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A Novel 13 Base Pair Insertion in the Sonic Hedgehog ZRS Limb Enhancer (*LMBR1*) Causes Preaxial Polydactyly with Triphalangeal Thumb

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Abstract

Mutations in the *Sonic Hedgehog* limb enhancer, the zone of polarizing activity regulatory sequence (*LMBR1*, commonly called the ZRS), cause limb malformations. In humans, three classes of mutations have been proposed based on the limb phenotype; single base changes throughout the region cause preaxial polydactyly, single base changes at one specific site cause Werner mesomelic syndrome and large duplications cause polysyndactyly. This study presents a novel mutation— a small insertion. In a Swedish family with autosomal dominant preaxial polydactyly, we found a 13 base pair insertion within the ZRS, ZRS603ins13 (NG-009240.1:g.106934_106935ins13). Computational transcription factor binding site predictions suggest that this insertion creates new binding sites and a mouse enhancer assay shows that this insertion causes ectopic gene expression. This study is the first to discover a small insertion in an enhancer

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that causes a human limb malformation and suggests a potential mechanism that could explain the ectopic expression caused by this mutation.

Keywords

enhancer; limb; polydactyly; *SHH*; *LMBR1*; *ZRS*

Preaxial polydactyly is often thought to be the result of problems in development related to patterning of the limb along the anterior-posterior (AP) axis. The AP axis is regulated by cells in a small posterior region of the limb bud called the zone of polarizing activity (ZPA). Cells in the ZPA express Sonic Hedgehog (SHH; MIM# 600725), which controls both digit number and digit identity.

Normal Shh expression level and restriction to the posterior ZPA region is governed by a long range *cis*-regulatory element called the ZPA regulatory sequence (ZRS; MIM# 605522). The ZRS is located about 1 megabase away from *SHH*, within intron 5 of the limb region 1 homolog gene (*LMBR1*; MIM# 605522). This region has a high degree of sequence conservation from humans to fish and controls *Shh* expression in the limb (Lettice, et al., 2003). Mutations in the ZRS have been found to cause preaxial polydactyly in many animals including dogs, cats, chickens and humans. These represent more than 20 different mutations over a region of approximately 2 kilobases. Experiments in mice and chickens that have preaxial polydactyly due to ZRS mutations have shown that these mutations can alter regulatory function, causing ectopic anterior expression of *Shh* and increased posterior mRNA expression levels. Other mutations, not found in mouse models, have been tested in a mouse enhancer assay and shown to cause anterior expression of the reporter gene (reviewed in VanderMeer and Ahituv, 2011), suggesting that anterior ZRS activity is a common mechanism in ZRS mutations.

Since the identification of the ZRS as a long-range enhancer for *SHH*, 13 different point mutations and 10 duplications including the ZRS have been identified in humans with limb malformations (reviewed in VanderMeer and Ahituv, 2011). These malformations represent a wide range of phenotypes. While large duplications including the ZRS and surrounding sequences cause complex Haas-type polysyndactyly (webbing between digits along with extra digits), point mutations in the ZRS cause relatively milder phenotypes including preaxial polydactyly with or without triphalangeal thumbs and Werner mesomelic syndrome.

Here we report a new mutation in the ZRS that causes preaxial polydactyly with triphalangeal thumb in a Swedish family. Previously reported preaxial polydactyly mutations have single base pair (bp) changes, but this mutation is an insertion of 13 bp in the highly conserved ZRS. Computational analyses of ZRS mutations in previous studies suggest that disruption of transcription factor binding sites (TFBS) may be a mechanism for changing AP limb patterning (Albuisson, et al., 2010; Gurnett, et al., 2007). In this case, our computational analysis suggests that rather than losing binding sites, the insertion may change patterning by creating novel TFBS for multiple transcription factors known to be critical for normal limb development. Analysis of the mutated ZRS sequence in a mouse enhancer assay shows that it causes expanded activity from the ZPA to an ectopic anterior limb region.

