

NIH Public Access

Author Manuscript

J Acquir Immune Defic Syndr. Author manuscript; available in PMC 2012 June 25

Published in final edited form as: *J Acquir Immune Defic Syndr.* 2005 December 1; 40(4): 494–497.

High Maternal HIV-1 Viral Load During Pregnancy Is Associated With Reduced Placental Transfer of Measles IgG Antibody

Carey Farquhar, MD, MPH^{*,†}, Ruth Nduati, MBChB, MMed, MPH[§], Nancy Haigwood, PhD[¶], William Sutton, MS[¶], Dorothy Mbori-Ngacha, MBChB, MMed, MPH[§], Barbra Richardson, PhD^{‡,#}, and Grace John-Stewart, MD, PhD^{*}

*Department of Medicine, University of Washington, Seattle, WA

[†]Department of Epidemiology, University of Washington, Seattle, WA

[‡]Department of Biostatistics, University of Washington, Seattle, WA

§Department of Paediatrics, University of Nairobi, Nairobi, Kenya

[¶]Seattle Biomedical Research Institute, Seattle, WA

[#]Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, WA

Abstract

Background—Studies among HIV-1–infected women have demonstrated reduced placental transfer of IgG antibodies against measles and other pathogens. As a result, infants born to women with HIV-1 infection may not acquire adequate passive immunity *in utero* and this could contribute to high infant morbidity and mortality in this vulnerable population.

Methods—To determine factors associated with decreased placental transfer of measles IgG, 55 HIV-1–infected pregnant women who were enrolled in a Nairobi perinatal HIV-1 transmission study were followed. Maternal CD4 count, HIV-1 viral load, and HIV-1–specific gp41 antibody concentrations were measured antenatally and at delivery. Measles IgG concentrations were assayed in maternal blood and infant cord blood obtained during delivery to calculate placental antibody transfer.

Results—Among 40 women (73%) with positive measles titers, 30 (75%) were found to have abnormally low levels of maternofetal IgG transfer (<95%). High maternal HIV-1 viral load at 32 weeks' gestation and at delivery was associated with reductions in placental transfer (P < 0.0001 and P = 0.0056, respectively) and infant measles IgG concentrations in cord blood (P < 0.0001 and P = 0.0073, respectively). High maternal HIV-1–specific gp41 antibody titer was also highly correlated with both decreased placental transfer (P = 0.0080) and decreased infant IgG (P < 0.0001).

Conclusions—This is the first study to evaluate the relationship between maternal HIV-1 viremia, maternal HIV-1 antibody concentrations, and passive immunity among HIV-1–exposed infants. These data support the hypothesis that high HIV-1 viral load during the last trimester may impair maternofetal transfer of IgG and increases risk of measles and other serious infections among HIV-1–exposed infants.

Copyright © 2005 by Lippincott Williams & Wilkins

Reprints: Carey Farquhar, University of Washington, 325 Ninth Avenue, Box 359909, Seattle, WA 98104-2499 (cfarq@u.washington.edu).

Written informed consent was obtained from all study participants and the study received ethical approval from the institutional review boards of the University of Washington and the University of Nairobi.

The authors have no commercial or noncommercial associations that might pose a conflict of interest.

Keywords

placental antibody transfer; maternal HIV-1 viral load; measles IgG; HIV-1–specific gp41 antibody; mother-to-child HIV-1 transmission

During the past decade the HIV epidemic has resulted in increased infant mortality in sub-Saharan Africa and has reversed gains made in part through childhood immunization programs. Infants born to HIV-1–infected mothers have a ~2- to 3-fold increased risk of death, and HIV-1–exposed, uninfected infants tend to have higher rates of hospitalization, severe pneumonia, and measles.^{1–4} A Malawian study reported a ~4-fold increased risk of measles among HIV-1–infected infants and a ~2-fold increased risk among HIV-1–exposed uninfected infants compared with infants born to HIV-1–uninfected women.¹ Measles also occurs at a younger age among HIV-1–infection is more likely to result in death with maternal HIV-1 infection regardless of infant HIV-1 status.^{5–7}

Maternal immunosuppression and HIV-1 viremia may influence infectious morbidity among infants by reducing passively acquired humoral immunity against important pathogens, including measles. It has previously been reported that placental transfer of IgG against measles virus is lower among HIV-1–infected than HIV-1–uninfected women⁸ and that concentrations of measles IgG prior to immunization are reduced among HIV-1–exposed infants compared with infants born to HIV-1–seronegative mothers.^{9,10} However, determinants of reduced placental transfer among HIV-1–infected women are not well characterized.

