



Review

Functions and physiological roles of two types of estrogen receptors, ER α and ER β , identified by estrogen receptor knockout mouse

Hye-Rim Lee¹, Tae-Hee Kim², Kyung-Chul Choi^{1*}

¹Laboratory of Veterinary Biochemistry and Immunology, College of Veterinary Medicine, Chungbuk National University, Cheongju, Korea

²Department of Obstetrics and Gynecology, College of Medicine, Soonchunhyang University, Bucheon, Korea

Estrogens, a class of steroid hormones, regulate the growth, development, and physiology of the human reproductive system. Estrogens also involve in the neuroendocrine, skeletal, adipogenesis, and cardiovascular systems. Estrogen signaling pathways are selectively stimulated or inhibited depending on a balance between the activities of estrogen receptor (ER) α or ER β in target organs. ERs belong to the steroid hormone superfamily of nuclear receptors, which act as transcription factors after binding to estrogen. The gene expression regulation by ERs is to modulate biological activities, such as reproductive organ development, bone modeling, cardiovascular system functioning, metabolism, and behavior in both females and males. Understanding of the general physiological roles of ERs has been gained when estrogen levels were ablated by ovariectomy and then replenished by treatment with exogenous estrogen. This technique is not sufficient to fully determine the exact function of estrogen signaling in general processes in living tissues. However, a transgenic mouse model has been useful to study gene-specific functions. ER α and ER β have different biological functions, and knockout and transgenic animal models have distinct phenotypes. Analysis of ER α and ER β function using knockout mouse models has identified the roles of estrogen signaling in general physiologic processes. Although transgenic mouse models do not always produce consistent results, they are the useful for studying the functions of these genes under specific pathological conditions.

Key words: Estrogen, estrogen receptors, knockout mice

Received 3 May 2012; Revised version received 21 May 2012; Accepted 25 May 2012

Estrogen receptors (ERs) belong to the steroid hormone superfamily of nuclear receptors (NRs) [1,2]. Other types of steroid receptors among the NRs include the estrogen-related receptors (EER), progesterone receptors (PR), androgen receptors (AR), glucocorticoid, and mineral corticoid receptors. Steroid receptors (SRs), act as ligand-dependent transcription factors, and their activity is associated with the cell cycle [3]. Analyses of the association between steroid receptor activity and the cell cycle revealed that SRs, especially ERs, regulate cell proliferation [4]. Moreover, estrogen is an important sex

hormone produced primarily in the ovaries in females and testes in males. This steroid hormone regulates the growth, development, and physiology of the reproductive system in humans [5]. Estrogen also affects the neuroendocrine, skeletal, adipogenic, and cardiovascular systems [6,7]. The biological functions of estrogen are mediated by binding to the ERs: estrogen receptor alpha (ER α) and estrogen receptor beta (ER β). Estrogen signaling is selectively stimulated or inhibited depending upon a balance between ER α and ER β activities in target organs. ER α was cloned from human breast

*Corresponding author: Kyung-Chul Choi, Laboratory of Veterinary Biochemistry and Immunology, College of Veterinary Medicine, Chungbuk National University, 52 Naesudongro (Gaesin-dong), Cheongju, Chungbuk 361-763, Korea
Tel: +82-43-261-3664; Fax: +82-43-267-3150; E-mail: kchoi@cbu.ac.kr

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

cancer MCF-7 cells in 1986 and ER β was cloned from rat prostate in 1996 [8].

ER α and ER β are encoded by distinct genes located on different chromosomes. The human ER α gene is located on chromosome 6 while the ER β gene is on chromosome 14 [9]. The full-length human ER α protein has 595 amino acids and a molecular size of 66 kDa while the full-length human ER β protein has 530 amino acids and a molecular size of 54 kDa. Similar to other NRs, ERs have five domains with distinct functions [10]. The N-terminal of the A/B domains of ERs consist of activation function 1 (AF1), which contributes to the transcriptional activity of ERs and is an essential domain for interaction with co-regulators. AF1 is the least conserved region with only 30% identity between ER α and ER β . Functional studies have shown that ER β has low levels of AF1 activity. The A/B domains also contain amino acids that are targets of post-transcriptional modifications including splicing to stimulate AF1 activity [9]. The C domain encodes a centrally located DNA binding domain (DBD) essential for sequence-specific binding of ERs to DNA and regulating the expression of target genes [11]. The D domain, a hinge region, includes amino acid sequences that stimulate nuclear localization signaling and facilitate post-translational modification of ERs, resulting in the activation of ER signaling in cells. Finally, the E/F domain, located in the C-terminal region, contains a ligand-binding domain (LBD) that serves as an interaction site with co-regulators and ligand-dependent activation function 2 (AF2). AF1 and AF2 control the transcriptional regulatory activity of ERs because activation of ERs are stimulated during cellular responses to the environment [12]. The E/F domains of ER α and ER β share a 53% sequence identity and affect cellular responses through ligand-dependent ER activation. The F domain also affects the activity of ER α and ER β . The differences between the F domain of the ERs may contribute to the ability of ERs to selectively control transcriptional activities of specific target genes [10].

