

Published in final edited form as:

Curr Opin Pharmacol. 2012 August ; 12(4): 458–463. doi:10.1016/j.coph.2012.02.003.

Ras Family of Small GTPases In Immunity And Inflammation

Derek S. Johnson and Youhai H. Chen

Department of Pathology and Laboratory Medicine, University of Pennsylvania School of Medicine, Philadelphia, P.A., 19104, USA

Abstract

The Ras superfamily of small GTPases is a group of more than 150 small G proteins, all of which share some degree of homology to the founding member Ras. These small GTPases function as molecular switches within cells, impacting nearly all cellular processes. The Ras superfamily can be further divided into several smaller subfamilies, with those proteins that most closely resemble Ras belonging to the Ras subfamily. While heavily studied within the field of cancer biology, the Ras family of proteins also plays cardinal roles in immunity and inflammation. Here we review the roles of these molecular switches in regulating immune cell homeostasis and functions.

What Are Small GTPases?

Small GTPases are molecular switches that are capable of being in an “on” state in which they are bound to Guanosine-Triphosphate (GTP), or an “off” state in which they are bound to Guanosine-Diphosphate (GDP) (Figure 1). Their binding to either of these nucleotides causes changes in the conformation of the GTPase, which enables them to interact with and activate/deactivate different effector molecules. Small GTPases have an intrinsic (albeit very weak) ability to hydrolyze GTP to GDP, hence their possession of an internal “timer” which will set off eventually. More typically these switches are activated by Guanine-Nucleotide-Exchange-Factors (GEFs) which “exchange” GTP into a binding pocket previously occupied by GDP, hence turning “on” the GTPase. GTPase-Activating-Proteins (GAPs) activate the intrinsic GTPase activity of these proteins (speeding up the timer), and convert bound GTP into GDP and turn “off” the switch. It is the interplay between the GTPases, their GEFs and GAPs that coordinate the functioning and modulation of many diverse biological processes.

The Ras Superfamily

Ras is a GTPase that was initially found to be mutated in a wide variety of cancers. Over time many similar GTPases were discovered to have a related 3D structure to Ras, and the family now stands at over 150 total members. This large superfamily can be further divided into at least six families; Ras, Rho, Ran, Rab, Rheb, and ARF. While each of these families of small GTPases has been extensively studied in the context of carcinogenesis, we are only beginning to scratch the surface of how these related families of proteins are able to control

© 2012 Elsevier Ltd. All rights reserved.

Corresponding Authors: Youhai H. Chen, yhc@mail.med.upenn.edu. Derek S. Johnson, dereksp@mail.med.upenn.edu. Department of Pathology and Laboratory Medicine, University of Pennsylvania School of Medicine, 713 Stellar-Chance Labs, 422 Curie Blvd., Philadelphia, PA 19104-6160. Phone: 215 898 4671. FAX: 215 573 3434.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

non-oncogenic processes such as immunity and inflammation. There is tremendous overlap in the functions performed by each of these families of GTPases, but the primary functions for each family are summarized in Figure 2. The Ras family, the first and most diverse of the subfamilies, can play a vital role in the regulation of immunity and inflammation.

Ras

Ras is the founding and prototypical member of the small GTPases. It regulates multiple cellular processes including cell survival, growth, and differentiation. Upon stimulation by a growth factor or other extracellular stimuli, a receptor tyrosine kinase (RTK) activates a Ras GEF such as Son of Sevenless (SOS), which activates Ras by loading it with GTP allowing it to bind its downstream effectors (Figure 3). Following activation, Ras triggers three primary effector arms, the Raf/Mek/Erk pathway, the PI3K pathway, and the RalGDS signaling pathway. The RalGDS pathway activates Ral A and Ral B, also members of the Ras GTPase family (to be discussed below).

ERK

When bound to Raf, Ras initiates the Raf/Mek/ERK MAPK signaling cascade. In this signaling cascade, each upstream molecule acts as a kinase to phosphorylate and activate a subsequent downstream molecule, with an eventual result being the modulation of transcription via the phosphorylation of a variety of transcription factors. ERK plays a crucial role in cell proliferation and survival. It is activated in lymphocytes downstream of TCR and BCR stimulation [1]. ERK activation can inhibit Fas-mediated apoptosis in T cells [2]. The production of the vital cytokine IL-2 by T cells is dependent upon ERK activation [3][4], as is the regulation of glucose metabolism following TCR stimulation [5]. Diacylglycerol, an important second messenger in the activation of Ras and ERK signaling, plays crucial roles in T cell responses and Fc γ receptor-mediated phagocytosis [6][7][8]. Finally, ERK signaling is also important in TLR-mediated chemokine production in dendritic cells [9].

