

Determinants of reproductive success across sequential episodes of sexual selection in a firefly

A. South* and S. M. Lewis

Department of Biology, Tufts University, Medford, MA 02155, USA

Because females often mate with multiple males, it is critical to expand our view of sexual selection to encompass pre-, peri- and post-copulatory episodes to understand how selection drives trait evolution. In *Photinus* fireflies, females preferentially respond to males based on their bioluminescent courtship signals, but previous work has shown that male paternity success is negatively correlated with flash attractiveness. Here, we experimentally manipulated both the attractiveness of the courtship signal visible to female *Photinus greeni* fireflies before mating and male nuptial gift size to determine how these traits might each influence mate acceptance and paternity share. We also measured pericopulatory behaviours to examine their influence on male reproductive success. Firefly males with larger spermatophores experienced dual benefits in terms of both higher mate acceptance and increased paternity share. We found no effect of courtship signal attractiveness or pericopulatory behaviour on male reproductive success. Taken together with previous results, this suggests a possible trade-off for males between producing an attractive courtship signal and investing in nuptial gifts. By integrating multiple episodes of sexual selection, this study extends our understanding of sexual selection in *Photinus* fireflies and provides insight into the evolution of male traits in other polyandrous species.

Keywords: bioluminescence; courtship signal; Lampyridae; mating success; paternity success; spermatophore

1. INTRODUCTION

One of the principal forces driving the evolution of morphological, behavioural and physiological traits is sexual selection. This evolutionary phenomenon was first described by Darwin [1], who originally conceived this as a selective force that arises from differential mating success owing to intrasexual competition or intersexual choice. However, molecular methods of ascertaining paternity have revealed that females commonly mate with multiple males [2–5]. Therefore, a male's reproductive success is determined by his ability to compete for and court females, to successfully mate, and to maintain paternity share when competing with other mating males. Morphological and behavioural traits traditionally considered as courtship signals might influence not only mating success, but also subsequent selection episodes such as male paternity success. Thus, a complete understanding of how sexual selection can drive trait evolution within polyandrous mating systems requires an integrative approach that encompasses courtship, pericopulatory (immediately before and during mating) and post-copulatory sexual selection episodes.

Different predictions have been made for the relationship between traits affecting male success across distinct episodes of selection. The phenotype-linked fertility hypothesis [6] predicts a positive association between male traits that mediate fitness across different selective episodes, and this relationship has some empirical

support [7–10]. This could arise from a positive association between a male's courtship signals and his fertilizing ability [11], or through reinforcement of initial female mating preferences via cryptic female choice [12]. Other work proposes a negative relationship between male success during pre- and post-copulatory sexual selection, which could be owing to trade-offs among male traits [13,14] or to sexual conflict [15]. Additional work is clearly needed to improve our understanding of how particular traits mediate male success during sequential episodes of sexual selection.

Across many animal taxa, males provide nuptial gifts to females during courtship and mating [16–18] and these gifts can potentially influence male paternity share. In many insects, males transfer their sperm in spermatophores, biochemically diverse packages that have been shown to influence male reproductive success [19–23]. Recent meta-analyses also show that spermatophore gifts can increase female fecundity [24,25]. Thus, variation in male nuptial gifts is likely to be an important factor influencing episodes of sexual selection.

Fireflies (Coleoptera: Lampyridae) are an especially interesting group for investigating how male traits influence reproductive success across distinct selection episodes. In *Photinus* fireflies, precopulatory sexual selection is based on a bioluminescent flash dialogue between flying males and stationary females [26] (reviewed in Lewis & Cratsley [27]). Females preferentially give flash responses to particular males based on temporal characteristics of male courtship signals [28–30]. Males that elicit higher response rates from females can locate females more quickly, and thus have higher mating success [31]. Furthermore, both sexes mate multiple times over their approximately two

* Author for correspondence (adamwsouth@gmail.com).

Electronic supplementary material is available at <http://dx.doi.org/10.1098/rspb.2012.0370> or via <http://rspb.royalsocietypublishing.org>.

week adult lifespan [32,33], and therefore a male's reproductive success will depend on both his mating and his paternity success. *Photinus* males produce an elaborate spermatophore that is transferred to females during mating [34]. Male-derived proteins are subsequently incorporated into developing oocytes [35], and females gain a fitness benefit from receiving multiple spermatophores via increased lifetime fecundity [36]. Gift production is costly for males, and spermatophore size declines across successive matings [37]. Males also vary in their post-copulatory reproductive success (based on their paternity share of offspring produced by doubly mated females [14,38]), and this might depend on spermatophore traits.

Previous work in *Photinus greeni* fireflies has demonstrated a negative relationship between a male's precopulatory courtship attractiveness and his subsequent post-copulatory paternity success [14]. Those males that were least attractive to females during courtship interactions nonetheless sired significantly more offspring compared with the most attractive males. Although this was a correlative study, it suggests the possibility of trade-offs between male traits affecting courtship attractiveness and other traits that influence paternity success, such as pericopulatory behaviours, spermatophore size or composition.

In this study, we build upon previous investigations of *P. greeni* fireflies to examine the relative importance of male flash attractiveness and spermatophore size in determining male close-range acceptance by females as well as paternity success in competitive mating situations. We experimentally manipulated flash attractiveness via photic playback, and also altered spermatophore size by manipulating male mating history. By using artificial signals, this experimental design allowed us to eliminate possible within-male trait correlations and isolate the effects of courtship signals and spermatophore-related traits. If male flash signals operate not only in the context of ensuring mating success, but also to increase paternity success, we predicted higher paternity share when females were exposed to more attractive courtship signals before mating. Independently manipulating male mating history allowed us to test the prediction that male paternity success was due to spermatophore-related traits. We predicted a positive relationship between spermatophore size and male reproductive success. In addition to measuring male paternity share, we also recorded pericopulatory behaviours and female mate acceptance after contact. Finally, we examined whether there were changes in sperm quantity between males' first and second spermatophores which might affect paternity success. This design thus allowed us to examine the separate effects of flash signals, pericopulatory behaviours and spermatophore size across multiple episodes of sexual selection. By spanning sequential episodes of sexual selection, this study provides novel insights into the evolution of male traits.