In the absence of hormones, ERs remain in an inactivation form due to association with heat shock protein 90 (Hsp90). Hsp90 regulates various processes in eukaryotic cells including protein stabilization, binding affinity of receptors to ligands, and signaling cascades [13]. Hsp90 inhibits the degradation of unbound ERs and prevents inactivated ERs from binding ligand [14]. After binding to ligand, ERs are

phosphorylated, form homo- or heterodimers, and then translocate into the nucleus [15]. ERs modulate the transcription of target genes by binding to estrogen response elements (EREs) in the DNA sequence [16]. Binding of ERs to EREs promotes DNA bending and looping, thereby permitting interaction with the transcriptional machinery and co-regulator proteins. These co-regulators include co-activators, co-repressors, co-integrators, histone acetyltransferases and deacetylases, and general transcriptional factors [12]. ER/co-regulator complexes act specifically on target genes in particular organs according to extracellular stimuli [17]. Analysis of ER α and ER β tissue distribution suggests that ERs have high specificity on the target tissue [18]. ER α is highly expressed in the uterus, prostate stroma, ovarian theca cells, Leydig cells in testes, epididymis, breast, and liver [19]. ER β is highly expressed in prostate epithelium, testes, ovarian granulosa cells, bone marrow, and brain [20]. As mentioned above, ER α and ER β have different downstream transcriptional activities, resulting in their tissue-specific biological actions [21].

The reproductive system in ER knockout mice

ER α and ER β have different biological functions. Not surprisingly, eliminating the expression of either factor results in distinct phenotypes in mice. Analysis of ER α and ER β using knockout mouse models has demonstrated the general roles of estrogen signaling. ER α (α ERKO) and ER β (β ERKO) knockout mice have been used to examine the various roles of ER signaling [22,23].

Fertility

Rodent reproductive physiology is associated with steroid hormones, including estrogen. Estrogen is basically known for its effect on the reproduction systems of both females and males [24]. Understanding of the general physiological roles of estrogen has resulted in ovariectomies to ablate estrogen production or restoring estrogen levels with exogenous E2 [25]. Currently, the exact function of estrogen signaling in biological processes is unclear. However, transgenic mice have been useful for discovering gene-specific ER functions [26]. Many experiments with ERKO mice were able to elucidate ERs function in specific pathological conditions, absence of ER expression. [27,28]. Surprisingly, both male and female α ERKO mice are infertile, whereas fertility differs among β ERKO mice according to gender. Female β ERKO mice have reduced fertility

while males have normal fertility [29].

Reproductive organs

Generally, maturation of the reproductive organs promoted by E2 is important for successful pregnancy and appropriate sexual behavior [30]. Infertility in α ERKO female mice is due to a failure to respond to estrogen in the uterus, which is a central organ for reproduction and pregnancy [31]. ER α is essential for uterine maturation but not development [32]. Female α ERKO mice have a decreased number of glands in the endometrium, whereas female β ERKO mice have normal uterine and vaginal tissues despite their reduced fertility rates. Another reason for infertility in the knockout mice is abnormal ovarian function. The ovary is affected by changes in estrogen levels that induce cyclic ovulation [33]. Estrogen stimulates follicular responses to gonadotropins and activities of enzymes such as aromatase for steroidogenesis [34]. The synthesis and secretion to E2 is known for action of follicular stimulating hormone (FSH) on granulosa cells and luteinizing hormone (LH) on theca cell. Both FSH and LH maintain the menstrual cycle and affect ovulation [35]. An abnormal ovarian phenotype causes infertility in α ERKO mice and reduced fertility in β ERKO mice. α ERKO mice appear incapable of ovulation as their follicles fail to differentiate. Ovarian dysregulation is also observed in β ERKO mice including an increased number of unruptured follicles and reduced numbers of oocytes [36].

