

Published in final edited form as:

Bioorg Med Chem Lett. 2012 July 15; 22(14): 4593–4598. doi:10.1016/j.bmcl.2012.05.103.

Re-engineering aryl methylcarbamates to confer high selectivity for inhibition of *Anopheles gambiae* vs human acetylcholinesterase

Joshua A. Hartsel^a, Dawn M. Wong^a, James M. Mutunga^b, Ming Ma^a, Troy D. Anderson^b, Ania Wysinski^b, Rafique Islam^c, Eric A. Wong^d, Sally L. Paulson^b, Jianyong Li^e, Polo C. H. Lam^f, Maxim Totrov^f, Jeffrey R. Bloomquist^{b,c}, and Paul R. Carlier^a

^aDepartment of Chemistry, Virginia Tech, Blacksburg, VA, 24061, USA

^bDepartment of Entomology, Virginia Tech, Blacksburg, VA, 24061, USA

^cDepartments of Entomology and Nematology, Emerging Pathogens Institute, University of Florida, Gainesville, FL 32610, USA

^dDepartment of Animal and Poultry Science, Virginia Tech, Blacksburg, VA, 24061, USA

^eDepartment of Biochemistry, Virginia Tech, Blacksburg, VA 24061, USA

^fMolsoft LLC, 11199 Sorrento Valley Road, San Diego, CA, 92121, USA

Abstract

To identify potential human-safe insecticides against the malaria mosquito we undertook an investigation of the structure activity relationship of aryl methylcarbamates inhibitors of acetylcholinesterase (AChE). Compounds bearing a β -branched 2-alkoxy or 2-thioalkyl group were found to possess good selectivity for inhibition of *Anopheles gambiae* AChE over human AChE; up to 530-fold selectivity was achieved with carbamate **11d**. A 3D QSAR model is presented that is reasonably consistent with log inhibition selectivity of 34 carbamates. Toxicity of these compounds to live *Anopheles gambiae* was demonstrated using both tarsal contact (filter paper) and topical application protocols.

Keywords

acetylcholinesterase; species-selectivity; malaria; mosquito; mechanism-based inactivator

Malaria exacts an enormous toll in the developing world, killing nearly 700,000 in 2010, most of whom were children under 5 years old.¹ Malaria therapeutic drugs directed at the parasite *Plasmodium sp.* play a critical role in reducing mortality,² but an important complementary approach involves controlling the vector of the malaria parasite, the

© 2012 Elsevier Ltd. All rights reserved.

Correspondence to: Paul R. Carlier.

Supplementary Material

Synthetic protocols and analytical characterization data for inhibitors; enzyme expression, assay, and mosquito toxicity protocols; and overlays of select inhibitors on the 3D QSAR lipophilic field visualization for *A.g/h* inhibition selectivity may be found in the online version, at doi.xxx.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

mosquito *Anopheles gambiae*. Among vector control measures, the use of insecticide-treated nets (ITNs) has been particularly successful in reducing malaria mortality.³ To date only two biological targets have been successfully exploited in the design of insecticides against adult vector mosquitoes: the voltage-gated Na⁺ ion channel, and acetylcholinesterase (AChE).⁴ Current generation ITNs rely on pyrethroids, which modulate the former target. However, widespread use of these nets has led to the emergence of pyrethroid-resistant mosquitoes, jeopardizing this important disease control measure.⁵

To confront this growing threat we sought to identify a class of insecticidal AChE inhibitors that would only weakly inhibit human AChE (*hAChE*). One intriguing strategy to achieve such selective inhibition relies upon the presence of a free cysteine in the active site gorge of *Anopheles gambiae* AChE (*AgAChE*); human (*hAChE*) has no such free cysteine.⁶ Recently, Pang and co-workers reported that the thiol-reactive compound **1** inhibited >95% of *AgAChE* activity after a 1 h exposure at 6 μM ; under these conditions *hAChE* activity was unchanged.^{6d}

We chose to pursue a different strategy: namely, redesign of the classic insecticidal aryl methylcarbamate pharmacophore⁷ (e.g. **2–5**, Figure 1) to achieve species-selectivity. This goal appeared feasible since *hAChE* is only 49% identical to *AgAChE*.^{6c} In addition, precedent for high species-selectivity for a carbamate is serendipitously provided by the Alzheimer's therapeutic drug rivastigmine **6**, which is 1,500-fold more potent towards *hAChE* than *Torpedo californica* AChE.⁸ In previous work^{6c} we assessed the inhibitory selectivity of several carbamates (including **2**) by measuring IC₅₀ values following a 10 minute incubation. However since carbamates inhibit AChE by covalent modification of the catalytic serine, in this study we used the Ellman Assay⁹ to monitor time-dependent inhibition of the enzyme, by measuring enzyme velocities as a function of incubation time at fixed inhibitor concentrations. These velocities (v/v_0) were used to calculate pseudo first-order rate constants k_{obs} (min^{-1}) for inactivation by plotting $\ln(v/v_0)$ vs incubation time t . For each inhibitor k_{obs} values were determined at three or more inhibitor concentrations ([I]). Plots of k_{obs} vs [I] were constructed and the slope of the linear fit provided the apparent second-order rate constants k_i ($\text{mM}^{-1} \text{min}^{-1}$) for inactivation (See Supplementary Material for more detail).^{8,10} Inhibition of both recombinant *AgAChE* and *hAChE* by commercial carbamate insecticides **2–5** is described in Table 1.

