

Research Article

Toxicity of Metals to a Freshwater Snail, *Melanoides tuberculata*

M. Shuhaimi-Othman, R. Nur-Amalina, and Y. Nadzifah

School of Environmental and Natural Resource Sciences, Faculty of Science and Technology, National University of Malaysia (UKM), Selangor, 43600 Bangi, Malaysia

Correspondence should be addressed to M. Shuhaimi-Othman, shuhaimi@ukm.my

Received 18 October 2011; Accepted 5 January 2012

Academic Editors: T. Brock, K. Kannan, J. Ruelas-Inzunza, and B. C. Suedel

Copyright © 2012 M. Shuhaimi-Othman et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Adult freshwater snails *Melanoides tuberculata* (Gastropod, Thiaridae) were exposed for a four-day period in laboratory conditions to a range of copper (Cu), cadmium (Cd), zinc (Zn), lead (Pb), nickel (Ni), iron (Fe), aluminium (Al), and manganese (Mn) concentrations. Mortality was assessed and median lethal times (LT₅₀) and concentrations (LC₅₀) were calculated. LT₅₀ and LC₅₀ increased with the decrease in mean exposure concentrations and times, respectively, for all metals. The LC₅₀ values for the 96-hour exposures to Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn were 0.14, 1.49, 3.90, 6.82, 8.46, 8.49, 68.23, and 45.59 mg L⁻¹, respectively. Cu was the most toxic metal to *M. tuberculata*, followed by Cd, Zn, Pb, Ni, Fe, Mn, and Al (Cu > Cd > Zn > Pb > Ni > Fe > Mn > Al). Metals bioconcentration in *M. tuberculata* increases with exposure to increasing concentrations and Cu has the highest accumulation (concentration factor) in the soft tissues. A comparison of LC₅₀ values for metals for this species with those for other freshwater gastropods reveals that *M. tuberculata* is equally sensitive to metals.

1. Introduction

Metals are released from both natural sources and human activity. The impact of metals on the environment is an increasing problem worldwide. The impact of metals on aquatic ecosystems is still considered to be a major threat to organisms health due to their potential bioaccumulation and toxicity to many aquatic organisms. Although metals are usually considered as pollutants, it is important to recognize that they are natural substances. Zinc, for example, is an essential component of at least 150 enzymes; copper is essential for the normal function of cytochrome oxidase; iron is part of the haemoglobin in red blood cells; boron is required exclusively by plants [1]. Malaysia, as a developing country, is no exception and faces metals pollution caused especially by anthropogenic activities such as manufacturing, agriculture, sewage, and motor vehicle emissions [2–5]. Metals are nonbiodegradable. Unlike some organic pesticides, metals cannot be broken down into less harmful components. Managing metal contamination requires an understanding of the concentration dependence of toxicity. Dose-response relationships provide the basis for the assessment of hazards and risks presented by environmental chemicals. Toxicity

testing is an essential tool for assessing the effect and fate of toxicants in aquatic ecosystems and has been widely used as a tool to identify suitable organisms as a bioindicator and to derive water quality standards for chemicals. There are many different ways in which toxicity can be measured, and most commonly the measure (end point) is death [1, 6, 7]. Metals research in Malaysia, especially using organisms as a bioindicator, is still scarce. Therefore, it is important to conduct studies with local organisms that can be used to gain data on metal toxicity, to determine the organism's sensitivity and to derive a permissible limit for Malaysian's water that can protect the local aquatic communities.

The freshwater molluscs of the Malaysian region are common, and most extant species are relatively easy to collect. The snails are rich fauna, while bivalve are the second. More than 150 aquatic nonmarine mollusc species have been recorded from the Malaysian region. *Melanoides tuberculata* (Müller 1774) is from class Gastropoda with shells higher than wide (elongate), conical, usually light brown in colour, and it is a cosmopolitan species [8]. *M. tuberculata* is a species of freshwater snail with an operculum, a parthenogenetic, aquatic gastropod mollusc in the family Thiaridae. The average shell length is about

20–27 mm and this species is native to subtropical and tropical northern Africa and southern Asia (Indo-Pacific region, Southern Asia, Arabia, and northern Australia), but they have established populations throughout the globe. The snail has an operculum that can protect it from desiccation and can remain viable for days on dry land [9]. It is a warm-climate species, prefers a temperature range of 18 to 32°C, and is primarily a burrowing species that tends to be most active at night. This snail feeds primarily on algae (microalgae) and acts as an intermediate host for many digenetic trematodes. *M. tuberculata* is a viviparous, gonochoric species with polyploid strains that reproduces by apomictic parthenogenesis. Because meiosis usually does not occur, offspring are identical to their mother. Females can be recognized by their greenish coloured gonads while males have reddish gonads. Under good conditions, females will produce fertilized eggs that are transferred to a brood pouch where they remain until they hatch. *M. tuberculata* will begin reproducing at a size as small as 5 to 10 mm in length and broods may contain over seventy offspring embryos which develop in the mother [10–12].

Molluscs have long been regarded as promising bioindicator and biomonitoring subjects. They are abundant in many terrestrial and aquatic ecosystems, being easily available for collection. They are highly tolerant to many pollutants and exhibit high accumulations of them, particularly heavy metals [13, 14]. Little information exists in the literatures concerning the toxic effects of metals for this snail. So far, only a few studies have been reported on metal toxicity to *M. tuberculata* [15, 16] and most of the studies were on the accumulation of metals [14, 17, 18]. Therefore, the purpose of this study was to determine the acute toxicity of eight metals (Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn) to the freshwater mollusc *M. tuberculata* and to examine the bioconcentration of these metals in the body after four days of exposure.

2. Materials and Methods

Snails *M. tuberculata* were collected from canals in the university in Bangi, Selangor, Malaysia. Identification of the species was based on Panha and Burch [8]. Prior to toxicity testing, the snails were acclimatized for one week under laboratory conditions (28–30°C with 12 h light : 12 h darkness) in 50-L stocking tanks using dechlorinated tap water (filtered by several layers of sand and activated carbon; T.C. Sediment Filter (TK Multitrade, Seri Kembangan, Malaysia)) aerated through an air stone. During acclimation the snails were fed on lettuce. The standard stock solution (100 mg L⁻¹) of Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn was prepared from analytical grade metallic salts of CuSO₄·5H₂O, CdCl₂·2.5H₂O, ZnSO₄·7H₂O, Pb(NO₃)₂, NiSO₄·6H₂O, FeCl₃, Al₂(SO₄)₃·18H₂O, and MnSO₄·H₂O, respectively (Merck, Darmstadt, Germany). The stock solutions were prepared with deionized water in 1 L volumetric flasks. Acute Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn toxicity experiments were performed for a four-day period using adult snails (shell length approximately 1.5–2.0 cm, mean wet weight 22.5 ± 1.6 mg) obtained from stocking tanks.

