

# The benefit of being a social butterfly: communal roosting deters predation

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Aposematic passion-vine butterflies from the genus Heliconius form communal roosts on a nightly basis. This behaviour has been hypothesized to be beneficial in terms of information sharing and/or antipredator defence. To better understand the adaptive value of communal roosting, we tested these two hypotheses in field studies. The information-sharing hypothesis was addressed by examining following behaviour of butterflies departing from natural roosts. We found no evidence of roost mates following one another to resources, thus providing no support for this hypothesis. The anti-predator defence hypothesis was tested using avian-indiscriminable Heliconius erato models placed singly and in aggregations at field sites. A significantly higher number of predation attempts were observed on solitary models versus aggregations of models. This relationship between aggregation size and attack rate suggests that communally roosting butterflies enjoy the benefits of both overall decreased attack frequency as well as a prey dilution effect. Communal roosts probably deter predators through collective aposematism in which aggregations of conspicuous, unpalatable prey communicate a more effective repel signal to predators. On the basis of our results, we propose that predation by birds is a key selective pressure maintaining Heliconius communal roosting behaviour.

**Keywords:** communal roosting; collective aposematism; dilution effect; aggregation; predation; Heliconius

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#### 1. INTRODUCTION

In 1867, the naturalist J. A. Allen first described the spectacular aggregations of migratory monarchs, and was thus the first to report the phenomenon of communal roosting in butterflies [1,2]. Shortly thereafter W. H. Edwards, the entomologist who inspired Bates and Wallace to visit the Amazon, reported communal roosting in the tropical Heliconius passion-vine butterflies [3]. Since the mid-nineteenth century, this unusual behaviour has generated a great deal of scientific and popular interest. After 140 years of work on butterfly roosting, however, it still remains unclear what the benefit of being a social butterfly is. Here, we test the major hypotheses for why Heliconius butterflies roost communally, and present experimental data assessing the adaptive function of this behaviour.

Communal roosting is observed in many types of animals, including birds, bats and primates [4-7], and is especially widespread in insects, having been observed in bees, wasps, beetles, dragonflies, butterflies and moths [8-10]. In butterflies, communal roosting is described as repeatedly gather in a particular location in their home range to roost for the night (figure 1a). Butterflies arrive at their roost sites as early as 3 h before sunset, and depart from roosts within the first 2 h after sunrise. Roost mates are generally conspecifics, but sometimes different

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species—often Müllerian co-mimics—roost together [13]. The evolution of communal roosting behaviour in *Heliconius* is believed to be facilitated by unpalatability, slow reproductive rate [14], limited learned home range [12,15] and long lifespan owing to pollen consumption [16,17].

Heliconius butterflies are well known for their brightly coloured wing patterns and their unpalatability owing to cyanogenic glycosides [18]. Because of these features, Heliconius butterflies serve as a textbook example of warning signalling—also known as aposematism. Aposematism is a major theme in the evolution of animal phenotypes, where its principal function is to provide warning signals associated with unprofitability to predators, such as toxicity, unpalatability or capture costs [19-21]. Aposematism is widespread in invertebrates and is often achieved through visual signalling via conspicuous colour patterns. Collective aposematism is a phenomenon in which aposematic prey form aggregations to enhance the effects of warning signals [22,23]. Despite the substantial amount that is known about Heliconius natural history, little is known about their communal roosting behaviour and its possible relationship to collective aposematism.

[12,13,24-29]; however, relatively few experimental studies have been performed to address the function of this behaviour. Although the adaptive consequences of roosting remain unclear, it is unlikely that aggregations are involved with thermoregulation [10], kin selection [13] or mating (females usually mate once in their lifetime, within hours or days after eclosion) [30,31]. The favoured explanations have been narrowed to two major hypotheses: information sharing and/or anti-predator defence. The information-sharing hypothesis proposes that new roost mates, presumably related individuals, follow experienced

There is a broad literature on *Heliconius* roosting

<sup>&#</sup>x27;a behavior in which individuals aggregate quiescently in close proximity to each other at a site for more than a few hours' ([9] p. 90). This behaviour is known primarily from the heliconiines, acraeines, ithomiines and danaiines [11,12]. Many species of Heliconius in particular have been observed to form communal roosts in which adults

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Figure 1. (a) Natural and (b) artificial roosts of *Heliconius erato*. (c) Beak pinch at end of model abdomen, and triangular beak mark imprint on wax wing (indicated by arrow). (d) Dorsal view of abdominal beak pinch.

members from the roost to food sources [27,32]. This form of information sharing is a common behaviour in other communal animals such as birds [33,34]. Conversely, the anti-predator hypothesis suggests predators avoid *Heliconius* aggregations as a result of collective aposematism or predator confusion [12,13,35]. In other aposematic insects, gregarious behaviour contributes to collective enhancement of warning signalling, resulting in more effective predator deterrence [22,36–38]. Another potential anti-predator mechanism is the prey dilution effect, often known as 'safety in numbers', which posits that the probability of a single individual being attacked in a group is lower with increasing density [39–42].

In field studies in Panama and Costa Rica, we tested these hypotheses to determine why *Heliconius* passionvine butterflies assemble in communal roosts. To test whether *Heliconius* butterflies rely on roosts as information-sharing centres, we examined following behaviour by butterflies during departures from natural aggregations. The anti-predator defence hypothesis was tested using avian-indiscriminable artificial butterfly models placed singly and in aggregations in the forest. Following the predation study, we investigated whether naturally occurring roost sizes correspond with optimal roost sizes inferred from experimental data.