2. MATERIAL AND METHODS

(a) *Study organism and design*

The effects of spermatophore size, flash signal attractiveness and pericopulatory behaviours on mate acceptance and paternity share were determined using *P. greeni* fireflies collected from Lincoln, MA, USA (46°26' N, 71°18' W). After collection (see the electronic supplementary material for details), virgin fireflies were weighed to the nearest 0.01 mg

and maintained under a natural light cycle. Fireflies were housed separately in containers with access to water only, as adults of this species do not feed. Females were randomly allocated to one of four double-mating treatments. Each female was mated once to a male producing a large spermatophore and once to a male producing a small spermatophore after being exposed to a courtship signal that was either attractive or unattractive. All matings were conducted in the laboratory under a natural light cycle.

(b) *Manipulating male flash signals*

The courtship signal of *P. greeni* males consists of paired pulses separated by approximately 1.0–1.5 s [39]. Photic playback experiments using artificial flash signals covering the normal intraspecific range have shown that *P. greeni* females prefer male signals with a shorter interval between the two pulses [30,40,41]. Specifically, paired flashes with a 1.0 s interpulse interval (IPI) regularly elicit response flashes from *P. greeni* females, but females rarely respond to signals with a 1.4 s IPI [30]. Therefore, in the current study, we created artificial courtship signals that were either attractive to females (1.0 s IPI) or unattractive (1.4 s IPI) using a light emitting diode (LED) controlled by a programmable microprocessor. The LED (572 nm, Ledtronics Inc. Torrance, CA, USA) produced flashes that matched the wavelength of male *P. greeni* flashes [42]. Prior to mating, females were exposed to 25 artificial courtship signals that differed only in IPI depending on the treatment; pulse duration was held constant at 80 ms, with 10 s between consecutive signals.

(c) *Manipulating male spermatophore size*

Male spermatophore size was manipulated by controlling male mating history. In many insects, males transfer spermatophores that decrease in size with successive matings (Lepidoptera: [43–45]; Coleoptera: [46,47]; Orthoptera: [48]), a pattern that is especially prevalent in capital breeders such as *Photinus* fireflies. For example, in the related firefly *Photinus ignitus*, spermatophore weight decreases by 36 per cent between a males' first and second matings [37]. Therefore, it is reasonable to assume that *P. greeni* males will produce relatively larger spermatophores during their first mating, and smaller spermatophores when they mate for a second time. In this experiment, we used virgin *P. greeni* males to obtain large (L) spermatophores, and used pre-mated males that were mated again the following night to obtain small (S) spermatophores.

Accompanying these changes in spermatophore size, sperm quantity may also change across successive matings: decreased numbers of sperm have been reported for some taxa [49], while others show increases [50]. Based on the potential for sperm quantity to influence male paternity success, we compared sperm quantity between *P. greeni* males' first versus second spermatophores. Twelve virgin males were each mated with two different virgin females on sequential nights. Each mating was interrupted after 45 min to ensure spermatophore transfer, after which females were frozen in 95 per cent EtOH. Females were dissected and male spermatophores were placed in 10 µl distilled water, then gently opened to ensure that all sperm were released. *Photinus* firefly sperm is packaged into bundles, each containing a fixed number of sperm [34] and sperm bundles were counted under 60× magnification (Olympus BX40, Olympus, Center Valley, PA, USA). Differences in the number of sperm bundles between males' first and second

spermatophores were compared using a paired *t*-test (SPSS v. 18, SPSS Inc., Chicago, IL, USA).

(d) Experimental treatments: female double matings

Male mating success and paternity share were measured when females were mated to two different males on successive nights. Females were assigned to one of four treatments as described below (see the electronic supplementary material, figure S1). For treatment 1, we describe the procedures and introduce the notation used for the remaining three treatments. *Treatment 1.* Night 1: Attractive signal + Large spermatophore. Night 2: Unattractive signal + Small spermatophore ($A + L \rightarrow U + S$; $n = 10$ females). On the first night, these females were shown attractive courtship signals (25 paired flashes with 1.0 s IPI) and were then mated to a large spermatophore male. After 24 h, these females were shown unattractive courtship signals (25 paired flashes with 1.4 s IPI) and were then remated to a male with a small spermatophore. *Treatment 2.* Night 1: Attractive signal + Small spermatophore. Night 2: Unattractive signal + Large spermatophore ($A + S \rightarrow U + L$; $n = 11$ females). *Treatment 3.* Night 1: Unattractive signal + Large spermatophore. Night 2: Attractive signal + Small spermatophore ($U + L \rightarrow A + S$; $n = 11$ females). *Treatment 4.* Night 1: Unattractive signal + Small spermatophore. Night 2: Attractive signal + Small spermatophore ($U + S \rightarrow A + L$; $n = 10$ females).

Thus, comparisons of treatments 1 versus 2 and treatments 3 versus 4 show effects of altering male spermatophore size, while comparing treatments 1 versus 3 and treatments 2 versus 4 shows the effect of altering courtship signals.

Experiments began at approximately 2000 h each night, when each female in a clear plastic container was placed 24 cm from the output LED and exposed to her assigned artificial courtship signal. Females perceived and gave flash responses to these artificial signals. After 25 signal repetitions, a single male was immediately (within 10 s) introduced into the container and placed near the female. In most cases, this prevented the male from emitting any courtship flashes of his own, and almost completely eliminated any courtship dialoguing between the sexes. Once a male contacts a female, he dorsally mounts her and inserts his aedeagus into her genital opening (copulation stage I; [33]). Spermatophore transfer takes place during stage II of copulation, after the male swivels 180° to assume an abdomen-to-abdomen position with the female [34]. Successful copulations (those that reached stage II) were recorded and allowed to terminate naturally (copulations can last up to 8 h; [33]). If stage II copulation did not occur within 15 min, beetles were set aside and checked every 5 min to determine whether mating had occurred.

Female fireflies are known to remate at 24 h intervals in the field [33], so females in all treatments were presented with their second mating opportunity 24 h after their first mating; 77 per cent of females remated at this time. Of the remaining 10 females, nine remated at 48 h after their first mating and one remated at 72 h. We observed a total of 121 male–female pairs of which 28 failed to mate (13 involved virgin females and the remaining 15 were females that had already mated once). Experiments were continued until we obtained a minimum of 10 doubly mated females within each treatment.

Following their second mating, females were maintained in the laboratory on a natural light cycle until their death. Females were provided moss for oviposition only after their second mating (i.e. no egg laying occurred between matings), and eggs were collected at 2 days intervals and placed into

sterile petri dishes with $1 \times$ phosphate-buffered saline. Eggs were incubated at 29°C until hatching, and first instar larvae were collected and frozen in 95 per cent EtOH at –80°C for later DNA extraction and paternity assignment (see below). Males and females were also frozen in 95 per cent EtOH. The total number of larvae that emerged from a given family varied between 0 and 103. Female fecundity (lifetime no. of offspring) was compared between the four treatments with a two-way ANOVA (SPSS, Inc.), with second mating male spermatophore size and courtship signal as fixed factors.