Following a range finding test, five Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn nominal concentrations were chosen (Table 1). Metal solutions were prepared by dilution of a stock solution with dechlorinated tap water. A control with dechlorinated tap water only was also used. The tests were carried out under static conditions with renewal of the solution every two days. Control and metal-treated groups each consisted of two replicates of five randomly allocated snails in a 500 mL glass beaker containing 400 mL of the appropriate solution. No stress was observed for the snails in the solution, indicated by 100% survival for the snails in the control water until the end of the study. A total of 10 animals per treatment/concentration were used in the experiment and a total of 410 animals were employed in the investigation [42, 43]. Samples of water for metal analysis taken before and immediately after each solution renewal were acidified to 1% with ARISTAR nitric acid (65%) (BDH Inc, VWR International Ltd., England) before metal analysis by flame or furnace Atomic Absorption Spectrophotometer (AAS—Perkin Elmer model AAnalyst800, Massachusetts, USA) depending on the concentrations.

During the toxicity test, the snails were not fed. The experiments were performed at room temperature of 28–30°C with photoperiod 12 h light : 12 h darkness, using fluorescent lights (334–376 lux). Water quality parameters (pH, conductivity, and dissolved oxygen) were measured every two days using portable meters (model Hydrolab Quanta, Hach, Loveland, USA) and water hardness samples were fixed with ARISTAR nitric acid and measured by flame atomic absorption spectrophotometer (AAS—Perkin Elmer model AAnalyst 800). Mortality was recorded every 3 to 4 hours for the first two days and then at 12 to 24 hour intervals throughout the rest of the test period. The criterion used to determine mortality were failure to respond to gentle physical stimulation. The death was further confirmed by putting the snail on the glass petri dish for few minutes and if it did not show any movement, it was considered dead. Any dead animals were removed immediately.

At the end of day four, the live snails were used to determine bioconcentration of the metals in the whole body (soft tissues) according to the concentrations used. The snails were cleaned with dechlorinated tap water, and soaked in boiling water for approximately 3 min. Tissues of the molluscs were removed from the shell, rinsed with deionized water, and each sample contained three replicates of three to five animals in a glass test tube (depending on how many live animals were left) and was oven-dried (80°C) for at least 48 hours before being weighed [14]. Each replicate was digested (whole organism) in 1.0 mL ARISTAR nitric acid (65%) in a block thermostat (80°C) for 2 hours. Upon cooling, 0.8 mL of hydrogen peroxide (30%) was added to the solutions. The test tubes were put back on the block thermostat for another 1 hour until the solutions became clear. The solutions were then made up to 25 mL with the addition of deionized water in 25 mL volumetric flasks. Efficiency of the digestion method was evaluated using mussel and lobster tissue reference material (SRM 2976 and TORT-2, National Institute of Standard and Technology, Gaithersburg, USA and National Research Council Canada, Ottawa, Ontario,

TABLE 1: Median lethal times (LT₅₀) for *M. tuberculata* exposed to different concentrations for Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn.

Nominal (and measured) concentration (mg L ⁻¹)	LT ₅₀ (h)	95% Confidence limits
Cu		
0.075 (0.081)	163.42	63.60–419.91
0.1 (0.145)	134.97	53.22–342.28
0.32 (0.292)	98.89	44.35–220.50
0.56 (0.549)	75.87	26.88–214.15
0.87 (0.915)	55.42	25.36–121.11
Cd		
0.56 (0.611)	283.44	85.46–940.12
1.0 (1.21)	114.89	52.78–250.09
5.6 (4.87)	57.21	30.29–108.05
10 (10.82)	22.34	11.03–45.27
32 (33.49)	7.82	4.63–13.23
Zn		
1.0 (1.09)	216.96	630.53–1541.63
5.6 (5.30)	96.71	52.82–177.09
10 (8.19)	61.83	39.85–95.94
32 (32.45)	32.44	22.13–47.55
56 (49.60)	12.34	8.25–18.45
Pb		
1.0 (1.02)	250.72	430.55–2057.69
5.6 (5.42)	179.32	38.42–837.02
10 (10.95)	88.25	30.24–257.71
18 (17.16)	40.36	13.91–117.15
32 (31.18)	11.17	6.71–18.57
Ni		
5.6 (5.51)	105.96	59.28–189.40
10 (9.02)	92.38	48.25–176.87
32 (31.53)	58.11	39.82–84.80
75 (67.11)	36.84	21.66–62.67
100 (97.84)	16.26	12.50–21.17
Fe		
5.6 (5.27)	134.97	53.22–342.28
8.7 (8.86)	102.06	40.17–259.29
10 (11.76)	79.72	28.18–225.53
32 (33.47)	34.71	13.36–90.15
56 (58.17)	20.04	7.5–53.51
Al		
56 (88.38)	80.87	111.48–58.66
100 (160.83)	57.91	87.09–38.50
320 (362.83)	42.75	65.99–27.70
560 (884.34)	18.57	40.94–8.42
1000 (1229.91)	8.40	18.52–3.81
Mn		
10 (12.98)	119.53	62.44–228.82
32 (31.60)	67.81	35.06–131.15
56 (57.81)	35.07	19.04–64.60
87 (85.61)	16.97	9.00–32.00
100 (97.01)	8.35	5.05–13.79

Canada, resp.). Efficiencies obtained were within 10% of the reference values. To avoid possible contamination, all glassware and equipment used were acid-washed (20% HNO₃) (Dongbu Hitek Co. Ltd., Seoul, Korea, 68%), and the accuracy of the analysis was checked against blanks. Procedural blanks and quality control samples made from

standard solutions for Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn (Spectrosol, BDH, England) were analyzed in every ten samples in order to check for sample accuracy. Percentage recoveries for metals analyses were between 85–105%.

Median lethal times (LT₅₀) and concentrations (LC₅₀) for the snails exposed to metals were calculated using measured

metal concentrations. FORTRAN programs based on the methods of Litchfield [44] and Litchfield and Wilcoxon [45] were used to compute the LT_{50} and LC_{50} . Data were analyzed using time/response (TR) and concentration/response (CR) methods by plotting cumulative percentage mortality against concentration and time, respectively, on logarithmic-probit paper. Concentration factors (CFs) were calculated for whole animals as the ratio of the metals concentrations in the tissues to the metals concentration measured in the water.