As assessed by k_i ratio, **2** (propoxur) is 16-fold selective for inhibition of *AgAChE*, greater than the 2-fold selectivity we reported previously based on IC₅₀ values.^{6c} However none of these carbamates **2–5** exhibits high (i.e. >100-fold) *Ag/h* selectivity, and thus it is not surprising that to date the World Health Organization has not authorized the use of any carbamate on ITNs.¹¹ We thus undertook the synthesis of a wide range of aryl carbamates to look for structural determinants of *Ag/h* selectivity. In this effort we focused on the use of methylcarbamates since they are known to have favorable insecticidal properties.^{7a} In addition basic or quaternary nitrogen functionality were avoided, since these are known to reduce insecticidal potency.^{7a,7b} One promising series to emerge featured β -branched 2-alkoxy and 2-thioalkyl substituents.¹² These compounds were trivially synthesized by alkylation of catechol (**7**) or 2-thiophenol (**8**), followed by carbamoylation (Scheme 1). *O*-Alkylated products **9a–f** were obtained in low to moderate yield due to competing dialkylation; *S*-alkylation to **10a–d,g** occurred in higher yield. Carbamoylation was achieved in moderate to excellent yield by deprotonation of the phenols **9** and **10** with KO t -Bu and trapping with MeNHC(O)Cl. New compounds **11a–f** and **12d** were recently described by the authors in a patent.¹² Inactivation rate constants for these compounds are given in Table 2.

As can be seen in Table 2, β -branched 2-alkoxyphenyl methylcarbamates **11a–d** are all more selective than **2**, which is an α -branched 2-alkoxyphenyl methylcarbamate. The

highest selectivity, 530-fold, is observed for **11d**, which bears a 2-(2-ethylbutoxy) group. To illustrate the high selectivity of this compound, Figure 2 provides plots of k_{obs} vs [**11d**] at both *Ag*AChE and *h*AChE.

Compounds **11e** and **11f** represent conformationally constrained analogs of **11d**; these modifications dramatically decreased *Ag/h* selectivity, by reducing *Ag*AChE k_i , without significantly affecting the *h*AChE k_i values. Thus on binding to *Ag*AChE, the ethyl groups in **11d** appear to be splayed apart, rather than adopting the *syn*-pentane-like conformation required by **11e–f**. 2-Thioalkylphenyl methylcarbamates **12a–d** also bear unconstrained β -branched alkyl groups, and offer 48- to 130-fold selectivity for inhibition of *Ag*AChE over *h*AChE. The highest selectivity in this series is seen for **12d**, which like **11d** bears a 2-ethylbutyl substituent on the heteroatom. To further assess the scope of isosteric substitution, novel carbamate **15d** was prepared, which features an X = CH₂ spacer unit (Scheme 1). As can be seen in Table 2, carbamate **15d** offers 100-fold selectivity for inhibition of *Ag*AChE over *h*AChE. Reviewing the performance of **11d** and **12d**, it is clear that the replacement of O by S renders **12d** significantly more inhibitory at both *Ag*AChE and *h*AChE. In contrast, for **11d** and **15d**, the replacement of O by CH₂ reduced the *Ag*AChE k_i value 3-fold, but left the *h*AChE k_i value relatively unchanged. But regardless of the spacer unit (O vs. S vs. CH₂) the 2-ethylbutyl group confers 100-fold selectivity for *Ag*AChE. Finally, to assess the effect of minimal 2-substitution on *Ag/h* inhibition potency and selectivity, compounds **12g**, **16** and **17** were prepared, that featured –SMe, –Me and –H in the *ortho*-position. As can be seen, *Ag*AChE k_i values of these compounds decrease dramatically relative to **11a–d**, **12a–d**, and **15d**, but *h*AChE k_i values remain largely unchanged.

In our 2008 report^{6c} we also discussed the inhibition selectivity of 3-substituted aryl methylcarbamates, particularly those bearing *t*-Bu (**20m**) and trialkylsilyl substituents (**20s,t**). To ascertain whether the 3-position had potential to confer high selectivity for *Ag*AChE, a variety of 3-substituted aryl methylcarbamates were prepared, as shown in Scheme 2. 3-Alkylphenols **19l, n–r** were prepared by methylation of the corresponding 2° or 3° benzylic alcohols, using the Shishido protocol (**19n, p**)¹³ or our recent modification of this method (**19l,o,q,r**).¹⁴ These benzylic alcohols **18l,n–r** were in turn prepared from commercially available acetophenones or benzoate esters, by addition of the corresponding Grignard reagents or CF₃SiMe₃.¹⁵ Phenols **19i,j,s,t** were prepared by Li/Br exchange on **21** and trapping with the appropriate electrophile. Phenol **19u** was prepared from **22** by a standard Weinreb amide/Wolff-Kishner approach. 3-Alkoxyphenol **19v** was prepared by alkylation of resorcinol **23**. Phenol **19w** was prepared from **21** by Li/Br exchange, trapping with the required Weinreb amide, and Wolff-Kishner reduction.

As can be seen in Table 3, most of the 3-substituted aryl methylcarbamates do not offer appreciable selectivity for inhibition of *Ag*AChE over *h*AChE. In particular **20s** and **20t** are only about 10-fold selective, as opposed to the previously reported 130-fold selectivity.^{6c} Similarly **20m** is only 12-fold selective rather than the 38-fold figure reported earlier. We trace these variations, and that of **2** mentioned above, to an unanticipated but reproducible effect of our earlier inhibitor dilution protocol, which in this work has been replaced with a protocol delivering a constant final assay concentration of 0.1% (v/v) DMSO.¹⁶ Compounds **20n, p, r, t, v**, and **w** have not been described previously. *Ag*AChE inactivation rate constants k_i are sensitive to the identity of the 3-substituent, varying from 17 (**20h**) to 10,000 mM⁻¹ min⁻¹ (**20l**).¹⁷ In particular, α -branching of the substituent plays an important role: *Ag*AChE k_i increases significantly as the substituent R³ is varied from methyl/1° alkyl (**20h,i,j,u,w**) to 2° alkyl (**20k,l**) or 3° alkyl (**20m–r**). It appears that 2° alkyl groups confer slightly faster inactivation than related 3° alkyl groups (cf. **20k** vs **20m,n**; **20l** vs **20o,p,r**). However, in general a similar structural dependence is seen for *h*AChE k_i values, resulting

in lower *Ag/h* inhibition selectivities than those seen for select 2-substituted aryl methylcarbamates (e.g. **11c**, **11d**, **12d**, **15d**, Table 2). The highest *Ag/h* inhibition selectivity in this class is seen for 3-isopentylphenyl methylcarbamate **20w** (53-fold). We note that the γ -branched isopentyl group of **20u** bears a resemblance to the β -branched alkoxy/thioalkyl and γ -branched 2-ethylbutyl groups that confer selectivity in the 2-substituted aryl methylcarbamate class (Table 2, **11a–d**, **12a–d**, **15d**). Yet interestingly, **20v**, which bears a 2-ethylbutoxy group at the C3 position, is poorly (6-fold) selective.