(e) Measuring pericopulatory behaviours

Because they occur in the dark, close-range male–female interactions that happen after contact but before copulation have not, to our knowledge, previously been described for any firefly species. These behaviours were videorecorded with a Sony TRV80 video camera under infrared illumination (Sony Night-shot, Tokyo, Japan). Filming started when males were first placed into the mating arena, and stopped once successful mating had occurred or after 15 min had elapsed. Digitized videos (30 frames per second) were analysed frame-by-frame using iMovie (Apple, Inc., Cupertino, CA, USA) to describe and quantify pericopulatory behaviours of both sexes (described in the electronic supplementary material).

In our behavioural analysis, we included unsuccessful matings only when we observed sex-specific rejection behaviours (see the electronic supplementary material, table S1 and figure S2). We excluded any pairs that failed to make contact and four additional pairs where the male successfully mounted the female but was unable to successfully copulate despite females adopting a receptive posture. We used exact logistic regression to determine how female mate acceptance (yes or no) was affected by male pericopulatory behaviours, spermatophore size and artificial flash attractiveness (each as a categorical predictor) using SAS PROC Logistic (SAS Inc., Cary, NC, USA). We used conditional exact tests in this analysis owing to sparseness of data, as the usual asymptotic methods are unreliable for such datasets [51]. In addition, we assessed whether male pericopulatory behaviours changed between a male's first and second matings using a Goodman–Kruskal test (STAT XACT v. 6, Cytel Inc., Cambridge, MA, USA).

(f) Measuring male paternity share

To determine paternity for offspring produced by doubly mated *P. greeni* females, we used random amplified polymorphic DNA (RAPD) markers [52] following the methods described in Demary & Lewis [14] (see the electronic supplementary material for details). RAPD markers require no prior knowledge of genomic DNA sequence [53] and have been used to assign paternity in multiple taxa when possible sires are known [54,55]. Paternity was determined for each larval offspring of doubly mated females based on the presence of polymorphic bands shared uniquely with either of the two potential fathers.

For females in each treatment, we calculated second-male paternity share (P_2) as the proportion of offspring sired by this male. A total of 650 larvae were genotyped for this study. Some mating treatments had fewer than 10 families because we only included doubly mated females that produced greater than or equal to nine offspring. Final sample sizes were as follows: five families in treatment 1, 11 families in treatment 2, 10 families in treatment 3 and 10 families in treatment 4.

To separately examine the effects of male spermatophore size, courtship signal attractiveness and their interaction on male paternity, we used a generalized linear model approach [56], where the proportion of offspring sired by the second male was modelled using binomial errors and a logit link function using SAS PROC GenMod (SAS Inc.). In addition, we examined whether second-male paternity within each family (P_2) was influenced by other male morphological or behavioural traits. To do this, we used logistic regressions where P_2 was modelled as a binomial response variable (this was possible because 86% of families showed P_2 of either 0 or 1, and the five families showing mixed paternity were assigned the closer P_2 value). Two separate logistic regressions were run with second males' body weight (continuous) and second males' pericopulatory behaviour (categorical) as predictors.

3. RESULTS

(a) Female and male pericopulatory behaviours

Photinus greeni females showed specific behaviours associated with rejecting a male as a mate (electronic supplementary material, table S2), but the likelihood of female rejection was not significantly affected by a male's pericopulatory behaviour (logistic regression, likelihood ratio $\chi^2_6 = 8.79$, $p = 0.1855$). Male pericopulatory behaviours did not change between each male's first and second matings (2×3 tests of association for leg behaviours, Goodman–Kruskal estimate = 0.0058, $p = 0.9$; for antennal behaviours, Goodman–Kruskal estimate = 0.0010, $p = 1.0$).

(b) Spermatophore size and courtship signals: influence on male mating success

The likelihood that female *P. greeni* fireflies would mate with a male differed significantly between treatments, with females significantly more likely to mate with virgin males that had larger spermatophores (figure 1, exact logistic regression, conditional exact test score = 6.11, $p = 0.0152$). Female mating status (virgin versus already mated) did not alter the likelihood of female acceptance (conditional exact test score = 2.40, $p = 0.1310$), and there was no effect of courtship signal attractiveness on the likelihood of female acceptance (conditional exact test score = 0.93, $p = 1.0$). Additionally, there was no interaction between the effects of spermatophore size and courtship signal attractiveness on mating success (conditional exact test score = 6.63, $p = 0.1310$).

(c) Spermatophore size and courtship signals: influence on male paternity share

Photinus greeni males with larger spermatophores sired a significantly greater proportion of females' offspring than did males with small spermatophores (figure 2; generalized linear model, spermatophore size estimate = 2.72, likelihood ratio: $\chi^2_{32} = 12.41$, $p = 0.0004$). This effect was particularly pronounced when males producing large spermatophores were the second ones to mate (treatments 2 and 4; figure 2*b,d*). Within all experimental treatments, paternity showed a strikingly bimodal distribution (figure 2); when they mated with previously mated females, some males in each treatment sired all the subsequent offspring ($P_2 = 1$) while others sired none ($P_2 = 0$). Mixed-paternity broods were seen in only five out of 36 families.

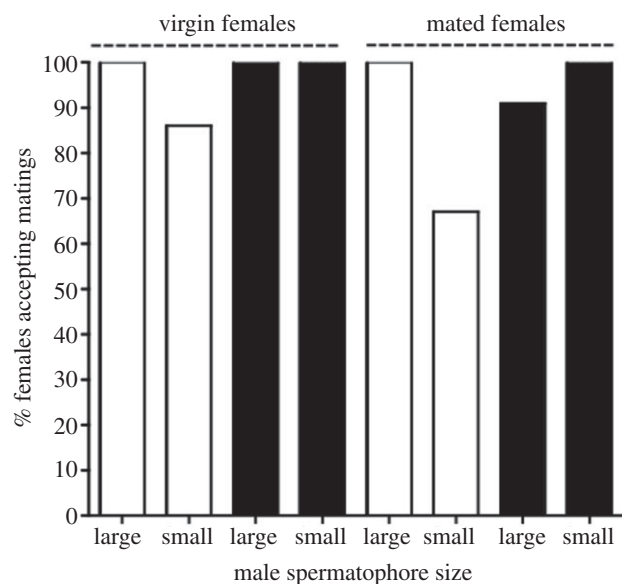


Figure 1. Percentage of females accepting *Photinus greeni* males as mates depending on male spermatophore size (large versus small), artificial flash signal attractiveness (unattractive versus attractive) and female mating status (virgin versus mated). Total number of pairs observed was 102. Open bars, attractive artificial flash; filled bars, unattractive artificial flash.