3. Results and Discussion

In all data analyses, the actual (measured concentration) rather than nominal Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn concentrations were used (Table 1). The mean water quality parameters measured during the test were pH 6.68 ± 0.22 , conductivity $180.0 \pm 46.0 \mu\text{S cm}^{-1}$, dissolved oxygen $6.1 \pm 0.27 \text{ mg L}^{-1}$, and total hardness (Mg^{2+} and Ca^{2+}) $18.72 \pm 1.72 \text{ mg L}^{-1}$ as CaCO_3 .

One hundred percent of control animals maintained in dechlorinated tap water survived throughout the experiment. The median lethal times (LT_{50}) and concentrations (LC_{50}) increased with a decrease in mean exposure concentrations and times, respectively, for all metals (Tables 1 and 2). However, the lethal threshold concentration could not be determined since the toxicity curves (Figures 1 and 2) did not become asymptotic to the time axis within the test period. Figures 1 and 2 show that Cu was the most toxic metal to *M. tuberculata*, followed by Cd, Zn, Pb, Ni, Fe, Mn, and Al. Other studies show different trends of toxicity with different snails. According to Luoma and Rainbow [7] the rank order of toxicity of metals will vary between organisms. With *Lymnaea luteola*, Khangarot and Ray [28, 30] showed that the order of toxicity was $\text{Cd} > \text{Ni} > \text{Zn}$; with *Viviparus bengalensis*, Gupta et al. [27] and Gadkari and Marathe [34] found that the order of toxicity was $\text{Zn} > \text{Cd} > \text{Pb} > \text{Ni}$; and with *Juga plicifera*, Nebeker et al. [20] found that Cu was more toxic than Ni.

The present study showed that LC_{50} s for 48 and 96 hours of Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn were 0.39, 11.85, 13.15, 10.99, 36.46, 21.78, 306.89, and 120.43 mg L^{-1} , and 0.14, 1.49, 3.90, 6.82, 8.46, 8.49, 68.23 and 45.59 mg L^{-1} , respectively (Table 1). A few studies had reported on the acute toxicity of metals to *M. tuberculata*. Bali et al. [15] and Mostafa et al. [16] showed that 96 h- LC_{50} of Cu to *M. tuberculata* were 0.2 and 3.6 mg L^{-1} , respectively, which were higher than the present study. In comparison with other freshwater gastropods (Table 3), this study showed that in general LC_{50} s for *M. tuberculata* were lower or similar compared to other freshwater snails. Direct comparisons of toxicity values obtained in this study with those in the literature were difficult because of differences in the characteristics (primarily water hardness, pH, and temperature) of the test waters. With similar water hardness (soft water) and using adult snails, Nebeker et al. [20] reported that 96 h- LC_{50} of Cu for *Fluminicola virens* was 0.08 mg L^{-1} , and of Zn for *Physa Gyrina* was 1.27 mg L^{-1} , which was lower than the present study. The toxicity reported by other studies (Table 3)

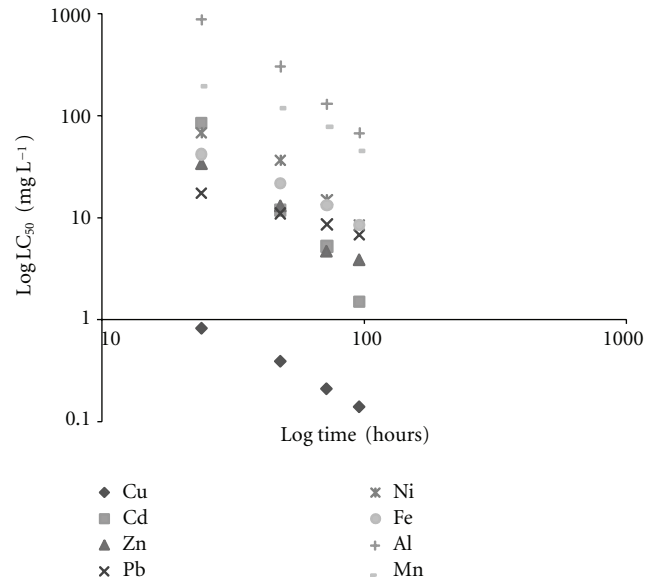


FIGURE 1: The relationship between median lethal concentration (LC_{50}) and exposure times for *M. tuberculata*.

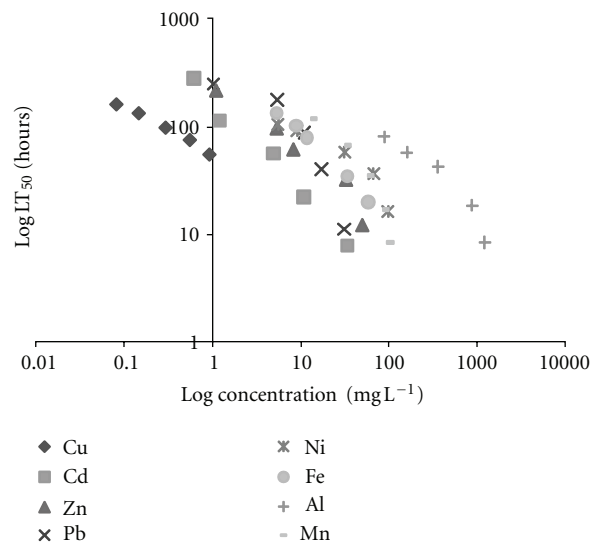


FIGURE 2: The relationship between median lethal time (LT_{50}) and exposure concentrations for *M. tuberculata*.

differs from that reported in this study owing to the different species, ages, and sizes of the organisms as well as varied test methods (water quality and water hardness) as this can affect toxicity [46–49]. In the present study, the water hardness used was considered low ($18.7 \text{ mg L}^{-1} \text{ CaCO}_3$), and the water was categorized as soft water ($<75 \text{ mg L}^{-1}$ as CaCO_3).

In comparison with other taxa, *M. tuberculata* shows less sensitivity to metals. LC_{50} s reported for other taxa from this laboratory such as Crustacea (prawn *Macrobrachium lanchesteri* [50] and ostracod *Stenocypris major* [51]), fish (*Rasbora sumatrana* and *Poecilia reticulata* [52]), and Annelida (*Nais elinguis* [53]) were lower than the LC_{50} values of *M. tuberculata* in the present study. Von Der Ohe and Liess [54] showed that 13 taxa belonging to Crustacea were among

TABLE 2: Median lethal concentrations (LC₅₀) for *M. tuberculata* at different exposure times for Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn.