#### 2. MATERIAL AND METHODS

#### (a) Field sites

All data collection was completed in Costa Rica and Panama. Field sites for this work were chosen based on the abundance

and accessibility of *Heliconius* butterflies and communal roosts. The Panama sites were part of the Smithsonian Tropical Research Institute; we used areas in Gamboa and Soberanía National Park along Pipeline Road. Data were collected in Panama from June through September of 2010 and 2011 during the rainy season. The 2010 visit resulted in natural roost data collected from *Heliconius erato*, and the 2011 visit resulted in the predation experiment data. In Costa Rica, we worked at the Organization for Tropical Studies' La Selva Tropical Biological Station in Sarapiquí. La Selva was visited in April and May of 2011, during the end of the dry season into the beginning of the rainy season. This site was used for collecting natural roost data from *Heliconius sara* and model predation experiments.

## (b) Natural roost observations

To assess the information-sharing hypothesis, which predicts that butterflies use roosts to learn the locations of foraging sites from other roost mates, we observed following behaviour of *H. erato* and *H. sara* individuals departing natural roosting aggregations. Following was confirmed only if a butterfly was observed departing the roost with another roost mate to subsequently arrive at a flowering plant with that same roost mate. The roosts were within 30 m of the first visited flowering plant, and it was feasible to follow butterflies to these plants. We began all observations approximately 30 min before sunrise, before butterflies left to forage, and stayed until only one individual remained. All *H. erato* roost members were given unique identification numbers using a Sharpie marker, and were sexed and age determined based on wing wear [43]. Butterflies were captured and marked after departing

their roosts to prevent them from associating the roost with danger [44]. Average roost sizes were determined by recording the number of individuals in each roost nightly.

#### (c) Models

We used artificial butterfly models to test the hypothesis that Heliconius roosts provide an anti-predatory benefit. Wing images were designed in Adobe Illustrator using highresolution scans of ventral Heliconius erato petiverana wings as a reference. Model butterfly wings were printed on Whatman filter paper, which produces reflectance spectra close in brightness to actual wings, using an Epson Stylus Pro 4880 printer with UltraChrome K3 ink. A 3-hydroxy-DLkynurenine (3-OHK) pigment solution of 1.0 mg 3-OHK dissolved in 100 µl of methanol was applied to the yellow bands on the hindwing to provide accurate UV reflectance, because printed yellows do not accurately mimic Heliconius yellow in the shape of their reflectance spectra. 3-OHK is the same yellow wing pigment as found in the wings of the butterflies themselves [45,46]. Portions of the artificial wings were dipped in clear wax to allow imprints of beak and bite marks, then Krylon matte finishing spray was applied lightly (before spectra measurements were taken) to coat the 3-OHK with a waterproofing element. Model abdomens were made of Newplast Plasticine.

Birds, in particular jacamars, flycatchers and tanagers, are major Heliconius predators [27,36,47-50]. Therefore, artificial butterfly models were designed and assessed using tetrachromatic bird colour-vision models in order to ensure that avian predators would find colour stimuli presented by the models indiscriminable from actual butterflies. Reflectance spectra of yellow, pink and black from the models and the ventral surface of natural H. erato petiverana wings were measured using an Ocean Optics USB2000 fibre optic spectrometer. A deuterium-halogen tungsten lamp (DH-2000, Ocean Optics) was used as a standardized light source, and measurements were taken using a bifurcating fibre cable (R400-7-UV-vis, Ocean Optics). The axis of the illuminating and detecting fibre was at an elevation of 45° to the plane of the wing and pointed left with respect to the body axis for every measurement. A white spectralon standard (WS-1-SL, Labsphere) was used to calibrate the spectrometer. Spectra measurements from the fibre optic spectrometer were processed using MATLAB software (see Briscoe et al. [46]). The quantum catches for stimuli [51] were calculated, and discriminability between artificial models and natural wing reflectance spectra was determined using tetrachromatic bird-vision models from Vorobyev & Osorio [52]. The comparisons were made using the blue tit (Parus caeruleus) and chicken (Gallus gallus) cone sensitivities, which represent the UV- and violet-type avian visual systems, respectively. Low light intensity and open habitat irradiance spectra were used [53]. All spectral comparisons represented by an average of wing measurements (n = 12) fell below the threshold of one just noticeable difference (figure 2 and table 1); therefore, the reflectance spectra of models and actual butterfly wings were inferred to be indiscriminable to birds.