Mammary gland

In mammals, the mammary gland is important for feeding offspring through the production milk. Development of the mammary gland involves ductal elongation from a nipple and formation of branches of the glands with fat pads. During the estrous cycle, mammary gland development proceeds in a manner similar to that observed during pregnancy through the actions of ERs and PRs [37]. Since α ERKO female mice cannot ovulate, mammary glands of these mice do not develop beyond an immature state, and only contain rudimentary ducts characteristic of embryonic and fetal stages. Insufficient maturation of the mammary gland is caused by a loss of estrogen signaling due to the absence of ER expression. In female α ERKO mice, the release of prolactin from the anterior pituitary gland is reduced and breast tissue for lactation is structurally impaired.

However, the β ERKO female have ability to nurse offspring since they develop normal mammary glands capable of lactation [38].

Male reproduction

The balance of two steroid hormones, androgen and estrogen, plays an important role in male reproductive organs, and is essential for normal prostate development and prostatic homeostasis [39]. However, estrogen signaling also plays a role in maintaining the male reproductive system. ER α is expressed in prostatic tissues during fetal development. ER β expression is initiated after ER α during adulthood. ERs regulate cell growth and differentiation in adult tissues [40]. Thus, an appropriate balance between ER α and ER β is required for normal development of male reproductive tissues. [39]. In male α ERKO mice, the testes are smaller than those in wild-type animals and the fertility rates are significantly reduced [41]. Low fertility in α ERKO male mice is due to reduced sperm counts and low sperm quality. In contrast, the β ERKO males have been shown to produce a sufficient number of sperm to maintain fertility. These findings imply that ER α is more important than ER β for reproduction system development and sperm maturation in male mice.

Bone development and maintenance in ER knockout mice

Bone plays a role in the support of soft organs and maintains calcium level homeostasis in blood. This tissue is continuously remodeled to maintain a suitable length and density for the body. Calcium, vitamin D, and steroid hormones such as estrogen are critical for bone homeostasis [42,43]. These factors control gene expression in bone cells including osteoblasts, osteoclasts, and osteocytes [44]. In addition, various factors are regulated by estrogen during the formation the bone, which modulate osteoblast differentiation and the development of osteoporosis [45]. Estrogen is known to stimulate the release of growth factors such as insulin-like growth factor 1 (IGF-1) that influence bone resorption. ERKO mice have been very valuable for elucidating the association between mechanisms underlying bone functions and ER signaling [46]. ER α is expressed at low levels in bone. In α ERKO mice, bone length and size are significantly decreased compare to normal mice. The bones of male α ERKO mice also show a reduction in mineral density [47]. However, skeletons of β ERKO mice are

Table 1. Phenotypes of estrogen receptor knockout (ERKO) mice

	Phenotype	
	α ERKO	β ERKO
Reproductive system	Infertile Incapability ovulation Reduced prolactin Mammary gland immature	Reduced fertility Ovulation deregulation Reduced oocyte
Bone	Decreased length and size	No difference compared to wild-type mice
Metabolism	Insulin resistance and obesity	No difference compared to wild-type mice
Brain and behavior	Aggressive behavior and depression in females	