The proband (Figure 1A, individual III/1) has bilateral symmetrical triphalangeal thumbs with an extra hypoplastic radial thumb containing two phalanges and one metacarpal – preaxial polydactyly type II (PPD2; MIM# 174500) (Figure 1B, C). The triphalangeal thumb

was not opposable and had a finger-like appearance. There was no involvement of the lower limbs in the proband or in any other affected family member. The proband belongs to a large 6 generation family with 78 individuals, 18 of whom are reported to be affected (Supp. Figure S1). DNA samples were available from 3 affected and 3 unaffected individuals in three generations and clinical examinations were performed on the proband's parents and sister (Figure 1A).

We screened a 2.1 kilobase (kb) region that encompasses the ZRS and a nearby region where mutations were shown to cause polydactyly in dogs (pZRS) (Park, et al., 2008) (chr7:156,583,564–156,585,727; UCSC Genome Browser; <http://genome.ucsc.edu>; hg19; g.104811–105583 in reference sequence AC007097.4). All affected individuals were found to be heterozygous for a 13 bp insertion (TAAGGAAGTGATT, Figure 2A) starting at position 603 of the ZRS sequence (Lettice, et al., 2003), position g.106934 within with reference sequence NG_009240.1 for the ZRS, here named ZRS603ins13 (approved variant nomenclature NG_009240.1:g.106934_106935ins13). This mutation has been added to the *LMBR1* mutation database (www.LOVD.nl/LMBR1). None of the three unaffected members tested (Figure 1A; individuals I/2, II/2, III/2) carried this insertion.

To understand how this insertion could affect the expression of *SHH* in the developing limb, we screened the mutant ZRS sequence for TFBS differences from the wildtype human ZRS sequence. TFBS motifs from the UniPROBE database (Newburger and Bulyk, 2009) as well as TFBS compiled from the literature for important limb developmental transcription factors not represented in the database were used for this screen (see Supp. Methods). The insertion creates two major sites that match the binding preferences for several transcription factors, including some that are important in limb development (Figure 2D, Supp. Figure S2). Among the detected TFBS known to be involved in limb development, are *Engrailed1* (*EN1*), *TBX5*, *TBX6*, *SOX8*, and multiple *HOX* genes including *HOXA9*, *HOXB8*, and *HOXC* genes expressed in the developing limb (Nelson, et al., 1996) (Figure 2D). No limb-related TFBS were predicted to be disrupted by the insertion

We next set out to determine the effect of the ZRS603ins13 insertion on the ZPA-specific enhancer function of the ZRS using a transgenic mouse enhancer assay. Wildtype and mutant alleles of the 2.1 kb human ZRS and pZRS region were cloned into the *HSP68-LacZ* enhancer assay vector. This vector has an *Hsp68* minimal promoter (a promoter that is not sufficient to drive reporter expression without the presence of a functional enhancer) followed by the LacZ reporter gene. Transgenic mice carrying these vectors were screened at mouse embryonic (E) day 11.5 – the stage at which previous studies have seen ectopic reporter gene expression from ZRS mutation assays (see Supp. Methods). The wildtype ZRS sequence recapitulates the normal *Shh* expression pattern in E11.5 mouse embryos with expression of the reporter gene in the posterior region of the limb bud in seven out of eight independent transgenic mice (Figure 2B, Supp. Figure S3). With the mutation, four out of six LacZ-positive embryos had limb expression and all four of these embryos showed ectopic anterior expression (Figure 2C, Supp. Figure S3) instead of the normal posterior-restricted ZPA expression pattern (Figure 2B). The expression is limited to the mesoderm of the limb bud and absent from the apical ectoderm ridge (AER) with staining at the anterior and posterior sides. While the four embryos did not show the same intensity of staining (Supp. Figure S3), each embryo had anterior expression that was nearly as strong as the posterior expression. It is worth noting that two of the seven limb-expressing wildtype ZRS embryos also showed a small degree of anterior limb expression that was much weaker than the posterior expression, in contrast to the ZRS603ins13 embryos where the anterior and posterior expression were similar (Supp. Figure S3). The variation between embryos is likely due to integration site or copy number differences between embryos and these differences highlight the qualitative nature of such transgenic assays. The reproducible

anterior expression in ZRS603ins13 embryos is consistent with the enhancer expression pattern observed in similar assays with ZRS mutations that have been shown to cause polydactyly (Furniss, et al., 2008; Lettice, et al., 2003; Lettice, et al., 2008). No other consistent expression patterns were observed in these embryos.