There was no significant effect of courtship signal attractiveness on male paternity share (figure 2; generalized linear model, courtship signal estimate = 0.153, likelihood ratio: $\chi^2_{32} = 0.03$, $p = 0.8559$), and there was no significant interaction between spermatophore size and signal attractiveness (interaction estimate = -0.936 , likelihood ratio, $\chi^2_{32} = 0.28$, $p = 0.5955$). Also, male paternity share was not influenced by either male body weight (logistic regression; likelihood ratio: $\chi^2_1 = 0.46$, $p = 0.4993$) or male pericopulatory behaviours (logistic regression; likelihood ratio: $\chi^2_5 = 6.19$, $p = 0.1853$).

As expected, because every doubly mated female received one large and one small spermatophore, lifetime offspring production did not vary between experimental treatments (two-way ANOVA, spermatophore size: $F_{1,38} = 0.127$, $p = 0.723$, courtship signal: $F_{1,38} = 0.482$, $p = 0.492$, interaction of spermatophore size and courtship signal: $F_{1,38} = 3.593$, $p = 0.066$).

Sperm quantity declined significantly between *P. greeni* males' first and second spermatophores (figure 3, paired $t_{11} = 12.33$, $p < 0.005$; mean difference \pm s.e. = 75.3 ± 6.1).

4. DISCUSSION

Theoretical models of sperm competition provide different theories for how males might invest into different phases of sexual selection. Parker [57] predicted a trade-off between investment in ejaculate quality and subsequent paternity success versus investment into other reproductive traits that modulate mating success. A number of empirical studies across a broad range of taxa document such a negative relationship [58–61]. However, Sheldon [6] advanced an alternative theory, the phenotype-linked fertility hypothesis. This theory predicts a concordance between investment into ejaculate and secondary sexual traits that

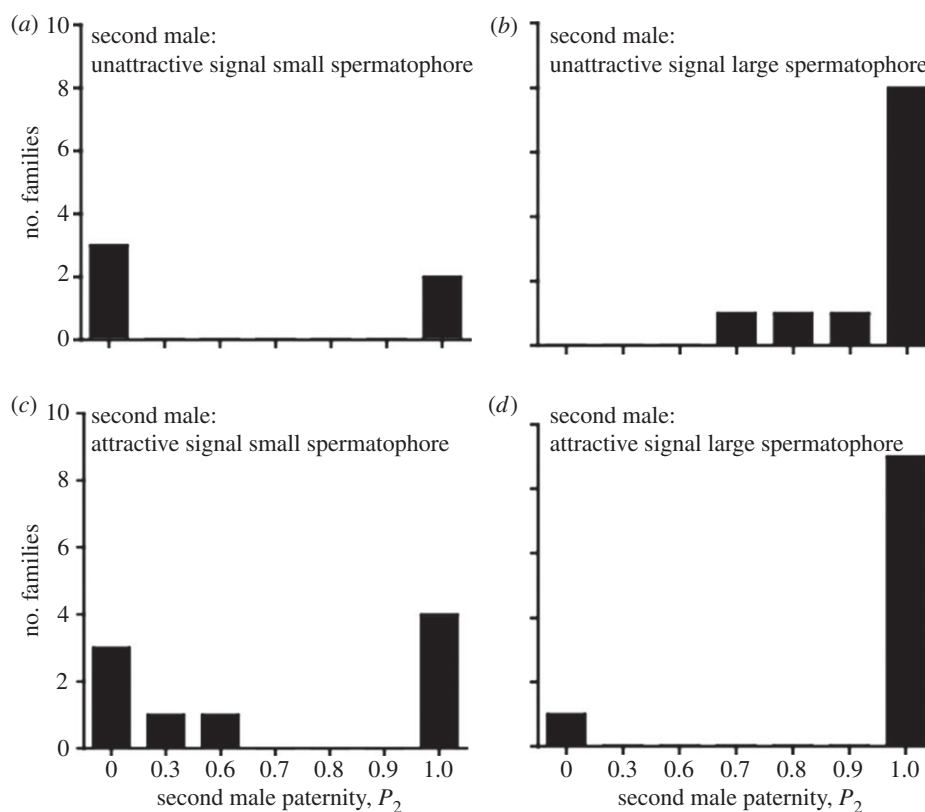


Figure 2. Frequency histograms showing the effect of artificial courtship signal (attractive or unattractive) and spermatophore size (large or small) on second male paternity share (P_2 equals the proportion of offspring produced by doubly mated *Photinus greeni* females that were sired by the second mating male). Treatment descriptions indicate conditions for the second mating male: (a) unattractive courtship signal and small spermatophore; (b) unattractive courtship signal and large spermatophore; (c) attractive flash and small spermatophore; and (d) attractive flash and large spermatophore. Virgin females in all treatments were mated to two males at 24–72 h intervals.

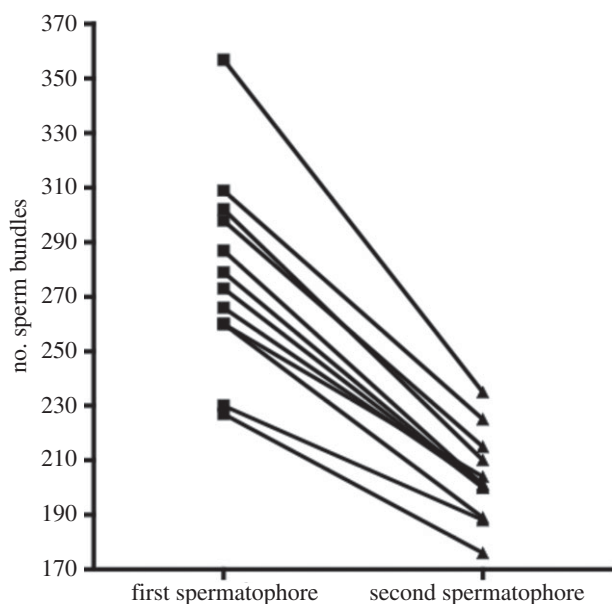


Figure 3. Sperm quantity (number of bundles) contained within spermatophores produced by *Photinus greeni* males during their first versus second matings ($n = 12$ males).

mediate mating success, a model that also has some empirical support [62–64]. Such concordance could arise because male courtship traits could covary with the traits that are responsible for paternity success. For example, male guppies (*Poecilia reticulata*) that are more attractive based on their coloration have greater fertilization success

relative to rivals owing to superior sperm competitive ability [64,65]. However, many such studies are observational or correlative in nature. By experimentally manipulating male traits and including both pericopulatory and post-copulatory sexual selection episodes, our study provides insight into how traits can influence multiple episodes of sexual selection, as well as into potential trade-offs between traits.