### (d) Predation experiments

Butterfly models were tied to branches with thread in appropriate roosting habitats and in natural roosting postures [10,13] (figure 1b). At our Costa Rica field site, a total of 320 aggregations containing five butterflies each and 320

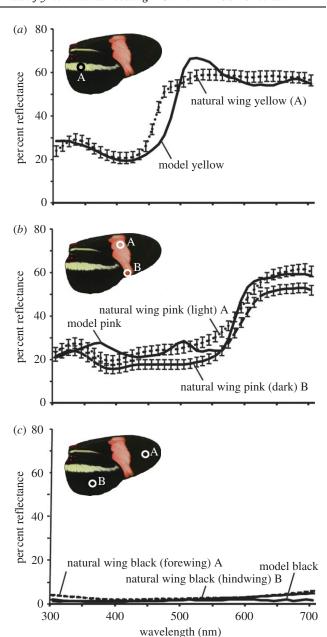


Figure 2. Reflectance spectra of natural and artificial butterfly wings used in discriminability calculations. Shown are the mean values and standard errors (each n = 12). Graphs include a ventral image of Heliconius erato petiverana with circled regions indicating where wing reflectance measurements were taken. (a) Yellow reflectance spectra. (b) Pink reflectance spectra, comparisons made using lightest A and darkest B pink regions on the wing. (c) Black reflectance spectra, comparisons made using both forewing A and hindwing B regions.

single butterflies were used for the first predation experiment. The models were placed in 80 different forest sites, each containing four roosts and four single individuals. All 80 sites were at least 250 m apart to control for the home range territories of primary Heliconius avian predators. Predator home range sizes vary between 100 and 250 m, and have been determined by other researchers through radio tracking (flycatchers [54]), harmonic distance method and core area use (tanagers [55]), minimum convex polygon modelling (flycatchers [54]) and observation (tanagers [56]; jacamars [57]; C. E. G. Pinheiro 2011, personal communication; L. E. Gilbert 2011, personal communication). Tree Tanglefoot

Table 1. Results from discriminability calculations using low light intensity and open habitat irradiance. Comparisons were made using the blue tit (*Parus caeruleus*) and chicken (*Gallus gallus*) cone sensitivities, and are based on mean values (n = 12). Spectra comparisons fell below the threshold of one just noticeable difference (JND). FV, forewing ventral; HV, hindwing ventral; L, light; D, dark.

| JND comparisons | FV-L pink versus model pink | FV-D pink versus model pink | HV yellow versus model yellow | FV black versus model black | HV black versus model black |
|-----------------|-----------------------------|-----------------------------|-------------------------------|-----------------------------|-----------------------------|
| P. caeruleus    | 0.597                       | 0.374                       | 0.736                         | 0.336                       | 0.440                       |
| G. gallus       | 0.556                       | 0.424                       | 0.858                         | 0.315                       | 0.533                       |

was applied to the base of plant stems containing artificial butterflies to avoid removal or attack of the models by ants and other small arthropods, and it was also effective in preventing small vertebrates such as lizards from reaching the butterfly models [58].

The models were left at their sites for a total of 96 h (4 days), and each model was examined daily for predation evidence and replaced if attacked. None of the 80 sites were used twice in the study because predator forgetting time varies between bird species, and is affected by prey conspicuousness and distastefulness [59-61], both difficult to measure for this project. A butterfly was considered attacked if damage to the abdomen and wings appeared in the form of beak marks and/or large indentations in the abdomen (figure 1c,d; see also [62,63]). Small chew-like marks, probably from mandibular insects such as grasshoppers, were not considered in the data analyses. If a model was attacked twice on two separate days, then it was counted only as a single attack. If multiple butterflies in a roost were attacked, this was also counted as a single attack (i.e. each roost was treated as a unit), because in nature when a roost is disturbed most or all individuals depart from the roosting site [13,29] (S. D. Finkbeiner 2010, personal observation), thereby reducing the probability of further predation attempts on individual roost mates. Predation differences were analysed using a Wilcoxon signed-rank test with continuity correction, using each site as a sample.

To assess the *per capita* attack risk of individual butterflies, single (focal) individuals were randomly selected from each roost to compare attack rates under the conservative assumption that one attack leads to the dispersal of the other roosting butterflies. For roosts of one, this single individual was the focal individual. The binomial response of attack (yes, no) was modelled as dependent upon roost size using generalized linear models. In these analyses, site was included as a random effect to account for potential non-independence among replicate roosts within sites.

As a control for our models, we compared the attack rate on models with real wings (and Plasticine abdomens) to the attack rate on models with artificial wings, using five forest sites separated 250 m apart, each with four models with real wings and four models with artificial wings. They were left for 96 h and checked daily for attacks. We found no difference in attacks between real-wing models and artificial-wing models (Wilcoxon signed-rank test: W = 249.5, p = 0.836, n = 20).

The second predation study, conducted in Panama, investigated the association between roost size and predation frequency. This was tested using the same artificial butterfly models described earlier. One hundred forest sites were chosen, approximately 250 m apart, and each of these sites contained two roosts of 2, two roosts of 5 and two roosts of 10 butterflies. This totalled 600 artificial roosts: 200 roosts

for each treatment. Roosts were removed after 4 days, each of the 100 sites was used only once and attacked models were replaced when necessary. If a roost was attacked more than once or if more than one butterfly in the roost was attacked, then it was counted only as one attack; thus each roost was considered as a single unit in the analysis. These predation data were analysed using a Kruskal-Wallis multiple comparison test with a post hoc Bonferroni correction, using each site as a sample. Individual per capita attack risk was determined by comparing attack rates between randomly selected single (focal) individuals in each roost treatment, and the binomial response of attack was modelled as dependent upon roost size using generalized linear models, with site included as a random effect. The post hoc tests for the significance of pairwise comparisons were made using a Tukey test.

We determined at what time of day roosts are most susceptible to attack to gain further information about the roost predators. This study was conducted with 100 artificial roosts in Panama: 52 roosts of two placed randomly along two trails in Soberanía National Park, with the remaining 48 roosts already being observed for predation data (16 more roosts of 2, 5 and 10 each). The artificial roosts were checked for attacks every 3 h from 06.00 (15 min prior to sunrise) to 18.00 (just before sunset) for 9 days.