These findings indicate that g.106934_106935ins13 (ZRS603ins13), the small insertion mutation within the ZRS, is likely the cause of preaxial polydactyly and triphalangeal thumb in this family. The mutation is shown to be present in three generations of affected individuals in a fully penetrant inheritance pattern with an invariable phenotype. This inheritance pattern has also been seen with most other ZRS mutations (Albuisson, et al., 2010; Farooq, et al., 2010; Semerci, et al., 2009). However some families with ZRS point mutations have variable phenotypes among affected individuals (Gurnett, et al., 2007; Lettice, et al., 2003) and at least one family is known to have reduced penetrance with phenotypically normal carriers (Gurnett, et al., 2007).

The known human point mutations in the ZRS are dispersed over approximately 600 bp and cause a range of preaxial polydactyly phenotypes (VanderMeer and Ahituv, 2011). There is no clear relationship between the location of the mutation and the severity of the phenotype, but it is thought that the mutations may disrupt TFBS. The ZRS603ins13 mutation results in the creation of two motifs that are predicted to bind multiple transcription factors that are known to be expressed in early limb development including *TBX5*, *TBX6*, *Engrailed1* (*ENI*), *SOX8*, and multiple *HOX* genes. *TBX5* mutations have been shown to cause Holt-Oram syndrome (MIM# 142900), a congenital defect syndrome with limb malformations that include triphalangeal thumbs. *ENI* (MIM# 131290) is a homeodomain-containing transcription factor required for ventral development of the limb and its deletion in mouse models leads to AP limb patterning defects. A consensus binding site for many *HOX* genes that are all expressed at various stages in the developing limb bud (Nelson, et al., 1996) was also found in this insertion. While these transcription factors are not known to directly regulate *SHH* expression in the limb, they are present at the stages in development where abnormal binding to an important regulatory element could stimulate improper expression or interfere with the normal binding of transcription factors that should be regulating *SHH*. Additional limb-associated TFBS created by this mutation that were not detected by our computational analysis could exist and may be related to ZRS activity in the developing limb.

Previous studies have reported bioinformatic analyses suggesting that point mutations in the human ZRS may destroy binding sites for *CDX* (Gurnett, et al., 2007), *MEIS1* and *SOX9* (Albuisson, et al., 2010), but no studies have reported the addition of novel TFBS created by the mutation. Interestingly, an increased affinity for proteins has been seen for two mutated ZRS sequences (Farooq, et al., 2010; Zhao, et al., 2009), though the identity of the particular element(s) that bind preferentially to the mutant sequence was only shown for one case (in this case, HnRNP U; Zhao, et al., 2009). Our study did not find any limb-associated TFBS to be disrupted by the insertion, though it is possible that there are binding sites for known transcription factors that could not be detected by the TFBS prediction program used for this analysis and there are limb-associated transcription factors whose DNA binding motifs are not known.

We also show that this mutation causes the ZRS to drive expression of a reporter gene in the anterior portion of the limb bud where *Shh* is not normally expressed. This type of ectopic anterior expression pattern is consistent with observations in mouse and chicken models of preaxial polydactyly and has also been reported in mouse enhancer assays of human ZRS point mutations that were inserted into a mouse ZRS sequence. Additionally, the *LacZ* reporter expression of the g.106934_106935ins13 mutation appears to be in a larger