The high *Ag/h* inhibition selectivities demonstrated by **11c**, **11d**, **12d**, and **15d** (Table 2) must find their origin in differences in the primary sequences of *AgAChE* and *hAChE*, and the possible role of particular amino acid substitutions is actively under study.

But at present we have constructed 3D QSAR models for Log (*AgAChE* k_i), Log (*hAChE* k_i), and Log *AgAChE/hAChE* selectivity. Multiple conformations of all compounds described in Tables 1–3 (except for **20w**) were generated and aligned in 3D using the Atomic Property Fields (APF) method¹⁸ as implemented in ICM.¹⁹ APF molecular alignment accuracy was favorably compared to other methods in a recent benchmarking study.²⁰ Following simplified APF 3D QSAR methodology, the pair-wise ligand APF scores were used to build a partial least square (PLS) models and derive property fields predictive of Log k_i values at both enzymes, as well as Log selectivity. Both enantiomers of chiral compounds **11c**, **12c**, **20l**, and **20p** were included in the initial modeling, and the worst fitting enantiomer in each pair was discarded. Thus the final 3D QSAR model presented below is chiral, and was selected arbitrarily from one of two equally valid enantiomeric solutions. The leave-one-out cross-validated R^2 values of PLS models of Log (*AgAChE* k_i), Log (*hAChE* k_i), and Log *AgAChE/hAChE* selectivity were 0.75, 0.81, and 0.66, respectively. The predicted value of **20w** was then used as an external test in the plot of predicted vs observed Log *AgAChE/hAChE* selectivity in Figure 3. As can be seen, the prediction for **20w** falls fairly close to the regression line (predicted and actual Log selectivities are 1.35 and 1.72, respectively). Because the compounds in Tables 1–3 vary primarily in the structure of the lipophilic 2- or 3- substituents, the lipophilic atomic property fields of the 3D QSAR models are useful to illustrate regions of 3D space which, when occupied by a lipophilic moiety, enhance or reduce inhibition potency or selectivity. The 3D QSAR lipophilic property field visualization for Log selectivity is presented in Figure 4.

Overall, the 3D QSAR lipophilic field indicates that in the subsite where the aromatic ring docks, *AgAChE* has a slightly larger ligand pocket than *hAChE*, so that steric bulk approximately two bonds away from the phenyl core is tolerated by *AgAChE* but not *hAChE*, conferring selectivity. Regions in which lipophilic occupancy improves *Ag/h* inhibition selectivity are colored blue; regions in which lipophilic occupancy decreases inhibition selectivity are colored red. The aligned structure of **20w** is depicted in stick, and the placement of the terminal methyl groups in the selectivity-conferring distal blue region of the 3D QSAR lipophilicity field is evident; note that **20w** is moderately (53-fold) selective. Highly selective (100–500 fold) inhibitors bearing β -branched alkoxy/thioalkyl or γ -branched alkyl groups at the 2-position (e.g. **11c–d**, **12d**, **15d**) were also examined in this way, and in each case the terminal methyl groups fell principally in the distal blue region and did not impinge the distal red region significantly (see Supplementary Material for Figures). In contrast, the cycloalkyl moieties of constrained analogs **11e** and **11f** were found to extend into the distal red region, consistent with the reduced selectivities (36- and 2-fold, respectively; see Supplementary Material). Finally, the low (6-fold) selectivity of **20v**, which bears a β -branched alkoxy substituent at the 3-position, is not well explained by the 3D QSAR lipophilicity field. The predicted selectivity of **20v** is relatively close to the regression line (predicted and actual log selectivities are 1.26 and 0.76, respectively) and the

accuracy of this prediction must be due to contribution from other property fields, likely penalizing the presence of an oxygen atom at C3.

Finally, toxicity of select carbamates to adult *Anopheles gambiae* was assessed using two standard protocols: tarsal contact of live mosquitoes with carbamate-treated filter papers,²¹ and topical application²² of ethanolic solutions of carbamates to the dorsal thorax of anesthetized mosquitoes (Table 4). With regard to the tarsal contact assay, commercial carbamates **2–5** gave LC₅₀ values in the range of 16 – 42 ug/mL, and several of the low-selectivity aryl carbamates prepared in this work offered similar toxicities (**20i,l,m,n,p**). Compounds **20i,m** also proved comparable to **2–5** in the topical application assay. However, the highly species-selective carbamates (**11c, 11d, 12d**) were less toxic than commercial carbamates **2–5** in both tarsal contact and topical application assays.

Although there is no simple relationship between the measured AgAChE k_i value and toxicity to live *Anopheles gambiae*, the expected negative correlations between LC₅₀/LD₅₀ and k_i were seen (see Supplementary Material). In the 2-alkoxyphenyl series (**2, 4, 11b–d**), **4** (carbofuran) is the most rapid inactivator and has the lowest LC₅₀/LD₅₀ values in the tarsal contact and topical application assays. Yet, **2** (propoxur) is 25-fold more toxic than **11d** in the topical application assay, despite their similar AgAChE k_i values (266 ± 9 and 255 ± 12 mM⁻¹ min⁻¹, respectively). Similarly, in the 3-alkylphenyl series (**20h–r,w**), **20i** is the most rapid AgAChE inactivator (AgAChE $k_i = 10,000 \pm 100$ mM⁻¹ min⁻¹), and in the tarsal contact assay it is among the most toxic inhibitors. However **20i**, which is 11-fold slower than **20l** for inactivation of AgAChE, has very similar toxicity to **20l** in the tarsal contact assay. Thus AgAChE inhibitory potency is not the only determinant of mosquito toxicity. Ability to penetrate the mosquito exoskeleton, as well as traditional ADME issues are likely at work. Work to address possible metabolic and transport liabilities of highly selective carbamates continues and will be reported in due course.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We thank the National Institutes of Health (AI082581), the Foundation for the National Institutes of Health (GCGH-1497) through the Grand Challenges in Global Health Initiative, and Virginia Tech (the College of Science, the Department of Chemistry, and Fralin Life Science Institute) for financial support of this work.