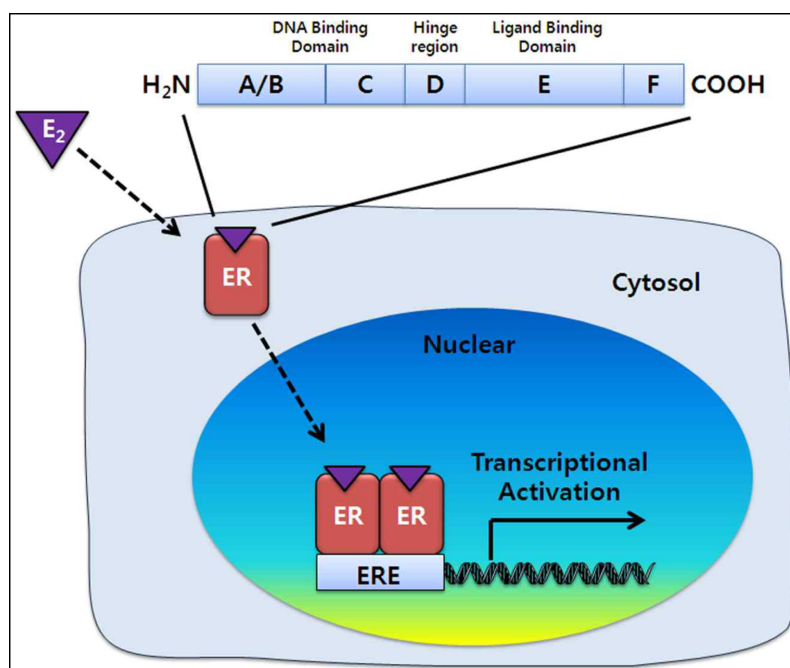


Figure 1. Mechanisms of estrogen receptor signaling pathway. Estrogen (E_2) is able to bind to estrogen receptors (ERs). The ERs then dimerize and translocate into the nucleus. These complexes bind to estrogen response elements (EREs). ERs contain five domains with distinct functions. The A/B domain contains transcriptional activation function 1 (AF1). The C domain has the DNA binding domain and the D domain is a hinge region. The E/F domain encodes the AF2 region.

phenotypically identical to those of wild-type animals, indicating the low importance of $ER\alpha$ in osteoporosis progression [48].

Cardiovascular tissues and metabolism in ER knockout mice

The rate of cardiovascular system disease for women increases with age after menopause. Although the direct effect of estrogen on the cardiovascular system is unclear, reduced estrogen levels are associated with the disruption of lipid or glucose regulation [49]. The ability of estrogen to decrease cholesterol levels is due to the elimination of low-density lipoprotein (LDL) from blood circulation. In

addition, various studies have recently demonstrated that estrogen affects blood vessel structure [5,50]. ERs are expressed in endothelial and smooth muscle cells of vascular tissues, suggesting that ER signaling involves in function of smooth muscle cells. Moreover, ERKO mice possess inherently vascular disease [51]. However, $ER\alpha$ is expressed in tissues involved in glucose and lipid metabolism such as the brain, adipose tissue, liver, and heart [52]. Imbalanced $ER\alpha/ER\beta$ ratios may lead to the development of metabolic diseases [53]. α ERKO mice are diabetic and insulin resistant. β ERKO mice exhibited insulin responsibility and have not excessive fat in the body [54].

Brain and behavior

Estrogen is fundamental for regulating the growth and differentiation of axons and dendrites in the brain [55]. Functions performed by ER α and ER β in the brain have been explored by studies of α ERKO and β ERKO mice. Interestingly, previous studies demonstrated differences in localization and expression levels of both ER isoforms. In addition, the results of these studies have shown that the function of ER α seems to be important for reproduction while that of ER β may affect cognitive processes [5,6,56]. However, the level of ER expression in the brain rapidly changes in response to injury [57]. ER α primarily contributes in masculinization while ER β contributes to the defeminization of sexual behavior [58]. ERKO females display aggressive behavior and depression. Additionally, estrogen signaling contributes to brain integrity and exerts different effects on behavior [59].

Summary

Estrogen regulates the growth, development, and physiology of the human reproductive system. This hormone also influences neuroendocrine, skeletal, adipose, and cardiovascular systems. Estrogen is an important sex hormone produced primarily by the ovaries in females and testes in males. The biological actions of estrogen are mediated by binding to the ERs in target organs. ER α and ER β are transcriptional factors that regulate the expression of specific genes in different tissues on a ligand dependent manner. Analysis of ER α and ER β functions using knockout mouse models has demonstrated the roles of estrogen signaling in different physiological processes. α ERKO and β ERKO mice are also valuable for examining the effect of ER signaling in specific target organs. Although transgenic mice do not always produce consistent results, these models are useful for evaluating the functions of genes under specific pathological conditions.

Acknowledgments

This work was supported by a National Research Foundation of Korea (NRF) grant (no. 2011-0015385) funded by the Ministry of Education, Science and Technology (MEST) of the South Korean government. In addition, this work was also supported by the Priority Research Centers Program (2011-0031403) through the NRF funded by the MEST.

