

In Vitro Activity of TP-271 against Mycobacterium abscessus, Mycobacterium fortuitum, and Nocardia Species

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The *in vitro* activities of TP-271, a novel fluorocycline antimicrobial, against 22 isolates of *Mycobacterium abscessus*, 22 isolates of *Mycobacterium fortuitum*, and 19 isolates of *Nocardia* spp. were studied by a microtiter broth dilution method. The $MIC_{90}s$ for *M. abscessus*, *M. fortuitum*, and *Nocardia* spp. were 0.5 µg/ml, 0.03 µg/ml, and 8 µg/ml, respectively. TP-271 was significantly more active than the respective control drug in virtually all tests.

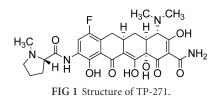
Mycobacterium abscessus and Mycobacterium fortuitum are rapidly growing mycobacteria that are associated primarily with opportunistic infections in immunocompromised subjects (2). Patients with bronchiectasis, especially those with cystic fibrosis, are at increased risk of *M. abscessus* infection. Other manifestations of *M. abscessus* infection include localized skin infections, postoperative wound infections, and infection of implanted medical devices. *M. fortuitum* is a relatively rare pathogen, even in immunocompromised individuals, but has been associated with skin infections and hospital-acquired postoperative infections (2). The most common clinical form of *Nocardia* infection is pulmonary nocardiosis, with and without dissemination, followed by skin and soft tissue infection (5).

Infections due to these microorganisms are occurring with increasing frequency and are often difficult to treat, especially the mycobacterial infections. Successful treatment is often hindered by the need for combination therapy, resistance to multiple drugs, necessity of long treatment duration, and lack of sufficiently active oral drugs (2, 3, 6).

While *Nocardia* remains susceptible to trimethoprim-sulfamethoxazole in the majority of cases, effective alternative oral therapy is lacking. Mortality from pulmonary nocardiosis remains high, in the range of 15 to 40% (5, 11).

TP-271 is a novel, fully synthetic fluorocycline antimicrobial related to tetracycline (Fig. 1) (1, 10). It has been shown to have potent broad-spectrum in vitro and in vivo activity against multiple community-acquired organisms, including Staphylococcus spp., Streptococcus spp., Haemophilus influenzae, Moraxella catarrhalis, Legionella pneumophila, and Acinetobacter baumannii, as well as biothreat organisms (Bacillus anthracis, Francisella tularensis, Burkholderia pseudomallei, and Burkholderia mallei) (4). The activity of TP-271 was shown to be unaffected by the Gram-positive tetracycline-specific pump tet(K) and ribosomal protection mechanism tet(M) and minimally affected by the most common Gram-negative efflux mechanisms, tet(A) and tet(B) (7). Promising oral activity was demonstrated by TP-271 in neutropenic mouse models of pneumonia caused by methicillin-resistant Staphylococcus aureus and Streptococcus pneumoniae and in immunocompetent models of pneumonia caused by S. pneumoniae in mice and *H. influenzae* in rats (4).

As a first step in its assessment as a novel therapy to treat infections caused by *M. abscessus*, *M. fortuitum*, and *Nocardia* spp., we evaluated the *in vitro* activity of TP-271 in comparison to those of several standard agents against clinical isolates.



TP-271 was obtained from Tetraphase Pharmaceuticals, Inc. (Watertown, MA), moxifloxacin (MOX) from Bayer Pharmaceuticals (West Haven, CT), and amikacin (AMK) from (Bristol-Myers Squibb, Princeton, NJ). Doxycycline (DOX) and tetracycline (TET) were purchased from Sigma Chemical Co. (St. Louis, MO).

