At least three human homeoboxes on chromosome 12 belong to the same transcription unit

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ABSTRACT

Mammalian homeoboxes show a clustered chromosomal organization. In the mouse, at least seven homeoboxes on chromosome 6 and at least six on chromosome 11 identify the murine $\underline{Hox-1}$ and $\underline{Hox-2}$ loci, respectively. A number of homeoboxes on chromosome 7 define the human $\underline{HOX-1}$ locus and homeoboxes on chromosome 17 define the human $\underline{HOX-2}$ locus. We studied the genomic organization of three homeobox sequences of the $\underline{HOX-3}$ locus on chromosome 12 and analyzed transcripts from this region. Structural characterization and sequencing of several cDNA clones reveal that the three homeobox sequences present in this chromosomal region identify a single transcription unit. Primary transcripts are alternatively processed to give mature messengers with a common 5' noncoding exon encoding different proteins containing one of the three homeodomains.

INTRODUCTION

The homeobox is a DNA sequence conserved in several genes involved in the determination of Drosophila body segment (1). Homeobox-containing genes have been identified in a wide range of species and a number of such genes have been isolated from sea urchin (2), frog (3-8), mouse (9-23) and man (24-28). The high degree of conservation of the homeodomains identified in these species suggests that vertebrate and fly homeobox genes may perform corresponding functions in development (29). Consistent with the hypothesis of a common role in the control of development, Drosophila and vertebrate homeobox genes share some features. Firstly, most mammalian homeobox sequences show a clustered chromosomal organization. At least 13 murine homeoboxes are clustered in two 60 kb regions on chromosomes 6 and 11 (30). The seven boxes present on chromosome 6 define collectively the murine Hox-1 locus, whereas the six on chromosome 11 define the murine Hox-2 locus. Two other class I homeobox sequences have been mapped on chromosomes 15 and 12, which should define murine <u>Hox-3</u> and <u>Hox-4</u> loci, respectively. We have isolated human homeoboxes (26-28) representing the human cognates of murine homeoboxes in the <u>HOX-1</u>, <u>HOX-2</u> and <u>HOX-3</u> loci (31). Secondly, structural analysis of several Xenopus, murine and human cDNA clones revealed that in common with most Drosophila homeobox genes the homeodomain

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lies in the last 3' exon with an intron just upstream from it. Finally, most <u>Drosophila</u> and vertebrate homeobox genes are expressed in multiple transcripts with a complex transcriptional organization.

We have determined the chromosomal organization of three human homeobox sequences of the $\underline{HOX-3}$ locus on chromosome 12. Analysis of cDNA clones representing transcripts from this region reveals that these three homeoboxes are transcribed in a single primary transcript, alternatively spliced to give different mature messengers containing different homeoboxes in their 3' exons.

MATERIALS AND METHODS

cDNA and genomic clones

Three cDNA libraries were prepared from poly(A)+RNA of full-term placenta in $\lambda gt11$ (32). A genomic library in pcos2EMBL cosmid vector (33) was kindly provided by Anna-Maria Frischauf. These libraries were screened according to standard procedures (34). DNA fragments of interest were subcloned in pEMBL8 (35). DNA sequencing was performed according to Sanger et al. (36) and Maxam and Gilbert (37).

