

Published in final edited form as:

J Med Genet. 2011 October ; 48(10): 698–702. doi:10.1136/jmedgenet-2011-100303.

7q21-rs6964587 and breast cancer risk: an extended case–control study by the Breast Cancer Association Consortium

Roger L Milne¹, Justo Lorenzo-Bermejo^{2,3}, Barbara Burwinkel^{4,5}, Núria Malats¹, Jose Ignacio Arias⁶, M Pilar Zamora⁷, Javier Benítez⁸, Manjeet K Humphreys⁹, Montserrat García-Closas^{10,11}, Stephen J Chanock¹⁰, Jolanta Lissowska¹², Mark E Sherman¹⁰, Arto Mannermaa^{13,14}, Vesa Kataja^{15,16}, Veli-Matti Kosma^{13,14}, Heli Nevanlinna¹⁷, Tuomas Heikkinen¹⁷, Kristiina Aittomäki¹⁸, Carl Blomqvist¹⁹, Hoda Anton-Culver²⁰, Argyrios Ziogas²⁰, Peter Devilee^{21,22}, Christie J van Asperen²³, Rob A E M Tollenaar²⁴, Caroline Seynaeve²⁵, Per Hall²⁶, Kamila Czene²⁶, Jianjun Liu²⁷, Astrid K Irwanto²⁷, Daehee Kang²⁸, Keun-Young Yoo²⁸, Dong-Young Noh²⁸, Fergus J Couch²⁹, Janet E Olson³⁰, Xianshu Wang²⁹, Zachary Fredericksen³⁰, Børge G Nordestgaard^{31,32}, Stig E Bojesen^{31,32}, Henrik Flyger³³, Sara Margolin³⁴, Annika Lindblom³⁵, Peter A Fasching^{36,37}, Ruediger Schulz-Wendtland³⁸, Arif B Ekici³⁹, Matthias W Beckmann³⁶, Shan Wang-Gohrke⁴⁰, Chen-Yang Shen^{41,42}, Jyh-Cherng Yu⁴³, Huan-Ming Hsu⁴³, Pei-Ei Wu^{41,42}, Graham G Giles⁴⁴, Gianluca Severi⁴⁴, Laura Baglietto⁴⁴, Dallas R English⁴⁵, Angela Cox⁴⁶, Ian Brock⁴⁶, Graeme Elliott⁴⁶, Malcolm W R Reed⁴⁷, Jonathan Beesley⁴⁸, Xiaoqing Chen⁴⁸, kConFab Investigators⁴⁹, AOCs Group^{48,49}, Olivia Fletcher⁵⁰, Lorna Gibson⁵¹, Isabel dos Santos Silva⁵¹, Julian Peto⁵¹, Bernd Frank^{4,52}, Joerg Heil⁵, Alfons Meindl⁵³, Jenny Chang-Claude⁵⁴, Rebecca Hein⁵⁴, Alina Vrieling⁵⁴, Dieter Flesch-Janys^{55,56}, Melissa C Southey⁵⁷, Letitia Smith⁵⁷, Carmel Apicella⁴⁵, John L Hopper⁴⁵, Alison M Dunning⁵⁸, Karen A Pooley⁵⁸, Paul D P Pharoah^{9,58}, Ute Hamann⁵⁹, Beate Pesch⁶⁰, Yon-Dschun Ko⁶¹, The GENICA Network^{59,60,61,62,63,64}, Douglas F Easton⁹, and Georgia Chenevix-Trench⁴⁸

¹Genetic and Molecular Epidemiology Group, Human Cancer Genetics Program, Spanish National Cancer Research Centre (CNIO), Madrid, Spain ²Institute of Medical Biometry and Informatics, University Hospital Heidelberg, Heidelberg, Germany ³Division of Molecular Genetic Epidemiology, German Cancer Research Center (DKFZ), Heidelberg, Germany ⁴Molecular Epidemiology Group, DKFZ, Heidelberg, Germany ⁵Department of Obstetrics and Gynecology, University Hospital Heidelberg, Heidelberg, Germany ⁶Servicio de Cirugía General y Especialidades, Hospital Monte Naranco, Oviedo, Spain ⁷Servicio de Oncología Médica, Hospital Universitario La Paz, Madrid, Spain ⁸Human Genetics Group, Human Cancer Genetics Program, CNIO, Madrid, Spain ⁹Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK ¹⁰Division of Cancer Epidemiology and Genetics, National Cancer Institute, Rockville, Maryland, USA ¹¹Sections of Epidemiology and Genetics, Institute of Cancer Research and Breakthrough Breast Cancer Research Centre, London, UK ¹²Department of Cancer Epidemiology and Prevention, M Sklodowska-Curie Memorial Cancer Center and Institute of Oncology, Warsaw, Poland ¹³School of Medicine, Institute of Clinical Medicine, Pathology and

Correspondence to Dr Roger L Milne, Genetic and Molecular Epidemiology Group, Human Cancer Genetics Program, Spanish National Cancer Research Centre (CNIO), Calle Melchor Fernández Almagro, 3, 28039 Madrid, Spain; rmilne@cnio.es.

