABAergic mechanism is necessary for coupling dissociable ventral and dorsal regional oscillators within the circadian clock. *Curr Biol* 15: 886–893.
7. de la Iglesia HO, Cambras T, Schwartz WJ, Diez-Noguera A (2004) Forced desynchronization of dual circadian oscillators within the rat suprachiasmatic nucleus. *Curr Biol* 14: 796–800.
8. Nagano M, Adachi A, Nakahama K, Nakamura T, Tamada M, et al. (2003) An abrupt shift in the day/night cycle causes desynchrony in the mammalian circadian center. *J Neurosci* 23: 6141–6151.
9. Nakamura W, Yamazaki S, Takasu NN, Mishima K, Block GD (2005) Differential response of Period 1 expression within the suprachiasmatic nucleus. *J Neurosci* 25: 5481–5487.
10. Yamaguchi S, Isejima H, Matsuo T, Okura R, Yagita K, et al. (2003) Synchronization of cellular clocks in the suprachiasmatic nucleus. *Science* 302: 1408–1412.
11. Aton SJ, Herzog ED (2005) Come together, right...now: synchronization of rhythms in a mammalian circadian clock. *Neuron* 48: 531–534.
12. Liu AC, Welsh DK, Ko CH, Tran HG, Zhang EE, et al. (2007) Intercellular coupling confers robustness against mutations in the SCN circadian clock network. *Cell* 129: 605–616.
13. Gonze D, Bernard S, Waltermann C, Kramer A, Herzel H (2005) Spontaneous synchronization of coupled circadian oscillators. *Biophys J* 89: 120–129.
14. Bernard S, Gonze D, Cajavec B, Herzel H, Kramer A (2007) Synchronization-induced rhythmicity of circadian oscillators in the suprachiasmatic nucleus. *PLoS Comput Biol* 3: e68.
15. Abraham U, Granada AE, Westermark PO, Heine M, Kramer A, et al. (2010) Coupling governs entrainment range of circadian clocks. *Mol Syst Biol* 6: 438.
16. Aton SJ, Colwell CS, Harmar AJ, Waschek J, Herzog ED (2005) Vasoactive intestinal polypeptide mediates circadian rhythmicity and synchrony in mammalian clock neurons. *Nat Neurosci* 8: 476–483.
17. Maywood ES, Reddy AB, Wong GK, O'Neill JS, O'Brien JA, et al. (2006) Synchronization and maintenance of timekeeping in suprachiasmatic circadian clock cells by neuropeptidergic signaling. *Curr Biol* 16: 599–605.
18. Li JD, Burton KJ, Zhang C, Hu SB, Zhou QY (2009) Vasopressin receptor V1a regulates circadian rhythms of locomotor activity and expression of clock-controlled genes in the suprachiasmatic nuclei. *Am J Physiol Regul Integr Comp Physiol* 296: R824–830.
19. Daido H (1992) Quasientrainment and slow relaxation in a population of oscillators with random and frustrated interactions. *Phys Rev Lett* 68: 1073–1076.
20. Daido H (2000) Algebraic relaxation of an order parameter in randomly coupled limit-cycle oscillators. *Phys Rev E Stat Phys Plasmas Fluids Relat Interdiscip Topics* 61: 2145–2147.
21. Hong H, Strogatz SH (2011) Kuramoto model of coupled oscillators with positive and negative coupling parameters: an example of conformist and contrarian oscillators. *Phys Rev Lett* 106: 054102.
22. Gu C, Wang J, Liu Z (2009) Free-running period of neurons in the suprachiasmatic nucleus: Its dependence on the distribution of neuronal coupling strengths. *Phys Rev E Stat Nonlin Soft Matter Phys* 80: 030904.
23. Ruoff P, Vinsjevik M, Monnerjahn C, Rensing L (1999) The Goodwin oscillator: on the importance of degradation reactions in the circadian clock. *J Biol Rhythms* 14: 469–479.
24. Locke JC, Westermark PO, Kramer A, Herzel H (2008) Global parameter search reveals design principles of the mammalian circadian clock. *BMC Syst Biol* 2: 22.
25. Ullner E, Buceta J, Diez-Noguera A, Garcia-Ojalvo J (2009) Noise-induced coherence in multicellular circadian clocks. *Biophys J* 96: 3573–3581.
26. Gu C, Wang J, Liu Z (2011) Mechanism of phase splitting in two coupled groups of suprachiasmatic-nucleus neurons. *Phys Rev E Stat Nonlin Soft Matter Phys* 83: 046224.

average of $\frac{d\theta}{dt}$ is defined as the angular frequency and the period is obtained by $T = \frac{2\pi}{\theta}$. To determine entrainment of VL or DM to

the T_L , we estimated the period of VL or DM and estimated its absolute difference from T_L . If the absolute difference in period is less than 0.25 h, the corresponding subgroup (VL or DM) is considered to be entrained. To numerically calculate the equations, we use the fourth order Runge-Kutta method with time step of 0.1 h. Initial 20000 time steps are neglected to avoid the effect of transients. The number of oscillators is $N=100$, and the simulations are performed five times, with initial conditions selected randomly from a uniform distribution in the range (0–1) for x , y , and z . We have also calculated the case of $N=400$ and time step of 0.01 h. Two additional simulations are performed with selective assignment of coupling in which larger values are assigned to either VL or DM without changing the intrinsic distribution of g_i .