References and notes

1. World Malaria Report 2011. The World Health Organization; http://www.who.int/malaria/world_malaria_report_2011/9789241564403_eng.pdf
2. (a) Garner P, Graves PM. Plos Medicine. 2005; 2:287.(b) Burrows JN, Chibale K, Wells TNC. Curr Top Med Chem. 2011; 11:1226. [PubMed: 21401508] (c) Sutherland CJ, Ord R, Dunyo S, Jawara M, Drakeley CJ, Alexander N, Coleman R, Pinder M, Walraven G, Targett GAT. Plos Medicine. 2005; 2:338.(d) Wells TNC, Alonso PL, Gutteridge WE. Nature Rev Drug Disc. 2009; 8:879.
3. (a) Lindblade KA, Eisele TP, Gimnig JE, Alaii JA, Odhiambo F, ter Kuile FO, Hawley WA, Wannemuehler KA, Phillips-Howard PA, Rosen DH, Nahlen BL, Terlouw DJ, Adazu K, Vulule JM, Slutsker L. JAMA. 2004; 291:2571. [PubMed: 15173148] (b) Bayoh MN, Mathias DK, Odiere MR, Mutuku FM, Kamau L, Gimnig JE, Vulule JM, Hawley WA, Hamel MJ, Walker ED. Malaria J. 2010;9.(c) Rietveld, A.; Kurdova-Mintcheva, R. Eliminating Malaria: Learning from the Past, Looking Ahead. World Health Organization; 2011. (d) Lim SS, Fullman N, Stokes A, Ravishankar N, Masiye F, Murray CJL, Gakidou E. Plos Medicine. 2011;8.
4. Nauen R. Public Health - Bayer Environ Sci J. 2006; 18:8.

5. N'Guessan R, Boko P, Odjo A, Knols B, Akogbeto M, Rowland M. *Trop Med Int Health*. 2009; 14:389. [PubMed: 19228349]
6. (a) Radi Z.; Taylor, P. *Toxicology of Organophosphate and Carbamate Compounds*. Gupta, RC., editor. Elsevier; Amsterdam: 2006. p. 161 (b) Pang YP. *Bioorg Med Chem Lett*. 2007; 17:197. [PubMed: 17046256] (c) Carlier PR, Anderson TD, Wong DM, Hsu DC, Hartsel J, Ma M, Wong EA, Choudhury R, Lam PCH, Totrov MM, Bloomquist JR. *Chemico-Biol Interact*. 2008; 175:368. (d) Pang YP, Ekstrom F, Polsinelli GA, Gao Y, Rana S, Hua DH, Andersson B, Andersson PO, Peng L, Singh SK, Mishra RK, Zhu KY, Fallon AM, Ragsdale DW, Brimijoin S. *PLOS One*. 2009; 4:e6851. [PubMed: 19714254]
7. (a) Kolbezen MJ, Metcalf RL, Fukuto TR. *J Agric Food Chem*. 1954; 2:864. (b) Metcalf RL, Fukuto TR. *J Agric Food Chem*. 1967; 15:1022. (c) Metcalf RL. *Bull Wld Hlth Org*. 1971; 44:43.
8. Bar-On P, Millard CB, Harel M, Dvir H, Enz A, Sussman JL, Silman I. *Biochemistry*. 2002; 41:3555. [PubMed: 11888271]
9. Ellman GL, Courtney KD, Andres VJ, Featherstone RM. *Biochem Pharm*. 1961; 7:88. [PubMed: 13726518]
10. Jiang H, Liu S, Zhao P, Pope C. *Insect Biochem Mol Biol*. 2009; 39:646. [PubMed: 19607916]
11. The World Health Organization Pesticide Evaluation Scheme (WHOPES) has approved **2** (propoxur) and **3** (bendiocarb) for spraying on walls and other vertical surfaces inside dwellings, but not for deployment on ITNs: <http://www.who.int/whopes/en/>
12. Carlier, Paul R.; Bloomquist, Jeffrey R.; Paulson, Sally L.; Wong, Eric A. Species-Selective Insecticidal Carbamates for Mosquito Control. U.S. Patent 8,129,428. issued March 6, 2012
13. Tanaka H, Shishido Y. *Bioorg Med Chem Lett*. 2007; 17:6079. [PubMed: 17919904]
14. Hartsel JA, Craft DT, Chen Q-H, Ma M, Carlier PR. *J Org Chem*. 2012; 77:3127. [PubMed: 22394317]
15. Prakash GKS, Panja C, Vaghoo H, Surampudi V, Kultyshev R, Mandal M, Rasul G, Mathew T, Olah GA. *J Org Chem*. 2006; 71:6806. [PubMed: 16930030]
16. Previously, inhibitor stock solutions in DMSO were diluted consecutively with buffer, causing variable final assay DMSO concentrations that were proportionate to the final inhibitor concentration. How the previous dilution protocol led to different results is under study and will be reported in due course; it is noteworthy that the selectivities of some highly selective inhibitors (e.g. **11c**, **11d**) were relatively insensitive to DMSO concentration.
17. The inactivation rate of *AgAChE* by 20l is very high, faster than the rate of inactivation of *AgAChE* by chlorpyrifos oxon (Alout H, Djogbenou L, Berticat C, Chandre F, Weill M. *Comp Biochem Physiol B-Biochem Mol Biol*. 2008; 150:271. [PubMed: 18455457]) and similar to the rate of inactivation of *hAChE* by nerve agents (Bartling A, Worek F, Szinicz L, Thiermann H. *Toxicology*. 2007; 233:166. [PubMed: 16904809]).
18. (a) Totrov M. *Chem Biol Drug Des*. 2008; 71:15. [PubMed: 18069986] (b) Grigoryan A, Kufareva I, Totrov M, Abagyan R. *Journal of Computer-Aided Molecular Design*. 2010; 24:173. [PubMed: 20229197]
19. Abagyan, RA. ICM software manual. Molsoft LLC; San Diego, CA: 2010. <http://www.molsoft.com/man/index.html>
20. Giganti D, Guillemain Hln, Spadoni J-L, Nilges M, Zagury J-Fo, Montes M. *J Chem Inf Model*. 2010; 50:992. [PubMed: 20527883]
21. Guidelines for Testing Mosquito Adulticides for Indoor Residual Spraying and Treatment of Mosquito Nets. World Health Organization; Geneva: 2006. WHO/CDS/NTD/WHOPES/GCDPP/2006.3
22. Pridgeon JW, Pereira RM, Becnel JJ, Allan SA, Clark GC, Linthicum KJ. *J Med Entomol*. 2008; 45:82. [PubMed: 18283946]

