MLL-AF4 driven leukemogenesis: what are we missing?

Ronald W Stam¹

¹Department of Pediatric Oncology/Haematology, Erasmus MC-Sophia Children's Hospital, Dr. Molewaterplein 50, Room: Ee15-14a, 3015 GE Rotterdam, The Netherlands Cell Research (2012) **22**:948-949. doi:10.1038/cr.2012.16; published online 31 January 2012

Cell Research (2012) 22.948-949. doi:10.1038/ci.2012.10, published online 51 January 20

Pre-leukemic MLL-AF4 fusions arise prenatally and typically lead to overt acute lymphoblastic leukemia (ALL) at or shortly after birth. In a recent study, Bueno and colleagues explored the effects of MLL-AF4 expression in human embryonic stem cells (hESCs), with a focus on early hemato-endothelial development.

Infant pro-B acute lymphoblastic leukemia (ALL) harboring MLL-AF4 fusion proteins instigated by chromosomal translocation t(4;11), represents an aggressive, high-risk type of childhood leukemia, characterized by a very brief disease latency and undisputedly originates in utero. Despite of recent advances and multiple important breakthroughs, MLL-AF4-driven leukemogenesis remains difficult to model and mouse models that accurately recapitulate the disease phenotype and latency are still lacking. Among several remarkable studies recently published, Montes et al. [1] demonstrated that enforced expression of MLL-AF4 in cord blood-derived hematopoietic stem cells (HSCs) increased the clonogenic potential of CD34⁺ progenitors and promoted proliferation, but appeared insufficient to induce leukemia. This study undeniably questions whether MLL-AF4 fusion proteins are capable

of driving leukemogenesis on their own or whether additional genetic events are required such as RAS mutations [2]. However, recent whole genome sequencing analysis in primary MLLrearranged infant ALL samples revealed the presence of remarkably few somatic mutations [3]. Contributing to the complexity of the matter, Bursen et al. [4] recently showed that introducing the reciprocal fusion protein AF4-MLL (resulting from the same balanced t(4;11)translocation), but not MLL-AF4, into murine hematopoietic stem/progenitor cells induced ALL in mice without the requirement of MLL-AF4. Nonetheless, these experiments have not yet been performed in human HSCs. Moreover, MLL-AF4 and AF4-MLL knockdown experiments have shown that t(4;11)positive cell lines display addiction to MLL-AF4 (which appeared essential for leukemic cell proliferation and survival), but not to AF4-MLL [5]. Thus, the AF4-MLL fusion protein may well be important or essential in the early transformation process and the MLL-AF4 fusion is certainly required for the maintenance of the leukemia. However, although the studies by Montes et al. [1] and Bursen et al. [4] seem to support that MLL-AF4 by itself is not sufficient to induce leukemogenesis in HSCs, others were able to induce lymphoid leukemias using MLL-AF4 knockin models in mice [2, 6]. Yet, in these latter studies both the disease phenotye and latency of the observed leukemias appeared to deviate from the highly immature pro-B cell phenotype characteristically found in humans.

npg

While the above described contradictions make it difficult to draw solid conclusions on the oncogenic potential of the MLL-AF4 fusion protein itself, there is another important question to be asked and answered: Are these MLL-AF4-driven leukemogenesis studies targeting the right cells? In this issue of Cell Research, Bueno et al. [7] elegantly attempted to address this question by creating a human-specific cellular system to study early hemato-endothelial development in MLL-AF4-expressing human embryonic stem cells (hESCs). A recent report showed that bone marrowderived mesenchymal stem cells from primary t(4;11)-positive pro-B infant ALL patients harbor and express the MLL-AF4 fusion gene [8]. Thus, MLL-AF4 may well arise prenatally in prehematopoietic mesodermal or hemangioblastic precursors sprouting from differentiating hESCs [9] rather than in more committed HSCs. From this perspective, Bueno et al. [7] introduced MLL-AF4 expression in hESCs and monitored the consequences. Interestingly, enforced MLL-AF4 expression in hESCs led to the accelerated emergence and elevated frequencies of hemogenic precursors. Moreover, in these hESCs, MLL-AF4 appeared to act as a global transcriptional activator, positively regulating homeobox gene expression, which is in line with what is usually (but