#### 3. RESULTS

# (a) Observations of roost departure following

To assess whether Heliconius butterflies rely on communal roosts as information-sharing centres, we observed following behaviour during morning roost departures. Of 256 H. erato departures by at least 66 unique individuals from nine different roosts, only one instance of following from the roost to a flowering plant was observed (Wilcoxon signed-rank test: W = 256, p < 0.0001, n =256, based on the null hypothesis that there is no difference between the number of butterflies that do and do not follow). Additionally, out of 74 H. sara butterfly departures from at least 25 unique individuals and three different roosts, we observed no incidence of following. When butterflies departed roosts in the morning occasionally more than one individual would leave at the same time, but they were never observed to follow one another. Most of the time the butterflies left individually, even when there was a disturbance event.

Lantana and Psychotria plants, considered to be key Heliconius resources, were common at our study sites, and it is important to note that spatial distribution and density of nectar and pollen sources may influence whether following happens. We observed that many roosting butterflies shared the same flower resources and followed each other

Table 2. Data representing attacks on butterfly models at field sites in Costa Rica and Panama, categorized by aggregation size. Roost attack risk was calculated by dividing attacked roosts by the overall number of roosts used in that treatment. Individual attack risk was determined by comparing attack rates between randomly selected single (focal) individuals in each roost treatment, based on the assumption that one attack leads to the dispersal of the other roosting butterflies. Probability values between pairwise comparisons are indicated by asterisks.

| aggregation size                                       | total observations | roosts attacked       | attack risk (per roost, %) | attack risk (per individual, %) |
|--|--------------------|-----------------------|----------------------------|---------------------------------|
| Costa Rica<br>single individuals<br>roost of five      | 320<br>320         | 68<br>25              | 21.3<br>7.8                | 21.3<br>3.4 <sup>*</sup> ***    |
| Panama<br>roost of two<br>roost of five<br>roost of 10 | 200<br>200<br>200  | 21\}*<br>*{8\}*<br>24 | 10.5<br>4.0<br>12.0        | 9.5<br>1.5<br>4.5               |

<sup>\*</sup>p < 0.05.

between flowering plants, as previously described by Waller & Gilbert [27]. As well, on multiple occasions, we observed a new recruit following an established roost member to the aggregation, suggesting following behaviour may play a role in roost recruitment.

#### (b) Observations on roost size and proximity

The average H. erato roost size was observed to be 4.3 individuals (s.d. = 1.6, n = 233 observations across nine natural roosts) from roosts in Gamboa, Panama. It was common to find multiple roosts within 15 m of each other, some as close as 3 m apart, and in line-of-sight from one another in a given part of the home range. We observed this in both H. erato and H. sara. When butterflies of *H. erato* were exercising pre-roosting behaviour, the butterflies often interacted with one another before convening at their preferred roosts.

# (c) Effect of roost size on predation frequency

To determine whether communal roosts provide an anti-predatory benefit, we first tested whether there is a difference in predation between single butterfly models or models placed in aggregations of five. We observed a very strong difference in attack frequencies between roosting and solitary butterflies. Of 320 artificial H. erato roosts, only 25 were attacked compared with 68 attacks on 320 single individuals (Wilcoxon signed-rank test: W = 4141.5, p = 0.000262, n = 80 sites; table 2). Individual attack risk also differed significantly between roosts of 1 (single butterflies) and roosts of 5 ( $F_{1,559}$  = 36.85, p < 0.0001, n = 640 roosts total), with 21.3 per cent of focal individuals being attacked in roosts of 1 but only 3.4 per cent of focal individuals being attacked in roosts of 5 (table 2).

Considering the difference we found in predation between single butterflies and roosts, we decided to investigate whether the predator deterrence effect was similar across roosts of different sizes (table 2). We found significantly higher predation on roosts of 2 and 10 than on roosts of 5 (Kruskal–Wallis test:  $\chi^2 = 8.7356$ , p =0.0127, n = 100 sites). The post hoc Bonferroni correction showed that roosts of 10 were attacked more than roosts of 5 (p = 0.016), and roosts of 2 were attacked more than roosts of 5 (p = 0.028). There was no significant difference in predation frequency between roosts of 2 and 10 (p = 1.000). Individual attack rate also differed significantly between the three roost sizes ( $F_{2,498} = 5.51$ , p = 0.0043, n = 600 roosts total), with attack rates on focal individuals in roosts of 2, 5 and 10 being 9.5 per cent, 1.5 per cent and 4.5 per cent, respectively (table 2). Adjusting significance levels to account for multiple comparisons, Tukey tests showed that a single butterfly in a roost of 2 has a higher attack risk than a single butterfly in a roost of 5 (p = 0.00064), but there is no difference in individual attack risk between roosts of 5 and 10 butterflies (p = 0.22), or between roosts of 2 and 10 butterflies (p = 0.13).

# (d) Timing and nature of predation

We sought to determine at which time of day roosts are most susceptible to attack. We found that all attacks occurred during the morning hours, as previously observed by Mallet [13]. Ten out of 12 attacks, from 100 different artificial roosts, occurred between the hours of 06.00 and 09.00, and two attacks occurred between the hours of 09.00 and 12.00. Triangular beak marks observed in the model abdomens supports previous findings [48-50] that birds are the primary predators of Heliconius. This is further supported because all attacks on models occurred between the hours of 06.00 and 12.00 when birds are most active [56,64].