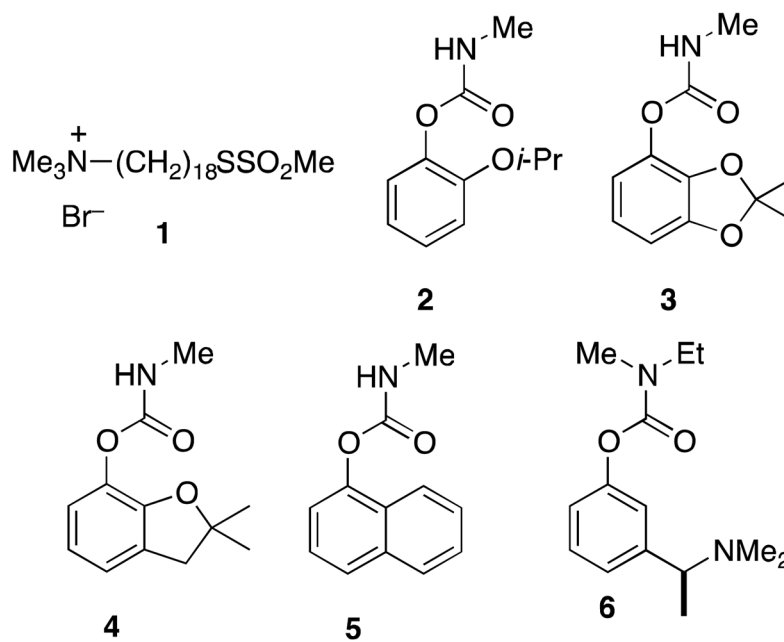


Figure 1. Thiol-reactive *AgAChE*-selective inhibitor **1**, insecticidal aryl methylcarbamates **2–5**, and *hAChE*-selective carbamate rivastigmine **6**.

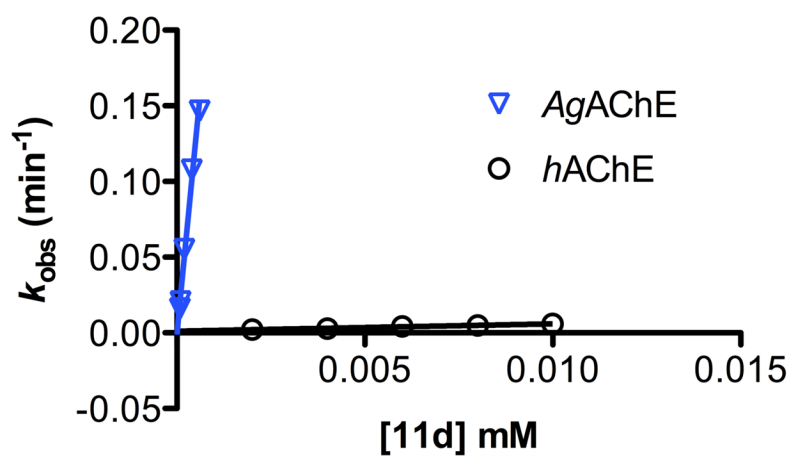


Figure 2. Plot of k_{obs} vs **[11d]** at both AgAChE and hAChE. Second-order rate constants for inactivation k_i ($\text{mM}^{-1} \text{min}^{-1}$) derive from the slope of each line.

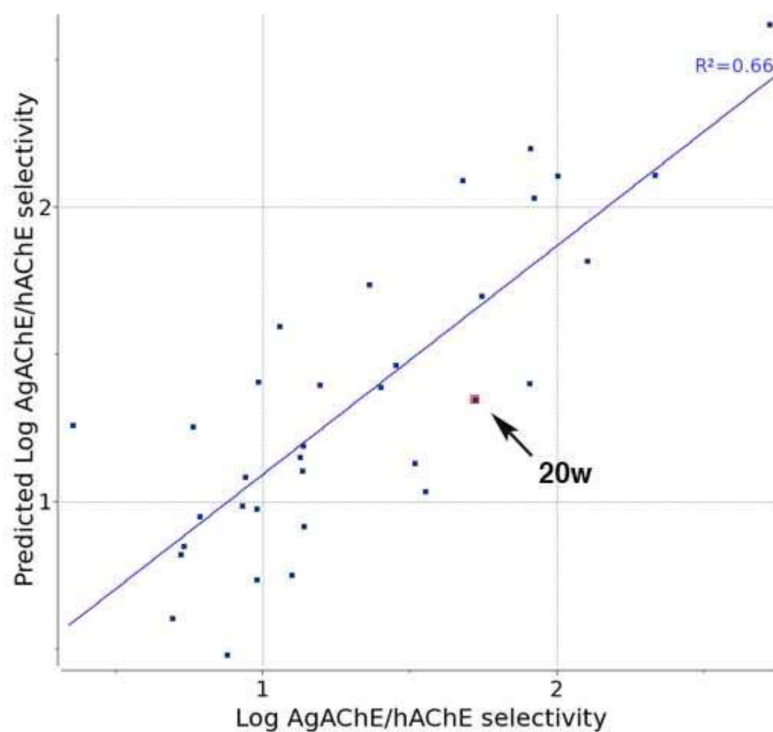


Figure 3. Predicted vs observed Log *AgAChE/hAChE* selectivity of inhibitors in Tables 1–3 except **20w** (33 points); an R^2 value of 0.66 is calculated. Note that prediction for each compound was made with a PLS model trained on a dataset excluding that particular compound, i.e. this is a leave-one-out prediction validation. Subsequently predicted vs. actual Log selectivity was determined for **20w** (indicated by arrow).