PI3K

Class I PI3K enzymes are composed of a regulatory (p85) and catalytic (p110) subunit. The catalytic subunit has a Ras binding domain and can be activated by GTP-bound Ras. Upon activation, PI3K converts PIP2 to PIP3, which can serve as a docking motif for proteins containing PH domains. Within the immune system, PI3K plays crucial roles in the activation and functioning of all immune cells. PI3K regulates cytokine responsiveness and functions both in effector and regulatory T cells, and mice with PI3K deficient T cells have several immune defects [10]. The downstream PI3K target mTOR determines whether T cells become activated or undergo anergy [11]. PI3K has been shown to be crucial for signalosome formation, without which T cells have compromised antigenic responses. PI3K is also crucial for proper functioning of B cells [7][12], NK cells [13], dendritic cells (both myeloid and plasmacytoid), mast cells [14], macrophages, and neutrophils [10].

Rap

The Rap proteins share approximately 50% sequence homology to Ras, and have an identical amino acid sequence in their effector loops. While there are multiple Rap proteins, we will only discuss here the two most well studied isoforms, Rap1a and Rap1b. It was initially hypothesized that Rap1 functioned as a direct inhibitor of Ras by competition for substrate binding. This idea has since been revised as it has become clear that Rap1 has its own set of effector proteins and exists in completely different signaling pathways than does Ras. Rap1 has been implicated in the activation of integrin-mediated adhesion [15][16], establishment of polarity [17][18], cell proliferation, and the control of cell-cell interactions

[19]. The innate immune system has a requirement for Rap1 in both macrophages [20] and dendritic cells [21]. In T cells, Rap1 is crucial for chemokine-induced polarization [19][22], and in B cells it regulates cell spreading and adhesion [23]. Rap1 is a potent activator of LFA1 [20], and at least 3 different second messengers can activate Rap: i.e., cAMP, calcium, and diacylglycerol.

Rap1a

Due to having 90% sequence homology, Rap1a and Rap1b were traditionally assumed to serve redundant roles in the body. However experiments with knockout and transgenic mice have significantly increased our understanding of Rap1 function, as well as the specific functions carried out by Rap1a and 1b. As mentioned above Rap1a was initially thought to exist to antagonize Ras signaling. This initial hypothesis was supported by the observation that large quantities of activated Rap1a are found in anergic T cells, and that activation of CD28 by antibody binding inhibits the induction of Rap1a by TCR stimulation. Additionally in Jurkat T cells, active Rap1a expression inhibits activation of ERK and induction of IL-2 gene expression [24]. The assumption from these results was that suppression of Rap1 expression is required for a maximal T cell response. Nevertheless the theory that Rap1a inhibits lymphocyte activation has been called into question by the generation of transgenic mice expressing constitutively active Rap1a. The Rap1a transgenic mice expressing an active form of Rap1a did not have problems with either Ras signaling or T cell activation [25]. Indeed, instead of displaying an anergic response, active Rap1a-overexpressing lymphocytes possessed an enhanced TCR response. Additionally activation of Rap1a caused T cells to bind more strongly to fibronectin, and induced strong activation of B1 and B2 integrins, a process that generally requires antigen receptor binding to induce. In contrast Rap1A null mice had lymphocytes that were defective in adhesion to fibronectin and ICAM coated plates. Rap 1A null T cells showed impaired polarization following CD3 stimulation, but these mice had otherwise healthy lymphocyte function [26]. Macrophages from Rap1a knockouts had increased haptotaxis but reduced chemotaxis, and had an increase in FcR-mediated phagocytosis. Neutrophils from these mice had reduced superoxide production in response to fMLP stimulation, which is likely due to the fact that Rap1a interacts with the p22 subunit of NADPH oxidase in neutrophils [27].