Competing interests None.

Ethics approval Ethics approval was provided by the relevant local institutional review boards for each of the 23 studies that contributed data.

Provenance and peer review Not commissioned; externally peer reviewed.

Additional materials and the funding statement are published online only. To view these files please visit the journal online (<http://jmg.bmj.com>).

Forensic Medicine, University of Eastern Finland, Biocenter Kuopio, Kuopio, Finland
¹⁴Department of Clinical Pathology, Kuopio University Hospital, Kuopio, Finland ¹⁵Department of Oncology, Kuopio University Hospital, Kuopio, Finland ¹⁶School of Medicine, Institute of Clinical Medicine, Oncology, University of Eastern Finland, Kuopio, Finland ¹⁷Department of Obstetrics and Gynecology, Helsinki University Central Hospital, Helsinki, Finland ¹⁸Department of Clinical Genetics, Helsinki University Central Hospital, Helsinki, Finland ¹⁹Department of Oncology, Helsinki University Central Hospital, Helsinki, Finland ²⁰Department of Epidemiology, University of California Irvine, Irvine, California, USA ²¹Department of Human Genetics, Leiden University Medical Centre, Leiden, The Netherlands ²²Department of Pathology, Leiden University Medical Centre, Leiden, The Netherlands ²³Department of Clinical Genetics, Leiden University Medical Centre, Leiden, The Netherlands ²⁴Department of Surgery, Leiden University Medical Centre, Leiden, The Netherlands ²⁵Department of Medical Oncology, Rotterdam Family Cancer Clinic, Erasmus MC-Daniel den Hoed Cancer Center, Rotterdam, The Netherlands ²⁶Department of Medical Epidemiology and Biostatistics, Karolinska Institute, Stockholm, Sweden ²⁷Human Genetics Laboratory, Genome Institute of Singapore, Singapore ²⁸Seoul National University College of Medicine, Seoul, Korea ²⁹Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, Minnesota, USA ³⁰Department of Health Sciences Research, Mayo Clinic, Rochester, Minnesota, USA ³¹Copenhagen General Population Study, Herlev University Hospital, University of Copenhagen, Copenhagen, Denmark ³²Department of Clinical Biochemistry, Herlev University Hospital, University of Copenhagen, Copenhagen, Denmark ³³Department of Breast Surgery (HF), Herlev University Hospital, University of Copenhagen, Copenhagen, Denmark ³⁴Department of Oncology–Pathology, Karolinska Institute, Stockholm, Sweden ³⁵Department of Molecular Medicine and Surgery, Karolinska Institute, Stockholm, Sweden ³⁶Department of Gynecology and Obstetrics, University Breast Center, University Hospital Erlangen, Erlangen, Germany ³⁷Department of Medicine Division of Hematology and Oncology, David Geffen School of Medicine, University of California, Los Angeles, Los Angeles, California, USA ³⁸Institute of Diagnostic Radiology, University Hospital Erlangen, Erlangen, Germany ³⁹Institute of Human Genetics, Friedrich Alexander University Erlangen-Nuremberg, Erlangen, Germany ⁴⁰Department of Obstetrics and Gynecology, University of Ulm, Ulm, Germany ⁴¹Institute of Biomedical Sciences, Academia Sinica, Taipei, Taiwan ⁴²Taiwan Biobank, Taipei, Taiwan ⁴³Department of Surgery, Tri-Service General Hospital, Taipei, Taiwan ⁴⁴Cancer Epidemiology Centre, Cancer Council Victoria, Melbourne, Australia ⁴⁵Centre for Molecular, Environmental, Genetic and Analytic Epidemiology, University of Melbourne, Melbourne, Australia ⁴⁶Institute for Cancer Studies, Sheffield University Medical School, Sheffield, UK ⁴⁷Academic Unit of Surgical Oncology, Sheffield University Medical School, Sheffield, UK ⁴⁸Queensland Institute of Medical Research, Brisbane, Australia ⁴⁹Peter MacCallum Cancer Centre, Melbourne, Australia ⁵⁰Breakthrough Breast Cancer Research Centre, The Institute of Cancer Research, London, UK ⁵¹London School of Hygiene and Tropical Medicine, London, UK ⁵²Division of Clinical Epidemiology and Aging Research, DKFZ, Heidelberg, Germany ⁵³Department of Gynaecology and Obstetrics, Technical University of Munich, Munich, Germany ⁵⁴Division of Cancer Epidemiology, DKFZ, Heidelberg, Germany ⁵⁵Department of Cancer Epidemiology/Clinical Cancer Registry, University Clinic Hamburg-Eppendorf, Hamburg, Germany ⁵⁶Institute for Medical Biometrics and Epidemiology, University Clinic Hamburg-Eppendorf, Hamburg, Germany ⁵⁷Department of Pathology, The University of Melbourne, Melbourne, Australia ⁵⁸Department of Oncology, University of Cambridge, Cambridge, UK ⁵⁹Molecular Genetics of Breast Cancer, DKFZ, Heidelberg, Germany ⁶⁰Institute for Prevention and Occupational Medicine of the German Social Accident Insurance (IPA), Bochum, Germany ⁶¹Department of Internal Medicine, Evangelische Kliniken Bonn gGmbH, Johanniter Krankenhaus, Bonn, Germany ⁶²Dr Margarete Fischer-Bosch Institute of Clinical Pharmacology, Stuttgart, Germany ⁶³University of Tübingen, Tübingen, Germany ⁶⁴Institute of Pathology, University of Bonn, Bonn, Germany