In this study, we hypothesized that women with advanced HIV-1 infection would have the greatest reductions in transplacental transfer of measles IgG. In a cohort of HIV-1–infected mothers and their infants, we evaluated the relationship between placental IgG transfer, infant measles antibody concentrations, and several modifiable and nonmodifiable factors, including maternal CD4⁺ T-cell count, HIV-1 viral load, and IgG directed against HIV-1 gp41.

MATERIALS AND METHODS

HIV-1–infected pregnant women were enrolled in a prospective study in Nairobi from 1999–2004 and mother–infant pairs with maternal CD4 counts <200 or >500 cells/µL were selected on baseline immune status and specimen availability.¹¹ At 32 weeks' gestation, HIV-1 RNA levels and CD4 T-cell counts were obtained and at 34 weeks' gestation, short-course oral zidovudine was initiated according to the Thai Centers for Disease Control protocol.¹² Women provided blood at delivery for HIV-1 RNA, measles IgG, and HIV-1– specific gp41 IgG. Infant cord blood was obtained at delivery for measles IgG and HIV-1– specific IgG, and a peripheral blood sample was taken from the neonate at birth and 1 month of age to determine HIV-1 infection status.

Measles IgG antibody concentrations were measured using a commercial enzyme-linked immunosorbent assay (ELISA) kit (Enzygnost, Germany) and HIV-1–specific antibody titers were performed with ELISA and recombinant gp41 (Chemicon International, Temecula, CA), as described elsewhere.¹³ Maternal CD4 counts were measured with flow cytometry (Becton Dickinson, Franklin Lakes, NJ) and HIV-1 RNA viral load quantified in maternal plasma using the Gen-Probe (San Francisco, CA) viral load assay, a transcription-mediated amplification method sensitive for detection of HIV-1 subtypes A, C, and D.¹⁴ Infant plasma specimens were tested with the same Gen-Probe assay, and infant whole

J Acquir Immune Defic Syndr. Author manuscript; available in PMC 2012 June 25.

blood filter paper specimens were assayed for HIV-1 DNA using polymerase chain reaction to amplify HIV-1 *gag* sequences as previously described.¹⁵

HIV-1 viral load data were log transformed and placental transfer was defined as the ratio of infant-to-maternal measles IgG concentrations in blood at delivery for mother–infant pairs with positive maternal antibody titers. Nonparametric statistical tests were used to compare characteristics between women with CD4 counts <200 cells/µL and those with CD4 counts >500 cells/µL. Univariate and multivariate linear regressions were performed to evaluate associations in the cohort, with CD4 T-cell count (<200 vs. >500) included in the model as an indicator variable.

RESULTS

Maternal and infant characteristics for 55 mother–infant pairs are presented in Table 1. Neonates weighed 3 kg on average and were full term; one infant (2%) born to a woman with CD4 <200 cells/µL was HIV-1 infected at birth, with HIV-1 RNA detected in plasma at <48 hours' postpartum. Overall, measles IgG antibody was detected in 40 maternal (73%) and 38 infant (69%) cord blood specimens. Among the 40 mother–infant pairs with positive maternal titers, 30 women (75%) had infant-to-maternal antibody ratios that were below normal (<95%) and median placental transfer for the entire cohort was ~50% of that expected for HIV-1–uninfected women.¹⁶ When comparing women with CD4 counts <200 and those with CD4 counts >500, we found significant differences in maternal HIV-1 viral load at the antenatal and delivery timepoints (P< 0.0001 for both) (Table 1). There were no differences between groups when comparing median maternal or infant measles IgG concentrations, placental transfer, or HIV-1 gp41 IgG midpoint titers (Table 1).

