

## Corrections

### NEUROSCIENCE

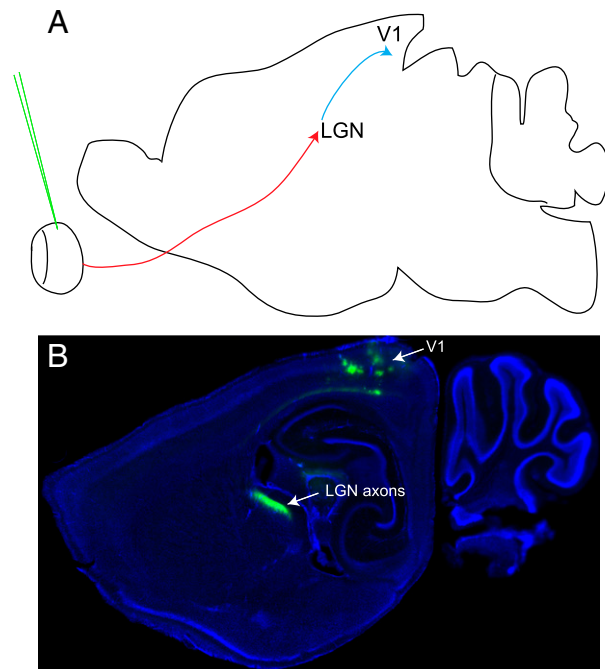
Correction for “Anterograde or retrograde transsynaptic labeling of CNS neurons with vesicular stomatitis virus vectors,” by Kevin T. Beier, Arpiar Saunders, Ian A. Oldenburg, Kazunari Miyamichi, Nazia Akhtar, Liqun Luo, Sean P. J. Whelan, Bernardo Sabatini, and Constance L. Cepko, which appeared in issue 37, September 13, 2011, of *Proc Natl Acad Sci USA* (108:15414–15419; first published August 8, 2011; 10.1073/pnas.1110854108).

The authors note the following: “Shortly after publication of the above manuscript, the replication-competent virus stock, vesicular stomatitis virus (VSV) (LCMV-G), used for the experiments in the paper, was found to be contaminated with wild type VSV [referred to as VSV (VSV-G)]. In addition, the replication-incompetent stock, VSV  $\Delta$ G (LCMV-G) stock, only used for the data in Fig. 3B, was also contaminated by the same wild type VSV. When a pure stock of VSV (LCMV-G) was made, it was found to give inefficient anterograde transmission, in contrast to the efficient anterograde transmission seen with the mixed stock. Most cells infected with the pure VSV (LCMV-G) stock at an initial inoculation site were glia, although the virus did transmit anterogradely to a small number of neurons. The original VSV (VSV-G) stock used for the experiments published in the paper, which contaminated the VSV (LCMV-G) stock, was found to give efficient and specific anterograde transmission. The anterograde transmission was for all injections made directly into the brain, including from the caudate putamen to all of the anterograde target locations published in our paper. Moreover, it did not give retrograde labeling (e.g., see Fig. 2A). However, neither the original VSV (VSV-G) stock, nor an independent VSV (VSV-G) stock obtained from another lab, were found to give anterograde tracing from the eye, or from the nose, to the brain, when inoculated into either of these peripheral locations. The anterograde tracing from the eye or nose to the brain was only seen in our initial studies using the mixed stock, and in our repeated set of experiments with this same mixed stock. We have since plaque purified viruses from this mixed stock. We tested 21 plaque purified viruses for their ability to give anterograde tracing from a peripheral injection site by injecting into the eye and examining the brains. Several stocks from plaque-purified viruses gave such anterograde transmission, and all of these viruses encoded VSV-G only (i.e., not LCMV-G) (for an example, see Fig. A). These same stocks also give the anterograde tracing patterns seen from inoculations of the caudate-putamen (similar to patterns shown in Fig. 3). We are now studying these VSV-G viruses to determine why they are such effective anterograde tracers, and why individual stocks differ when injected into peripheral sites. The differences may be due to the fact that injecting a peripheral site demands long distance travel from the initially infected cells to the brain, or it may have more to do with some other aspect of the sites being peripheral to the brain.

“For those neuroscientists wishing to perform anterograde, polysynaptic tracing, we recommend using the VSV (VSV-G) stock that we have plaque purified. For monosynaptic tracing, the  $\Delta$ G VSV genome can be used with any of the viral G proteins

published in PNAS: VSV-G (for anterograde), LCMV-G (for anterograde), or RABV-G (for retrograde). It may be the case that VSV-G is more efficient for anterograde monosynaptic tracing than LCMV-G. More testing needs to be done, particularly in vivo, to determine the relative efficiencies of these G proteins for monosynaptic tracing.

“We stand by the conclusions of the paper that VSV is an effective transsynaptic tracer, and that the G protein determines the direction of transmission. The RABV-G gives retrograde transmission, while the VSV-G and the LCMV-G direct anterograde transmission, with the VSV-G giving more efficient transmission than LCMV-G as a replication-competent virus.”



**Fig. A.** Anterograde pattern of spread of VSV (VSV-G) from a virus stock derived from a purified plaque. Anterograde transmission was tested by injection into the vitreous body of the eye. (A) A diagram of a parasagittal section of the brain illustrates the expected pattern of transmission for an anterograde transsynaptic virus injected into the eye. It would be expected to label several brain centers involved in visual processing, including the lateral geniculate nucleus (LGN) and visual cortex area 1 (V1). The red arrow indicates the path of the retinal ganglion cell axons to their direct targets, the cells of the LGN. The blue arrow indicates the path of the LGN axons to their V1 targets. The green needle indicates the injection site. (B) A parasagittal section from an injected brain at 7 d post infection, showing labeling of the LGN and V1. All injections that gave brain labeling using this virus stock showed a similar pattern (5/10 animals injected).