Abstract

Background—Using the Breast Cancer Association Consortium, the authors previously reported that the single nucleotide polymorphism 7q21-rs6964587 (*AKAP9*-M463I) is associated with breast cancer risk. The authors have now assessed this association more comprehensively using 16 independent case–control studies.

Methods—The authors genotyped 14 843 invasive case patients and 19 852 control subjects with white European ancestry and 2595 invasive case patients and 2192 control subjects with Asian ancestry. ORs were estimated by logistic regression, adjusted for study. Heterogeneity in ORs was assessed by fitting interaction terms or by subclassifying case patients and applying polytomous logistic regression.

Results—For white European women, the minor T allele of 7q21-rs6964587 was associated with breast cancer risk under a recessive model (OR 1.07, 95% CI 1.00 to 1.13, $p = 0.04$). Results were inconclusive for Asian women. From a combined analysis of 24 154 case patients and 33 376 control subjects of white European ancestry from the present and previous series, the best-fitting model was recessive, with an estimated OR of 1.08 (95% CI 1.03 to 1.13, $p = 0.001$). The OR was greater at younger ages (p trend = 0.01).

Conclusion—This may be the first common susceptibility allele for breast cancer to be identified with a recessive mode of inheritance.

INTRODUCTION

In a previous publication, we reported that the M463I variant in the A-kinase anchoring protein 9 gene (*AKAP9*) on chromosome 7 (7q21-rs6964587) was associated with breast cancer risk, based on a study of 9523 patients with breast cancer and 13 770 control subjects from seven independent European and Australian studies.¹ The estimated OR for rare TT homozygotes compared to GG homozygotes was 1.17 (95% CI 1.08 to 1.27, $p = 0.0003$). We aimed to assess this association more comprehensively by extending the study of this single nucleotide polymorphism to an additional 17 438 female patients with invasive breast cancer and 22 044 female control subjects from 16 independent studies participating in the Breast Cancer Association Consortium, by testing additional genetic models and by considering different breast cancer subtypes defined by immunohistochemical tumour markers.

MATERIALS AND METHODS

Eleven studies were conducted in Europe, two in the USA, one in Australia and two in Asia (table 1). All studies provided information on disease status and ethnic group (white European, Asian, other), as well as age at diagnosis and family history of breast cancer for case patients; all except one (Karolinska Breast Cancer Study) provided age at interview for control subjects. Patients with ‘genetically enriched’ breast cancer were defined as those aged younger than 40 years at diagnosis, with bilateral breast cancer and/or with at least one first-degree relative affected with breast cancer, corresponding to ‘familial’ cases in the original publication.¹ Oestrogen receptor (ER) status and progesterone receptor (PR) status were provided for a subset of 12 385 (77% positive) and 11 347 (63% positive) white European case patients, respectively, while human epidermal growth factor receptor 2 (HER2) status was provided for 5322 (15% positive) white European case patients (table 1). These variables were also obtained for 6228 (77% positive), 5400 (67% positive) and 3614 (18% positive) white European case patients, respectively, from studies that contributed data to the previously published analysis.¹ This histopathology information was generally abstracted from medical reports. Subjects who reported having ethnicity other than white European were excluded, with the exception of those from the two Asian studies (Seoul

Breast Cancer Study and Taiwanese Breast Cancer Study), for which only subjects of Asian origin were included. All subjects gave written informed consent, and each study was approved by relevant local institutional review boards.