### 4. DISCUSSION

In this study, we assessed the two major hypotheses for explaining why Heliconius butterflies participate in communal roosting behaviour. We found no support for the information-sharing hypothesis, because there was little evidence of roost mates following each other to resources upon departure from roosts. These findings are in agreement with Mallet [13], who observed a similar lack of following and even a predictable tendency for roost mates to visit different flowers.

In contrast, we found very strong support for the antipredator defence hypothesis. Our field experiments in Costa Rica using avian-indiscriminable butterfly models showed predation attempts on singly placed models were nearly three times higher than predation attempts on roosts of five models, and the predation risk for a

<sup>\*\*</sup>p < 0.001.

<sup>\*\*\*</sup>p < 0.0005.

single butterfly is over six times the per capita predation risk for an individual butterfly in a roost of 5 (table 2). A second field experiment in Panama showed the same trend, with attack rates more than twice as high on roosts of 2 versus 5, and individual risk over six times higher in a roost of 2 than a roost of 5 (table 2). Surprisingly, however, the Panama experiment also showed that attacks on roosts of 10 were three times as high as on roosts of 5 (table 2), thus suggesting the predator deterrence effect may be weak or non-existent in large aggregations. In the Panama experiment, the greatest individual fitness benefit was seen in roosts of 5, with an individual predation risk of 1.5 per cent; however, individuals in roosts of 10 benefited only slightly less (not significant) than those in roosts of 5 (individual risk of 4.5% in roosts of 10). Therefore, even though roosts of 10 did not enjoy a significantly decreased predation rate compared with roosts of 2 or 5, a simple prey dilution effect [39] would still favour large roost sizes.

Our field studies suggest that the most beneficial minimum roost size, with respect to group advantage, may be around 5 individuals. This is because our experimental aggregations of five models experienced the lowest overall attack rates and also offered the lowest per capita attack risk for individuals. Interestingly, this experimentally determined minimum roost size corresponds closely to naturally observed *H. erato* roost sizes (4.3). This correspondence implies that predator deterrence, coupled with a prey dilution effect, could help explain roost sizes observed in natural populations. An optimal or minimum roost size may be important for predator deterrence when butterfly densities are too low for assembling in larger aggregations; however, roost sizes are probably influenced by foraging and resource availability as well.

Because medium-sized roosts provide an anti-predatory benefit through collective aposematism, it is unclear why larger aggregations appear to lose their ability to deter predators. It is possible that solitary individuals or very small roosts are too small to communicate an effective warning signal, whereas very large roosts may be conspicuous enough to attract naïve predators. In support of this idea, Salcedo [29] noted that the most frequent predator disturbances on *H. sara* roosts occurred on the largest aggregation studied (10-16 individuals), suggesting that oversized aggregations of Heliconius may increase the frequency of predator attacks. Although there is much evidence that predator wariness and aggregation-induced phobias increase with the size of aposematic prey aggregations [61], greater aggregation distinctiveness increases detectability costs and, in some cases, larger aggregation sizes of defended animals result in higher predation [23,65]. There may be other costs to forming larger groups such that small movements made by other butterflies could produce incidental disturbances or 'false alarms', causing unexpected or premature roost departures, but this would not explain higher predation on larger roosts from our experimental data.

We propose that it is no coincidence that multiple roosts are found in the same location in a home range and often in line-of-sight from one another. We have observed this in *H. erato*, *H. sara*, *H. melpomene* and *H. charithonia*, with some roosts neighbouring the roosts of heterospecifics (in Costa Rica). Others have observed this *Heliconius* behaviour of preferentially forming smaller

aggregations as well [13,66] (C. Boggs 2011, personal communication). Salcedo [29] proposes these may be early-stage aggregations made up of individuals who have not yet located a larger roost. In contrast, however, our field observations of pre-roosting interactions between butterflies from different roosts suggest that the butterflies are aware of other roosts in their home range, yet still choose to join smaller aggregations, despite plenty of substrate (i.e. dry branches and twigs) to sustain much larger aggregations. On the basis of the assumption that individual fitness should increase with larger aggregations owing to a reduced per capita predation risk, again it is unclear why local butterflies do not simply choose to form very large roosts. There is the possibility that an interaction among roosts is introduced, so that if individuals in one group are attacked the predator is inhibited from attacking individuals in other groups [42]. This repeated warning display could therefore facilitate rapid learning in naïve birds whose feeding areas may include multiple Heliconius roosting sites [67,68]. It is also possible that very large roosts could attract high enough levels of predation to cancel out the prey dilution benefit, although experimental work is required to test this idea.

Here, we show that *Heliconius* butterflies do not rely on communal roosts as information-sharing centres. Instead, our field studies indicate that this behaviour confers an anti-predatory benefit at both the individual level (prey dilution) and group level (collective aposematism), and attack risks vary between individuals in different aggregation sizes. This correspondence suggests that predation may be the key selective pressure maintaining communal roosting in *Heliconius*, thus providing insight into the types of ecological pressures that contribute to the evolution of social behaviour in historically solitary animals.

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## **REFERENCES**

- 1 Scudder, S. H. & Allen, J. A. 1869 A preliminary list of the butterflies of Iowa. *Trans. Chicago Acad. Sci.* 1, 326–337.
- 2 Brower, L. P. 1995 Understanding and misunderstanding the migration of the monarch butterfly (Nymphalidae) in North America: 1857–1995. J. Lepid. Soc. 49, 304–385.