References

1. Osz J, Brelivet Y, Peluso-Iltis C, Cura V, Eiler S, Ruff M, Bourguet W, Rochel N, Moras D. Structural basis for a molecular allosteric control mechanism of cofactor binding to nuclear receptors. *Proc Natl Acad Sci U S A* 2012; 109(10): E588-594.
2. Choi KC, Jeung EB. The biomarker and endocrine disruptors in mammals. *J Reprod Dev* 2003; 49(5): 337-345.
3. Weigel NL, Moore NL. Cyclins, cyclin dependent kinases, and regulation of steroid receptor action. *Mol Cell Endocrinol* 2007; 265-266: 157-161.
4. Lee HR, Hwang KA, Park MA, Yi BR, Jeung EB, Choi KC. Treatment with bisphenol A and methoxychlor results in the growth of human breast cancer cells and alteration of the expression of cell cycle-related genes, cyclin D1 and p21, via an estrogen receptor-dependent signaling pathway. *Int J Mol Med* 2012; 29(5): 883-890.
5. Swedenborg E, Power KA, Cai W, Pongratz I, Ruegg J. Regulation of estrogen receptor beta activity and implications in health and disease. *Cell Mol Life Sci* 2009; 66(24): 3873-3894.
6. Hughes ZA, Liu F, Marquis K, Muniz L, Pangalos MN, Ring RH, Whiteside GT, Brandon NJ. Estrogen receptor neurobiology and its potential for translation into broad spectrum therapeutics for CNS disorders. *Curr Mol Pharmacol* 2009; 2(3): 215-236.
7. Xiao J, Wang NL, Sun B, Cai GP. Estrogen receptor mediates the effects of pseudoprotodiocsin on adipogenesis in 3T3-L1 cells. *Am J Physiol Cell Physiol* 2010; 299(1): C128-138.
8. Welboren WJ, Sweep FC, Span PN, Stunnenberg HG. Genomic actions of estrogen receptor alpha: what are the targets and how are they regulated? *Endocr Relat Cancer* 2009; 16(4): 1073-1089.
9. Kong EH, Pike AC, Hubbard RE. Structure and mechanism of the oestrogen receptor. *Biochem Soc Trans* 2003; 31(Pt 1): 56-59.
10. Skafar DF, Zhao C. The multifunctional estrogen receptor-alpha F domain. *Endocrine* 2008; 33(1): 1-8.
11. Geserick C, Meyer HA, Haendler B. The role of DNA response elements as allosteric modulators of steroid receptor function. *Mol Cell Endocrinol* 2005; 236(1-2): 1-7.
12. Edwards DP. The role of coactivators and corepressors in the biology and mechanism of action of steroid hormone receptors. *J Mammary Gland Biol Neoplasia* 2000; 5(3): 307-324.
13. Sanchez ER. Chaperoning steroidal physiology: Lessons from mouse genetic models of Hsp90 and its cochaperones. *Biochim Biophys Acta* 2012; 1823(3): 722-729.
14. Beliakoff J, Whitesell L. Hsp90: an emerging target for breast cancer therapy. *Anticancer Drugs* 2004; 15(7): 651-662.
15. McDevitt MA, Glidewell-Kenney C, Jimenez MA, Ahearn PC, Weiss J, Jameson JL, Levine JE. New insights into the classical and non-classical actions of estrogen: evidence from estrogen receptor knock-out and knock-in mice. *Mol Cell Endocrinol* 2008; 290(1-2): 24-30.
16. Park MA, Hwang KA, Choi KC. Diverse animal models to examine potential role(s) and mechanism of endocrine disrupting chemicals on the tumor progression and prevention: Do they have tumorigenic or anti-tumorigenic property? *Lab Anim Res* 2011; 27(4): 265-273.
17. Zhao C, Dahlman-Wright K, Gustafsson JA. Estrogen signaling via estrogen receptor {beta}. *J Biol Chem* 2010; 285(51): 39575-39579.
18. Zhao C, Dahlman-Wright K, Gustafsson JA. Estrogen receptor beta: an overview and update. *Nucl Recept Signal* 2008; 6: e003.
19. Lane PH. Estrogen receptors in the kidney: lessons from genetically altered mice. *Gend Med* 2008; 5 Suppl A: S11-18.
20. Weiser MJ, Foradori CD, Handa RJ. Estrogen receptor beta in the brain: from form to function. *Brain Res Rev* 2008; 57(2): 309-320.
21. Hwang KA, Park SH, Yi BR, Choi KC. Gene alterations of ovarian cancer cells expressing estrogen receptors by estrogen and bisphenol A using microarray analysis. *Lab Anim Res* 2011; 27(2): 99-107.