In Table 2, we present associations between maternal and infant measles IgG concentrations, placental antibody transfer, and clinical and laboratory parameters of interest. Maternal measles antibody concentrations did not correlate with HIV-1 viral load at 32 weeks' gestation or at delivery (P = 0.98 and P = 0.83, respectively) (Table 2). There was also no association between maternal measles IgG and HIV-1 gp41 antibody titers (P = 0.13) (Table 2). However, we did observe strong inverse associations between maternal viral load at 32 weeks' gestation and placental transfer and between maternal viral load at 32 weeks' gestation and infant cord blood measles IgG concentrations (P < 0.0001 for both) (Table 2). For every 1-log increase in maternal HIV-1 RNA load, the ratio between infant and maternal IgG was reduced by 44% and infant measles IgG levels were reduced by 8.5 mg/mL. A similar association was found between maternal HIV-1 viral load measured at the time of delivery and both placental transfer (P = 0.0056) and measles antibody concentrations in infant cord blood (P = 0.0073) (Table 2). Women with higher HIV-1–specific gp41 antibody titer at delivery also had reduced maternofetal antibody transfer and lower infant measles IgG concentrations (P = 0.0080 and P < 0.0001, respectively). Delivery HIV-1-specific gp41 titers were positively correlated with maternal viral load at both 32 weeks of pregnancy and at delivery (P = 0.0010 and P = 0.0002, respectively).

When both maternal HIV-1 RNA levels and HIV-1–specific antibody titer were included in a multivariate model, HIV-1 viral load was found to be a strong independent predictor of placental transfer and infant measles antibody levels (P= 0.0002 and P= 0.0008, respectively) (Table 2; see footnote). In multivariate analysis, maternal HIV-1 antibody also remained correlated with infant measles IgG (P= 0.0026) and there was a trend for an association between maternal HIV-1 gp41 antibody titers and placental antibody transfer (P = 0.090) (Table 2; see footnote). Although in univariate analysis there was a trend for women with CD4 <200 to have reduced placental transfer compared with women with CD4 >500 (P= 0.06), no association was found between CD4 group, placental transfer, and infant

J Acquir Immune Defic Syndr. Author manuscript; available in PMC 2012 June 25.

Page 4

IgG in the multivariate model. We were not able to evaluate CD4 count as a continuous variable (a more powerful analysis) because the cohort did not include women with CD4 between 200–500.

DISCUSSION

Maternal antibodies transferred via the placenta play a major role in protecting newborns against measles and other infections. In this study, highmaternal HIV-1 viral load and high HIV-1–specific antibody concentrations were associated with significant reductions in maternofetal transfer of measles IgG. It is not known whether HIV-1 impairs active transport of maternal IgG across placental syncytiotrophoblasts and into fetal circulation. Infections such as malaria can cause pathologic damage to placental tissue and may decrease IgG transport by destroying IgG-specific Fc receptors (hFcRn) that mediate transplacental transfer.¹⁷ There is less evidence that HIV-1 infection results in histologic changes in the placenta, and other explanations have been considered, including competition by HIV-1–specific antibodies for a finite number of hFcRn receptors, such as has been observed with herpes simplex virus, tetanus toxoid, and *Streptococcus pneumoniae*.⁸ Correlation between high maternal HIV-1–specific gp41 antibody and reduced placental IgG transfer in our cohort would be consistent with this mechanism; high concentrations of HIV-1–specific antibodies in maternal blood during the last trimester could block hFcRn receptors, inhibiting active transport of measles antibodies from mother to infant.

Our finding that HIV-1 viral load and HIV-1–specific antibody levels were inversely associated with placental measles antibody transfer suggests that advanced maternal disease may place HIV-1–exposed infants at increased risk for measles infection and poor outcome, irrespective of infant HIV-1 status. It is possible that controlling maternal HIV-1 viremia with effective antiretroviral therapy prior to the last 4 weeks of gestation, the period when the majority of IgG is transferred from mother to child, could influence placental transfer among infants born to HIV-1–infected mothers. Other interventions may also improve HIV-1–exposed infant antibody levels early in life and merit further investigation. These include passive immunization for select, high-risk infants, as well as active immunization at 6 months for all asymptomatic HIV-1–exposed infants.