Rap1b

Since Rap1b is highly expressed in B cells, Rap1b null mice were created to understand the role of Rap1b in B cells [28]. Rap1b knockout mice showed reduced T-dependent, but normal T-independent humoral responses. B cells from these mice showed reduced migration in response to chemokines, and reduced homing to lymph nodes, while lung endothelial cells had delayed healing in a wound-healing assay [29]. These mice had fewer pre-B cells in the bone marrow, although splenic B cell proliferation was not affected. Additionally, mice have been generated that lack the Rap1 Gap SPA1 and loss of this enzyme (resulting in increased Rap1 activity) in hematopoietic and peripheral T cells resulted in antigen-induced T cell anergy. Young mice had defective T cell memory responses, and stronger Rap1 activation upon stimulation with anti-CD3 antibody. These mice had an increased rate of myeloproliferative disease and an increase in the CD44-high population of T cells. Rap1b is also the primary isoform expressed in NK cells, and may play a role in cytokine and chemokine production in these cells [30].

From these studies it is clear that, *in vivo*, Rap1 is crucial for proper functioning and development of T and B lymphocytes and that while both isoforms seem to affect chemotaxis, Rap1a has a larger effect on adhesion and neutrophil activity, while rap1b plays a crucial role in NK and humoral responses. Thus we can conclude that Rap1 is a regulator of cell adhesion, cell proliferation, and cell junction formation.

Ral

The Ral (Ras-like) family of small GTPases is composed of two isoforms, Ral A and Ral B, which share 85% protein sequence identity between them and approximately 50% homology with Ras. Ral proteins have six known GEFs, four of which are of the RalGDS family (RalGDS, RGL1, RGL2, RGL3) and are primarily activated by upstream Ras signaling. Ral A and Ral B are highly pleiotropic and impact many diverse signaling pathways within the cell. The Ral proteins have many downstream binding partners that they interact with in order to modulate cellular conditions and respond to extracellular signals. The primary of these effectors are Ralbp1, the exocyst complex, and phospholipase D. In addition the RalGDS family itself has the capacity to act as a scaffold to activate the Ser/Thr Kinase AKT and promote cell survival and proliferation [31].

RalBP1

Also known as RLIP76 and RIP (Ral interacting protein), RalBP1 is a non-ABC multi-functional membrane transport protein that is responsible for the majority of glutathione electrophile conjugate export in mammalian cells. This transporter is responsible for a large amount of chemotherapeutic drug removal from cancerous cells, and provides protection from a variety of forms of oxidative damage or radiation-induced stress. RalBP1 has been linked to migration, endocytosis of multiple receptor ligand pairs including TGF β , and has GAP activity towards some RHO family GTPases. Inhibition of RalBP1 with an antibody directed towards an extracellular region of the molecule induced apoptosis in target cells. Also autoantibodies to the C terminal region of RalBP1 are associated with several immune mediated diseases such as Behçet disease, SLE, and carotid atherosclerosis [32]. Ralbp1 is crucial for the proper expression of the transcription factor hsf-1, which is necessary for the expression of many heat shock proteins, which are essential for cell survival under many infectious conditions.

Exocyst

The exocyst complex is a tethering complex that exists to tether intracellular vesicles to the plasma membrane prior to vesicular fusion during polarized exocytosis. It is an eight-member complex that exists in two parts, a three-member complex, which exists on the vesicle being trafficked, and a five-member complex that exists on the plasma membrane. Ral binds to and induces fusion between members of each of these complexes, Exo84 on the vesicular complex, and sec5 on the membrane complex. The exocyst complex plays a central role in polarized exocytosis. Additionally the exocyst has been shown to play a role in ciliogenesis, cytokinesis, wound healing, and cell migration [33]. Moreover one study has shown that the exocyst complex is needed for appropriate NK cell degranulation and proper NK cell cytotoxicity [34]. Recently it has been reported that RalB and Exo84 can assemble to autophagosomes and initiate autophagy [35]. RalB is also capable of inducing an interaction between sec5 and the non-canonical IKK TBK1, resulting in the activation of TBK1 and the downstream viral defense and survival pathways it controls [36]. A systems biology approach to assessing pieces of the drosophila phagosome identified components of the exocyst complex [37]. Being crucial for a variety of key processes involving cell polarization, the exocyst complex and its multiple components will likely be recognized to play an even more vital role within the immune system in the future.

PLD

Phospholipase D (PLD) is an enzyme that hydrolyses phosphatidylcholine into phosphatidic acid. PLD plays an important role in the internalization and recycling of receptors. Both RalA and RalB can interact with PLD, though it is the RalA interaction with phospholipase D positively regulates Fc γ R mediated phagocytosis [38]. PLD, via phosphatidic acid

production, can activate the central kinase mTOR, and this activation can be modulated by RalA [39].