To obtain the phase-response curve, we applied 1-h light pulses at different phases to the model, with intensity K_f and with different values for $p \geq p_c$. The corresponding advance or delay is detected from the output of the network. Advance corresponds to the capacity of the SCN network to follow a light-dark cycle with period less than the free running period, and delay is the capacity of the network to follow a light-dark cycle with period greater than the free running period [35].

Author Contributions

Conceived and designed the experiments: CG WJS PI. Performed the experiments: CG ZL. Analyzed the data: CG ZL WJS PI. Contributed reagents/materials/analysis tools: CG ZL WJS PI. Wrote the paper: CG ZL WJS PI.

27. Campuzano A, Vilaplana J, Cambras T, Diez-Noguera A (1998) Dissociation of the rat motor activity rhythm under T cycles shorter than 24 hours. *Physiol Behav* 63: 171–176.
28. Vilaplana J, Cambras T, Campuzano A, Diez-Noguera A (1997) Simultaneous manifestation of free-running and entrained rhythms in the rat motor activity explained by a multioscillatory system. *Chronobiol Int* 14: 9–18.
29. Usui S, Takahashi Y, Okazaki T (2000) Range of entrainment of rat circadian rhythms to sinusoidal light-intensity cycles. *Am J Physiol Regul Integr Comp Physiol* 278: R1148–1156.
30. Liang X, Tang M, Dhamala M, Liu Z (2009) Phase synchronization of inhibitory bursting neurons induced by distributed time delays in chemical coupling. *Phys Rev E Stat Nonlin Soft Matter Phys* 80: 066202.
31. Strogatz S (2000) From Kuramoto to Crawford: exploring the onset of synchronization in populations of coupled oscillators. *Physica D* 143: 20.
32. Daan S, Pittendrigh C (1976) A functional analysis of circadian pacemakers in nocturnal rodents. II. The variability of phase response curves. *J Comp Physiol A* 106: 253–256.
33. Ruoff P, Vinsjevik M, Monnerjahn C, Rensing L (2001) The Goodwin model: simulating the effect of light pulses on the circadian sporulation rhythm of *Neurospora crassa*. *J Theor Biol* 209: 29–42.
34. Granada A, Hennig RM, Ronacher B, Kramer A, Herzel H (2009) Phase response curves elucidating the dynamics of coupled oscillators. *Methods Enzymol* 454: 1–27.
35. Pendergast JS, Friday RC, Yamazaki S (2010) Photic entrainment of period mutant mice is predicted from their phase response curves. *J Neurosci* 30: 12179–12184.
36. Cambras T, Vilaplana J, Campuzano A, Canal-Corretger MM, Carulla M, et al. (2000) Entrainment of the rat motor activity rhythm: effects of the light-dark cycle and physical exercise. *Physiol Behav* 70: 227–232.
37. Granada AE, Cambras T, Diez-Noguera A, Herzel H (2011) Circadian desynchronization. *Interface Focus* 1: 153–166.
38. Schwartz MD, Wotus C, Liu T, Friesen WO, Borjigin J, et al. (2009) Dissociation of circadian and light inhibition of melatonin release through forced desynchronization in the rat. *Proc Natl Acad Sci U S A* 106: 17540–17545.
39. Casiraghi LP, Oda GA, Chiesa JJ, Friesen WO, Golombek DA (2012) Forced desynchronization of activity rhythms in a model of chronic jet lag in mice. *J Biol Rhythms* 27: 59–69.
40. Rohling JH, vanderLeest HT, Michel S, Vansteensel MJ, Meijer JH (2011) Phase resetting of the mammalian circadian clock relies on a rapid shift of a small population of pacemaker neurons. *PLoS One* 6: e25437.
41. Meijer JH, Schwartz WJ (2003) In search of the pathways for light-induced pacemaker resetting in the suprachiasmatic nucleus. *J Biol Rhythms* 18: 235–249.
42. Vivanco P, Ojalora BB, Rol MA, Madrid JA (2010) Dissociation of the circadian system of *Octodon degus* by T28 and T21 light-dark cycles. *Chronobiol Int* 27: 1580–1595.
43. Abrams DM, Mirollo R, Strogatz SH, Wiley DA (2008) Solvable model for chimera states of coupled oscillators. *Phys Rev Lett* 101: 084103.
44. Pikovsky A, Rosenblum M (2008) Partially integrable dynamics of hierarchical populations of coupled oscillators. *Phys Rev Lett* 101: 264103.
45. Ott E, Antonsen TM (2008) Low dimensional behavior of large systems of globally coupled oscillators. *Chaos* 18: 037113.
46. Lee WS, Ott E, Antonsen TM (2009) Large coupled oscillator systems with heterogeneous interaction delays. *Phys Rev Lett* 103: 044101.
47. Martens EA, Laing CR, Strogatz SH (2010) Solvable model of spiral wave chimeras. *Phys Rev Lett* 104: 044101.
48. Indic P, Schwartz WJ, Paydarfar D (2008) Design principles for phase-splitting behaviour of coupled cellular oscillators: clues from hamsters with ‘split’ circadian rhythms. *J R Soc Interface* 5: 873–883.
49. Rosenblum MG, Pikovsky AS, Kurths J (1996) Phase synchronization of chaotic oscillators. *Phys Rev Lett* 76: 1804–1807.
50. Aylett M, Marples G, Jones K (1999) Home blood pressure monitoring: its effect on the management of hypertension in general practice. *Br J Gen Pract* 49: 725–728.