Time (hour)	LC ₅₀ (mg L ⁻¹)	95% Confidence limits
Cu		
24	0.82	0.49–4.21
48	0.39	0.23–0.88
72	0.21	0.12–0.33
96	0.14	0.09–0.20
Cd		
24	85.03	13.94–518.57
48	11.85	2.70–51.99
72	5.24	0.96–28.43
96	1.49	0.34–6.53
Zn		
24	33.97	21.59–65.31
48	13.15	6.93–26.06
72	4.73	2.28–8.10
96	3.90	1.81–6.67
Pb		
24	17.39	12.06–29.47
48	10.99	6.04–19.68
72	8.57	4.37–14.79
96	6.82	2.89–12.67
Ni		
24	68.35	48.18–102.23
48	36.46	20.76–70.91
72	15.04	5.23–28.97
96	8.46	3.53–14.02
Fe		
24	42.12	25.74–133.99
48	21.78	10.52–88.85
72	13.29	4.09–29.47
96	8.49	1.58–15.25
Al		
24	880.78	553.91–2147.55
48	306.89	184.29–487.20
72	130.22	35.51–226.38
96	68.23	2.24–123.87
Mn		
24	194.52	112.85–335.27
48	120.43	58.08–249.72
72	78.35	36.20–169.56
96	45.59	20.17–103.04

the most sensitive to metal compounds and concluded that taxa belonging to Crustacea are similar to one another and to *Daphnia magna* in terms of sensitivity to organics and metals and that Molluscs have an average sensitivity to metals. Mitchell et al. [9] reported that the snail has a tightly sealing operculum that allows it to withstand desiccation and apparently also increases its tolerance to chemicals.

Bioconcentration of Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn in surviving *M. tuberculata* is as shown in Figure 3. Bioconcentration data for live snails were obtained from five Cd (0.61, 1.21, 4.87, 10.82 and 33.49 mg L⁻¹), Fe (5.27, 8.86, 11.76, 33.47, and 58.17 mg L⁻¹), and Mn (12.98, 31.60, 57.81, 85.61 and 97.01 mg L⁻¹) concentration exposures;

four Pb (1.02, 5.42, 10.95 and 17.16 mg L⁻¹) concentration exposures; three Cu (0.081, 0.145 and 0.292 mg L⁻¹), Zn (1.09, 5.30 and 8.19 mg L⁻¹), Ni (5.51, 9.02 and 31.53 mg L⁻¹), and Al (88.38, 160.83 and 362.83 mg L⁻¹) concentration exposures. In general, the Cu, Cd, Pb, Zn, Ni, Fe, Al, and Mn bioconcentration in *M. tuberculata* increases with increasing concentration exposure. Similar results were reported by Moolman et al. [18] on Cd and Zn accumulation by two freshwater gastropods (*M. tuberculata* and *Helisoma duryi*). Hoang and Rand [55] showed that whole body Cu concentration of juvenile apple snails (*Pomacea paludosa*) was significantly correlated with soil and water Cu concentrations. In other experiments, Hoang et al. [56] showed that

TABLE 3: Comparison of LC₅₀ values of freshwater gastropod *M. tuberculata* with other freshwater mollusc.

Metal	Species	Water hardness (mg L ⁻¹)	Live stage	Test duration	LC ₅₀ (mg L ⁻¹)	Reference
Copper	<i>M. tuberculata</i>	18.7	Adult	96 h	0.14	This study
	<i>M. tuberculata</i>			48 h	3.6	[16]
	<i>M. tuberculata</i>		Juvenile	24 h	0.2	[15]
	<i>B. glabrata</i>	44	Adult	48 h	0.18	[19]
	<i>F. virens</i>	21	Adult	96 h	0.08	[20]
	<i>J. plicifera</i>	21	Adult	96 h	0.015	[20]
	<i>B. glabrata</i>	100	—	96 h	0.04	[21]
	<i>P. paludosa</i>	68	60 d	96 h	0.14	[22]
	<i>P. jenkinsi</i>	—	Adult	96 h	0.08	[23]
Cadmium	<i>M. tuberculata</i>	18.7	Adult	96 h	1.49	This study
	<i>Amnicola</i> sp.	50	Adult	96 h	8.4	[24]
	<i>P. fontinalis</i>	—	—	96 h	0.08	[25]
	<i>A. hypnorum</i>	45	Adult	96 h	0.09	[26]
	<i>B. glabrata</i>	100	—	96 h	0.3	[21]
	<i>V. bengalensis</i>	180	—	96 h	1.2	[27]
	<i>L. luteola</i>	195	Adult	96 h	1.5	[28]
Zinc	<i>M. tuberculata</i>	18.7	Adult	96 h	3.90	This study
	<i>P. gyrina</i>	36	Adult	96 h	1.27	[20]
	<i>L. acuminata</i>	375	—	96 h	10.49	[29]
	<i>L. luteola</i>	195	Adult	96 h	11.0	[30]
	<i>V. bengalensis</i>	180	—	96 h	0.64	[27]
	<i>P. heterostropha</i>	20	Adult	96 h	1.11	[31]
	<i>P. heterostropha</i>	100	Adult	96 h	3.16	[31]
Lead	<i>M. tuberculata</i>	18.7	Adult	96 h	6.82	This study
	<i>L. emarginata</i>	150	—	48 h	14.0	[32]
	<i>E. livescens</i>	150	—	48 h	71.0	[32]
	<i>Filopaludina</i> sp.	—	Adult	96 h	190	[33]
	<i>V. bengalensis</i>	165	—	96 h	2.54	[34]
	<i>A. hypnorum</i>	60.9	—	96 h	1.34	[35]
Nickel	<i>M. tuberculata</i>	18.7	Adult	96 h	8.46	This study
	<i>Amnicola</i> sp.	50	Adult	96 h	14.3	[24]
	<i>J. plicifera</i>	59	Adult	96 h	0.24	[20]
	<i>L. luteola</i>	195	Adult	96 h	1.43	[28]
	<i>V. bengalensis</i>	180	—	96 h	9.92	[27]
	<i>L. acuminata</i>	375	—	96 h	2.78	[29]
Iron	<i>M. tuberculata</i>	18.7	Adult	96 h	8.49	This study
	<i>P. gyrina</i>	109	—	96 h	12.09	[36]
	<i>Planorbarius</i> sp.	—	—	48 h	7.32	[37]
	<i>S. libertina</i>	—	—	48 h	76.0	[38]
Aluminium	<i>M. tuberculata</i>	18.7	Adult	96 h	68.23	This study
	<i>Physa</i> sp.	47	—	96 h	55.5	[39]
	<i>A. limosa</i>	PH 3.5	—	96 h	1.0	[40]
	<i>A. limosa</i>	PH 4.5	—	96 h	0.40	[40]
Manganese	<i>M. tuberculata</i>	18.7	Adult	96 h	45.59	This study
	<i>B. globosus</i>	53	—	96 h	100.0	[41]