The method used by each study to genotype 7q21-rs6964587 is provided in table 1. All studies complied with the Breast Cancer Association Consortium genotyping quality control standards by including at least 2% of samples in duplicate and a common set of 93 CEPH (Centre d'Etude du Polymorphisme Humain) DNAs used by the HapMap Consortium (HAPMAPPT01; Coriell Institute for Medical Research, Camden, New Jersey, USA).

The association of 7q21-rs6964587 with breast cancer risk was assessed by estimating genotype-specific and per-allele ORs using multivariate logistic regression, with study as a categorical covariate. Dominant and recessive models were also considered. Additional adjustment for age made no substantial difference to the results. The best-fitting genetic model was identified using Akaike's Information Criterion (AIC), which is defined as $AIC = -2 * (\ln(\text{likelihood})) + 2 * (\text{number of parameters})$.¹⁸ Between-study heterogeneity in ORs was assessed using a likelihood ratio test (LRT) comparing the model with interaction terms for the per-allele, dominant or recessive (df = 1), or genotype-specific (df = 2), log-OR by study to the model with no interaction terms. Differences in ORs by ethnicity and age were evaluated using a similar LRT. Differences in ORs between case patient groups defined by ER, PR and HER2 status were tested for white Europeans by an LRT comparing polytomous logistic regression models with and without the per-allele, dominant or recessive (all df = 1) or genotype-specific (df = 2) log-OR constrained to be equal for the two corresponding case patient groups. This LRT was also used to test the enrichment of the putative risk genotype(s) in *AKAP9*-rs6964587 in selected case patients, even though the OR estimate for genetically enriched case patients cannot be interpreted as a RR.¹⁹ All statistical tests were two-sided. The term 'statistically significant' implies $p < 0.05$. All analyses were carried out using Stata: Release 10 (StataCorp).

RESULTS

A minimum genotype concordance of 98% for duplicated samples and 95% for the CEPH samples was observed in all 16 studies, as were minimum genotype calls of 95% for case patients and control subjects. Based on Pearson's χ^2 test applied to control subjects, statistical evidence of departure from Hardy-Weinberg equilibrium was observed for two studies (Genetic Epidemiology Study of Breast Cancer by Age 50 (GESBCS) and Mayo Clinic Breast Cancer Study; $p = 0.03$ and 0.02 , respectively); for both studies, cluster plots were double-checked visually and determined to be of high quality, and all their genotype data were therefore included in the final analysis.

Initial analyses were based on 14 843 case patients and 19 852 control subjects with white European ancestry and 2595 case patients and 2192 control subjects with Asian ancestry. The estimated frequency of the minor (T) allele at 7q21-rs6964587 in control subjects was 0.39 for white Europeans (range among studies 0.37 to 0.42) but lower for Asians (0.17 in both studies) (supplementary table 1). The OR estimate for white Europeans was 1.01 (95% CI 0.98 to 1.04, $p = 0.5$) per T allele, 0.97 (95% CI 0.92 to 1.02, $p = 0.2$) for genotype GT versus GG and 1.05 (95% CI 0.98 to 1.12, $p = 0.2$) for TT versus GG. The corresponding estimates for Asians were 1.07 (95% CI 0.96 to 1.19, $p = 0.2$), 1.06 (95% CI 0.93 to 1.20, $p = 0.4$) and 1.16 (95% CI 0.83 to 1.62, $p = 0.4$), although ORs did not differ by ethnicity ($p > 0.4$). In contrast to the previous analysis,¹ there was no evidence of association for white Europeans under a dominant model (OR 0.99, 95% CI 0.94 to 1.03, $p = 0.6$). However, there was evidence of increased risk under a recessive model (OR 1.07, 95% CI 1.00 to 1.13, $p = 0.04$). Study-specific OR estimates are provided in figure 1 and in supplementary figure 1.

As observed in the previous publication¹ for the models tested, the recessive OR estimate was greater when case patients with genetically enriched breast cancer were compared to controls (OR 1.13, 95% CI 1.03 to 1.24, $p = 0.01$). We reanalysed the data from Frank *et al*¹ based on 9311 case patients and 13 524 control subjects of white European ancestry and obtained consistent estimates under the recessive model (OR 1.10, 95% CI 1.02 to 1.19, $p = 0.01$).