(a) Spermatophore size influences male paternity share and pericopulatory success

This study demonstrates that in *P. greeni* fireflies, male spermatophore size positively affects two distinct episodes of sexual selection. Relative to males with smaller spermatophores, males with large spermatophores gained fitness benefits through increased paternity share, and also through their higher likelihood of successfully mating once they contacted a female. A possible mechanism for the effect on paternity share is that larger spermatophores contain more sperm, which could provide a numerical advantage in sperm competition [4]. Males with larger spermatophores had higher paternity share regardless of whether they were a female's first or second mate. Thus, it seems that large spermatophores provide a benefit not only in sperm offence, but also in sperm defence, a pattern also documented in the almond moth, *Cadra cautella* [66].

This study also demonstrated that *P. greeni* males which had not previously mated and thus would transfer relatively larger spermatophores were significantly more likely to be

accepted as mates. Although previous studies have also found that seminal nuptial gifts can influence both male mating success and paternity success [19–23], possible mechanisms for how male spermatophore size might affect mate acceptance are unclear. Although females clearly rejected certain males, female acceptance of *P. greeni* males based on their pericopulatory behaviours seems unlikely, as we found no behavioural differences between males' first and second matings. In the moth *Utetheisa ornatrix*, females choose among males during close-range courtship on the basis of a pheromonal signal that is correlated with chemical defence titres within the male spermatophore [67]. Although similar close-range chemical cues might allow *Photinus* females to distinguish between virgin and previously mated males, studies to date provide no evidence for signalling in *Photinus* fireflies via either cuticular hydrocarbons [68] or volatile pheromones [69]. Because *Photinus* fireflies are chemically defended [70], it is tempting to speculate that firefly pericopulatory mate acceptance might be based on signals correlated with lucibufagin content of male spermatophores, but this remains to be explored.

(b) *Influence of male courtship signals is limited to precopulatory female choice*

Previous work on *Photinus* fireflies has shown that in the field, a male's mating success is determined primarily by how attractive his courtship flash is to females (reviewed by Lewis & Cratsley [27]). In *P. greeni*, females preferentially respond to courtship signals with faster pulse rates by emitting their own response flashes [30]. *Photinus* males use these female response flashes to locate females [26], and males that can elicit more female responses have higher mating success [28]. In the current study, however, we found that *P. greeni* male courtship signals have no direct influence on later sexual selection episodes, as they did not affect either the likelihood of female acceptance after contact or males' paternity share.

Demary & Lewis [14] found a negative relationship in *P. greeni* between a male's attractiveness based on his courtship signal and his subsequent paternity share. To explore this relationship further, the current study used artificial courtship signals to control for other possible differences among males. Because our results show no direct effect of courtship signal on male paternity success, taken together, these findings suggest that males may be subject to energetic trade-offs constraining them either to produce an attractive, fast-pulsed courtship signal or to invest in larger nuptial gifts. This adds to a growing body of evidence suggesting trade-offs between secondary sexual traits that mediate mating success and ejaculate quality (e.g. plumage in red-backed fairy-wrens *Malurus melanocephalus*, [59]; pheromones and dominance behaviour in Australian field crickets; *Teleogryllus oceanicus* [71], and level of sexual ornamentation in guppies *Poecilia reticulata*, [58]).

(c) *Bimodal distribution of paternity share*

In the current study, the vast majority of females produced offspring that were sired solely by either their first mate ($P_2 = 0$) or their second mate ($P_2 = 1$); very few broods showed mixed paternity. Such starkly bimodal distributions of P_2 have now been documented across many taxa (reviewed by Simmons [4]; *Poecilia reticulata* guppies [72]; *Ephippiger ephippiger* bushcrickets [73];

Teleopsis dalmanni stalk-eyed flies [74]; butterflies and moths [75]; and *Cadra cautella* moths [66]). Despite this ubiquity, the mechanisms generating such bimodal paternity share are not well understood. In *T. dalmanni*, Corley *et al.* [74] suggest that differences in male fertility, patterns of sperm usage and ejaculate expenditure as a function of female reproductive value could explain extreme variations in paternity share. A recent study on *Teleogryllus commodus* field crickets [76] suggests a role for both sperm competition and cryptic female choice in determining reproductive success, highlighting the complexity of these post-copulatory interactions.

Bimodality of P_2 could also be influenced by male-derived substances such as seminal fluid proteins, which are important in sperm competition (see [77]). In *P. greeni* fireflies, another mechanism generating bimodal paternity share might involve different sperm storage organs for housing first versus second males' sperm. *Photinus greeni* females have two sperm storage chambers, and differences in stored sperm viability have been documented between these [78]. The idea that females might shunt sperm to different storage sites has been previously suggested as a mechanism for females to retain control over paternity [79,80]. Females that choose their mates based on spermatophore size might gain direct benefits through enhanced fecundity and longevity. If females can assess spermatophore size after mating, they may be able to bias fertilizations towards males with larger spermatophores. Such post-copulatory choice could provide females with indirect benefits, as spermatophore size has been found to be heritable in some insects [81]. Thus, female choice at different episodes could provide either direct or indirect benefits. Additional work is needed to examine how cryptic female choice and sperm competition might interact to determine patterns of paternity.

We are grateful to Gwen Kohl, Katie LeVan, Ray Kameda, Mitch Duffy and Jessica Walden for assistance with field and laboratory work. We thank Quincy Browne and Hillary Cohen for help with the video analysis, Durwood Marshall for expert advice on statistical analysis and two anonymous reviewers for helpful comments on the manuscript. This work was funded by National Science Foundation award no. IOB-0543738.