RNA isolation and analysis

Eight-week human embryos were obtained virtually intact by legal curettage abortions (26). Total RNA was extracted from embryos and term placenta by the guanidinium thiocyanate technique (38) and poly(A)+-selected by one passage on oligo (dT)-cellulose columns. Poly(A)+ RNA was run on 1.0% agarose-formaldehyde gels, transferred to nitrocellulose (Schleicher & Schuell, BA-85) or nylon (Amersham, Hybond N) membranes by Northern capillary blotting and hybridized to 10⁷ c.p.m. of DNA probe labelled by nick translation to a specific activity of 3-8 x 10⁸ c.p.m. per μĝ. Pre-hybridization and hybridization were carried out as described (26). stringent conditions (15 mM NaCl/1.5 mM After washing under sodium citrate/0.1% SDS at 65°C), the blots were exposed for 1-7 days at -70°C to Kodak XR-5 films in X-omatic intensifying screen cassette. Probes were then removed by boiling in 0.1% SDS buffered solution, and the filters rehybridized to a human actin probe for normalization. Probes used in Northern blot experiments shown in Fig. 6 are as follows: a) probe labelled "5' exon" contains nucleotides 400-635 of the first exon of Fig. 2; b) probe "3' cp25" contains nucleotides 660-865 of the last exon of Fig. 2: c) probe "3' cp19" contains nucleotides 297-493 of the last exon of Fig. 3; d) probe "cp8 specific" contains nucleotides 437-664 of the second exon of Fig. 3.

RESULTS

We have reported the isolation of several human cDNA clones containing homeobox sequences, which represent transcripts from four different genomic sequences (25). Representative clones of the four groups were termed <u>c1</u> (=<u>HOX-2.3</u>), <u>c10</u> (=<u>HOX-2.1</u>), <u>c8</u> and <u>c13</u>.

In order to study the chromosomal organization of human homeobox genes we isolated genomic sequences present in overlapping cosmid clones using our cDNA clones as probes. A 45 kb DNA region of the human $\underline{HOX-3}$ locus on

E E E	с8 ср11 ср Е Е Е Е Е Е Е Е Е Е Н П Е Е Е Е Е Е Е Е
2kb	
Antp	10 ArglysArg8lyArg8lnThrTyrThrArgTyr8lnThrLeu8luLeu8luLys8luPhe C6CAAAC6C6G8AA6AGGAA6ATACACCC66TACCA6AACTATA6A6CTA6A6AA66A6TTT
c8	Arg <u>Arg</u> Arg&lyArg&ln <u>ile</u> Tyr <mark>&er</mark> ArgTyr&lnThrLeu&luLeu&luLys&luPhe C&GA&GC&C&CCA&ATC1ACTC&C>ACCA&ACCT&GAACT&GAAGGAATTT
CP11	<u>91 y</u> lysarg <u>Bar</u> arg <u>1hr9ar</u> tyrthrarg tyrði nihrleuði uleuði ulysði uPhe Gecaagusgi Cocgaaccagttacaggggi accagaci ci ggaact Cgagaaagaai tC
CP 19	<u>Pro</u> lysarg <u>Ber</u> arg <u>àleàle</u> tyrthrarg <u>01n</u> 01n <u>Vel</u> leuGluleu01ulysGluPhe CCCAAGCGC1CGAGGGCAGCCIATACCCGGCAGCAAGTCCTGGAATTAGAGAAAGAGITT
Antp	30 HisPheAsnArgTyrLeuThrArgArgArg11e01uI1eA1aHisA1aLeuCysLeu CACIICAAICSCIACTIGACCCGICGGAAGGATCGCGAGATCGCCCAGGCCIGIGCCCC
cB	HisPheAsnArgTyrLeuThrArgArgArgArg11e01uI1eA1a <u>Asn</u> A1aLeuCysLeu CAC11CAAICGCTACCTAACGCGGCGCCCGGCGCATCGAGATCGCCAACGCGCTTTGCCTG
CP11	HisPheAsnArgTyrLeuThrArgArgArgArg1le8luIleAla <u>AsnAsn</u> LeuCysLeu CaciiiAacceciaccicacicececaecaecaeaaaacaiceccaacaaciic
CP 19	His <u>Tyr</u> AsnArgTyrLeuThrArgArgArgArgIle0luIleAlaHis <u>Ser</u> LeuCysLeu Callacaaccoclgacccgaaggaggaggalcgaglcgagl
	50 60
Antp	1hrGluArgGlnIleLysIle1rpPheGlnAsnArgArgMetLysIrpLysLysGluAsnLys AcggaglgCcCAgaIaaagaIIIGgIICCAgaaICGGCGCAIGAagIGGAAgAAgAAgAACAAg
c8	Thr BluArgBln I lelys I le TrpPheBlnAsnArgArgMetLys TrpLysLysBlu <u>BerAsn</u> Accgagcgacaga i caaaa i ctgg i tccagaaccggcggatgaag i ggaaaaaagaa i ci aa t
CP11	<u>Asn</u> gluargGlnilelysileTrpPheGlnAsnArgArgMetLysTrpLysLys <u>AspBer</u> Lys An Igagayacaga I caagat Ctggttccagaaccggatgaaggagaagaagaagattccaaa
CP 19	<u>Ser</u> biuArgGinileLysIleTrpPheGinAsnArgArgMetLysTrpLysLys <u>AsphisArg</u> 1016/06/66/069104441016611004100661060416449166446647640644