When we combined the data from the 21 studies of white Europeans in the present replication series and those in Frank *et al*¹ (24 154 case patients and 33 376 control subjects in total), we obtained OR estimates of 1.01 (95% CI 0.97 to 1.05, $p = 0.6$) for genotype GT versus GG and 1.09 (95% CI 1.03 to 1.15, $p = 0.002$) for TT versus GG. While a log-additive model could not be rejected (per-allele OR 1.04, 95% CI 1.01 to 1.06, $p = 0.006$, AIC = 74 083.4), the best-fitting model was recessive (AIC = 74 080.6), giving an estimated OR of 1.08 (95% CI 1.03 to 1.13, $p = 0.001$). The combined recessive OR estimate was higher (OR 1.14, 95% CI 1.06 to 1.22, $p = 0.0005$), but not statistically significantly so ($p = 0.09$), when case patients with genetically enriched breast cancer were compared to controls. However, the recessively increased risk was stronger for younger women, with estimated ORs of 1.22 (95% CI 1.02 to 1.45), 1.11 (95% CI 1.03 to 1.19) and 1.01 (95% CI 0.93 to 1.09) for women aged <40, 40–59 and 60 years or older, respectively (p trend = 0.01). There was no evidence of heterogeneity in ORs under any model by study ($p = 0.2$), and results were consistent across analyses excluding each study, one by one (supplementary table 2), suggesting that no single study was driving them. There was also no evidence of heterogeneity in ORs by tumour ER, PR or HER2 status ($p = 0.2$) (supplementary table 3).

DISCUSSION

The present study has found independent evidence of an association between 7q21-rs6964587 and breast cancer risk for white women of European origin. This combined analysis of more than 57 000 white European women suggests that homozygotes for the T allele have an average 8% increased risk compared to G allele homozygotes, with no evidence of an increased risk for heterozygotes, and this increased risk was greater for younger women. The results were inconclusive for Asian women, which was not surprising given the smaller sample size and that the T allele is less frequent; the estimated power to detect a recessive OR of 1.08 at 5% statistical significance was 6%. Given that the replication study was 50% larger than the previous study,¹ that no evidence of log-additive or dominant association was found ($p = 0.5$) and that the results of the previous study were consistent with the association being recessive, it seems reasonable to assume that the previously reported increased risk for genotype GT versus GG was due to chance.

This may be the first common variant found to be associated with breast cancer risk under a recessive mode of inheritance. However, because T allele homozygotes are relatively uncommon, further large studies will be needed to estimate the associated RR reliably. The 7q21-rs6964587 variant is a potentially deleterious²⁰ non-synonymous coding single nucleotide polymorphism in *AKAP9*. It is in strong linkage disequilibrium ($r^2 = 0.97^1$) with 7q21-rs6960867 (*AKAP9*-N2792S), which has also been suggested to be potentially deleterious.²⁰ However, it is not clear that either variant is causal. They are located in a region of high linkage disequilibrium that spans beyond *AKAP9*, and so the association, if real, may be due to a causal relationship with a variant in another gene nearby. Again, further studies will be required to identify a causal variant.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We thank all the individuals who took part in these studies and all the researchers, clinicians, technicians and administrative staff who have enabled this work to be carried out. In particular, we thank Charo Alonso, Tais Moreno, Guillermo Pita, Primitiva Menendez, Anna González-Neira, Jonathan Morrison, Louise Brinton, Neonila Szeszenia-Dabrowska, Beata Peplonska, Witold Zatonski, Pei Chao, Michael Stagner, Eija Myöhänen, Helena Kemiläinen, Kirsimari Aatonen, Hanna Jäntti, Irja Erkkilä, the Finnish Cancer Registry, Irene Masunaka, PEA Huijts, E Krol-Warmerdam, J Blom, Ursula Eilber, Tanya Koehler, Simon Cross, Helen Cramp, Dan Conlley, Heather Thorne, Eveline Niedermayr, the Australian Ovarian Cancer Study Management Group (D Bowtell, G Chenevix-Trench, A deFazio, D Gertig, A Green and P Webb), the Australian Cancer Study Management Group (A Green, P Parsons, N Hayward, P Webb and D Whiteman), Eileen Williams, Elaine Ryder-Mills, Kara Sargus, Rita K Schmutzler, Claus R Bartram, Tracy Slanger, Elke Mutschelknauss, Ramona Salazar, Sabine Behrens, Renate Birr, Belinda Kaspereit, Nicole Knese, Maggie Angelakos, Judi Maskiell, Gillian Dite, the SEARCH (Study of Epidemiology and Risk Factors in Cancer Heredity) and European Prospective Investigation into Cancer and Nutrition (EPIC) teams, Christina Justenhoven, Hiltrud Brauch, Volker Harth, Sylvia Rabstein, Thomas Brüning and Hans-Peter Fischer.