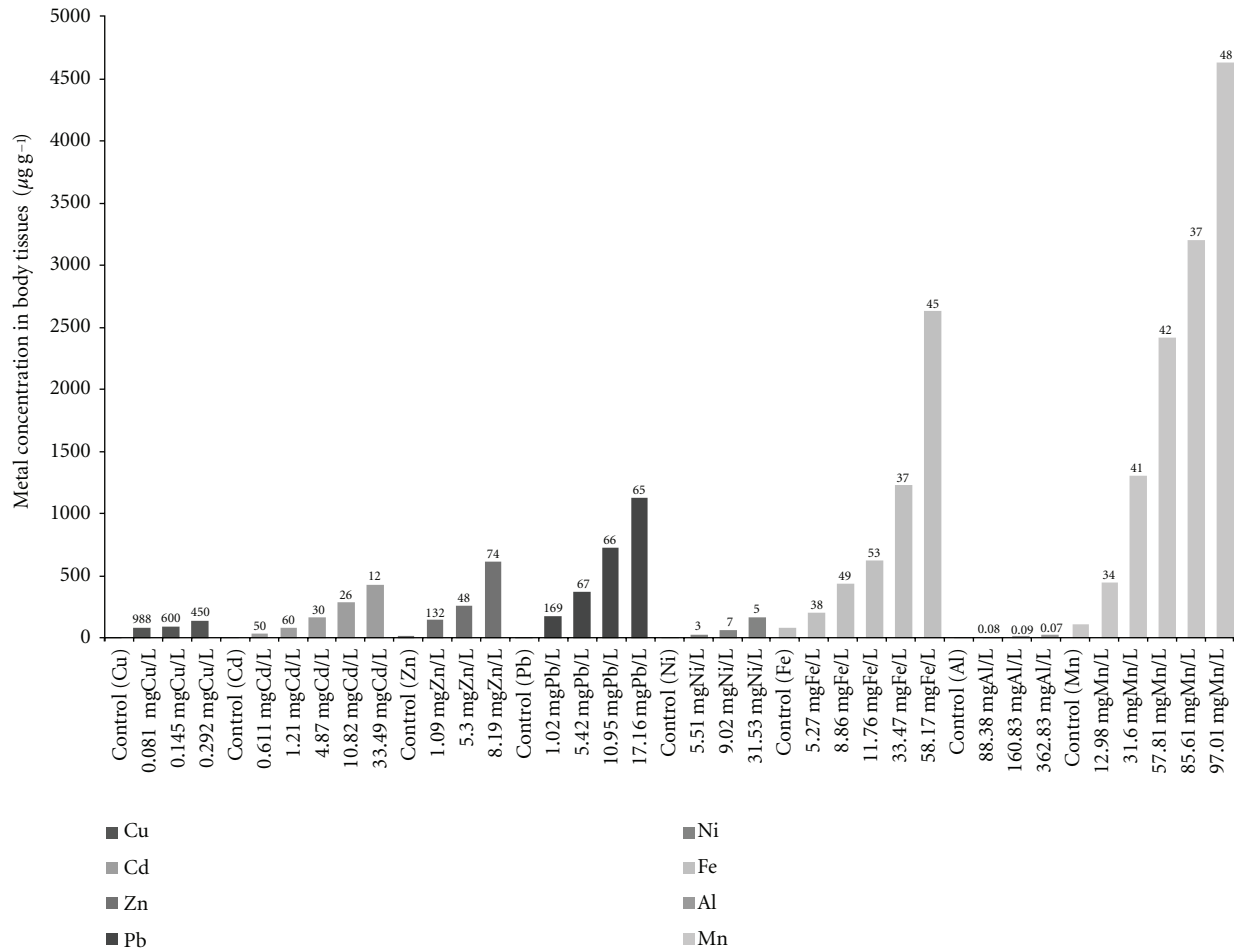


FIGURE 3: Bioconcentration of Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn (mean) in *M. tuberculata* soft tissues ($\mu\text{g g}^{-1}$ dry weight) after a four-day exposure to different concentrations of Cu, Cd, Zn, Pb, Ni, Fe, Al, and Mn. Concentration factor (CF) is indicated at the top of each bar.

whole body Cu concentrations of juvenile snails (*P. paludosa*) increased with exposure time and concentration and reached a plateau (saturation) after 14 days of exposure. These results are in agreement with the statement of Luoma and Rainbow [7] who state that the uptake of trace metals from solution by an aquatic organism is primarily concentration dependent. The higher the dissolved concentration of the trace metal, the higher the uptake of the metal from solution into the organism will be, until the uptake mechanism becomes saturated.

The present study shows that in general the highest concentration factor (CF) was noted for Cu (988), Pb (169), and Zn (132), and the lowest CF was for Al (0.07) (Figure 3). Similar results were reported by Lau et al. [14] who reported that *M. tuberculata* collected from the wild (Sarawak River) accumulated higher amounts of Cu, Zn, and As in the soft tissues compared to other metals. Adewunmi et al. [17] showed that Cu, Pb, and Cd were the highest metal accumulated in tissues of freshwater snails in dams and rivers in southwest Nigeria, and metal concentrations in the snails were varied with the seasons, especially for Cu which was higher in the dry season compared to the rainy season. According to Luoma and Rainbow [7] the factors

that affect the rate of uptake of metals affect the toxicity of metal. This is in agreement with the results from the present study which shows that Cu, which was the most toxic to the snail, also has the highest CF in the soft tissues of *M. tuberculata*. In explaining the toxicity of Cu, Hoang and Rand [55] demonstrate that the potential toxicity of Cu carbonate to snails may be explained by the carbonate content in the snails. The carbonate requirement for snails is more than for fish because snails require it for shell development. Copper may enter snails as Cu carbonate. After entering snails, Cu carbonate may be disassociated through biological and chemical reactions. Carbonate would be available for shell development and Cu would be accumulated in soft tissue. Hoang et al. [56] also reported that with the juvenile apple snail (*Pomacea paludosa*), most of the accumulated Cu was located in soft tissue (about 60% in the viscera and 40% in the foot) and the shell contained <4% of the total accumulated copper. However, a comparison of the uptake rate in aquatic organisms showed that in general the order of the uptake rate constant is $\text{Ag} > \text{Zn} > \text{Cd} > \text{Cu} > \text{Co} > \text{Cr} > \text{Se}$ [7]. This discrepancy is probably due to short time of exposure (four days) to metals in this study. Other factors which may influence the bioaccumulation of heavy metals in

aquatic organisms has been suggested, such as their feeding habit [57], growth rate and age of the organism [14, 58], and the bioavailability of the metals, which greatly depends on hardness of water, pH, and the acid-volatile sulphide of the water [59]. Hoang and Rand [55] showed that the apple snails (*Pomacea paludosa*) accumulated more Cu from soil-water than from water-only treatments and this suggests that apple snails accumulate Cu from soil (-sediment)/water systems. Organisms with higher growth rates also usually have lower metal concentrations in their bodies as the rate of increase in the weight of its tissue and shell will be higher than the accumulated metals [14]. According to Lau et al. [14], the shell of *M. tuberculata* would be most suitable for monitoring Cu in the aquatic environment, which has an approximately thirtyfold magnification capability and with standard errors of less than 10%. Zn would be best monitored by using the shell of *M. tuberculata*, whose magnification capability was approximately 35 times and its error was at approximately 15%. Both tissue and shell of *M. tuberculata* could also be used for monitoring arsenic as it has good magnification capabilities with moderate irregularity approximately 23%. However, it is important to note that the Lau et al. [14] study was conducted in the field (long-term exposure), while the present study was conducted in the laboratory with short-term exposure, and differences in accumulation trend and strategies (higher accumulation in soft tissues or shell) may exist.