<u>Figure 1</u>: Genomic map of a region of the human $\underline{HOX-3}$ locus containing three homeobox sequences (filled boxes). E=EcoRI. Below the map the nucleotide sequence of the three homeoboxes is shown together with the conceptual translation. The aminoacid changes relative to a consensus sequence, represented by the <u>Drosophila</u> <u>Antennapedia</u> (<u>Antp</u>) homeodomain, are underscored.

chromosome 12 was isolated using clone $\underline{c8}$ as probe. Fig. 1 shows the map of this chromosomal region. Two new homeobox sequences have been mapped in this region downstream from the $\underline{c8}$ homeobox and provisionally termed $\underline{cp11}$ and $\underline{cp19}$. Their sequence is also shown in Fig. 1.

We screened three different cDNA libraries prepared from human placenta with genomic probes containing these three <u>HOX-3</u> homeoboxes. Several cDNA clones were isolated belonging to at least four classes. We report here the structure of representatives of three classes. Fig. 2 shows the nucleotide sequence of clone <u>cp25</u>, a placental messenger containing the <u>c8</u> homeobox.



<u>Figure 2</u>: Nucleotide sequence of genomic regions corresponding to exons of the cDNA clone <u>cp25</u>. An arrowhead marks the 5' terminus of the first exon as determined by RNase protection experiments. Dashes indicate nucleotide identity between genomic and cDNA sequences. The conserved pentapeptide Ile-Tyr-Pro-Trp-Met is underlined as are polyadenylation signals. The homeodomain is boxed. The 3' terminus of clone <u>c8.5111</u> (28) is indicated.

It has a 3' untranslated (3'-UT) region 82 bp longer than clone <u>c8.5111</u> (28), isolated from adult fibroblasts, because it uses an alternative polyadenylation signal. It consists of 3 exons with a first long intron of about 11 kb and a second of 800 bp just upstream from the homeobox as is the case for <u>c8.5111</u> (28). The coding region of predicted 153 aminoacid residues initiates in the second exon, which contains the conserved <u>pre-box</u> pentapeptide (27), and terminates in the 3' exon 34 aminoacid residues