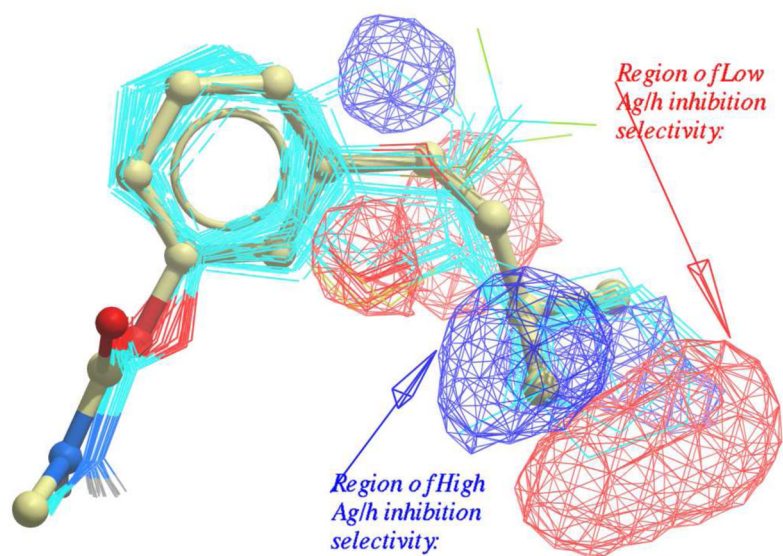
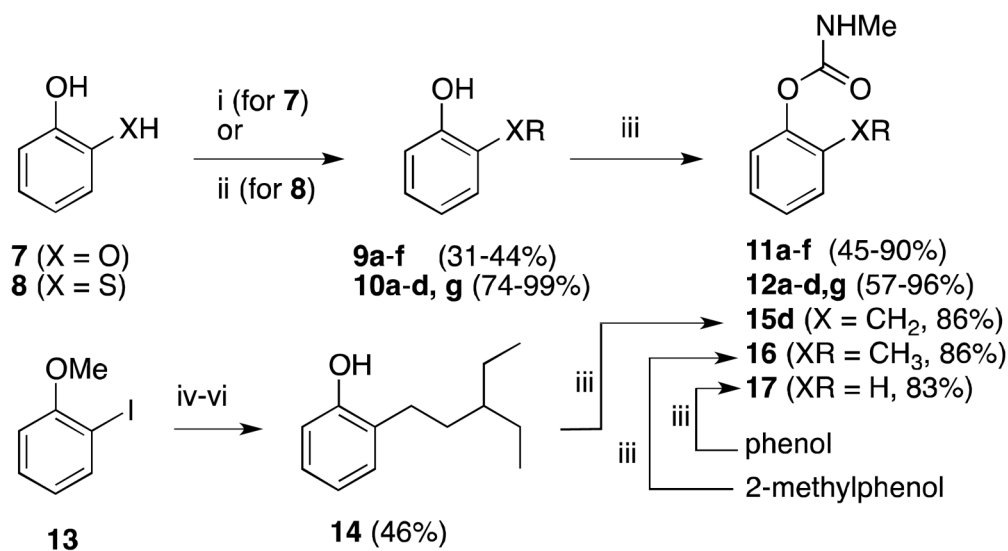


Figure 4. 3D QSAR lipophilic field visualizations for the *Ag/h* inhibition selectivity of the compounds in Tables 1–3. Lipophilic occupancy of blue regions improves *Ag/h* inhibition selectivity, and lipophilic occupancy of red regions decreases selectivity. **20w** is shown in stick; other compounds are shown in CPK-colored wire (carbon is cyan).