Aquatic molluscs possess very diverse strategies in the handling and storage of accumulated metals, which include being in the forms of metal-rich granules metallothioneins (MT) or metallothionein-like proteins [60–62]. Accumulation strategies of invertebrates vary intraspecifically between metals and interspecifically for the same metal in closely related organisms [62, 63]. Moolman et al. [18] showed that *M. tuberculata* had a much higher uptake of Zn in the Zn and in the mixed Cd/Zn exposures compared to *Helisoma duryi*, and Zn was readily accumulated with increasing metal concentrations. Lau et al. [14] also demonstrated that Zn concentrations in *M. tuberculata* were significantly higher than those in the molluscs *Brotia costula* and *Clithon* sp. The present study shows that the CF of Zn was higher than the Cd in the soft tissues of *M. tuberculata*. With the juvenile apple snail, Hoang et al. [56] showed that the snails accumulated Cu during the exposure phase and eliminated Cu during the depuration phase. Metals accumulated in animals can be stored without excretion leading to high body concentrations (accumulators), or the metal levels in the body can be maintained at a low constant body concentration (regulators) by balancing the uptake with controlled rates of excretion [64].

4. Conclusions

This study showed that *M. tuberculata* was equally sensitive to metals compared to other freshwater gastropods. Cu was the most toxic metal to *M. tuberculata* followed by Cd, Zn, Pb, Ni, Fe, Mn, and Al. A comparison of the bioconcentration of metals in soft tissues of *M. tuberculata* showed that among the eight metals studied; Cu, Pb, and Zn

were the most accumulated and Al was least accumulated. *M. tuberculata* is widely distributed in urban and suburban areas which makes it easy to sample and very useful in ecotoxicology studies. This study indicates that *M. tuberculata* could be a potential bioindicator organism of metals pollution and in toxicity testing.

Acknowledgments

This study was funded by the Ministry of Science and Technology, Malaysia (MOSTI) under e-Science Fund code nos. 06-01-02-SF0217 and 06-01-02-SF472. The authors do not have any direct financial relation with the commercial identity mentioned in this paper.

References

- [1] C. H. Walker, S. P. Hopkin, R. M. Silby, and D. B. Peakall, *Principles of Ecotoxicology*, CRC Press, Boca Raton, Fla, USA, 3rd edition, 2006.
- [2] N. A. M. Shazili, K. Yunus, A. S. Ahmad, N. Abdullah, and M. K. A. Rashid, "Heavy metal pollution status in the Malaysian aquatic environment," *Aquatic Ecosystem Health and Management*, vol. 9, no. 2, pp. 137–145, 2006.
- [3] DOE (Department of Environment, Malaysia), *Malaysia Environment Quality Report 2008*, Department of Environment, Ministry of Natural Resources and Environment, Kuala Lumpur, Malaysia, 2009.
- [4] S. Z. Zulkifli, F. Mohamat-Yusuff, T. Arai, A. Ismail, and N. Miyazaki, "An assessment of selected trace elements in intertidal surface sediments collected from the Peninsular Malaysia," *Environmental Monitoring and Assessment*, vol. 169, no. 1–4, pp. 457–472, 2010.
- [5] C. K. Yap and B. H. Pang, "Assessment of Cu, Pb, and Zn contamination in sediment of north western Peninsular Malaysia by using sediment quality values and different geochemical indices," *Environmental Monitoring and Assessment*, vol. 183, no. 1–4, pp. 23–39, 2011.
- [6] W. J. Adams and C. D. Rowland, "Aquatic toxicology test methods," in *Handbook of Ecotoxicology*, D. J. Hoffman, B. A. Rattner, G. A. Burton Jr., and J. Cairns Jr., Eds., CRC Press, Boca Raton, Fla, USA, 2nd edition, 2003.
- [7] S. N. Luoma and P. S. Rainbow, *Metal Contamination in Aquatic Environment: Science and Lateral Management*, Cambridge University Press, New York, NY, USA, 2008.
- [8] S. Panha and J. B. Burch, "Mollusca," in *Freshwater Invertebrates of the Malaysian Region*, C. M. Yule and Y. H. Sen, Eds., pp. 225–253, Academy of Science Malaysia, Kuala Lumpur, Malaysia, 2004.
- [9] A. J. Mitchell, M. S. Hobbs, and T. M. Brandt, "The effect of chemical treatments on red-rim melania *Melanoides tuberculata*, an exotic aquatic snail that serves as a vector of trematodes to fish and other species in the USA," *North American Journal of Fisheries Management*, vol. 27, no. 4, pp. 1287–1293, 2007.
- [10] G. Pererea and J. G. Walls, *Apple Snails in the Aquarium*, 1996.
- [11] A. J. Benson, "*Melanoides tuberculatus*," USGS Nonindigenous Aquatic Species Database, Gainesville, Fla, USA, 2008, <http://nas.er.usgs.gov/queries/FactSheet.aspx?speciesID=1037>.
- [12] C. C. Appleton, A. T. Forbes, and N. T. Demetriades, "The occurrence, bionomics and potential impacts of the invasive freshwater snail *Tarebia granifera* (Lamarck, 1822)