posterior region than the normal ZPA expression. While the reporter assay used here is not a quantitative measure of enhancer activity, this increased posterior expression has been reported as a trend in similar enhancer mutation analyses. In addition, mouse and chicken models of preaxial polydactyly have been shown to have increased posterior *Shh* expression due to ZRS mutations (Blanc, et al., 2002; Dunn, et al., 2011). It has been suggested that the degree of ectopic expression in the mouse transgenic reporter assay is related to the severity of the human phenotype caused by the ZRS mutation (Lettice, et al., 2008). The inherent expression differences due to integration site and copy number variability makes this relationship difficult to define, but the ectopic anterior expression from this 13 bp insertion mutation appears to be stronger than what was seen with other ZRS point mutations that cause preaxial polydactyly (Furniss, et al., 2008; Lettice, et al., 2008). The relationship between mutations in the ZRS and the resulting phenotype are complicated and suggest that the regulatory potential of the ZRS is not limited to enhancer activity – driving expression of *Shh* in the posterior limb bud – but also may include some repressor activity to prevent the expression of *Shh* in the anterior limb. The mutation reported in this study and the effect it has in creating novel TFBS suggests that the binding of additional transcription factors and the ablation of TFBS and loss of transcription factor binding may lead to similar outcomes.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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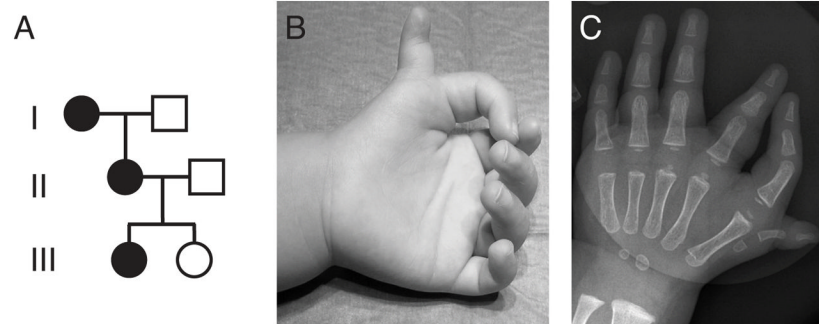


Figure 1. Family with preaxial polydactyly and triphalangeal thumb. A. Pedigree of the family showing subjects available for DNA screening. B,C. Photograph of the right hand and radiograph of the left hand of the proband (patient III/1) showing preaxial polydactyly with triphalangeal thumb.

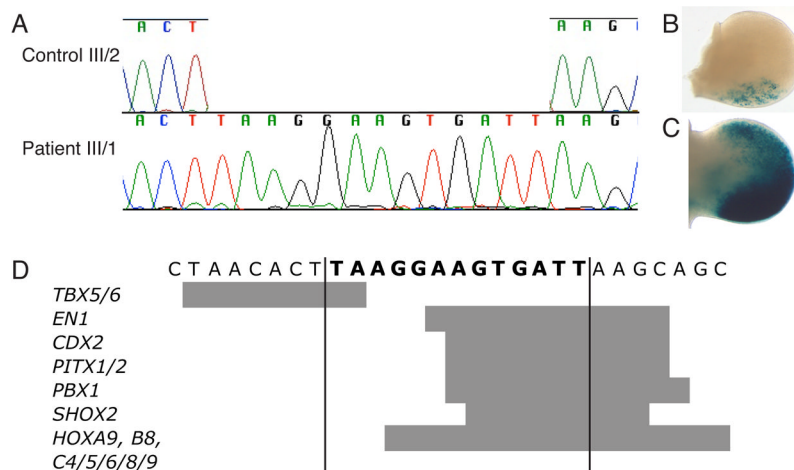


Figure 2. The insertion changes expression of a reporter gene and creates predicted transcription factor binding sites. **A.** Chromatogram of the ZRS603ins13 (NG_009240.1:g.106934_106935ins13) insertion mutation. Sequencing a cloned allele of the ZRS from patient III/1 shows an insertion of 13 bp compared to the wildtype sequence represented by the sequence from an unaffected sibling (III/2). **B.** ZRS E11.5 mouse transgenic assay of human wtZRS showing posterior *LacZ* expression. **C.** ZRS E11.5 mouse transgenic assay of ZRS603ins13 showing strong posterior and ectopic anterior *LacZ* expression. **D.** The 13bp insertion, in bold, creates binding sites for multiple limb-associated transcription factors. Grey bars indicate the location of the TFBS.