TIPE2

Finally a recent study from our lab has shown that a crucial inflammation regulator TIPE2 operates via inhibition of the RalGDS family of GEFs (and hence Ral activity) [40].

Conclusion

The Ras family of small GTPases plays crucial roles in many cellular processes, regulating survival, motility, polarization, attachment, proliferation, and many more. This puts them into a position of paramount importance in the proper functioning and operation of the immune system. It is anticipated that many new advances will be made in the future as we learn more about this family of molecular switches in immune cell homeostasis and function.

Acknowledgments

Supported by grants from the National Institutes of Health, USA (AI-077533, AI-050059, and GM-085112).

References

1. Donahue AC, Fruman DA. Leukocyte signaling Distinct signaling mechanisms activate the target of rapamycin in response to different B-cell stimuli. *European Journal of Immunology*. 2007; 37:2923–2936.
2. Holmstro TH, Schmitz I, So TS, Poukkula M, Johnson VL, Chow SC, Krammer PH, Eriksson JE. MAPK/ERK signaling in activated T cells inhibits CD95/Fas-mediated apoptosis downstream of DISC assembly. *EMBO Journal*. 2000; 19:5418–5428. [PubMed: 11032809]
3. Tsukamoto N, Hattori M, Yang H, Bos JL, Minato N. Rap1 GTPase-activating Protein SPA-1 Negatively Regulates Cell Adhesion. *The Journal of biological chemistry*. 1999; 274:18463–18469. [PubMed: 10373454]
4. Koike T, Yamagishi H, Hatanaka Y, Fukushima A, Chang J-wen, Xia Y, Fields M, Chandler P, Iwashima M. A Novel ERK-dependent Signaling Process That Regulates Interleukin-2 Expression in a Late Phase of T Cell Activation. *The Journal of biological chemistry*. 2003; 278:15685–15692. [PubMed: 12595531]
- 5*. Marko AJ, Miller RA, Kelman A, Frauwirth KA. Induction of Glucose Metabolism in Stimulated T Lymphocytes Is Regulated by Mitogen-Activated Protein Kinase Signaling. *PloS one*. 2010; 5:e15425. Identifies ERK as a key component required for glucose metabolism in T Cells. Provides insight into the role played by both CD28 and TCR in important metabolic events within T cells, and how they cooperate to allow a T cell to become fully active. [PubMed: 21085672]
6. Riese MJ, Grewal J, Das J, Zou T, Patil V, Chakraborty AK, Koretzky GA. Decreased Diacylglycerol Metabolism Enhances ERK Activation and Augments CD8 T Cell Functional Responses. *Journal of Biological Chemistry*. 2011; 286:5254–5265. NEW. [PubMed: 21138839]
7. Shin J, Brien TFO, Grayson JM. Differential Regulation of Primary and Memory CD8 T Cell Immune Responses by Diacylglycerol Kinases. *The Journal of Immunology*. 2012; 190:4049/ jimmunol.1102265.
8. Botelho RJ, Harrison RE, Stone JC, Hancock JF, Philips MR, Jongstra-bilen J, Mason D, Plumb J, Gold MR, Grinstein S. Localized Diacylglycerol-dependent Stimulation of Ras and Rap1 during Phagocytosis. *Journal of Biological Chemistry*. 2009; 284:28522–28532. [PubMed: 19700408]
9. Mitchell D, Olive C. Regulation of Toll-like receptor-induced chemokine production in murine dendritic cells by mitogen-activated protein kinases. *Molecular Immunology*. 2010; 47:2065–2073. [PubMed: 20451253]