22. Chen M, Wolfe A, Wang X, Chang C, Yeh S, Radovick S. Generation and characterization of a complete null estrogen receptor alpha mouse using Cre/LoxP technology. *Mol Cell Biochem* 2009; 321(1-2): 145-153.
23. Jayachandran M, Preston CC, Hunter LW, Jahangir A, Owen WG, Korach KS, Miller VM. Loss of estrogen receptor beta decreases mitochondrial energetic potential and increases thrombogenicity of platelets in aged female mice. *Age (Dordr)* 2010; 32(1): 109-121.
24. Singh SP, Wolfe A, Ng Y, DiVall SA, Buggs C, Levine JE, Wondisford FE, Radovick S. Impaired estrogen feedback and infertility in female mice with pituitary-specific deletion of estrogen receptor alpha (ESR1). *Biol Reprod* 2009; 81(3): 488-496.
25. Santollo J, Eckel LA. Effect of a putative ERalpha antagonist, MPP, on food intake in cycling and ovariectomized rats. *Physiol Behav* 2009; 97(2): 193-198.
26. Shao R. Understanding the mechanisms of human tubal ectopic pregnancies: new evidence from knockout mouse models. *Hum Reprod* 2010; 25(3): 584-587.
27. Bockamp E, Sprengel R, Eshkind L, Lehmann T, Braun JM, Emmrich F, Hengstler JG. Conditional transgenic mouse models: from the basics to genome-wide sets of knockouts and current studies of tissue regeneration. *Regen Med* 2008; 3(2): 217-235.
28. Sun J, Langer WJ, Devish K, Lane PH. Compensatory kidney growth in estrogen receptor-alpha null mice. *Am J Physiol Renal Physiol* 2006; 290(2): F319-323.
29. Chen M, Hsu I, Wolfe A, Radovick S, Huang K, Yu S, Chang C, Messing EM, Yeh S. Defects of prostate development and reproductive system in the estrogen receptor-alpha null male mice. *Endocrinology* 2009; 150(1): 251-259.
30. Cholieris E, Clipperton AE, Phan A, Kavaliers M. Estrogen receptor beta agonists in neurobehavioral investigations. *Curr Opin Investig Drugs* 2008; 9(7): 760-773.
31. Wintermantel TM, Elzer J, Herbison AE, Fritzemeier KH, Schutz G. Genetic dissection of estrogen receptor signaling in vivo. *Ernst Schering Found Symp Proc* 2006; (1): 25-44.
32. Lee GS, Kim HJ, Jung YW, Choi KC, Jeung EB. Estrogen receptor alpha pathway is involved in the regulation of Calbindin-D9k in the uterus of immature rats. *Toxicol Sci* 2005; 84(2): 270-277.
33. An BS, Choi KC, Hong EJ, Jung YW, Manabe N, Jeung EB. Differential transcriptional and translational regulations of calbindin-D9k by steroid hormones and their receptors in the uterus of immature mice. *J Reprod Dev* 2004; 50(4): 445-453.
34. Lee S, Kang DW, Hudgins-Spivey S, Krust A, Lee EY, Koo Y, Cheon Y, Gye MC, Chambon P, Ko C. Theca-specific estrogen receptor-alpha knockout mice lose fertility prematurely. *Endocrinology* 2009; 150(8): 3855-3862.
35. Glidewell-Kenney C, Hurley LA, Pfaff L, Weiss J, Levine JE, Jameson JL. Nonclassical estrogen receptor alpha signaling mediates negative feedback in the female mouse reproductive axis. *Proc Natl Acad Sci U S A* 2007; 104(19): 8173-8177.
36. Drummond AE, Fuller PJ. The importance of ERbeta signalling in the ovary. *J Endocrinol* 2010; 205(1): 15-23.
37. Silberstein GB, Van Horn K, Hrabeta-Robinson E, Compton J. Estrogen-triggered delays in mammary gland gene expression during the estrous cycle: evidence for a novel timing system. *J Endocrinol* 2006; 190(2): 225-239.
38. Hewitt SC, Korach KS. Oestrogen receptor knockout mice: roles for oestrogen receptors alpha and beta in reproductive tissues. *Reproduction* 2003; 125(2): 143-149.
39. McPherson SJ, Ellem SJ, Risbridger GP. Estrogen-regulated development and differentiation of the prostate. *Differentiation* 2008; 76(6): 660-670.