Correspondence: Ronald W Stam Tel: +31-10-7044654; Fax: +31-10-7044708 E-mail: r.stam@erasmusmc.nl

not always) observed in primary t(4:11)positive infant ALL samples [10]. Nevertheless, the MLL-AF4 fusion protein was not able to transform hESC-derived hematopoietic cells, but instead strongly impaired subsequent hematopoietic commitment in favor of an endothelial cell fate. Unfortunately, the latter brings us back to the same persistent conundrum: In order to successfully induce MLL-AF4-positive leukemia, did this system adopted by Bueno et al. require additional genetic hits such as the presence of the AF4-MLL fusion or RAS mutations? Or, despite the relevant rationale behind targeting hESC-derived pre-hematopietic precursors, did these cells not reflect the correct equivalents from which t(4;11)-positive pro-BALL in infants originate? Perhaps MLL-AF4positive infant ALL does arise in HSCs or early HSC progenitors, but not in those obtained from cord blood or the bone marrow. Given the strong body of evidence supporting that MLL-AF4 fusions arise during embryonic development [11, 12] when hematopoiesis still mainly takes place in the liver, the correct target cells, e.g., hematopoietic or specific lymphoid-monocytic stem cells [13], should possibly be searched for in the fetal liver.

Nonetheless, the study presented by Bueno *et al.* [7] provides unique insights into how MLL fusions regulate human embryonic hematopoietic specification and represents an intriguing experimental system to study the impact of MLL fusions from a developmental point of view. Hopefully this or similar approaches will further be exploited to unravel the riddle of MLL-AF4-driven leukemogenesis.

References

- Montes R, Ayllon V, Gutierrez-Aranda I, et al. Enforced expression of MLL-AF4 fusion in cord blood CD34+ cells enhances the hematopoietic repopulating cell function and clonogenic potential but is not sufficient to initiate leukemia. *Blood* 2011; **117**:4746-4758.
- 2 Tamai H, Miyake K, Takatori M, *et al.* Activated K-Ras protein accelerates human MLL/AF4-induced leukemolymphomogenicity in a transgenic mouse model. *Leukemia* 2011; **25**:888-891.
- 3 Andersson AK, Ma J, Wang J, et al. Whole genome sequence analysis of 22 MLL rearranged infant acute lymphoblastic leukemias reveals remarkably few somatic mutations: A report from the St Jude Children's Research Hospital - Washington University Pediatric Cancer Genome Project. 53rd ASH Annual Meeting and Exposition 2011; abstract #69, http://ash.confex.com/ash/2011/ webprogram/Paper40274.html
- 4 Bursen A, Schwabe K, Ruster B, et al. The AF4.MLL fusion protein is capable of inducing ALL in mice without requirement of MLL.AF4. *Blood* 2010; 115:3570-3579.
- 5 Kumar AR, Yao Q, Li Q, Sam TA,

Kersey JH. t(4;11) leukemias display addiction to MLL-AF4 but not to AF4-MLL. *Leuk Res* 2011; **35**:305-309.

- 6 Krivtsov AV, Feng Z, Lemieux ME, et al. H3K79 methylation profiles define murine and human MLL-AF4 leukemias. Cancer Cell 2008; 14:355-368.
- 7 Bueno C, Montes R, Melen GJ, *et al.* A human ESC model for MLL-AF4 leukemic fusion gene reveals an impaired early hematopoietic-endothelial specification. *Cell Res* 2012; **22**:986-1002.
- 8 Menendez P, Catalina P, Rodriguez R, *et al.* Bone marrow mesenchymal stem cells from infants with MLL-AF4+ acute leukemia harbor and express the MLL-AF4 fusion gene. *J Exp Med* 2009; **206**:3131-3141.
- 9 Vodyanik MA, Yu J, Zhang X, *et al*. A mesoderm-derived precursor for mesenchymal stem and endothelial cells. *Cell Stem Cell* 2010; 7:718-729.
- 10 Stam RW, Schneider P, Hagelstein JA, *et al.* Gene expression profilingbased dissection of MLL translocated and MLL germline acute lymphoblastic leukemia in infants. *Blood* 2010; **115**:2835-2844.
- 11 Ford AM, Ridge SA, Cabrera ME, et al. In utero rearrangements in the trithoraxrelated oncogene in infant leukaemias. Nature 1993; 363:358-3460.
- 12 Greaves MF, Maia AT, Wiemels JL, Ford AM. Leukemia in twins: lessons in natural history. *Blood* 2003; 102:2321-2333.
- 13 Greaves MF, Wiemels J. Origins of chromosome translocations in childhood leukaemia. *Nat Rev Cancer* 2003; 3:639-649.