10. Sasaki T, Oliveira-dos-santos AJ, Stanford WL, Koziaradzki I, Joza N, Mak TW. Function of PI3K in Thymocyte Development, T Cell Activation, and Neutrophil Migration. *Science*. 2000; 1040:1040–1046. [PubMed: 10669416]
11. Zheng Y, Collins SL, Lutz MA, Amy N, Kole TP, Zarek PE, Powell JD, Zheng Y, Collins SL, Lutz MA, et al. A Role for Mammalian Target of Rapamycin in Regulating T Cell Activation versus Anergy. *The Journal of Immunology*. 2007; 178:2163–2170. [PubMed: 17277121]
12. Heidt S, Roelen DL, Eijsink C, Kooten CV, Claas FHJ, Mulder A. Effects of Immunosuppressive Drugs On Purified Human B Cells: Evidence Supporting the Use of MMF and Rapamycin. *American Journal of Transplantation*. 2008; 86:1292–1300.
- 13*. Ackermann JA, Radtke D, Maurberger A, Winkler TH, Nitschke L. Grb2 regulates B-cell maturation, B-cell memory responses and inhibits B-cell Ca²⁺ signalling. *The EMBO Journal*. 2011; 30:1621–1633. Ties the Ras signaling adaptor Grb2 to B Cell receptor signaling. Further clarifies earlier work by showing that Grb2 is important for JNK and PI3K activation downstream of Ras, although its role in ERK activation in B cells may be less crucial. [PubMed: 21427701]
14. Wai, L-en; Fujiki, M.; Takeda, S.; Martinez, OM.; Krams, SM. Rapamycin, But Not Cyclosporine or FK506, Alters Natural Killer Cell Function. *Transplantation*. 2008; 85:145–149. [PubMed: 18192925]
15. Cirillo R, Ciccarelli A, Oriente A, Marone ANDG. Characterization of The Anti-Inflammatory Effect of Human Mast Cells. *The Journal of Immunology*. 1991; 147:4278–4285. [PubMed: 1721644]
16. Tsukamoto H, Irie A, Nishimura Y. B-Raf Contributes to Sustained Extracellular Signal-regulated Kinase Activation Associated with Interleukin-2 Production Stimulated through the T Cell Receptor. *The Journal of biological chemistry*. 2004; 279:48457–48465. [PubMed: 15339934]
17. Shimonaka M, Katagiri K, Nakayama T, Fujita N, Tsuruo T, Yoshie O, Kinashi T. Rap1 translates chemokine signals to integrin activation, cell polarization, and motility across vascular endothelium under flow. *The Journal of Cell Biology*. 2003; 161:417–427. [PubMed: 12707305]
18. Schwamborn JC, Püschel AW. The sequential activity of the GTPases Rap1B and Cdc42 determines neuronal polarity. *Nature Neuroscience*. 2004; 7:923–929.
19. Gérard A, Mertens AEE, Kammen RAVD, Collard JG. The Par polarity complex regulates Rap1 - and chemokine-induced T Cell polarization. *The Journal of Cell Biology*. 2007; 176:863–875. [PubMed: 17353362]
20. Katagiri K, Hattori M, Minato N, Irie S-kichi, Takatsu K, Kinashi T. Rap1 Is a Potent Activation Signal for Leukocyte Function-Associated Antigen 1 Distinct from Protein Kinase C and Phosphatidylinositol-3-OH Kinase. *Molecular and Cellular Biology*. 2000; 20:1956–1969. [PubMed: 10688643]
21. Caron E, Self AJ, Hall A. The GTPase Rap1 controls functional activation of α M β 2 macrophage by LPS and other inflammatory mediators. *Current Biology*. 2000; 10:974–978. [PubMed: 10985384]
22. Katagiri K, Ohnishi N, Kabashima K, Iyoda T, Takeda N, Shinkai Y, Inaba K, Kinashi T. Crucial functions of the Rap1 effector molecule RAPL in lymphocyte and dendritic cell trafficking. *Nature Immunology*. 2004; 5:1045–1051. [PubMed: 15361866]
23. Mcleod SJ, Shum AJ, Lee RL, Takei F, Gold MR, Immunol MRJ, Biol MRJ. The Rap GTPases Regulate Integrin-mediated Adhesion, Cell Spreading, Actin Polymerization, and Pyk2 Tyrosine Phosphorylation in B Lymphocytes. *The Journal of biological chemistry*. 2004; 279:12009–12019. [PubMed: 14701796]
24. Boussiotis VA, Freeman GJ, Berezovskaya A, Barber DL, Nadler LM. Maintenance of Human T Cell Anergy: Blocking of IL-2 Gene Transcription by Activated Rap1. *Science*. 1997; 124:124–128. [PubMed: 9311917]
25. Sebzda E, Bracke M, Tugal T, Hogg N, Cantrell DA. Rap1A positively regulates T cells via integrin activation rather than inhibiting lymphocyte signaling. *Nature Immunology*. 2002; 3:251–258. [PubMed: 11836528]