- 3 Edwards, W. H. 1881 On certain habits of *Heliconia charitonia* Linn., a species of butterfly found in Florida. *Papilio* 1, 209–215.
- 4 Ward, P. 1965 Feeding ecology of the black-faced dioch *Quellea quellea* in Nigeria. *Ibis* **107**, 173–214. (doi:10. 1111/j.1474-919X.1965.tb07296.x)
- 5 Soini, P. 1987 Ecology of the saddle-back tamarin Saguinus fuscicollis-illigeri on the Rio Pacaya, northeastern Peru. Folia Primatol. 49, 11–32. (doi:10.1159/000156305)
- 6 Lewis, S. E. 1995 Roost fidelity of bats: a review. *J. Mammal.* **76**, 481–496. (doi:10.2307/1382357)
- 7 Merkel, F. R. & Mosbech, A. 2008 Diurnal and nocturnal feeding strategies in common eiders. *Waterbirds* **31**, 580–586. (doi:10.1675/1524-4695-31.4.580)
- 8 Pearson, D. L. & Anderson, J. J. 1985 Perching heights and nocturnal communal roosts of some tiger beetles (Coleoptera: Cicindelidae) in southeastern Peru. *Biotropica* 17, 126–129. (doi:10.2307/2388504)
- 9 DeVries, P. J., Schull, J. & Greig, N. 1987 Synchronous nocturnal activity and gregarious roosting in the neotropical skipper butterfly *Celaenorrhinus fritzgaertneri* (Lepidoptera: Hesperiidae). *Biol. J. Linn. Soc.* **89**, 89–103. (doi:10.1111/j.1096-3642.1987.tb01345.x)
- 10 Salcedo, C. 2010 Environmental elements involved in communal roosting in *Heliconius* butterflies (Lepidoptera: Nymphalidae). *Environ. Entom.* 39, 907–911. (doi:10. 1603/EN09340)
- 11 Benson, W. W. & Emmel, T. C. 1973 Demography of gregariously roosting populations of the nymphaline butterfly *Marpesia berania* in Costa Rica. *Ecology* 54, 326–335. (doi:10.2307/1934340)
- 12 Turner, J. R. G. 1975 Communal roosting in relation to warning colour in two heliconiine butterflies (Nymphalidae). *J. Lepid. Soc.* 29, 221–226.
- 13 Mallet, J. 1986 Gregarious roosting and home range in *Heliconius* butterflies. *Natl. Geogr. Res.* 2, 198–215.
- 14 Erlich, P. R. & Gilbert, L. E. 1973 Population structure and dynamics of the tropical butterfly *Heliconius ethilla*. *Biotropica* 5, 69–82. (doi:10.2307/2989656)
- 15 Mallet, J. 1986 Dispersal and gene flow in a butterfly with home range behavior: *Heliconius erato* (Lepidoptera: Nymphalidae). *Oecologia* 68, 210–217. (doi:10.1007/ BF00384789)
- 16 Gilbert, L. E. 1972 Pollen feeding and reproductive biology of *Heliconius* butterflies. *Proc. Natl Acad. Sci. USA* **69**, 1403–1407. (doi:10.1073/pnas.69.6.1403)
- 17 Boggs, C. L., Smiley, J. T. & Gilbert, L. E. 1981 Patterns of pollen exploitation by *Heliconius* butterflies. *Oecologia* **48**, 284–289. (doi:10.1007/BF00347978)
- 18 Engler-Chaouat, H. S. & Gilbert, L. E. 2007 *De novo* synthesis versus sequestration: negatively correlated metabolic traits and the evolution of host plant specialization in cyanogenic butterflies. *J. Chem. Ecol.* 33, 25–42. (doi:10.1007/s10886-006-9207-8)
- 19 Cott, H. B. 1957 Adaptive coloration in animals. London, UK: Methuen.
- 20 Guilford, T. 1990 The evolution of aposematism. In Insect defenses: adaptive mechanisms and strategies of prey and predators (eds D. L. Evans & J. O. Schmidt), pp. 23-61. Albany, NY: State University of New York Press.
- 21 Santos, J. C., Coloma, L. A. & Cannatella, D. C. 2003 Multiple, recurring origins of aposematism and diet specialization in poison frogs. *Proc. Natl Acad. Sci. USA* **100**, 12 792–12 797. (doi:10.1073/pnas.2133521100)
- 22 Gamberale, G. & Tullberg, B. S. 1998 Aposematism and gregariousness: the combined effect of group size and coloration on signal repellence. *Proc. R. Soc. Lond. B* **265**, 889–894. (doi:10.1098/rspb.1998.0374)
- 23 Riipi, M., Alatalo, R. V., Lindström, L. & Mappes, J. 2001 Multiple benefits of gregariousnes cover detectability costs