[www.pnas.org/cgi/doi/10.1073/pnas.1207087109](http://www.pnas.org/cgi/doi/10.1073/pnas.1207087109)

## CELL BIOLOGY

Correction for “Wnt signaling and a Smad pathway blockade direct the differentiation of human pluripotent stem cells to multipotent neural crest cells,” by Laura Menendez, Tatiana A. Yatskievych, Parker B. Antin, and Stephen Dalton, which appeared in issue 48, November 29, 2011, of *Proc Natl Acad Sci USA* (108:19240–19245; first published November 14, 2011; 10.1073/pnas.1113746108).

The authors note that they omitted a reference to an article by Wang et al. The complete reference appears below.

Additionally, the authors note that on page 19245, left column, third full paragraph, lines 1–6, “WA09 (WiCell), RUES1, RUES2 (A. Brivanlou, The Rockefeller University, New York) hESCs, and the hiPSC lines Fib2-iPS4 and Fib2-iPS5 (George Daley, Children’s Hospital, Boston) were cultured on Geltrex-coated plates (Invitrogen) in chemically defined media containing Heregulin  $\beta$  (10 ng/mL), Activin A (10 ng/mL), LR-Igf (200 ng/mL), and Fgf2 (8 ng/mL) as described previously (24)” should instead appear as “WA09 (WiCell), RUES1, RUES2 (A. Brivanlou, The Rockefeller University, New York) hESCs, and the hiPSC lines Fib2-iPS4 and Fib2-iPS5 (George Daley, Children’s Hospital, Boston) were cultured on Geltrex-coated plates (Invitrogen) in chemically defined media containing Heregulin  $\beta$  (10 ng/mL), Activin A (10 ng/mL), LR-Igf (200 ng/mL), and Fgf2 (8 ng/mL) as described previously (25).”

25. Wang L, et al. (2007) Self-renewal of human embryonic stem cells requires insulin-like growth factor-1 receptor and ERBB2 receptor signaling. *Blood* 110:4111–4119.

[www.pnas.org/cgi/doi/10.1073/pnas.1207810109](http://www.pnas.org/cgi/doi/10.1073/pnas.1207810109)

## APPLIED BIOLOGICAL SCIENCES, ENGINEERING

Correction for “Supramolecular nanostructures that mimic VEGF as a strategy for ischemic tissue repair,” by Matthew J. Webber, Jörn Tongers, Christina J. Newcomb, Katja-Theres Marquardt, Johann Bauersachs, Douglas W. Losordo, and Samuel I. Stupp, which appeared in issue 33, August 16, 2011, of *Proc Natl Acad Sci USA* (108:13438–13443; first published August 1, 2011; 10.1073/pnas.1016546108).

The authors note that the following statement should be added to the Acknowledgments: “Funding was also provided by NIH Grants HL-095874 and P01HL-108795.”

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## MEDICAL SCIENCES

Correction for “Glucocerebrosidase gene-deficient mouse recapitulates Gaucher disease displaying cellular and molecular dysregulation beyond the macrophage,” by Pramod K. Mistry, Jun Liu, Mei Yang, Timothy Nottoli, James McGrath, Dhanpat Jain, Kate Zhang, Joan Keutzer, Wei-Lein Chuang, Wajahat Z. Mehal, Hongyu Zhao, Aiping Lin, Shrikant Mane, Xuan Liu, Yuan Z. Peng, Jian H. Li, Manasi Agrawal, Ling-Ling Zhu, Harry C. Blair, Lisa J. Robinson, Jameel Iqbal, Li Sun, and Mone Zaidi, which appeared in issue 45, November 9, 2010, of *Proc Natl Acad Sci USA* (107:19473–19478; first published October 20, 2010; 10.1073/pnas.1003308107).

The authors note that the author name Wei-Lein Chuang should instead appear as Wei-Lien Chuang. The corrected author line appears below. The online version has been corrected.

**Pramod K. Mistry, Jun Liu, Mei Yang, Timothy Nottoli, James McGrath, Dhanpat Jain, Kate Zhang, Joan Keutzer, Wei-Lien Chuang, Wajahat Z. Mehal, Hongyu Zhao, Aiping Lin, Shrikant Mane, Xuan Liu, Yuan Z. Peng, Jian H. Li, Manasi Agrawal, Ling-Ling Zhu, Harry C. Blair, Lisa J. Robinson, Jameel Iqbal, Li Sun, and Mone Zaidi**

[www.pnas.org/cgi/doi/10.1073/pnas.1207533109](http://www.pnas.org/cgi/doi/10.1073/pnas.1207533109)

## BIOPHYSICS AND COMPUTATIONAL BIOLOGY, CHEMISTRY

Correction for “Ligand binding to protein-binding pockets with wet and dry regions,” by Lingle Wang, B. J. Berne, and R. A. Friesner, which appeared in issue 4, January 25, 2011, of *Proc Natl Acad Sci USA* (108:1326–1330; first published January 4, 2011; 10.1073/pnas.1016793108).

The authors note that their conflict of interest statement was omitted during publication. The authors declare that R.A.F. is a founder of Schrodinger, Inc.

[www.pnas.org/cgi/doi/10.1073/pnas.1207504109](http://www.pnas.org/cgi/doi/10.1073/pnas.1207504109)