- (Gastropoda: Thiaridae) in South Africa," *Zoologische Mededelingen*, vol. 83, pp. 525–536, 2009.
- [13] U. Gardenfors, T. Westermark, U. Emanuelsson, H. Mutvei, and H. Walden, "Use of land-snail shells as environmental archives: preliminary results," *AMBIO*, vol. 17, no. 5, pp. 347–349, 1988.
- [14] S. Lau, M. Mohamed, A. Tan Chi Yen, and S. Su'Ut, "Accumulation of heavy metals in freshwater molluscs," *Science of the Total Environment*, vol. 214, no. 1–3, pp. 113–121, 1998.
- [15] H. S. Bali, S. Singh, and D. P. Singh, "Trial of some molluscicides on snails *Melanooides tuberculatus* and *Vivipara bengalensis* in laboratory," *Indian Journal of Animal Sciences*, vol. 54, no. 4, pp. 401–403, 1984.
- [16] B. B. Mostafa, F. A. El-Deeb, N. M. Ismail, and K. M. El-Said, "Impact of certain plants and synthetic molluscicides on some fresh water snails and fish," *Journal of the Egyptian Society of Parasitology*, vol. 35, no. 3, pp. 989–1007, 2005.
- [17] C. O. Adewunmi, W. Becker, O. Kuehnast, F. Oluwole, and G. Dörfler, "Accumulation of copper, lead and cadmium in freshwater snails in southwestern Nigeria," *Science of the Total Environment*, vol. 193, no. 1, pp. 69–73, 1996.
- [18] L. Moolman, J. H. J. Van Vuren, and V. Wepener, "Comparative studies on the uptake and effects of cadmium and zinc on the cellular energy allocation of two freshwater gastropods," *Ecotoxicology and Environmental Safety*, vol. 68, no. 3, pp. 443–450, 2007.
- [19] E. C. De Oliveira-Filho, R. Matos Lopes, and F. J. Roma Paumgarten, "Comparative study on the susceptibility of freshwater species to copper-based pesticides," *Chemosphere*, vol. 56, no. 4, pp. 369–374, 2004.
- [20] A. V. Nebeker, A. Stinchfield, C. Savonen, and G. A. Chapman, "Effects of copper, nickel and zinc on three species of oregon freshwater snails," *Environmental Toxicology and Chemistry*, vol. 5, no. 9, pp. 807–811, 1986.
- [21] C. Bellavere and J. Gorbi, "A comparative analysis of acute toxicity of chromium, copper and cadmium to *Daphnia magna*, *Biomphalaria glabrata*, and *Brachydanio rerio*," *Environmental Technology Letters*, vol. 2, no. 3, pp. 119–128, 1981.
- [22] E. C. Rogevich, T. C. Hoang, and G. M. Rand, "The effects of water quality and age on the acute toxicity of copper to the Florida apple snail, *Pomacea paludosa*," *Archives of Environmental Contamination and Toxicology*, vol. 54, no. 4, pp. 690–696, 2008.
- [23] A. J. Watton and H. A. Hawkes, "The acute toxicity of ammonia and copper to the gastropod *Potamopyrgus jenkinsi* (Smith)," *Environmental Pollution Series A*, vol. 36, no. 1, pp. 17–29, 1984.
- [24] R. Rehwooldt, L. Lasko, C. Shaw, and E. Wirhowski, "The acute toxicity of some heavy metal ions toward benthic organisms," *Bulletin of Environmental Contamination and Toxicology*, vol. 10, no. 5, pp. 291–294, 1973.
- [25] K. A. Williams, D. W. J. Green, and D. Pascoe, "Studies on the acute toxicity of pollutants to freshwater macroinvertebrates. I. Cadmium," *Archiv für Hydrobiologie*, vol. 102, no. 4, pp. 461–471, 1985.
- [26] G. W. Holcombe, G. L. Phipps, and J. W. Marier, "Methods for conducting snail (*Aplexa hypnorum*) embryo through adult exposures: effects of cadmium and reduced pH levels," *Archives of Environmental Contamination and Toxicology*, vol. 13, no. 5, pp. 627–634, 1984.
- [27] P. K. Gupta, B. S. Khangarot, and V. S. Durve, "Studies on the acute toxicity of some heavy metals to an Indian freshwater pond snail *Viviparus bengalensis* L.," *Archiv für Hydrobiologie*, vol. 91, no. 2, pp. 259–264, 1981.
- [28] B. S. Khangarot and P. K. Ray, "Sensitivity of freshwater pulmonate snails, *Lymnaea luteola* L., to heavy metals," *Bulletin of Environmental Contamination and Toxicology*, vol. 41, no. 2, pp. 208–213, 1988.
- [29] B. S. Khangarot, S. Mathur, and V. S. Durve, "Comparative toxicity of heavy metals and interaction of metals on a freshwater pulmonate snail *Lymnaea acuminata* (Lamarck)," *Acta Hydrochimica et Hydrobiologica*, vol. 10, no. 4, pp. 367–375, 1982.
- [30] B. S. Khangarot and P. K. Ray, "Zinc sensitivity of a freshwater snail, *Lymnaea luteola* L., in relation to seasonal variations in temperature," *Bulletin of Environmental Contamination and Toxicology*, vol. 39, no. 1, pp. 45–49, 1987.
- [31] C. B. Wurtz, "Zinc effects on fresh-water Mollusks," *Nautilus*, vol. 76, pp. 53–61, 1962.
- [32] J. Cairns Jr., D. I. Messenger, and W. F. Calhoun, "Invertebrate response to thermal shock following exposure to acutely sub lethal concentrations of chemicals," *Archiv für Hydrobiologie*, vol. 77, no. 2, pp. 164–175, 1976.
- [33] S. Lantataeme, M. Kruatruchue, S. Kaewsawangsap, Y. Chitramvong, P. Sretarugga, and E. S. Upatham, "Acute toxicity and bioaccumulation of lead in the snail, *Eilopaludina* (Siamopaludina) *Martensi martensi* (Frauenfeldt)," *Journal of the Science Society of Thailand*, vol. 22, no. 3, pp. 237–247, 1996.
- [34] A. S. Gadkari and V. B. Marathe, "Toxicity of cadmium and lead to a fish and a snail from two different habitats," *Indian Association Water Pollution Control Technology*, vol. 5, pp. 141–148, 1983.
- [35] D. J. Call, L. T. Brooke, N. Ahmad, and D. D. Vaishnav, *Aquatic Pollutant Hazard Assessments and Development of a Hazard Prediction Technology by Quantitative Structure-Activity Relationships*, U.S. EPA Cooperation Agreement No. CR 809234-01-0, Centre for Lake Superior Environmental Studies, University of Wisconsin, Superior, Wis, USA, 1981.
- [36] W. J. Birge, J. A. Black, A. G. Westerman et al., *Recommendations on Numerical Values for Regulating Iron and Chloride Concentrations for the Purpose of Protecting Warmwater Species of Aquatic Life in the Commonwealth of Kentucky*, University of Kentucky, Lexington, Ky, USA, 1985.
- [37] M. Furmanska, "Studies of the effect of copper, zinc, and iron on the biotic components of aquatic ecosystems," *Polskie Archiwum Hydrobiologii*, vol. 26, no. 1-2, pp. 213–220, 1979.
- [38] Y. Nishiuchi and K. Yoshida, "Toxicities of pesticides to some fresh water snails," *Bulletin of the Agricultural Chemicals Inspection Station*, vol. 12, pp. 86–92, 1972.
- [39] D. J. Call, L. T. Brooke, C. A. Lindberg, T. P. Markee, D. J. McCauley, and S. H. Poirier, *Toxicity of Aluminum to Freshwater Organisms in Water of pH 6.5-8.5*, Technical Report Project No.549-238-RT-WRD, Center for Lake Superior Environmental Studies, University of Wisconsin, Superior, Wis, USA, 1984.
- [40] G. L. Mackie, "Tolerances of five benthic invertebrates to hydrogen ions and metals (Cd, Pb, Al)," *Archives of Environmental Contamination and Toxicology*, vol. 18, no. 1-2, pp. 215–223, 1989.
- [41] P. Tomasik, C. M. Magadza, S. Mhizha, A. Chirume, M. F. Zaranyika, and S. Muchiriri, "Metal-metal interactions in biological systems. Part IV. Freshwater snail *Bulinus globosus*," *Water, Air, and Soil Pollution*, vol. 83, no. 1-2, pp. 123–145, 1995.
- [42] APHA (American Public Health Association), *Standard Method for the Examination of Water and Wastewater*, part