26. Duchniewicz M, Zemojtel T, Grossmann S, Scheele JS, Zwartkruis FJT, Duchniewicz M, Zemojtel T, Kolanczyk M, Grossmann S. Rap1A-Deficient T and B Cells Show Impaired Integrin-Mediated Cell Adhesion. *Molecular and Cellular Biology*. 2006; 26:643–653. [PubMed: 16382154]
27. Li Y, Yan J, De P, Chang H-chen, Li KWC, Nivanka C, Peng X, Kim C, Kapur R, Chen H, et al. Rap1a Null Mice Have Altered Myeloid Cell Functions Suggesting Distinct Roles for the Closely Related Rap1a and 1b Proteins. *The Journal of Immunology*. 2007; 179:8322–8331. [PubMed: 18056377]
28. Chu H, Awasthi A, Li GCW, Chrzanowska-wodnicka M, Chu H, Awasthi A, Li GCW, Chrzanowska-wodnicka M, Malarkannan S. Rap1b Regulates B Cell Development, Homing, and T Cell-Dependent Humoral Immunity. *The Journal of Immunology*. 2008; 181:3373–3383. [PubMed: 18714009]
29. Chrzanowska-wodnicka M, Kraus AE, Gale D, Li GCW, De W, Chrzanowska-wodnicka M, Kraus AE, Gale D, Li GCW, Vansluys J. Defective angiogenesis, endothelial migration, proliferation, and MAPK signaling in Rap1b-deficient mice. *Blood*. 2008; 111:2647–2656. [PubMed: 17993608]
30. Awasthi A, Samarakoon A, Chu H, Kamalakannan R, Quilliam LA, Chrzanowska-wodnicka M, Li GCW. Rap1b facilitates NK cell functions via IQGAP1-mediated signalosomes. *The Journal of Experimental Medicine*. 2010; 207:1923–38. [PubMed: 20733035]
31. Hao Y, Wong R, Feig LA, Hao Y, Wong R, Feig LA. RalGDS Couples Growth Factor Signaling to Akt Activation RalGDS Couples Growth Factor Signaling to Akt Activation. *Molecular and Cellular Biology*. 2008; 28:2851–2859. [PubMed: 18285454]
32. Margutti P, Matarrese P, Conti F, Colasanti T, Delunardo F, Garofalo T, Profumo E, Riganò R, Siracusano A, Salvati B, et al. Autoantibodies to the C-terminal subunit of RLIP76 induce oxidative stress and endothelial cell apoptosis in immune-mediated vascular diseases and atherosclerosis. *Blood*. 2008; 111:4559–4570. [PubMed: 17993611]
33. Rossé C, Hatzoglou A, Parrini M-carla, White MA, Chavrier P, Camonis J, Rosse C, Hatzoglou A, Parrini M-carla, White MA, et al. RalB Mobilizes the Exocyst To Drive Cell Migration. *Molecular and Cellular Biology*. 2006; 26:727–734. [PubMed: 16382162]
- 34*. Sánchez-ruiz J, Mejías R, García-belando M, Barber DF, González-garcía A, Gonza A. Ral GTPases Regulate Cell-Mediated Cytotoxicity in NK Cells. *The Journal of Immunology*. 2011; 187:2433–2441. This is the first time that Ral and the exocyst complex have been directly linked to an immune system polarized secretion process. While it may seem obvious that something highly expressed in the immune system that regulates polarized cellular events would regulate polarized immune function, it had not been conclusively shown until it was done here. [PubMed: 21810610]
- 35**. Bodemann BO, Orvedahl A, Cheng T, Ram RR, Ou Y-H, Formstecher E, Maiti M, Hazelett CC, Wauson EM, Balakireva M, et al. RalB and the exocyst mediate the cellular starvation response by direct activation of autophagosome assembly. *Cell*. 2011; 144:253–67. Ral and a part of the exocyst complex are tied to macroautophagy. This has extremely wide-ranging potential ramifications. Since Ral has now been tied to both mTOR and autophagy it may serve as a vital link between this critical kinase and critical cellular process. Moreover it truly brings the Ral family of proteins into the limelight from the perspective of the immune system as autophagy is increasingly being recognized as a vital mechanism of host defense. [PubMed: 21241894]
36. Chien Y, Kim S, Bumeister R, Loo Y-ming, Kwon SW, Johnson CL, Balakireva MG, Romeo Y, Kopelovich L, Gale M, et al. RalB GTPase-Mediated Activation of the I κ B Family Kinase TBK1 Couples Innate Immune Signaling to Tumor Cell Survival. *Cell*. 2006; 127:157–170. [PubMed: 17018283]
37. Stuart LM, Boulais J, Charriere GM, Hennessy EJ, Brunet S, Jutras I, Goyette G, Rondeau C, Letarte S, Huang H, et al. LETTERS A systems biology analysis of the Drosophila phagosome. *Nature*. 2007; 445:95–101. [PubMed: 17151602]
38. Corrotte M, Nguyen APT, Harlay ML, Vitale N, Bader M-F, Grant NJ. Ral isoforms are implicated in Fc gamma R-mediated phagocytosis: activation of phospholipase D by RalA. *Journal of Immunology*. 2010; 185:2942–50.
39. Xu L, Salloum D, Medlin PS, Saqcena M, Yellen P, Perrella B, Foster Da. Phospholipase D Mediates Nutrient Input to Mammalian Target of Rapamycin Complex 1 (mTORC1). *The Journal of Biological Chemistry*. 2011; 286:25477–86. [PubMed: 21622984]