M. abscessus isolates were provided by Barbara Body (LabCorp, Burlington, NC) and Barbara Brown-Elliott (University of Texas Health Science Center, Tyler, TX). M. fortuitum isolates were provided by Barbara Brown-Elliott, and Nocardia isolates were from the American Type Culture Collection (Manassas, VA), Barbara Body, and Betty Ann Forbes (Virginia Commonwealth University, Richmond, VA). The compounds were initially dissolved in water to make concentrated stock solutions that were frozen at -20°C. Stock solutions were subsequently diluted in Mueller-Hinton (MH) broth (Becton, Dickinson, Sparks, MD) prior to their use. An *in vitro* microtiter broth dilution method similar to that suggested by the Clinical and Laboratory Standards Institute (CLSI) (9) was utilized, with several modifications. Polystyrene 96-well round-bottom plates (Corning Inc., Corning, NY) were prepared with 50 µl of MH broth per well. The compounds were prepared at 4 times the maximum concentration at which they were tested and were added to the first well prior to being serially diluted 2-fold. The bacterial isolates were grown in MH broth for 3 to 5 days and then diluted in MH broth with 20% glycerol to 100 Klett units (equivalent to 5×10^7 CFU/ml) (Photoelectric Colorimeter; Manostat Corp., New York, NY) and stored at -70°C until used. The bacterial cultures were thawed and diluted to a

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 TABLE 1 MICs of TP-271, moxifloxacin, doxycycline, and tetracycline against Mycobacterium abscessus

| TABLE 2 MICs | of TP-271, | doxycycline, | and | amikacin against | |
|---------------|------------|--------------|-----|------------------|--|
| Mycobacterium | | | | - | |

| | MIC (µg/ml) | | | | |
|-------------------|-------------|--------------|--------------|-------------|--|
| Isolate | TP-271 | Moxifloxacin | Tetracycline | Doxycycline | |
| BB2 | 0.5 | 8 | >64 | >64 | |
| BB1 | 0.5 | 8 | >64 | | |
| BB3 | 0.06 | 1 | 8 | | |
| BB4 | 0.5 | 4 | 64 | | |
| BB6 | 0.06 | 1 | >64 | | |
| BB8 | 0.5 | 1 | >64 | | |
| MC6005 | 0.5 | 8 | >64 | | |
| MC6025 | 0.25 | 8 | >64 | | |
| B87 | 0.06 | 0.5 | | | |
| MC5812 | 0.125 | 2 | | | |
| MC5922 | 0.06 | 0.5 | | | |
| MC5785 | 0.06 | 0.25 | | | |
| AC5960 | 1 | 0.25 | | 4 | |
| MC6031 | 0.06 | 1 | | 8 | |
| AC5605 | 0.125 | 4 | | | |
| AC5908 | 0.125 | 4 | | | |
| MC5901 | 0.06 | 1 | | | |
| MC5931 | 0.5 | 4 | | | |
| 3B10 | 0.5 | 0.5 | | 2 | |
| MC6111 | 0.5 | 16 | | >64 | |
| MC6136 | 1 | >16 | | >64 | |
| MC6153 | 0.5 | 8 | | >64 | |
| MIC ₅₀ | 0.25 | 2 | | | |
| MIC ₉₀ | 0.50 | 8 | | | |

| | MIC (µg/ml) | | | | |
|-------------------|-------------|-------------|----------|--|--|
| Isolate | TP-271 | Doxycycline | Amikacir | | |
| 3349 | 0.015 | 32 | 1 | | |
| 3107 | 0.03 | 256 | 2 | | |
| 3499 | 0.0038 | £0.25 | 0.125 | | |
| 3357 | 0.03 | 64 | 0.25 | | |
| 3488 | 0.03 | 256 | 0.5 | | |
| 3126 | 0.015 | 64 | 0.25 | | |
| 3480 | 0.03 | 128 | 0.25 | | |
| 3316 | 0.015 | 32 | 0.06 | | |
| 3490 | 0.015 | 64 | 0.125 | | |
| 3276 | 0.06 | 2 | 0.25 | | |
| 3579 | 0.03 | 16 | 0.25 | | |
| 3481 | 0.06 | ≤0.25 | 0.25 | | |
| 3315 | 0.015 | 64 | 0.25 | | |
| 3484 | 0.015 | 64 | 2 | | |
| 3489 | ≤0.001 | 1 | 0.125 | | |
| 3491 | 0.0019 | ≤0.25 | 0.5 | | |
| 2797 | 0.015 | 128 | 0.5 | | |
| 3442 | 0.125 | ≤0.25 | 2 | | |
| 3423 | 0.125 | ≤0.25 | 2 | | |
| 3502 | 0.015 | 128 | 2 | | |
| 3562 | 0.0075 | ≤0.25 | 0.25 | | |
| 2491 | 0.015 | 128 | 1 | | |
| MIC ₅₀ | 0.015 | 32 | 0.25 | | |
| MIC ₉₀ | 0.06 | 128 | 2 | | |