In conclusion, immunization guidelines from the World Health Organization address reduced maternofetal IgG transfer in the setting of maternal HIV-1 infection and recommend measles immunization at age 6 months, in addition to 9 months, for HIV-1–infected infants.¹⁸ However, this policy has not been widely implemented and does not reduce risk among HIV-1–exposed infants whose status is negative or unknown. We hope data from the current study will raise awareness and promote new approaches to preventing morbidity and mortality in HIV-1–exposed infants.

Acknowledgments

Research was funded by a US National Institutes of Child Health and Development (NICHD) grant (R01 HD 23412), the University of Washington's Center for AIDS Research (CFAR, P30 AI 27757) Immunology Core, and a 2002 developmental grant from CFAR. C. Farquhar was a scholar in the AIDS International Training and Research Program (D43 TW00007) supported by the NIH Fogarty International Center and she currently receives support from NIH grant K23 HD41879. G. John-Stewart is an Elizabeth Glaser Pediatric AIDS Foundation (EGPAF) Scientist and D. Mbori-Ngacha has an EGPAF Leadership Award.

REFERENCES

1. Taha TE, Graham SM, Kumwenda NI, et al. Morbidity among human immunodeficiency virusinfected and -uninfected African children. Pediatrics. 2000; 106:E77. [PubMed: 11099620]

J Acquir Immune Defic Syndr. Author manuscript; available in PMC 2012 June 25.

- Nakiyingi JS, Bracher M, Whitworth JA, et al. Child survival in relation to mother's HIV infection and survival: evidence from a Ugandan cohort study. AIDS. 2003; 17:1827–1834. [PubMed: 12891069]
- 3. Ng'weshemi J, Urassa M, Isingo R, et al. HIV impact on mother and child mortality in rural Tanzania. J Acquir Immune Defic Syndr. 2003; 33:393–404. [PubMed: 12843752]
- 4. Spira R, Lepage P, Msellati P, et al. Natural history of human immunodeficiency virus type 1 infection in children: a five-year prospective study in Rwanda. Mother-to-Child HIV-1 Transmission Study Group. Pediatrics. 1999; 104:e56. [PubMed: 10545582]
- Embree JE, Datta P, Stackiw W, et al. Increased risk of early measles in infants of human immunodeficiency virus type 1-seropositive mothers. J Infect Dis. 1992; 165:262–267. [PubMed: 1730893]
- 6. Oshitani H, Suzuki H, Mpabalwani ME, et al. Measles case fatality by sex, vaccination status, and HIV-1 antibody in Zambian children. Lancet. 1996; 348:415. [PubMed: 8709770]
- Moss WJ, Monze M, Ryon JJ, et al. Prospective study of measles in hospitalized, human immunodeficiency virus (HIV)-infected and HIV-uninfected children in Zambia. Clin Infect Dis. 2002; 35:189–196. [PubMed: 12087526]
- de Moraes-Pinto MI, Almeida ACM, Kenj G, et al. Placental transfer and maternally acquired neonatal IgG immunity in human immunodeficiency virus infection. J Infect Dis. 1996; 173:1077– 1084. [PubMed: 8627057]
- Lepage P, Dabis F, Msellati P, et al. Safety and immunogenicity of high-dose Edmonston-Zagreb measles vaccine in children with HIV-1 infection: a cohort study in Kigali, Rwanda. Am J Dis Child. 1992; 146:550–555. [PubMed: 1621655]
- Perry RT, Mmiro F, Ndugwa C, et al. Measles infection in HIV-infected African infants. Ann N Y Acad Sci. 2000; 918:377–380. [PubMed: 11131730]
- Farquhar C, Rowland-Jones S, Mbori-Ngacha D, et al. Human leukocyte antigen (HLA) B*18 and protection against mother-to-child HIV type 1 transmission. AIDS Res Hum Retroviruses. 2004; 20:692–697. [PubMed: 15307911]
- Shaffer N, Chuachoowong R, Mock PA, et al. Short-course zidovudine for perinatal HIV-1 transmission in Bangkok, Thailand: a randomised controlled trial. Bangkok Collaborative Perinatal HIV Transmission Study Group. Lancet. 1999; 353:773–780. [PubMed: 10459957]
- Mossman SP, Pierce CC, Watson AJ, et al. Protective immunity to SIV challenge elicited by vaccination of macaques with multigenic DNA vaccines producing virus-like particles. AIDS Res Hum Retroviruses. 2004; 20:425–434. [PubMed: 15157361]
- Emery S, Bodrug S, Richardson BA, et al. Evaluation of performance of the Gen-Probe human immunodeficiency virus type 1 viral load assay using primary subtype A, C, and D isolates from Kenya. J Clin Microbiol. 2000; 38:2688–2695. [PubMed: 10878065]
- Panteleeff DD, John G, Nduati R, et al. Rapid method for screening dried blood samples on filter paper for human immunodeficiency virus type 1 DNA. J Clin Microbiol. 1999; 37:350–353. [PubMed: 9889216]
- Cáceres VM, Strebel PM, Sutter RW. Factors determining prevalence of maternal antibody to measles virus throughout infancy: a review. Clin Infect Dis. 2000; 31:110–119. [PubMed: 10913406]
- Galbraith RM, Fox H, Hsi B, et al. The human materno-foetal relationship in malaria. II. Histological, ultrastructural and immunopathological studies of the placenta. Trans R Soc Trop Med Hyg. 1980; 74:61–72. [PubMed: 7001685]
- Moss WJ, Clements CJ, Halsey NA. Immunization of children at risk of infection with human immunodeficiency virus. Bull World Health Organ. 2003; 81:61–70. [PubMed: 12640478]