- 40**. Gus-brautbar Y, Johnson D, Zhang L, Sun H, Wang P, Zhang S, Zhang L. Article The Anti-inflammatory TIPE2 Is an Inhibitor of the Oncogenic Ras [Internet]. *Molecular Cell*. 2012 This paper identifies TIPE2 as a regulator of Ral signaling, and thus identifies the Ral pathway as component of inflammation regulation. It additionally provides a strong mechanistic linkage between inflammation and cancer. 10.1016/j.molcel.2012.01.006

Highlights

Discuss the role and recent thought of the role of PI3K/ERK in immunity and inflammation.

Review the differing roles of Rap1a and Rap1b within the immune system.

Discuss the role of the Ral and RalGDS proteins in immunity and inflammation.

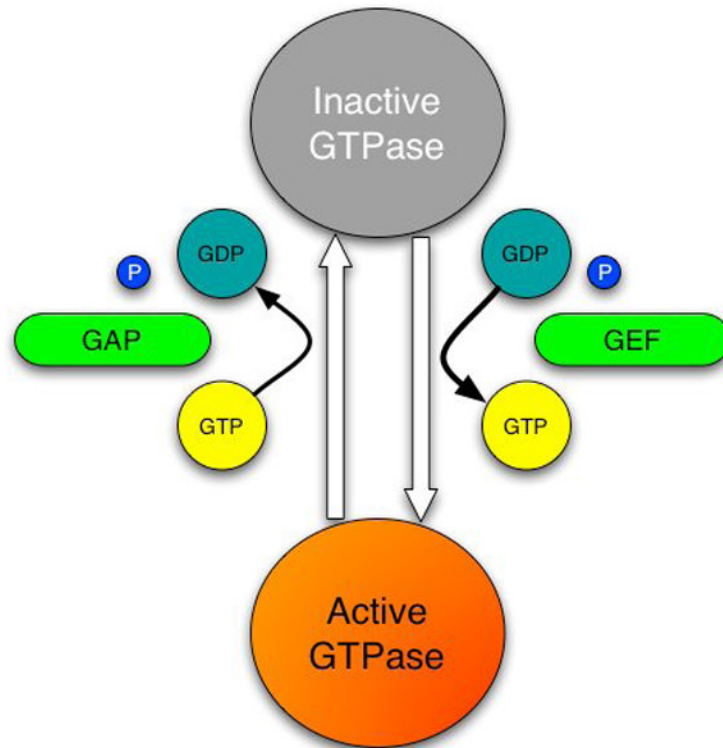


Figure 1. Overview of GTPase signaling

Small GTPase is “off” when bound to GDP; a GEF then removes GDP and allows GTP to bind to the GTPase, turning it “on”. All GTPases can eventually hydrolyze GTP to GDP and turn themselves off, though GAPs rapidly accelerate this process.