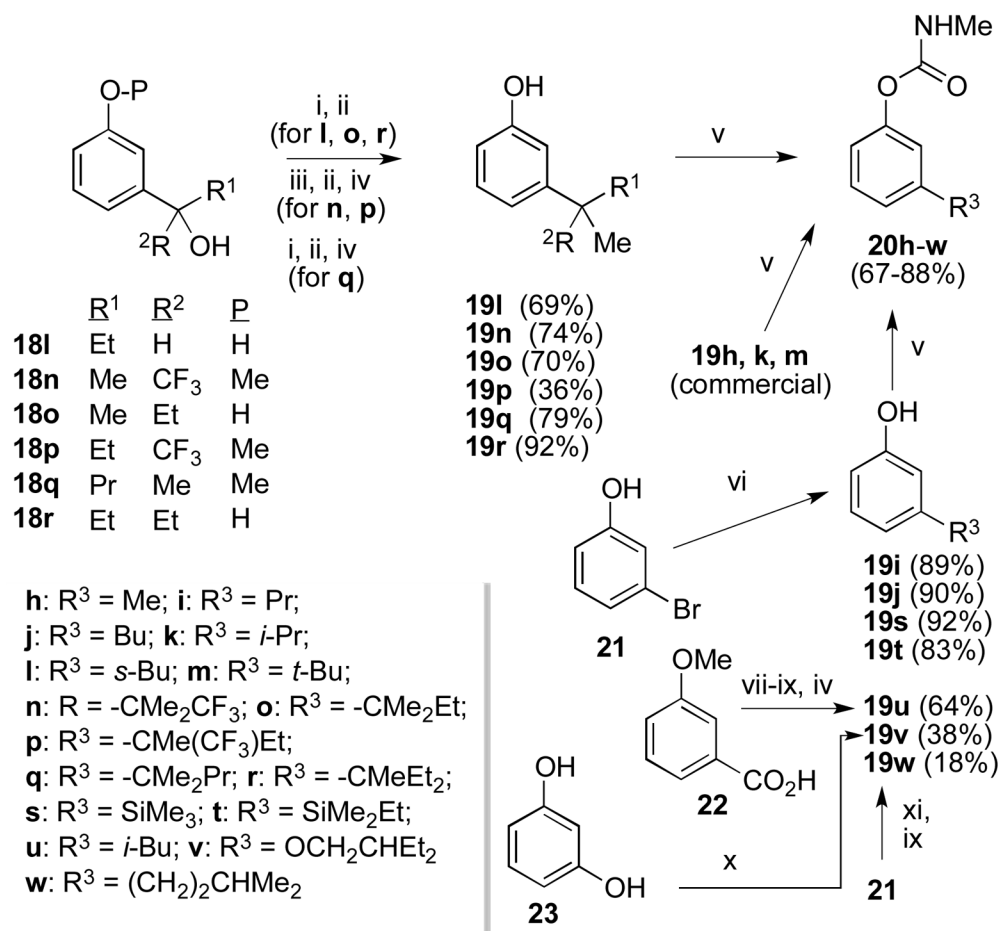


a: R = -CH₂C(Me)=CH₂, **b:** R = -CH₂CHMe₂; **c:** R = (±)-CH₂CHMeEt
d: R = -CH₂CHEt₂; **e:** R = -CH₂c-C₅H₉; **f:** R = -CH₂c-C₆H₁₁; **g:** R = Me

Scheme 1.

Synthesis of 2-substituted aryl methylcarbamates and unsubstituted control **17**

i) Cs₂CO₃, R-Br (or Cl, I, OTs), DMF, 80 °C, 16 h. ii) NaHCO₃, R-Br (or Cl, I, OTs), DMF, 50 °C, 16 h. iii) KO^tBu, THF; MeNHC(O)Cl. iv) *i*-PrMgCl, THF, -78 °C; MeO(Me)NC(O)CH₂CH(Et)₂. v) BBr₃, CH₂Cl₂, 0°C. vi) NaBH₄, CH₃OH; Pd/C, H₂, CH₃OH.

**Scheme 2.**

Synthesis of 3-substituted aryl methylcarbamates **20h-w**.

NHMe i) SOCl₂, 0 °C. ii) AlMe₃, CH₂Cl₂, 0 °C. iii) NaH, THF, MsCl. iv) BBr₃, CH₂Cl₂. v) KO*t*-Bu, THF; CH₃NHC(O)Cl. vi) 2.1 equiv *n*-BuLi, THF, -78 to 0 °C; 2 equiv R³-Br or R₃-Cl (for **19s, t**); aq. HCl. vii) DMAP, EDCl, HN(OMe)Me, CH₂Cl₂. viii) *i*-PrMgCl, Et₂O, reflux. ix) NH₂NH₂, EtOH; KOH, ethylene glycol, 160 °C. x) NaOMe, BrCH₂CHEt₂, MeOH. xi) 2.1 equiv *n*-BuLi, THF, -78 to 0 °C; Me(MeO)NC(O)CH₂CHMe₂.

Table 1*Ag* AChE and *h*AChE inactivation rate constants k_i of commercial agricultural carbamates

| Compound | <i>Ag</i> AChE ^a k_i (mM ⁻¹ min ⁻¹) | <i>h</i> AChE ^b k_i (mM ⁻¹ min ⁻¹) | <i>Ag/h</i> selectivity ^c |
|----------------|---|--|--------------------------------------|
| 2 (propoxur) | 266 ± 9 | 17.0 ± 0.4 | 16 ± 1 |
| 3 (bendiocarb) | 839 ± 22 | 111 ± 5 | 7.6 ± 0.4 |
| 4 (carbofuran) | 2,620 ± 150 | 428 ± 12 | 6.1 ± 0.4 |
| 5 (carbaryl) | 386 ± 10 | 15.4 ± 0.4 | 25 ± 1 |

^aRecombinant *Ag*AChE purified to 2,500 ± 100 U/mg (Supporting Information).^bCommercial recombinant *h*AChE (Sigma C1682).^cDefined as $k_i(\textit{Ag})/k_i(\textit{h})$; error in ratio calculated by standard propagation of error.

A_g AChE and h AChE inactivation rate constants k_i of 2-substituted aryl methylcarbamates **11a–f**, **12a–d**, **g**, **15d**, **16** and unsubstituted control **17**.

Table 2

| Cpd | X | R | A_g AChE ^a k_i (mM ⁻¹ min ⁻¹) | h AChE ^b k_i (mM ⁻¹ min ⁻¹) | A_g/h selectivity ^c |
|------------|-----------------|---|---|---|----------------------------------|
| 11a | O | -CH ₂ CMe=CH ₂ | 62.2 ± 2.0 | 0.74 ± 0.05 | 84 ± 7 |
| 11b | O | -CH ₂ CHMe ₂ | 52.8 ± 2.7 | 0.65 ± 0.20 | 81 ± 5 |
| 11c | O | (±)-CH ₂ CHMeEt | 125 ± 2.8 | 0.58 ± 0.05 | 216 ± 18 |
| 11d | O | -CH ₂ CHEt ₂ | 255 ± 12 | 0.48 ± 0.12 | 530 ± 130 |
| 11e | O | -CH ₂ c-C ₃ H ₉ | 9.8 ± 0.3 | 0.28 ± 0.02 | 36 ± 3 |
| 11f | O | -CH ₂ c-C ₆ H ₁₁ | 0.48 ± 0.02 | 0.21 ± 0.04 | 2.3 ± 0.4 |
| 12a | S | -CH ₂ CMe=CH ₂ | 414 ± 18 | 7.5 ± 0.3 | 55 ± 3 |
| 12b | S | -CH ₂ CHMe ₂ | 526 ± 11 | 6.5 ± 0.2 | 81 ± 3 |
| 12c | S | (±)-CH ₂ CHMeEt | 806 ± 15 | 16.8 ± 0.4 | 48 ± 1 |
| 12d | S | -CH ₂ CHEt ₂ | 1,850 ± 100 | 14.5 ± 1.5 | 130 ± 15 |
| 12g | S | Me | 68.9 ± 1.6 | 5.1 ± 0.3 | 14 ± 1 |
| 15d | CH ₃ | -CH ₂ CHEt ₂ | 75.3 ± 2.7 | 0.75 ± 0.03 | 100 ± 5 |
| 16 | none | -CH ₃ | 1.31 ± 0.07 | 0.24 ± 0.07 | 5.4 ± 1.5 |
| 17 | none | -H | 2.75 ± 0.26 | 0.20 ± 0.06 | 13 ± 4.4 |