References

1. Frank B, Wiestler M, Kropp S, Hemminki K, Spurdle AB, Sutter C, Wappenschmidt B, Chen X, Beesley J, Hopper JL, Meindl A, Kiechle M, Slanger T, Bugert P, Schmutzler RK, Bartram CR, Flesch-Janys D, Mutschelknauss E, Ashton K, Salazar R, Webb E, Hamann U, Brauch H, Justenhoven C, Ko YD, Brüning T, Silva Idos S, Johnson N, Pharoah PP, Dunning AM, Pooley KA, Chang-Claude J, Easton DF, Peto J, Chenevix-Trench G, Fletcher O, Burwinkel B. Australian Breast Cancer Family Study Investigators; Houlston R Gene Environment Interaction and Breast Cancer in Germany Group, Kathleen Cuninghame Foundation Consortium for Research into Familial Breast Cancer Investigators, Australian Ovarian Cancer Study Management Group. Association of a common AKAP9 variant with breast cancer risk: a collaborative analysis. *J Natl Cancer Inst.* 2008; 100:437–42. [PubMed: 18334708]
2. Schrauder M, Frank S, Strissel PL, Lux MP, Bani MR, Rauh C, Sieber CC, Heusinger K, Hartmann A, Schulz-Wendland R, Strick R, Beckmann MW, Fasching PA. Single nucleotide polymorphism D1853N of the ATM gene may alter the risk for breast cancer. *J Cancer Res Clin Oncol.* 2008; 134:873–82. [PubMed: 18264724]
3. Weischer M, Bojesen SE, Tybjaerg-Hansen A, Axelsson CK, Nordestgaard BG. Increased risk of breast cancer associated with CHEK2*1100delC. *J Clin Oncol.* 2007; 25:57–63. [PubMed: 16880452]
4. Milne RL, Ribas G, Gonzalez-Neira A, Fagerholm R, Salas A, Gonzalez E, Dopazo J, Nevanlinna H, Robledo M, Benitez J. ERCC4 associated with breast cancer risk: a two-stage case-control study using high-throughput genotyping. *Cancer Res.* 2006; 66:9420–7. [PubMed: 17018596]
5. Chang-Claude J, Eby N, Kiechle M, Bastert G, Becher H. Breastfeeding and breast cancer risk by age 50 among women in Germany. *Cancer Causes Control.* 2000; 11:687–95. [PubMed: 11065005]
6. Heikkinen T, Karkkainen H, Aaltonen K, Milne RL, Heikkilä P, Aittomäki K, Blomqvist C, Nevanlinna H. The breast cancer susceptibility mutation PALB2 1592delT is associated with an aggressive tumor phenotype. *Clin Cancer Res.* 2009; 15:3214–22. [PubMed: 19383810]
7. Margolin S, Werelius B, Fornander T, Lindblom A. BRCA1 mutations in a population-based study of breast cancer in Stockholm County. *Genet Test.* 2004; 8:127–32. [PubMed: 15345109]
8. Hartikainen JM, Tuhkanen H, Kataja V, Dunning AM, Antoniou A, Smith P, Arffman A, Pirskanen M, Easton DF, Eskelinen M, Uusitupa M, Kosma VM, Mannermaa A. An autosome-wide scan for linkage disequilibrium-based association in sporadic breast cancer cases in eastern Finland: three candidate regions found. *Cancer Epidemiol Biomarkers Prev.* 2005; 14:75–80. [PubMed: 15668479]
9. Olson JE, Ma CX, Pelleymounter LL, Schaid DJ, Pankratz VS, Vierkant RA, Fredericksen ZS, Ingle JN, Wu Y, Couch F, Sellers TA, Weinshilboum RM, Vachon CM. A comprehensive examination of CYP19 variation and breast density. *Cancer Epidemiol Biomarkers Prev.* 2007; 16:623–5. [PubMed: 17372263]
10. Giles GG, English DR. The Melbourne Collaborative Cohort Study. *IARC Sci Publ.* 2002; 156:69–70. [PubMed: 12484128]