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TCCBTTCCBABCGCTCCCCABAACABTCCTCCCCTBTAAGAGCCTAACCATTGCCAGGGAAACCTGCCCTGGGCGCTCCCTTCATTAGCAGTATTTTTTT
                                           CD
200
BEGEAGAGAGACAGAAGTIGABBBECATCICICICICICICCCBACCCICIDBCCCCCCAABBBBCABBAATBCABBABCABBABCIBABTIGAGBACIIBABTIGEGAGCT 400
500
600
CAGAGAGGCAGAGAGGGAGAGAGGGAGAGTGACAGCAGCGCTCGGTAAGTGTGTTTCCTTATTGGTT
                                                      35kb intron
                                                                          667
100
            ------
CCTECCTCECCEGECCECATEGECCAGAGEETTEGETEABTETATEGEGAAGAGEGECTEGACTCTEGETATECTTEGATEGEGEGECACTCCAGECTCTCCAG
   CCTCCTC66CTCA6CCT666CCCCTCCCCATCCAACATCCACTCCA6TCCTCATTCAACTTCCTCTTCCT6CGAAA6A66666C6CT6CCCC6T6ACCTAC 300
ACAGACTEAGACACGATCECCATEAATEGABACCTCTEGAAAAGCTCASGAGCCCGAGCCCCAGCGGGGCCCTEAGGGGGAGACCCTEGEGCEGB
                                                                          400
GGCTGAATCACTGCCTCCCGACAGTCCCCCCAATGCCCGGGCTTTBGAGGGGAGCCGGGGAGCTTCCCATCTCCTTTTBCAGGGGAGGGTTGTCAGTCTGCC
                                                                          500
GGGA 1516CAC1666666CAC1CCAACC1C16C1A8C1AACCCCACA1CACCACCCCCGCC1CCCASCACCACCACCACCACACACAAAAAAA11
                                                                          600
GGATACATTITIGAA TAAABCGA TICGBITICCTTATCCGGGGGACTGGBTIGCTCCBTBTGATTGGCCGGAGGABTCACATGGTGAAAGTAACTTTACAGGB
                                                                          200
                                                       HetileHetSerSerTyrLeuHei
TCGCTAGCTAGCTAGCAGGGCTTTATGGAGCAGAAAAACGACAAAGCGAGAAAAATTATTTTCCACTCCAGAAATTAATGATCATGAGCTCGTATTTGATG
                                                                          800
      -----
                                      20
                                             20
AspSerAssTyr 11eAspProLysPheProProCysBluBluTyr SerBlaAssSerTyr 11eProBluHisSerProBluTyr TyrBlyArgThrArgB
GACTCTAACTACATCGATCCGAAATTTCCTCCATGCGAAGAATATTCGCAAAATAGCTACATCCCTGAACACAGTCCGGAATATTACGGCCGG
                                                                   ACCAE
                                                                          900
80
                                      40
luSerGlyPheGlnHisHisHisGlnGluLeuTyrProProProProProProArgProSerTyrProGluArgGlnTyrSerCysThrSerLeuBlnGlyPr
AATCGGGATTCCAGCATCACCACCAGGAGCTGTACCCACCACCACCGCCTCGCGCCCTAGCTACCCTGAGCGCCCAGTATAGCTGCACCAGTCTCCAGGGGCC 1000
100
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                        120
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SorProSorProAlaProProAlaCysSerGlnProAlaProAspHisProSerSerAlaAlaSerLysGlnProlle<u>ValTyrProIrpHet</u>LysLysI
leHisValSerThe
TICACGITAGCACGGGTAGGCAACTTTGCTTTTT
                                          500 bp intron
                                                                         1234
                  180
                                       140
ValAssProAssTyrAssBiyBiyBiyBiyPoLysArgSerArgAlaAlatyrThrArgBisBisvalLeuGluLeuGluLysElu
CTCTTCTTCTTAT6CABT6AACCCCAATTATAAC56AA6666AACCCAA66CCCTCBA656CA6CTCTATACCC68CAA6TCCT66AATTAFAA6AAA6AB
                                                                          100
            180
                                190
PheHislyrAsnArglyrLeulhrArgArgArgArg11e6iulleAlaHisSerLeuCysLeuSer6iuArg6inlieLys11eTrpPhe6inAsnArgA
TTTCATTACAACCGCTACCTGACCCGAAGGAGGAGGAGGATCGAGATCGCCCCCCCTGTGGCCTCTCTGAGAGGCAGATCAAAATCTGGTTCCAAAACCGTC
                                                                          200
220
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BCATGAAATGGAAGAAGGACGACCGCCCCCCAACACCCAAASICAGGICAGCACCCCCGGCGGCGCGCGCGCGCGCCCCTICCGCAGCIACCCCGGG
                                                                          300
240
y Thr Ser GluAspHisSer GlnSer Ala Thr ProProGluGlnGlnArgAlaGluAsp11eThr ArgLevEND
TACTICIGAAGACCACICCCAGAGCGCCACGCCGGAGCAGCAGCAGCAGCAGGGCAGAGGACATIACCAGGIIAIAAAACATAACICACACCCCIGCCCCCACC 400
493
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Figure 3: Nucleotide sequence of genomic regions corresponding to exons of the cDNA clone <u>cp8</u>. An arrowhead marks the 5' terminus of the first exon. Dashes indicate nucleotide identity between genomic and cDNA sequences (<u>cp8</u> and <u>cp19</u>, upper and lower lines, respectively). The conserved Val-Tyr-Pro-Trp-Met pentapeptide is underlined. The homeodomain is boxed. Sequenced region does not include a polyadenylation signal.