_
_
~
_
_
_
0
-
~
-
<u> </u>
_
_
_
U U
_
<
_
01
L L
-
_
_
()
0,
0
U
_
~
0
-

TABLE 1

Cohort Characteristics

	W	aternal CD4 <200 (n = 25)	W	aternal CD4 >500 (n = 30)	
Maternal	Z	Median (IQR) or n (%) of Women	Z	Median (IQR) or n (%) of Women	<i>P</i> Value [*]
Age (y)	25	26 (22–29)	30	23 (21–26)	0.26
CD4 ⁺ T-cell count (cells/µL)					
32 weeks of pregnancy	25	151 (108–173)	30	624 (577–869)	<0.0001
HIV-1 RNA viral load (log10 copies/mL)					
32 weeks of pregnancy	24	5.3 (5.1–5.8)	30	4.4 (3.9–5.1)	<0.0001
Delivery	25	5.1 (4.7–5.7)	28	4.0 (3.2-4.6)	<0.0001
Antimeasles IgG antibody status					
Positive titer at delivery $\dot{\tau}$	25	21 (84%)	30	19 (63%)	0.0866
Concentration (mg/mL)	21	16.6 (11–28)	19	11.9 (8.9–22)	0.0930
HIV-1 gp41 IgG (midpoint titers)	25	1900 (1050–3000)	30	800 (308–1950)	0.14
Infant					
Gender (female)	25	15 (60%)	30	15 (50%)	0.36
Birth weight (kg)	25	3.0 (2.7–3.6)	30	3.0 (2.7–3.2)	0.53
Infant gestational age (wk)	25	40 (39–40)	30	39 (38–40)	0.17
Antimeasles IgG antibody status					
Positive infant cord blood titer \ddagger	25	17 (68%)	30	21 (70%)	0.87
Cord blood concentration (mg/mL)	17	8.9 (6.1–16.4)	21	7.3 (5.3–14.3)	0.45
HIV-1 gp41 IgG (midpoint titers)	25	800 (505–1150)	30	615 (312–1275)	0.47
Placental transfer of antimeasles $\lg G^{S}$	21	0.47 (0.18–0.80)	19	0.48 (0.37–1.0)	0.16
*				,	

J Acquir Immune Defic Syndr. Author manuscript; available in PMC 2012 June 25.