11. Huijts PE, Vreeswijk MP, Kroeze-Jansema KH, Jacobi CE, Seynaeve C, Krol-Warmerdam EM, Wijers-Koster PM, Blom JC, Pooley KA, Klijn JG, Tollenaar RA, Devilee P, van Asperen CJ. Clinical correlates of low-risk variants in FGFR2, TNRC9, MAP3K1, LSP1 and 8q24 in a Dutch cohort of incident breast cancer cases. *Breast Cancer Res.* 2007; 9:R78. [PubMed: 17997823]
12. Garcia-Closas M, Brinton LA, Lissowska J, Chatterjee N, Peplonska B, Anderson WF, Szeszenia-Dabrowska N, Bardin-Mikolajczak A, Zatonski W, Blair A, Kalaylioglu Z, Rymkiewicz G, Mazepa-Sikora D, Kordek R, Lukaszek S, Sherman ME. Established breast cancer risk factors by clinically important tumour characteristics. *Br J Cancer.* 2006; 95:123–9. [PubMed: 16755295]
13. Wedren S, Lovmar L, Humphreys K, Magnusson C, Melhus H, Syvanen AC, Kindmark A, Landegren U, Fermer ML, Stiger F, Persson I, Baron J, Weiderpass E. Oestrogen receptor alpha gene haplotype and postmenopausal breast cancer risk: a case control study. *Breast Cancer Res.* 2004; 6:R437–49. [PubMed: 15217512]
14. MacPherson G, Healey CS, Teare MD, Balasubramanian SP, Reed MW, Pharoah PD, Ponder BA, Meuth M, Bhattacharyya NP, Cox A. Association of a common variant of the CASP8 gene with reduced risk of breast cancer. *J Natl Cancer Inst.* 2004; 96:1866–9. [PubMed: 15601643]
15. Ziogas A, Gildea M, Cohen P, Bringman D, Taylor TH, Seminara D, Barker D, Casey G, Haile R, Liao SY, Thomas D, Noble B, Kurosaki T, Anton-Culver H. Cancer risk estimates for family members of a population-based family registry for breast and ovarian cancer. *Cancer Epidemiol Biomarkers Prev.* 2000; 9:103–11. [PubMed: 10667470]
16. Han S, Lee KM, Choi JY, Park SK, Lee JY, Lee JE, Noh DY, Ahn SH, Han W, Kim DH, Hong YC, Ha E, Yoo KY, Kang D. CASP8 polymorphisms, oestrogen and progesterone receptor status, and breast cancer risk. *Breast Cancer Res Treat.* 2008; 110:387–93. [PubMed: 17940865]
17. Ding SL, Yu JC, Chen ST, Hsu GC, Kuo SJ, Lin YH, Wu PE, Shen CY. Genetic variants of BLM interact with RAD51 to increase breast cancer susceptibility. *Carcinogenesis.* 2009; 30:43–9. [PubMed: 18974064]
18. Akaike H. A new look at the statistical model identification. *IEEE Trans Automat Contr.* 1974; 19:716–23.
19. Byrnes GB, Southey MC, Hopper JL. Are the so-called low penetrance breast cancer genes, ATM, BRIP1, PALB2 and CHEK2, high risk for women with strong family histories? *Breast Cancer Res.* 2008; 10:208. [PubMed: 18557994]
20. Ng PC, Henikoff S. Predicting the effects of amino acid substitutions on protein function. *Annu Rev Genomics Hum Genet.* 2006; 7:61–80. [PubMed: 16824020]

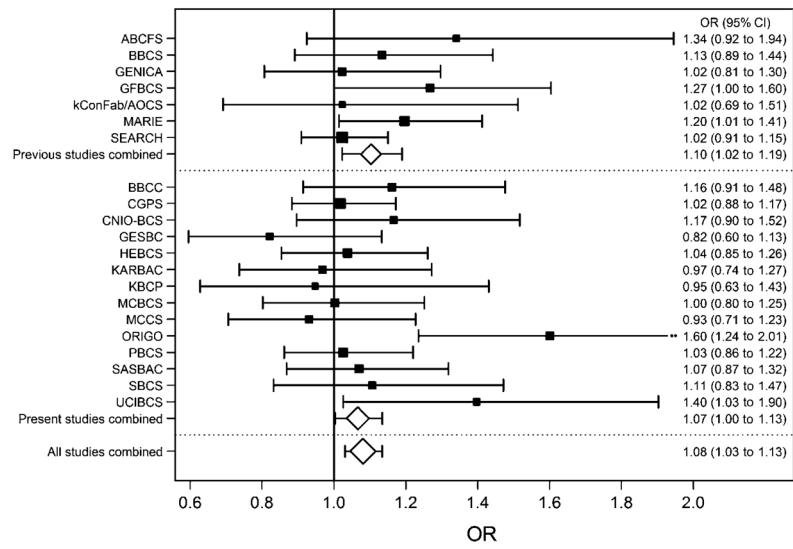


Figure 1. OR estimates and their associated 95% CIs under a recessive model, by study. The area of the box/diamond is inversely proportional to the standard error of the log-OR estimate.