REFERENCES

- 1 Darwin, C. 1871 *The descent of man and selection in relation to sex*. London, UK: John Murray.
- 2 Smith, R. L. 1984 *Sperm competition and the evolution of animal mating systems*. New York, NY: Academic Press.
- 3 Birkhead, T. R. & Moller, A. P. 1998 *Sperm competition and sexual selection*. London, UK: Academic Press.
- 4 Simmons, L. W. 2001 *Sperm competition and its evolutionary consequences in the insects*. Princeton, NJ: Princeton University Press.
- 5 Birkhead, T. R. & Pizzari, T. 2002 Postcopulatory sexual selection. *Nat. Rev. Genet.* **3**, 262–273. (doi:10.1038/nrg774)
- 6 Sheldon, B. C. 1994 Male phenotype, fertility, and the pursuit of extra-pair copulations by female birds. *Proc. R. Soc. Lond. B* **257**, 25–30. (doi:10.2307/1938476)
- 7 Lewis, S. M. & Austad, S. N. 1994 Sexual selection in flour beetles: the relationship between sperm precedence and male olfactory attractiveness. *Behav. Ecol.* **5**, 219–224. (doi:10.4319/lo.2009.54.6.2025)
- 8 Evans, J. P., Zane, L., Francescato, S. & Pilastro, A. 2003 Directional postcopulatory sexual selection revealed by

- artificial insemination. *Nature* **421**, 360–363. (doi:10.1038/nature01367)
- 9 Wagner, W. E. & Harper, C. J. 2003 Female life span and fertility are increased by the ejaculates of preferred males. *Evolution* **57**, 2054–2066. (doi:10.1111/j.1365-294X.2010.04701.x)
 - 10 Hosken, D. J., Taylor, M. L., Hoyle, K., Higgins, S. & Wedell, N. 2008 Attractive males have greater success in sperm competition. *Curr. Biol.* **18**, R553–R554. (doi:10.1016/j.cub.2008.04.028)
 - 11 Simmons, L. W. & Kotiaho, J. S. 2002 Evolution of ejaculates: patterns of phenotypic and genotypic variation and condition dependence in sperm competition traits. *Evolution* **56**, 1622–1631. (doi:10.4319/lo.1997.42.4.0687)
 - 12 Edvardsson, M. & Arnqvist, G. 2000 Copulatory courtship and cryptic female choice in red flour beetles *Tribolium castaneum*. *Proc. R. Soc. Lond. B* **267**, 559–563. (doi:10.3354/meps07572)
 - 13 Simmons, L. W. & Emlen, D. J. 2006 Evolutionary trade-off between weapons and testes. *Proc. Natl Acad. Sci. USA* **103**, 16 346–16 351. (doi:10.1007/BF00389086)
 - 14 Demary, K. C. & Lewis, S. M. 2007 Male courtship attractiveness and paternity success in *Photinus greeni* fireflies. *Evolution* **61**, 431–439. (doi:10.1111/j.1558-5646.2007.00033.x)
 - 15 Arnqvist, G. & Rowe, L. 2005 *Sexual conflict*. Princeton, NJ: Princeton University Press.
 - 16 Boggs, C. L. 1995 Male nuptial gifts: phenotypic consequences and evolutionary implications. In *Insect reproduction* (eds S. R. Leather & J. Hardie), pp. 215–242. New York, NY: CRC Press.
 - 17 Vahed, K. 1998 The function of nuptial feeding in insects: review of empirical studies. *Biol. Rev.* **73**, 43–78. (doi:10.1017/S0006323197005112)
 - 18 Gwynne, D. T. 2008 Sexual conflict over nuptial gifts in insects. *Annu. Rev. Entomol.* **53**, 83–101. (doi:10.1146/annurev.ento.53.103106.093423)
 - 19 Sakaluk, S. K. 1984 Male crickets feed females to ensure complete sperm transfer. *Science* **223**, 609–610. (doi:10.1098/rspb.2007.1752)
 - 20 Wedell, N. & Arak, A. 1989 The wartbiter spermatophore and its effect on female reproductive output (Orthoptera: Tettigoniidae, *Decticus verrucivorus*). *Behav. Ecol. Sociobiol.* **24**, 117–125. (doi:10.1007/BF00299643)
 - 21 Wedell, N. 1993 Spermatophore size in bushcrickets: comparative evidence for nuptial gifts as a sperm protection device. *Evolution* **47**, 1203–1212. (doi:10.2307/2409986)
 - 22 LaMunyon, C. W. & Eisner, T. 1994 Spermatophore size as determinant of paternity in an arctiid moth (*Utetheisa ornatrix*). *Proc. Natl Acad. Sci. USA* **91**, 7081–7084. (doi:10.1073/pnas.91.15.7081)
 - 23 Wedell, N. & Cook, P. A. 1998 Determinants of paternity in a butterfly. *Proc. R. Soc. Lond. B* **265**, 625–630. (doi:10.1098/rspb.1998.0340)
 - 24 Arnqvist, G. & Nilsson, T. 2000 The evolution of polyandry: multiple mating and female fitness in insects. *Anim. Behav.* **60**, 145–164. (doi:10.1006/anbe.2000.1446)
 - 25 South, A. & Lewis, S. M. 2011 The influence of male ejaculate quantity on female fitness: a meta-analysis. *Biol. Rev.* **86**, 299–309. (doi:10.1111/j.1469-185X.2010.00145.x)