40. Raskin K, de Gendt K, Duittoz A, Liere P, Verhoeven G, Tronche F, Mhaouty-Kodja S. Conditional inactivation of androgen receptor gene in the nervous system: effects on male behavioral and neuroendocrine responses. *J Neurosci* 2009; 29(14): 4461-4470.
41. Lee KH, Park JH, Bunick D, Lubahn DB, Bahr JM. Morphological comparison of the testis and efferent ductules between wild-type and estrogen receptor alpha knockout mice during postnatal development. *J Anat* 2009; 214(6): 916-925.
42. Imai Y, Kondoh S, Kouzmenko A, Kato S. Regulation of bone metabolism by nuclear receptors. *Mol Cell Endocrinol* 2009; 310(1-2): 3-10.
43. Li BY, Tong J, Zhang ZL. [Exogenous estrogen improved calcium homeostasis and skeletal mineralization in vitamin D receptor gene knockout female mice.]. *Sheng Li Xue Bao* 2006; 58(6): 573-576.
44. Venken K, Callewaert F, Boonen S, Vanderschueren D. Sex hormones, their receptors and bone health. *Osteoporos Int* 2008; 19(11): 1517-1525.
45. Auld KL, Berasi SP, Liu Y, Cain M, Zhang Y, Huard C, Fukayama S, Zhang J, Choe S, Zhong W, Bhat BM, Bhat RA, Brown EL, Martinez RV. Estrogen-related receptor α regulates osteoblast differentiation via Wnt/ β -catenin signaling. *J Mol Endocrinol* 2012; 48(2): 177-191.
46. Vico L, Vanacker JM. Sex hormones and their receptors in bone homeostasis: insights from genetically modified mouse models. *Osteoporos Int* 2010; 21(3): 365-372.
47. Chilibeck PD, Cornish SM. Effect of estrogenic compounds (estrogen or phytoestrogens) combined with exercise on bone and muscle mass in older individuals. *Appl Physiol Nutr Metab* 2008; 33(1): 200-212.
48. Syed FA, Fraser DG, Spelsberg TC, Rosen CJ, Krust A, Chambon P, Jameson JL, Khosla S. Effects of loss of classical estrogen response element signaling on bone in male mice. *Endocrinology* 2007; 148(4): 1902-1910.
49. Knowlton AA, Lee AR. Estrogen and the cardiovascular system. *Pharmacol Ther* 2012; 135(1): 54-70.
50. Kim KH, Moriarty K, Bender JR. Vascular cell signaling by membrane estrogen receptors. *Steroids* 2008; 73(9-10): 864-869.
51. Luksha L, Kublickiene K. The role of estrogen receptor subtypes for vascular maintenance. *Gynecol Endocrinol* 2009; 25(2): 82-95.
52. Barros RP, Gustafsson JA. Estrogen receptors and the metabolic network. *Cell Metab* 2011; 14(3): 289-299.
53. Riant E, Waget A, Cogo H, Arnal JF, Burcelin R, Gourdy P. Estrogens protect against high-fat diet-induced insulin resistance and glucose intolerance in mice. *Endocrinology* 2009; 150(5): 2109-2117.
54. Faulds MH, Zhao C, Dahlman-Wright K, Gustafsson JA. The diversity of sex steroid action: regulation of metabolism by estrogen signaling. *J Endocrinol* 2012; 212(1): 3-12.
55. Kalita K, Szymczak S. [Estrogen receptors in the brain]. *Neurol Neurochir Pol* 2003; 37 Suppl 3: 63-78.
56. Hill RA, Boon WC. Estrogens, brain, and behavior: lessons from knockout mouse models. *Semin Reprod Med* 2009; 27(3): 218-228.
57. Walf AA, Koonce C, Manley K, Frye CA. Proestrous compared to diestrous wildtype, but not estrogen receptor beta knockout, mice have better performance in the spontaneous alternation and object recognition tasks and reduced anxiety-like behavior in the elevated plus and mirror maze. *Behav Brain Res* 2009; 196(2): 254-260.
58. Kudwa AE, Michopoulos V, Gatewood JD, Rissman EF. Roles of estrogen receptors alpha and beta in differentiation of mouse sexual behavior. *Neuroscience* 2006; 138(3): 921-928.
59. Crews D, Fuller T, Mirasol EG, Pfaff DW, Ogawa S. Postnatal environment affects behavior of adult transgenic mice. *Exp Biol Med (Maywood)* 2004; 229(9): 935-939.