Table 1

Participating studies, number of subjects and genotyping methods used

Study acronym	Study name (reference)	Country	Controls (n)	Cases (n)	ER	PR	HER2	Genotyping method*
Women of European origin (previously published series) ¹								
ABCFS	Australian Breast Cancer Family Study	Australia	368	540	473 (67)	474 (70)	0	iPLEX
BBCS	British Breast Cancer Study	UK	2635	580	0	0	0	Sentrix
GENICA	Gene Environment Interaction and Breast Cancer in Germany	Germany	995	983	934 (78)	932 (70)	604 (27)	TaqMan
GFBCS	German Familial Breast Cancer Study	Germany	1115	1083	0	0	0	TaqMan
kConFab/AOCS	Kathleen Cunningham Foundation Consortium for Research into Familial Breast Cancer/Australian Ovarian Cancer Study	Australia	714	271	119 (71)	100 (73)	0	iPLEX
MARIE	Mammary Carcinoma Risk Factor Investigation	Germany	3187	1608	1646 (76)	1644 (64)	1472 (20)	TaqMan
SEARCH	Study of Epidemiology and Risk Factors in Cancer Heredity	UK	4510	4246	3056 (79)	2250 (67)	1538 (11)	TaqMan
Women of European origin (replication series)								
BBC	Bavarian Breast Cancer Cases and Controls ²	Germany	965	1251	995 (73)	992 (65)	880 (16)	TaqMan
CGPS	Copenhagen General Population Study ³	Denmark	6541	1931	1711 (82)	1149 (58)	0	TaqMan
CNIO-BCS	Spanish National Cancer Centre Breast Cancer Study ⁴	Spain	807	696	240 (75)	254 (57)	145 (26)	TaqMan
GESBC	Genetic Epidemiology Study of Breast Cancer by Age 50 ⁵	Germany	560	511	431 (60)	423 (57)	0	TaqMan
HEBCS	Helsinki Breast Cancer Study ⁶	Finland	1273	2233	2216 (81)	2215 (65)	1304 (16)	iPLEX
KARBAC	Karolinska Breast Cancer Study ⁷	Sweden	855	796	433 (83)	365 (76)	0	TaqMan
KBCP	Kuopio Breast Cancer Project ⁸	Finland	404	467	440 (76)	438 (62)	398 (13)	TaqMan
MCBCS	Mayo Clinic Breast Cancer Study ⁹	USA	1152	1045	1079 (83)	1074 (73)	735 (20)	TaqMan
MCCS	Melbourne Collaborative Cohort Study ¹⁰	Australia	760	663	605 (73)	606 (57)	396 (12)	TaqMan
ORIGO	Leiden University Medical Centre Breast Cancer Study ¹¹	Netherlands	1419	552	403 (76)	355 (59)	0	TaqMan
PBCS	NCI Polish Breast Cancer Study ¹²	Poland	2323	1941	1808 (65)	1802 (52)	1268 (11)	TaqMan
SASBAC	Singapore and Sweden Breast Cancer Study ¹³	Sweden	1458	1217	833 (82)	812 (74)	0	iPLEX
SBCS	Sheffield Breast Cancer Study ¹⁴	UK	822	727	505 (78)	185 (57)	196 (9)	TaqMan
UCIBCS	UCI Breast Cancer Study ¹⁵	USA	513	813	686 (80)	677 (70)	0	TaqMan
Total (white Europeans)			33 376	24 154	18 613 (77)	16 747 (64)	8936 (16)	
Studies of Asian women (replication series)								

Study acronym	Study name (reference)	Country	Controls (n)	Cases (n)	ER	PR	HER2	Genotyping method*
SEBCS	Seoul Breast Cancer Study ¹⁶	South Korea	1114	1689	0	0	0	TaqMan
TWBCS	Taiwanese Breast Cancer Study ¹⁷	Taiwan	1078	906	779 (63)	779 (56)	347 (33)	TaqMan
Total (Asians)			2192	2595	779 (63)	779 (56)	347 (33)	

* TaqMan, nuclease assay (TaqMan®), with reagents designed by Applied Biosystems (<http://www.appliedbiosystems.com/>) as Assays-by-DesignSM and genotyping performed using the ABI PRISM 7900HT, 7700 or 7500 Sequence Detection Systems according to manufacturer's instructions; Sentrix, customised Illumina Sentrix Bead Arrays (Illumina, San Diego, California, USA); iPLEX, matrix-assisted laser desorption/ionization time of flight mass spectrometry for the determination of allele-specific primer extension products using Sequenom's MassARRAY system and iPLEX technology (Sequenom, San Diego, California, USA), with oligonucleotide design carried out according to the guidelines of Sequenom and performed using MassARRAY Assay Design software (V.3.1).

ER, oestrogen receptor; HER2, human epidermal growth factor receptor 2; PR, progesterone receptor.

ER, number of cases with known ER status (percentage positive in parentheses); PR, number of cases with known PR status (percentage positive in parentheses); HER2, number of cases with known HER2 status (percentage positive in parentheses).