 - 26 Lloyd, J. E. 1966 Studies on the flash communication system in *Photinus* fireflies. *Univ. Mich. Misc. Publ.* **130**, 1–95.
 - 27 Lewis, S. M. & Cratsley, C. K. 2008 Flash signal evolution, mate choice, and predation in fireflies. *Annu. Rev. Entomol.* **53**, 293–321. (doi:10.1146/annurev.ento.53.103106.093346)
 - 28 Branham, M. A. & Greenfield, M. D. 1996 Flashing males win mate success. *Nature* **381**, 745–46. (doi:10.1038/381745b0)
 - 29 Cratsley, C. K. & Lewis, S. M. 2003 Female preference for male courtship flashes in *Photinus ignitus* fireflies. *Behav. Ecol.* **14**, 135–140. (doi:10.1093/beheco/14.1.135)
 - 30 Michaelidis, C., Demary, K. & Lewis, S. M. 2006 Male courtship signals and female signal assessment in *Photinus greeni* fireflies. *Behav. Ecol.* **17**, 329–335. (doi:10.1093/beheco/arj035)
 - 31 Demary, K., Michaelidis, C. & Lewis, S. M. 2006 Firefly courtship: behavioral and morphological predictors of male mating success in *Photinus greeni*. *Ethology* **112**, 485–492. (doi:10.1111/j.1439-0310.2005.01176.x)
 - 32 Wing, S. R. 1985 Prolonged copulation in *Photinus macdermotti* with comparative notes on *Photinus collustrans* (Coleoptera: Lampyridae). *Fla. Entomol.* **68**, 627–634. (doi:10.2307/3494865)
 - 33 Lewis, S. M. & Wang, O. 1991 Reproductive ecology of two species of *Photinus* fireflies (Coleoptera: Lampyridae). *Psyche* **98**, 293–307. (doi:10.1155/1991/76452)
 - 34 van Der Reijden, E., Monchamp, J. & Lewis, S. M. 1997 The formation, transfer, and fate of male spermatophores in *Photinus* fireflies (Coleoptera: Lampyridae). *Can. J. Zool.* **75**, 1202–1205. (doi:10.1139/z97-143)
 - 35 Rooney, J. A. & Lewis, S. M. 1999 Differential allocation of male-derived nutrients in two lampyrid beetles with contrasting life-history characteristics. *Behav. Ecol.* **10**, 97–104. (doi:10.1093/beheco/10.1.97)
 - 36 Rooney, J. A. & Lewis, S. M. 2002 Fitness advantage of nuptial gifts in female fireflies. *Ecol. Entomol.* **27**, 373–377. (doi:10.1046/j.1365-2311.2002.00420.x)
 - 37 Cratsley, C. K., Rooney, J. & Lewis, S. M. 2003 Limits to nuptial gift production by male fireflies, *Photinus ignitus*. *J. Insect Behav.* **16**, 361–370. (doi:10.1023/A:1024876009281)
 - 38 Rooney, J. A. 2000 Male reproductive investment in two fireflies, *Photinus ignitus* and *Ellychmia corrusca*: effects on male and female reproductive success. PhD. thesis, Tufts University, Medford, MA, USA.
 - 39 Lloyd, J. E. 1969 Flashes, behavior and additional species of Nearctic *Photinus* fireflies (Coleoptera: Lampyridae). *Coleop. Bull.* **23**, 29–40.
 - 40 Buck, J. B. & Buck, E. 1972 Photic signaling in the firefly *Photinus greeni*. *Biol. Bull.* **142**, 195–205. (doi:10.2307/1540224)
 - 41 Buck, J. B. & Case, J. F. 1986 Flash control and female dialog repertory in the firefly *Photinus greeni*. *Biol. Bull.* **170**, 176–197. (doi:10.2307/1541802)
 - 42 Case, J. F. 1984 Vision in mating behaviour of fireflies. In *Insect communication* (ed. T. Lewis), pp. 195–222. Orlando, FL: Academic.
 - 43 Oberhauser, K. S. 1988 Male monarch butterfly spermatophore mass and mating strategies. *Anim. Behav.* **36**, 1384–1388. (doi:10.1016/S0003-3472(88)80208-2)
 - 44 Svard, L. & Wiklund, C. 1989 Mass and production rate of ejaculates in relation to monandry/polyandry in butterflies. *Behav. Ecol. Sociobiol.* **24**, 395–402. (doi:10.1007/BF00293267)
 - 45 Royer, L. & McNeil, J. N. 1993 Male investment in the European corn borer, *Ostrinia nubilalis*: impact on female longevity and reproductive performance. *Funct. Ecol.* **7**, 209–215. (doi:10.2307/2389889)
 - 46 Savalli, U. M. & Fox, C. W. 1999 The effect of male mating history on paternal investment, fecundity and female re-mating in the seed beetle, *Callosbruchus maculatus*. *Funct. Ecol.* **13**, 169–177. (doi:10.1046/j.1365-2435.1999.00287.x)
 - 47 Wilson, N., Tufton, T. J. & Eady, P. E. 1999 The effect of single, double and triple matings on the lifetime fecundity of *Callobruchus analis* and *Callobruchus maculatus* (Coleoptera: Bruchidae). *J. Insect Behav.* **12**, 295–306. (doi:10.1023/A:1020883220643)