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	10 20 3	o
c13	MVM8BYMVN8KYVDPKFPPCEEYLQGGYL	G
cp19	* * * * * * * * * * * * * * * * *	P
		_
		0
C13	* * * *	W
cp19	EHSPEYYG RTRESGFOHHHOELYPPPPR	P
		_
c13	70 90 90 9 PFGGSGPBPGSALPARBHBGEPBBPGBHY	O A
cp19	8 Y P E R Q Y 8 C T 8 L Q G P 6 N 8 R 6 H 6 P A Q A 6 H H	н
	100 110 12	0
c13	A P B E P C P A P P A A P P A A L P B A R A Y B B B D P K	9
cp19	PEK898LCEPAPL88A8P8PAPPAC89PA	P
	130 140 15	0
c13	PPSGTALKQPAVVYPWMKKVHANSVNPNY	т
cp19	DHPSSAASKQPIVYPWMKKIHVSTVNPNY	N
c13	160 170 16 BBEPKRBRTAYTROQVLELEKEFHFNRYL	
	******* ***************	*
cp19	GGEPKRSRAAYTRQQVLELEKEFHYNRYL	T
	190 200 21	0
c13	RRRIEIAHTLCLSERGIKIWFQNRRMKW	ĸ
ce 19	***************************************	*
		<u> </u>
	220 230 24	0
c13	KDHKLPNTKGRSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSS	н
CP19	KDHRLPNTKVRSAPPAGAAPSTLSAATPG	т
	NPR	_
c13	250 260 27 LQPMAKDHHTDLTTLEnd	0
CP19	SEDHSUSATPPEQQRAGDITRLEnd	

<u>Figure 4</u>: Alignment of predicted protein products of <u>c13</u> (27) and <u>cp19</u> cDNA clones. The aminoterminal domain is boxed as is a domain including the conserved pentapeptide, the homeodomain and five aminoacid residues down-stream from the latter. Asterisks indicate aminoacid identity and a vertical bar aminoacid conservation. The carboxyl terminus is underlined. The <u>Drosophila Dfd</u> product carboxyl terminus is DLTAL (39). Dashed lines indicate a potential recognition sequence for tyrosine protein kinase (44) conserved in the <u>Dfd</u> product as well (39).



2kb

Figure 5: Structure of three cDNA clones isolated from placenta. Above the genomic map the stucture of clone $\underline{c8.5111}$ (28) isolated from fibroblasts is shown. Boxes represent exons and a filled box indicates the homeobox. Clone $\underline{cp25}$ differs from $\underline{c8.5111}$ only in the terminal 3'-UT region. These clones use two alternative polyadenylation signals 76 bp apart.

downstream from the homeodomain. By means of RNase protection experiments we determined the 5' terminus of the first exon, as shown in Fig. 2.

Fig. 3 shows the structure of clones <u>cp19</u> and <u>cp8</u> containing the <u>cp19</u> homeobox. Clone <u>cp19</u> is composed of 4 exons with a first intron of about 35 kb, a second of 600 bp and a third of 500 bp upstream from the homeobox. The coding region of predicted 262 aminoacid residues initiates in the third exon and terminates 43 aminoacid residues downstream from the homeodomain. The 3'-UT region is not present in full in this clone. The predicted protein product encoded by clone <u>cp19</u> appears to be homologous to the <u>c13</u> product (27). The amino terminal domain and a second region centered around the homeodomain are particularly conserved (Fig. 4). The <u>cp19</u> homeodomain itself and five aminoacids downstream from it are very similar to the murine <u>Hox-1.4</u> (21), the human <u>HOX-1.4</u>, <u>HOX-2.6</u> (unpublished) and <u>c13</u> (27) and frog <u>Xhox-1A</u> (5) homeodomains, related in turn to the <u>Drosophila Deformed</u> (<u>Dfd</u>) homeodomain.