- 8000, toxicity, Washington American Public Health Association, 18th edition, 1992.
- [43] J. D. Cooney, "Freshwater test," in *Fundamental of Aquatic Toxicology: Effects, Environmental fate and Risk Assessment*, G. M. Rand, Ed., pp. 71–102, Taylor & Francis, 2nd edition, 1995.
- [44] J. T. Lichfield, "A method for the rapid graphic solution of time-percentage effect curves," *Journal of Pharmacology and Experimental Therapeutics*, vol. 97, pp. 399–408, 1949.
- [45] J. T. Lichfield and F. Wilcoxon, "A simplified method of evaluating dose-effect experiments," *Journal of Pharmacology and Experimental Therapeutics*, vol. 96, pp. 99–113, 1949.
- [46] C. P. McCahon and D. Pascoe, "Use of *Gammarus pulex* (L.) in safety evaluation tests: culture and selection of a sensitive life stage," *Ecotoxicology and Environmental Safety*, vol. 15, no. 3, pp. 245–252, 1988.
- [47] I. Landner and R. Reuther, *Metals in Society and in the Environment*, Kluwer Publisher, Dordrecht, The Netherlands, 2004.
- [48] J. Gorski and D. Nugegoda, "Sublethal toxicity of trace metals to larvae of the blacklip abalone, *Haliotis rubra*," *Environmental Toxicology and Chemistry*, vol. 25, no. 5, pp. 1360–1367, 2006.
- [49] M. Ebrahimpour, H. Alipour, and S. Rakhshah, "Influence of water hardness on acute toxicity of copper and zinc on fish," *Toxicology and Industrial Health*, vol. 26, no. 6, pp. 361–365, 2010.
- [50] M. Shuhaimi-Othman, Y. Nadzifah, R. Nur-Amalina, and A. Ahmad, "Sensitivity of the freshwater prawn, *Macrobrachium lanchesteri* (Crustacea: Decapoda), to heavy metals," *Toxicology and Industrial Health*, vol. 27, no. 6, pp. 523–530, 2011.
- [51] M. Shuhaimi-Othman, Y. Nadzifah, R. Nur-Amalina, and A. Ahmad, "Toxicity of metals to a freshwater ostracod, *Stenocypris major*," *Journal of Toxicology*, vol. 2011, pp. 1–8, 2011.
- [52] M. Shuhaimi-Othman, N. Yakub, N. A. Ramle, and A. Abas, "Comparative metal toxicity to freshwater fish," submitted to *Journal of Toxicology and Industrial Health*, In press.
- [53] N. S. Umirah, *Toxicity of metals to aquatic worm Nais elinguis and midge Chironomus javanus*, Unpublished M.S. thesis, Universiti Kebangsaan Malaysia (UKM), Bangi, Malaysia, 2009.
- [54] P. C. Von Der Ohe and M. Liess, "Relative sensitivity distribution of aquatic invertebrates to organic and metal compounds," *Environmental Toxicology and Chemistry*, vol. 23, no. 1, pp. 150–156, 2004.
- [55] T. C. Hoang and G. M. Rand, "Exposure routes of copper: short term effects on survival, weight, and uptake in Florida apple snails (*Pomacea paludosa*)," *Chemosphere*, vol. 76, no. 3, pp. 407–414, 2009.
- [56] T. C. Hoang, E. C. Rogevich, G. M. Rand, and R. A. Frakes, "Copper uptake and depuration by juvenile and adult Florida apple snails (*Pomacea paludosa*)," *Ecotoxicology*, vol. 17, no. 7, pp. 605–615, 2008.
- [57] G. Mance, *Pollution Threat of Heavy Metal in Aquatic Environments*, Elsevier Applied Science, New York, NY, USA, 1990.
- [58] R. J. Pentreath, "The accumulation of organic mercury from sea water by the plaice, *Pleuronectes platessa* L.," *Journal of Experimental Marine Biology and Ecology*, vol. 24, no. 2, pp. 121–132, 1976.
- [59] J. M. Besser, C. G. Ingersoll, and J. P. Giery, "Effects of spatial and temporal variation of acid-volatile sulphide on the bioavailability of copper and zinc in freshwater sediment," *Environmental Technology and Chemistry*, vol. 15, pp. 286–293, 1996.
- [60] G. Roesijadi, "Metallothioneins in metal regulation and toxicity in aquatic animals," *Aquatic Toxicology*, vol. 22, no. 2, pp. 81–114, 1992.
- [61] A. Z. Mason and K. D. Jenkin, "Metal detoxification in aquatic organisms," in *Metal Speciation and Bioavailability in Aquatic Systems*, A. Tessier and D. R. Turner, Eds., pp. 479–608, Wiley, Chichester, UK, 1995.
- [62] P. S. Rainbow, "Trace metal concentrations in aquatic invertebrates: why and so what?" *Environmental Pollution*, vol. 120, no. 3, pp. 497–507, 2002.
- [63] P. S. Rainbow and R. Dallinger, "Metal uptake, regulation, and excretion in freshwater invertebrates," in *Ecotoxicology of Metals in Invertebrates*, R. Dallinger and P. S. Rainbow, Eds., pp. 119–131, Lewis Publishers, Boca Raton, Fla, USA, 1993.
- [64] P. S. Rainbow, "Heavy metals in marine invertebrates," in *Heavy Metals in the Marine Environment*, R. W. Furness and P. S. Rainbow, Eds., pp. 68–75, CRC Press, Boca Raton, Fla, USA, 1990.