- 48 Davies, P. M. & Dadour, I. R. 1989 A cost of mating by male *Requena verticalis* (Orthoptera: Tettigoniidae). *Ecol. Entomol.* **14**, 467–469. (doi:10.1111/j.1365-2311.1989.tb00949.x)
- 49 Sturm, R. 2011 The effect of remating on sperm number in the spermatophores of *Teleogryllus commodus* (Gryllidae). *Invert. Biol.* **130**, 362–367. (doi:10.1111/j.1744-7410.2011.00240.x)
- 50 Cook, P. & Wedell, N. 1996 Ejaculate dynamics in butterflies: a strategy for maximizing fertilization success? *Proc. R. Soc. Lond. B* **263**, 1047–1051. (doi:10.1098/rspb.1996.0154)
- 51 Derr, R. E. 2009 Performing exact logistic regression with the SAS system-revised. See <http://support.sas.com/rnd/app/papers/exactlogistic2009.pdf>.
- 52 Welch, J. & McClelland, M. 1990 Fingerprinting genomes using PCR with arbitrary primers. *Nucleic Acids Res.* **18**, 7213–7218. (doi:10.1093/nar/18.24.7213)
- 53 Liu, Z. J. & Cordes, J. F. 2004 DNA marker technologies and their applications in aquaculture genetics. *Aquaculture* **238**, 1–37. (doi:10.1016/j.aquaculture.2004.05.027)
- 54 Goto, S., Miyahara, F. & Ide, Y. 2002 Identification of male parents and half-sib progeny from Japanese black pine (*Pinus thunbergii* Parl.) clonal seed orchard using RAPD markers. *Breed. Sci.* **52**, 71–77. (doi:10.1270/jsbbs.52.71)
- 55 Santolamazza Carbone, S. & Cordero Rivera, A. 2003 Fertility and paternity in the Eucalyptus snout-beetle *Gonipterus scutellatus*: females might benefit from sperm mixing. *Ethol. Ecol. Evol.* **15**, 283–294. (doi:10.1080/08927014.2003.9522673)
- 56 Arnqvist, G. & Danielsson, I. 1999 Postmating sexual selection: the effect of male body size and recovery period on paternity and egg production rate in a water strider. *Behav. Ecol.* **10**, 358–365. (doi:10.1093/beheco/10.4.358)
- 57 Parker, G. A. 1998 Sperm competition and the evolution of ejaculates: towards a theory base. In *Sperm competition and sexual selection* (eds T. R. Birkhead & A. P. Møller), pp. 3–49. San Diego, CA: Academic Press.
- 58 Evans, J. P. 2010 Quantitative genetic evidence that males trade attractiveness for ejaculate quality in guppies. *Proc. R. Soc. B* **277**, 3195–3201. (doi:10.1098/rspb.2010.0826)
- 59 Rowe, M., Swaddle, J. P., Pruett-Jones, S. & Webster, M. S. 2010 Plumage coloration, ejaculate quality and reproductive phenotype in the red-backed fairy-wren. *Anim. Behav.* **79**, 1239–1246. (doi:10.1016/j.anbehav.2010.02.020)
- 60 Pitcher, T. E., Doucet, S. M., Beausoleil, J. M. J. & Hanley, D. 2009 Secondary sexual characters and sperm traits in coho salmon *Oncorhynchus kisutch*. *J. Fish Biol.* **74**, 1450–1461. (doi:10.1111/j.1095-8649.2009.02210.x)
- 61 Thomas, M. L. & Simmons, L. W. 2009 Male dominance influences pheromone expression, ejaculate quality, and fertilization success in the Australian field cricket, *Teleogryllus oceanicus*. *Behav. Ecol.* **20**, 1118–1124. (doi:10.1093/beheco/arp105)
- 62 Ruther, J., Matchhke, M., Garbe, L.-A. & Steiner, S. 2009 Quantity matters: male sex pheromone signals mate quality in the parasitic wasp *Nasonia vitripennis*. *Proc. R. Soc. B* **276**, 3303–3310. (doi:10.1098/rspb.2009.0738)
- 63 Rogers, D. W., Denniff, M., Chapman, T., Fowler, K. & Pomiankowski, A. 2008 Male sexual ornament size is positively associated with reproductive morphology and enhanced fertility in the stalk-eyed fly *Teleopsis dalmanni*. *BMC Evol. Biol.* **8**, 236–242. (doi:10.1186/1471-2148-8-236)
- 64 Pitcher, T. E. & Evans, J. P. 2001 Male phenotype and sperm number in the guppy (*Poecilia reticulata*). *Can. J. Zool.* **79**, 1891–1896.
- 65 Pilastro, A., Evans, J. P., Sartorelli, S. & Bisazza, A. 2002 Male phenotype predicts insemination success in guppies. *Proc. R. Soc. Lond. B* **269**, 1325–1330. (doi:10.1098/rspb.2002.2017)
- 66 McNamara, K. B., Elgar, M. A. & Jones, T. M. 2009 Large spermatophores reduce female receptivity and increase male paternity success in the almond moth, *Cadra cautella*. *Anim. Behav.* **77**, 931–936. (doi:10.1016/j.anbehav.2009.01.007)
- 67 Eisner, T. & Meinwald, J. 1995 The chemistry of sexual selection. *Proc. Natl Acad. Sci. USA* **92**, 50–55. (doi:10.1073/pnas.92.1.50)
- 68 South, A., LeVan, K., Leombruni, L., Orians, C. M. & Lewis, S. M. 2008 Examining the role of cuticular hydrocarbons in firefly species recognition. *Ethology* **114**, 916–924. (doi:10.1111/j.1439-0310.2008.01535.x)
- 69 Lloyd, J. E. 1972 Chemical communication in fireflies. *Environ. Entomol.* **1**, 265–266.
- 70 Eisner, T., Wierner, D. F., Haynes, L. W. & Meinwald, J. 1978 Lucibufagins: defensive steroids from the fireflies *Photinus ignitus* and *P. marginellus* (Coleoptera: Lampyridae). *Proc. Natl Acad. Sci. USA* **75**, 905–908. (doi:10.1073/pnas.75.2.905)
- 71 Thomas, M. L. & Simmons, L. W. 2009 Male dominance influences pheromone expression, ejaculate quality, and fertilization success in the Australian field cricket, *Teleogryllus oceanicus*. *Behav. Ecol.* **20**, 1118–1124. (doi:10.1093/beheco/arp105)
- 72 Evans, J. P. & Magurran, A. E. 2001 Patterns of sperm precedence and predictors of paternity in the Trinidadian guppy. *Proc. R. Soc. Lond. B* **268**, 719–724. (doi:10.1098/rspb.2000.1577)
- 73 Hockham, L. R., Jefferson, A. G. & Ritchie, M. G. 2004 Sperm competition and the level of polyandry in a bush-cricket with large nuptial gifts. *Behav. Ecol. Sociobiol.* **57**, 149–154. (doi:10.1007/s00265-004-0838-x)
- 74 Corley, L. S., Cotton, S., McConnell, E., Chapman, T., Fowler, K. & Pomiankowski, A. 2006 Highly variable sperm precedence in the stalk-eyed fly, *Teleopsis dalmanni*. *BMC Evol. Biol.* **6**, 53. (doi:10.1186/1471-2148-6-53)
- 75 Wedell, N. 2005 Sperm competition in butterflies and moths. In *Insect evolutionary ecology* (eds M. Fellowes, G. Holloway & J. Rolff), pp. 49–81. London, UK: CABI Publishing.
- 76 Hall, M. D., Bussiere, L. F., Demont, M., Ward, P. I. & Brooks, R. C. 2010 Competitive PCR reveals the complexity postcopulatory sexual selection in *Teleogryllus commodus*. *Mol. Ecol.* **19**, 610–619. (doi:10.1111/j.1365-294X.2009.04496.x)
- 77 Avila, F., Sirot, L. K., Laflamme, B. A., Rubinstein, C. D. & Wolfner, M. F. 2011 Seminal fluid proteins: identification and function. *Annu. Rev. Entomol.* **56**, 21–40. (doi:10.1146/annurev-ento-120709-144823)
- 78 Demary, K. C. 2005 Sperm storage and viability in *Photinus* fireflies. *J. Insect Physiol.* **51**, 837–841. (doi:10.1016/j.jinsphys.2005.04.001)
- 79 Hellriegel, B. & Ward, P. I. 1998 Complex female reproductive tract morphology: its possible use in postcopulatory female choice. *J. Theor. Biol.* **190**, 179–186. (doi:10.1006/jtbi.1997.0546)
- 80 Snow, L. S. & Andrade, M. C. B. 2005 Multiple sperm storage organs facilitate female control of paternity. *Proc. R. Soc. B* **272**, 1139–1144. (doi:10.1098/rspb.2005.3088)
- 81 Wedell, N. 2006 Male genotype affects female fitness in a paternally investing species. *Evolution* **60**, 1638–1645.