Mann–Whitney Utest was performed for continuous variables and Pearson χ^2 test for dichotomous variables.

 $\dot{\tau}_{f}^{2}$ Borderline results were considered negative and placental transfer was not determined for these mothers. Borderline results occurred in 4 women (8%) with CD4 <200 and in 7 women (23%) with CD4 >500.

 \star^{\pm} Borderline results occurred in 1 (4%) and 4 (13%) infants born to mothers with CD4 <200 and CD4 >500, respectively.

 $^{g}_{N}$ Defined as the ratio of infant-to-maternal antimeasles IgG concentrations at delivery.

IQR, interquartile range.

TABLE 2

Correlates of Maternal Measles IgG Concentrations, Infant Measles IgG, and Placental Transfer of Measles IgG for 40 Mother-Infant Pairs With Positive Maternal Antibody Titers^{*}

					Slope [95% CI	(P Value)			
	Ma	ternal Antimes IgG (mg/mL)	sles		Infant Antimea IgG (mg/mL	sles)		Placental Transf Antimeasles Ig	er of G†
Maternal age (y)	-0.23	[-1.1, 0.59]	(0.57)	-0.80	[-1.7, 0.07]	(0.077)	-0.38	[-0.80, 0.04]	(0.66)
Birth weight (kg)	-0.52	[-13.7, 3.3]	(0.22)	-3.4	[-13.2, 6.4]	(0.49)	0.10	[-0.35, 0.54]	(0.67)
Infant gestational age (wk)	-0.55	[-2.7, 1.6]	(0.61)	0.24	[-2.2, 2.6]	(0.84)	0.07	[-0.04, 0.18]	(0.23)
HIV-1 RNA at 32 weeks of pregnancy (log ₁₀ copies/mL)	-0.07	[-4.4, 4.2]	(0.98)	-8.5	[-12, -4.8]	$(<0.0001)^{\ddagger}$	-0.44	[-0.62, -0.25]	$(<0.0001)^{\ddagger}$
HIV-1 RNA at delivery (log ₁₀ copies/mL)	-0.42	[-4.4, 3.5]	(0.83)	-5.6	[-9.3, -1.8]	(0.0056) [§]	-0.26	[-0.44, -0.07]	$(0.0073)^{s}$
Maternal HIV-1 gp41 IgG (log ₁₀ midpoint titers)	-6.0	[-14.0, 1.9]	(0.13)	-14.4	[-21.1, -7.6]	(<0.0001)	-0.53	[-0.91, -0.15]	(0800) ∬
Infant HIV-1 gp41 IgG (log ₁₀ midpoint titers)	-6.5	[-14.5, 1.5]	(0.11)	-10.3	[-18.0, -2.6]	(0.0078)	-0.26	[-0.67, 0.16]	(0.22)
* CD4 count was included in univ	variate an	d multivariate re	sgression	models as	an indicator vari	iable (CD4 <20	00 = 0; C	D4 >500 = 1).	
$\dot{\tau}^{t}$ Defined as the ratio of infant-to	-maternal	antimeasles Ig	G concent	rations at	delivery.				
*									

⁴In multivariate analysis adjusting for maternal HIV-1 gp41 lgG titer, HIV-1 RNA at 32 weeks' gestation was inversely associated with infant antimeasles lgG concentrations and placental transfer (P = 0.0008 and P = 0.0002, respectively).

gIn multivariate analysis adjusting for maternal HIV-1 gp41 lgG titer, HIV-1 RNA at delivery was inversely associated with infant anti measles lgG concentrations and placental transfer (P=0.029 and P= 0.026, respectively).

fin multivariate analysis adjusting for HIV-1 RNA at 32 weeks' gestation, maternal HIV-1 gp41 IgG titer was inversely associated with infant antimeasles IgG concentrations (P= 0.0026) and there was a trend for an inverse association with placental transfer (P = 0.090).

In multivariate analysis adjusting for HIV-1 RNA at 32 weeks' gestation, infant HIV-1 gp41 IgG titer was inversely associated with infant antimeasles IgG concentrations (P = 0.042).