- in aposematic aggregations. *Nature* **413**, 512–514. (doi:10.1038/35097061)
- 24 Jones, F. M. 1930 The sleeping heliconias of Florida. *Nat. Hist.* **30**, 635-644.
- 25 Young, A. M. & Thomason, J. H. 1975 Notes on communal roosting of *Heliconius charitonius* (Nymphalidae) in Costa Rica. J. Lepid. Soc. 29, 243-255.
- 26 Mallet, J. L. B. 1980 A laboratory study of gregarious roosting in the butterfly *Heliconius melpomene*. MSc thesis, University of Newcastle-upon-Tyne, Newcastleupon-Tyne, UK.
- 27 Waller, D. A. & Gilbert, L. E. 1982 Roost recruitment and resource utilization: observations on a *Heliconius charitonia* L. roost in Mexico. *J. Lepid. Soc.* 36, 178–184.
- 28 Mallet, J. & Gilbert, L. E. 1995 Why are there so many mimicry rings? Correlations between habitat, behaviour and mimicry in *Heliconius* butterflies. *Biol. J. Linn. Soc.* 55, 159–180. (doi:10.1111/j.1095-8312.1995.tb01057.x)
- 29 Salcedo, C. 2011 Evidence of predation and disturbance events at *Heliconius* (Insecta: Lepidoptera: Nymphalidae) nocturnal aggregations in Panama and Costa Rica. *J. Nat. Hist.* **45**, 1715–1721. (doi:10.1080/00222933. 2011.559692)
- 30 Gilbert, L. E. 1976 Postmating female odor in *Heliconius* butterflies: male-contributed anti-aphrodisiac. *Science* **193**, 419–420. (doi:10.1126/science.935877)
- 31 Schulz, S., Estrada, C., Yildizhan, S., Boppré, M. & Gilbert, L. E. 2008 An antiaphrodisiac in *Heliconius melpomene* butterflies. *J. Chem. Ecol.* 34, 82–93. (doi:10.1007/s10886-007-9393-z)
- 32 Gilbert, L. E. 1975 Ecological consequences of a coevolved mutualism between butterflies and plants. In *Coevolution of animals and plants* (eds L. E. Gilbert & P. R. Raven), pp. 210–240. Austin, TX: University of Texas Press.
- 33 Ward, P. & Zahavi, A. 1973 The importance of certain assemblages of birds as 'information centers' for food finding. *Ibis* 115, 517–534. (doi:10.1111/j.1474-919X. 1973.tb01990.x)
- 34 Dall, S. R. X. 2002 Can information sharing explain recruitment to food from communal roosts? *Behav. Ecol.* **13**, 42–51. (doi:10.1093/beheco/13.1.42)
- 35 Gillett, S. D., Hogarth, P. J. & Noble, F. E. 1979 The response of predators to varying densities of *Gregaria* locust nymphs. *Anim. Behav.* 27, 592–596. (doi:10. 1016/0003-3472(79)90195-7)
- 36 Benson, W. W. 1971 Evidence for the evolution of unpalatability through kin selection in the Heliconiinae (Lepidoptera). *Am. Nat.* **105**, 213–226. (doi:10.1086/282719)
- 37 Edmunds, M. 1974 Defence in animals: a survey of antipredator defences. Harlow, UK: Longman.
- 38 Vulinek, K. 1990 Collective security: aggregation by insects as a defense. In *Insect defenses: adaptive mechanisms and strategies of prey and predators* (eds D. L. Evans & J. O. Schmidt), pp. 251–288. Albany, NY: State University of New York Press.
- 39 Bertram, B. C. R. 1978 Living in groups: predators and prey. In *Behavioural ecology: an evolutionary approach* (eds J. R. Krebs & N. B. Davies), pp. 64–96. Sunderland, MA: Sinauer Associates.
- 40 Hamilton, W. D. 1971 Geometry for the selfish herd. J. Theor. Biol. 31, 295-311. (doi:10.1016/0022-5193 (71)90189-5)
- 41 Turner, G. F. & Pitcher, T. J. 1986 Attack abatement: a model for group protection by combined avoidance and dilution. *Am. Nat.* **128**, 228–240. (doi:10.1086/284556)
- 42 Sillén-Tullberg, B. & Leimar, O. 1988 The evolution of gregariousness in distasteful insects as a defense against predators. *Am. Nat.* 132, 723–734. (doi:10. 1086/284884)