final concentration of approximately 1×10^5 CFU/ml (working stock) in MH broth. The inocula used were measured by titration and plated on MH II agar (Becton, Dickinson, Sparks, MD). Fifty microliters of the working stock was added to each well containing compound. The microtiter plates were covered with SealPlate adhesive sealing film (Exel Scientific, Wrightwood, CA) prior to being incubated at 37°C in ambient air prior to reading (4 days for *M. abscessus* and 2 to 3 days for M. *fortuitum*). Each isolate was tested in duplicate. The CLSI recommends a 4-day incubation period for *Nocardia*; however, some of our isolates grew slowly, requiring 7 days of incubation. For consistency, all *Nocardia* plates were incubated for 7 days. The MIC was defined as the lowest concentration of antimicrobial agent yielding no visible turbidity.

The MIC₅₀ and MIC₉₀ of TP-271 and MOX against *M. abscessus* isolates were 0.25 µg/ml and 0.5 µg/ml for TP-271 and 2 µg/ml and 8 µg/ml for MOX, respectively; the MIC range for TP-271 was 0.06 to 1 µg/ml, and that for MOX was 0.25 to >16 µg/ml (Table 1). TET and DOX were tested against subsets of these organisms and were found to be much less active than TP-271. The MIC ranges for TET and DOX were 8 µg/ml to >64 µg/ml (n = 8) and 2 µg/ml to 64 µg/ml (n = 7), respectively.

The MIC₅₀ and MIC₉₀ for TP-271, DOX, and AMK against *M. fortuitum* isolates were 0.015 µg/ml and 0.06 µg/ml, 32 µg/ml and 128 µg/ml, and 0.25 µg/ml and 2 µg/ml, respectively (Table 2). The MIC range for TP-271 was ≤ 0.001 to 0.125 µg/ml, the range for DOX was ≤ 0.25 to 256 µg/ml, and the range for AMK was 0.06 to 2 µg/ml.

The MIC₅₀ and MIC₉₀ for TP-271 and TET against *Nocardia* spp. were 2 μ g/ml and 8 μ g/ml and 32 μ g/ml and 64 μ g/ml,

respectively (Table 3). The MIC range for TP-271 was 0.06 to 16 μ g/ml, and the range for DOX was 2 to 64 μ g/ml.

Our results indicate that TP-271 has promising *in vitro* activity against *M. abscessus* and *M. fortuitum* isolates, with somewhat less

TABLE 3 MICs of TP-271 and tetracycline against Nocardia spp.

| | MIC (µg/ml) | | |
|-------------------|-------------|-------------|--|
| Isolate | TP-271 | Doxycycline | |
| 6 | 4 | 16 | |
| 1260 | 2 | 32 | |
| 7 | 16 | 64 | |
| 651 | 2 | 16 | |
| 2 | 0.06 | 64 | |
| 16 | 2 | 16 | |
| 8 | 2 | 32 | |
| 2039 | 1 | 16 | |
| 3840 | 1 | 16 | |
| 12 | 4 | 32 | |
| 1276 | 1 | 32 | |
| 14 | 4 | 32 | |
| 9B | 1 | 8 | |
| 71743 | 8 | 32 | |
| 5 | 8 | 32 | |
| 5 | 8 | 64 | |
| 32 | 4 | 32 | |
| 9A | 1 | 2 | |
| 11 | 8 | 64 | |
| MIC ₅₀ | 2 | 32 | |
| MIC ₉₀ | 8 | 64 | |

activity against *Nocardia* spp. The activity of TP-271 was much better than those of other orally available tetracycline-related compounds (8). Infections due to these organisms are not common. Treatment of these infections is challenging, particularly for *M. abscessus* due to the lack of effective agents and the immunosuppressed population affected. It appears that further evaluation of TP-271 in animal studies should be performed to assess its potential *in vivo* activity.

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