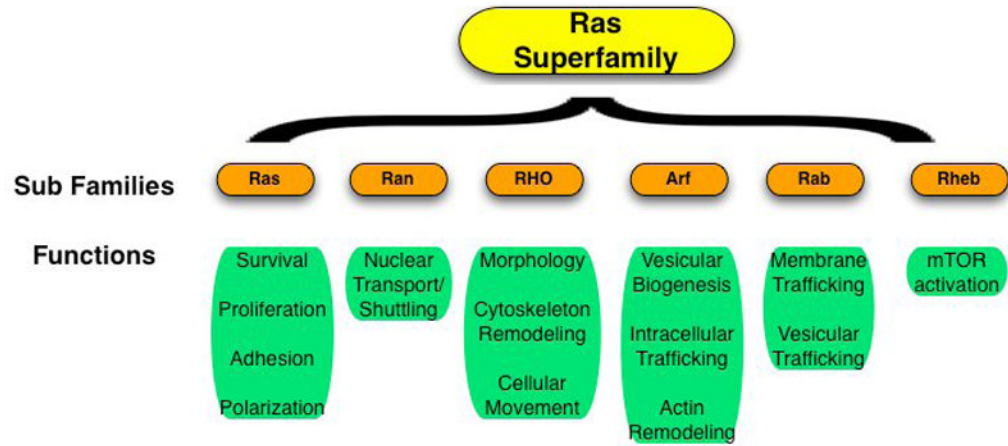


Figure 2. The Ras Superfamily

The Ras superfamily is broken down into 6 families, each listed with their primary functions.

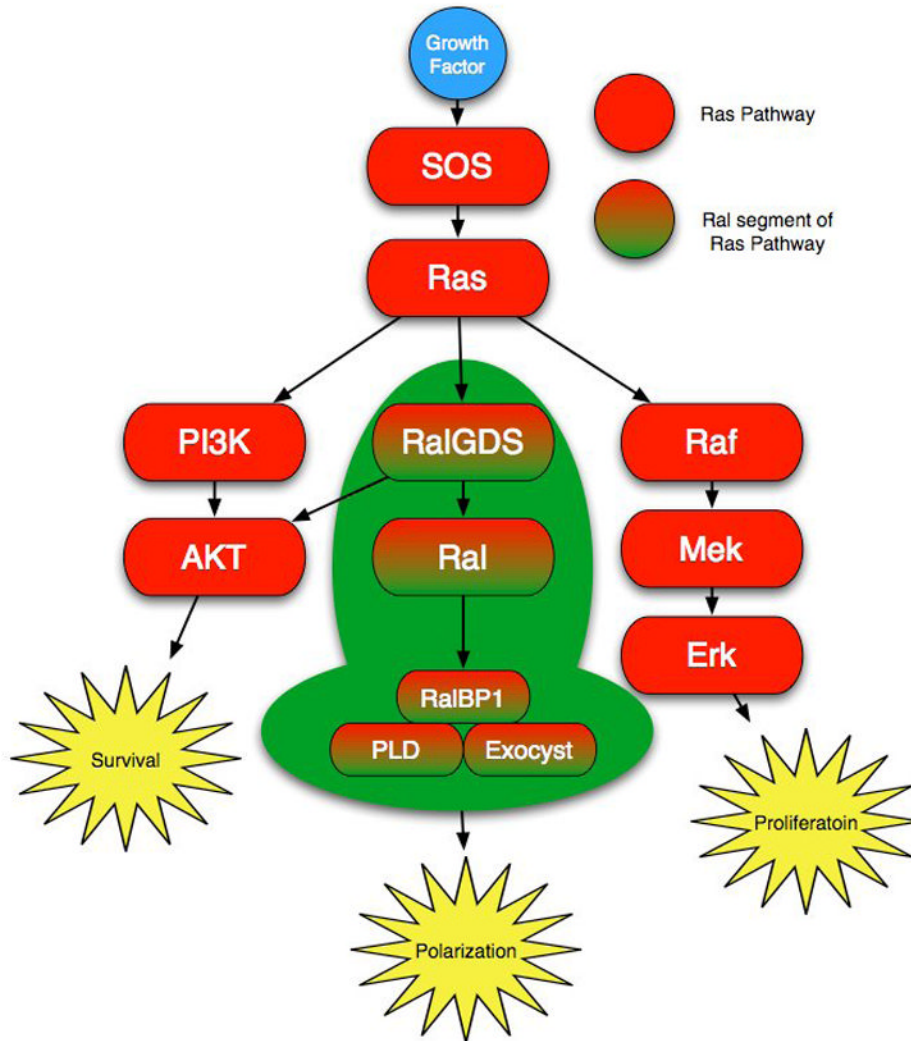


Figure 3. Overview of Ras Signaling

Ras signaling pathway is highlighted in Red. The Ral signaling segment of the Ras pathway is additionally marked in Green.