- 43 Karlsson, B. 1987 Variation in egg weight, oviposition rate and reproductive reserves with female age in a natural population of the speckled wood butterfly, *Pararge aegeria*. *Ecol. Entomol.* **12**, 473–476. (doi:10.1111/j. 1365-2311.1987.tb01029.x)
- 44 Mallet, J., Longino, J. T., Murawski, D., Murawski, A. & Simpson de Gamboa, A. 1987 Handling effects in *Heliconius*: where do all the butterflies go? J. Anim. Ecol. 56, 377–386. (doi:10.2307/5054)
- 45 Reed, R. D., McMillan, W. O. & Nagy, L. M. 2008 Gene expression underlying adaptive variation in *Heliconius* wing patterns: non-modular regulation of overlapping *cinnabar* and *vermilion* prepatterns. *Proc. R. Soc. B* 275, 37–45. (doi:10.1098/rspb.2007.1115)
- 46 Briscoe, A. D., Bybee, S. M., Bernard, G. D., Yuan, F., Sison-Mangus, M. P., Reed, R. D., Warren, A. D., Llorente-Bosquets, J. & Chiao, C.-C. 2010 Positive selection of a duplicated UV-sensitive visual pigment coincides with wing pigment evolution in *Heliconius* butterflies. *Proc. Natl Acad Sci. USA* 107, 3628–3633. (doi:10.1073/pnas.0910085107)
- 47 Chai, P. 1986 Field observations and feeding experiments on the responses of rufous-tailed jacamars (*Galbula ruficauda*) to free-flying butterflies in a tropical rainforest. *Biol. J. Linn. Soc.* **29**, 161–189. (doi:10.1111/j.1095-8312.1986.tb01772.x)
- 48 Pinheiro, C. E. G. 2003 Does Müllerian mimicry work in nature? Experiments with butterflies and birds (Tyrannidae). *Biotropica* **35**, 356–364. (doi:10.1111/j.1744-7429. 2003.tb00589.x)
- 49 Langham, G. M. 2004 Specialized avian predators repeatedly attack novel color morphs of *Heliconius* butterflies. *Evolution* **58**, 2783–2787.
- 50 Pinheiro, C. E. G. 2011 On the evolution of warning coloration, Batesian and Müllerian mimicry in Neotropical butterflies: the role of jacamars (Galbulidae) and tyrant-flycatchers (Tyrannidae). *J. Avian Biol.* 42, 277–281. (doi:10.1111/j.1600-048X.2011.05435.x)
- 51 Kelber, A., Vorobyev, M. & Osorio, D. 2003 Animal colour vision: behavioural tests and physiological concepts. *Biol. Rev.* 78, 81–118. (doi:10.1017/S146479 3102005985)
- 52 Vorobyev, M. & Osorio, D. 1998 Receptor noise as a determinant of colour thresholds. *Proc. R. Soc. Lond. B* **265**, 351–358. (doi:10.1098/rspb.1998.0302)
- 53 Bybee, S. M., Yuan, F., Ramstetter, M. D., Llorente-Bousquets, J., Reed, R. D., Osorio, D. & Briscoe, A. D. 2011 UV Photoreceptors and UV-yellow wing pigments in *Heliconius* butterflies allow a color signal to serve both mimicry and intraspecific communication. *Am. Nat.* 179, 38–51. (doi:10.1086/663192)
- 54 Ning, W., Zhang, Y. & Zheng, G. 2007 Home ranges and habitat vegetation characters in breeding season of Narcissus Flycatcher and Yellow-rumped Flycatcher.

- Front. Biol. China 2, 345-350. (doi:10.1007/s11515-007-0051-1)
- 55 Samuel, M. D., Pierce, D. J. & Garton, E. O. 1985 Identifying areas of concentrated use within home range. *J. Anim. Ecol.* 54, 711–719. (doi:10.2307/4373)
- 56 Buskirk, W. H., Powell, G. V. N., Wittenberger, J. F., Buskirk, R. E. & Powell, T. U. 1972 Interspecific bird flocks in tropical highland Panama. *The Auk* 89, 612–624.
- 57 Pinheiro, C. E. G., Bagno, M. A. & Brandão, R. A. 2003 Diet and foraging behavior of the rufous-tailed jacamar (*Galbula ruficauda*, Galbulidae) in central Brazil. *Ararajuba* 11, 241–243.
- 58 Dial, R. & Roughgarden, J. 1995 Experimental removal of insectivores from rain forest canopy: direct and indirect effects. *Ecology* **76**, 1821–1834. (doi:10.2307/1940714)
- 59 Servedio, M. R. 2000 The effects of predator learning, forgetting, and recognition errors on the evolution of warning coloration. *Evolution* **54**, 751–763. (doi:10. 1111/j.0014-3820.2000.tb00077.x)
- 60 Speed, M. P. 2000 Warning signals, receiver psychology and predator memory. *Anim. Behav.* **60**, 269–278. (doi:10.1006/anbe.2000.1430)
- 61 Ruxton, G. D., Sherratt, T. N. & Speed, M. P. 2004 Avoiding attack: the evolutionary ecology of crypsis, warning signals and mimicry. Oxford, UK: Oxford University Press.
- 62 Brodie, E. D. 1993 Differential avoidance of coral snake banded patterns by free-ranging avian predators in Costa Rica. *Evolution* 47, 227–235. (doi:10.2307/2410131)
- 63 Stobbe, N. & Schaefer, H. M. 2008 Enhancement of chromatic contrast increases predation risk for striped butterflies. *Proc. R. Soc. B* **275**, 1535–1541. (doi:10. 1098/rspb.2008.0209)
- 64 DesLauriers, J. V. & Francis, C. M. 1991 The effect of time of day on mist-net captures of passerines on spring migration. *J. Field Ornithol.* **62**, 107–116.
- 65 Lindstedt, C., Huttunen, H., Kakko, M. & Mappes, J. 2011 Disentangling the evolution of weak warning signals: high detection risk and low production costs of chemical defences in gregarious pine sawfly larvae. *Evol. Ecol.* 25, 1029–1046. (doi:10.1007/s10682-010-9456-4)
- 66 Salcedo, C. 2006 Spatial dynamics of night roosting in Heliconius erato petiverana (Lepidoptera: Nymphalidae). MSc thesis, University of Florida, Gainesville, FL.
- 67 Poulton, E. B. 1931 The gregarious sleeping habits of *Heliconius charitonius* L. *Proc. R. Entomol. Soc. Lond.* 6, 4–10.
- 68 Tuskes, P. M. & Brower, L. P. 1978 Overwintering ecology of the monarch butterfly, *Danaus plexippus L.*, in California. *Ecol. Entomol.* 3, 141–153. (doi:10.1111/j. 1365-2311.1978.tb00912.x)