Clone <u>cp8</u>, bearing the same coding region of clone <u>cp19</u>, differs from it only because it contains, unspliced, the second intron of <u>cp19</u> (Fig. 3). Northern blot experiments show that it is not a cloning artefact (see

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Figure 6: Northern blot analysis of polyadenylated RNA (3 μ g) from 8-week embryos (E) and placenta (P). P lanes are overexposed. Probes used are about 200 bp DNA fragments from the common 5' exon, the 3'-UT region of cp25, the 3'-UT region of cp19 and the region of the second exon of cp8 not present in cp19 (Figs. 3 and 6). Transcript sizes are given in kb.

below). Strikingly, both $\underline{cp8}$ and $\underline{cp19}$ clones initiate within the genomic region representing the first exon of $\underline{cp25}$. There is only one genomic region corresponding to this 533 bp 5' exon, located 13 kb upstream from the $\underline{c8}$ homeobox region (Fig. 5). The three cDNA clones share a common 5' exon spliced, at the same 5' splice site, to different exons to give mature mRNAs encoding two different proteins with alternative homeodomains.

Northern blot experiments with placenta polyadenylated RNA (Fig. 6) reveal an intricate expression pattern. The 5' exon detects 3 major transcript classes of 3.5 kb, 2.2 kb and 1.8-1.7 kb. The 3.5 kb transcript class contains the 3' untranslated region of <u>cp19</u>, whereas the 2.2 kb size class contains the 3'-UT region of <u>cp25</u>. The <u>cp25</u> 3'-UT region probe detects in placenta a fainter 1.8 kb band representing transcripts possibly lacking the 5' exon. The <u>cp19</u> 3'-UT region probe detects two additional 3.2 kb and 2.8 kb bands. A probe derived from the second intron of <u>cp19</u>, termed "<u>cp8</u> specific" probe in Fig. 6, detects the 3.5 kb band. <u>cp25</u>, <u>cp19</u> and <u>cp8</u> expression in 8-week total human embryos is also shown.

DISCUSSION

Murine <u>Hox-1</u> and <u>Hox-2</u> homeoboxes show a clustered genomic organization with at least six homeoboxes per locus, whereas no <u>Hox-3</u> homeobox has been

mapped other than <u>Hox-3.1</u> (18). The human <u>HOX-3</u> locus on chromosome 12 contains clustered homeoboxes as is the case of the human HOX-1 and HOX-2 loci and of a locus mapped on chromosome 2 (unpublished). We have identified so far 18 homeoboxes in these four loci (our unpublished results) with the same 5'-3' orientation in each locus. It is interesting to note that in all four human loci there is a Dfd-like homeobox: HOX-1.4, HOX-2.6, cp19 and c13. We have shown that the entire cp19 predicted protein bears similarity to the c13 protein (Fig. 4) and both, in turn, to the Dfd protein, as already noted (39). The same is true for the HOX-2.6 protein (unpublished). Therefore, there are at least four human protein products corresponding to Dfd. The significance of this redundancy is elusive and requires further investigation. Alignment of the four human loci in such a way that HOX-1.4, HOX-2.6, cp19 and c13 homeoboxes occupy corresponding positions leads to interesting conclusions on the organization and origin of various loci. For example, the triplet <u>c8</u>, <u>cpl1</u>, <u>cpl9</u> should correspond to human (or murine) 1.2, 1.3, 1.4 and 2.2, 2.1, 2.6, suggesting a large-scale duplication of an ancestral Hox region, followed by dispersion in various chromosomes.