^aRecombinant A_g AChE purified to 2,500 ± 100 U/mg (Supporting Information).

^bCommercial recombinant h AChE (Sigma C1682).

^cDefined as $k_i(A_g)/k_i(h)$; error in ratio calculated by standard propagation of error.

Table 3

Apparent bimolecular rate constants k_i for inactivation of *Ag*AChE and *h*AChE by 3-substituted aryl methylcarbamates

| Cpd | R ³ | <i>Ag</i> AChE ^a k_i (mM ⁻¹ min ⁻¹) | <i>h</i> AChE ^b k_i (mM ⁻¹ min ⁻¹) | <i>Ag/h</i> selectivity ^c |
|-----|--|---|--|--------------------------------------|
| 20h | -CH ₃ | 17.2 ± 0.3 | 0.52 ± 0.08 | 33 ± 5 |
| 20i | -Pr | 881 ± 17 | 63.7 ± 1.9 | 14.0 ± 0.5 |
| 20j | -Bu | 741 ± 5 | 32.1 ± 0.6 | 23.0 ± 0.5 |
| 20k | - <i>i</i> -Pr | 3,270 ± 70 | 383 ± 19 | 8.5 ± 0.5 |
| 20l | - <i>s</i> -Bu (±) | 10,000 ± 100 | 1,910 ± 110 | 5.3 ± 0.4 |
| 20m | - <i>t</i> -Bu | 1,510 ± 110 | 126 ± 3 | 12 ± 1 |
| 20n | -CMe ₂ CF ₃ | 1,580 ± 40 | 139 ± 6 | 11 ± 0.6 |
| 20o | -CMe ₂ Et | 7,600 ± 300 | 784 ± 28 | 9.7 ± 0.5 |
| 20p | -CMe(CF ₃)Et (±) | 3,270 ± 70 | 375 ± 6 | 8.7 ± 0.2 |
| 20q | -CMe ₂ Pr | 6,690 ± 240 | 1,360 ± 40 | 4.9 ± 0.2 |
| 20r | -CMeEt ₂ | 5,280 ± 170 | 553 ± 13 | 9.5 ± 0.4 |
| 20s | -SiMe ₃ | 648 ± 29 | 67.8 ± 2.6 | 9.6 ± 0.6 |
| 20t | -SiMe ₂ Et | 1,510 ± 70 | 110 ± 6 | 14 ± 1 |
| 20u | -CH ₂ CHMe ₂ | 155 ± 6 | 5.5 ± 0.30 | 28 ± 2 |
| 20v | -OCH ₂ CHEt ₂ | 28.8 ± 0.4 | 5.0 ± 0.6 | 5.8 ± 0.7 |
| 20w | -(CH ₂) ₂ CHMe ₂ | 272 ± 10 | 5.2 ± 2 | 53 ± 3 |

^aRecombinant *Ag*AChE purified to 2,500 ± 100 U/mg (Supporting Information).

^bCommercial recombinant *h*AChE (Sigma C1682).

^cDefined as $k_i(\text{Ag})/k_i(\text{h})$; error in ratio calculated by standard propagation of error.

Table 4

Toxicity of select carbamates to *Anopheles gambiae* using tarsal contact (filter paper) and topical application protocols.

| Compound | Tarsal Contact to treated filter paper LC ₅₀ ug/mL (95% CI) | Topical application LD ₅₀ ng/mosquito (95% CI) |
|-----------------------|---|--|
| 2 (propoxur) | 39 (32–45) | 3.2 (2.4–4.2) |
| 3 (bendiocarb) | 16 (14–17) | 0.74 (0.52–0.97) |
| 4 (carbofuran) | 16 (11–25) | 0.85 (0.7–1.1) |
| 5 (carbaryl) | 42 (32–55) | 4.1 (3–5) |
| 11b | 290 (271–309) | 19 (12–27) |
| 11c | 445 (396–503) | 33 (24–43) |
| 11d | 27% @ 1,000 ug/mL | 81 (64–94) |
| 12a | 317 (256–389) | 12 (7–19) |
| 12b | 212 (145–279) | 10 (7–14) |
| 12d | 27% @ 1,000 ug/mL | 10 (8–12) |
| 20h | 712 (539–887) | nd |
| 20i | 41 (35–46) | nd |
| 20j | 237 (217–257) | nd |
| 20l | 31 (29–34) | 1.6 (1.4–1.8) |
| 20m | 37 (14–60) | 4.5 (3.6–5.4) |
| 20n | 61 (43–124) | nd |
| 20o | 115 (95–147) | nd |
| 20p | 68 (64–72) | nd |
| 20q | 236 (210–259) | nd |
| 20r | 72% @ 250 ug/mL | nd |
| 20t | 169 (162–176) | 8 (6–10) |
| 20w | 342 (267–472) | nd |

^aMosquitoes were exposed (1 h) to dried filter papers previously treated with ethanolic solutions of carbamates; mortality was recorded after 24 h. LC₅₀ values derive from the concentrations of inhibitor used to treat the papers.

^bEthanolic solutions of the carbamate (0.2 uL) were applied to the dorsal thorax of anesthetized mosquitoes; mortality was recorded after 24 h.