Drosophila homeotic and segmentation genes belonging to the bithoraxand <u>Antennapedia</u>-complex have separate promoters and often different orientation. The human HOX-3 region we have studied seems to exhibit a different transcriptional organization. cDNA clones encoding proteins with different homeodomains share a common 5' noncoding exon. We have reported (40) preliminary characterization of a HOX-3 cDNA clone, cpl1, encoding a protein with a third homeodomain, namely the cpll homeodomain, and sharing the same 5' noncoding exon. Therefore, the whole genomic region from the common 5' exon to the last cp19 exon appears to be transcribed in a single primary transcript, alternatively spliced to give the mature mRNAs from which the cDNA clones derive. Primer extension experiments (not shown) and an estimate of transcript length from Northern blot analysis suggest that one or more short exons may in turn precede the common 5' exon. The finding that at least in one tissue the three homeoboxes belong to the same transcription unit and do not represent three separate genes is bound to influence considerably our view on homeobox genes. For Drosophila homeotic and segmentation genes nothing similar has been reported so far. Severa1 different <u>Ultrabithorax</u> (<u>Ubx</u>) proteins have been reported (41) to originate from alternative splicing. Different messengers with common 5' and 3' exons differ from each other for three internal short exonic sequences A similar, albeit simpler, organization may be alternatively spliced. present in Antennapedia (Antp) transcripts (41). In all these instances the different protein products contain the same homeodomain. In human placenta different proteins might be encoded in alternative mature transcripts obtained by means of an alternate use of homeobox regions. This should limit the number of different human homeobox genes, while increasing the complexity of their transcriptional organization. We are not yet in a position to assess how general this phenomenon is and, in particular, how many HOX-3 homeoboxes belong to the same transcription unit.

An alternative explanation for the origin of the cDNA clones studied

can be considered. In fact, exons specific for the three cDNA clones could be transcribed from three different promoters and the common 5' exon could be spliced post-transcriptionally with a trans-splicing mechanism as that found operating in trypanosomes (42). Although for the time being the explanation cannot be ruled out, we think this unlikely. First, the genomic region corresponding to the common 5' exon lies in cis, upstream from the 3' exons. Second, nuclear run-on transcription assays (43) show that the whole region is transcribed (not shown). Finally, we found several alternative cDNA clones containing various exons between the 5' and 3' exons. T+ appears likely that a considerable proportion of the genomic region under scrutiny will turn out to be present in mature transcripts. Northern blot analysis supports this picture and suggests that clone cp25 represents a placental messenger 2.2 kb long, that clone cp19 derives from a transcript of 2.8 kb and clone cp8 from a transcript of 3.5 kb. Additional placental mature transcripts map in this region and await a characterization.

Because we found this transcriptional organization in placental tissues, it seemed appropriate to wonder whether the observed mature messengers are predominant in these tissues or specific for them. Northern blot analysis of embryonic tissues reveals common mature transcripts and some differences. In particular, the embryonic spinal cord shows a complex expression pattern including all transcripts detected in placenta (not shown).

In summary, analysis of cDNA clones and polyadenylated RNAs from the HOX-3 locus suggests that transcription might start from a major promoter upstream from homeodomain-containing exons. Several polyadenylation signals are recognized giving rise to a number of polyadenylated RNAs, in turn spliced to generate mature messengers RNAs. We do not know whether this organization is unique to the HOX-3 locus or can be generalized to other homeobox loci. This transcription organization could readily account for the same 5'-3' orientation of all mammalian homeoboxes isolated so far and for the unique degree of conservation of the whole homeobox chromosomal region in mice and men (14). The presence of a major upstream promoter does not exclude that additional specific promoters in front of single homeobox regions exist in placenta or other tissues. Transcription from the upstream promoter might provide the substrate for a concerted fine tuning of the expression of homeobox messengers whereas transcription from specific secondary promoters might serve specific purposes.

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