

Worldwide Appraisal and Update (2010) of Telavancin Activity Tested against a Collection of Gram-Positive Clinical Pathogens from Five Continents

Rodrigo E. Mendes,^a Helio S. Sader,^a David J. Farrell,^a and Ronald N. Jones^{a,b}

JMI Laboratories, North Liberty, Iowa, USA,^a and Tufts University School of Medicine, Boston, Massachusetts, USA,^b

A total of 15,480 Gram-positive pathogens were collected from 89 sites in the United States, Europe, the Asia-Pacific region, and Latin America in 2010. Telavancin was active against indicated *Staphylococcus aureus* (MIC_{50/90}, 0.12/0.25 µg/ml), vancomycin-susceptible *Enterococcus faecalis* (MIC_{50/90}, 0.5/0.5 µg/ml), and beta-hemolytic (MIC_{50/90}, 0.06/0.12 µg/ml) and viridans group streptococcus (MIC_{50/90}, 0.03/0.06 µg/ml) isolates. These MIC results showed potency for telavancin equal to or greater than that of comparators. These *in vitro* data confirm a continued potent activity of telavancin when tested against contemporary Grampositive clinical isolates.

Several agents directed against Gram-positive pathogens have been developed for the treatment of complicated infections in the last decade. However, only linezolid and daptomycin, and more recently ceftaroline and telavancin, have been approved for clinical use (7). The latter was approved (2009) in the United States and Canada for the treatment of adults with complicated skin and skin structure infections (cSSSI) caused by susceptible organisms (16). In addition, telavancin was recently approved (2011) in all member states of the European Union, Norway, and Iceland for the treatment of adults with nosocomial pneumonia (NP), including ventilator-associated pneumonia (VAP), known or suspected to be caused by methicillinresistant *Staphylococcus aureus* (MRSA).

Similarly to vancomycin, telavancin inhibits bacterial cell wall peptidoglycan biosynthesis through binding to the acyl-D-alanyl-D-alanine (D-Ala-D-Ala) terminus of peptidoglycan precursors, consequently inhibiting cell wall extension (transglycosylation) (8). A second mechanism of action comprises the interaction of telavancin with the cell wall precursor (lipid II), causing membrane depolarization and increased membrane permeability. The induced depolarization was shown to be time and concentration dependent (10). This unique dual mechanism of action is likely to be responsible for the documented telavancin *in vitro* activity against MRSA, heterogeneous vancomycin-intermediate *S. aureus* (hVISA), and VISA strains (10), which have been demonstrated to be susceptible to membrane depolarization (12).

(The results included in this study have been partially presented at the 2011 Interscience Conference on Antimicrobial Agents and Chemotherapy.)

The overall objective of this study was to provide an updated evaluation of the potency and spectrum of activity of telavancin against contemporary (2010) Gram-positive isolates from a global surveillance program. A total of 15,480 Gram-positive, nonduplicated clinical isolates were collected from 89 medical sites in the United States (26 hospitals; 6,719 isolates), Europe (31 hospitals; 4,647 isolates), Latin America (10 hospitals; 1,814 isolates), and the Asia-Pacific region (22 hospitals; 2,300 isolates). Isolates were primarily recovered from patients with bacteremia (39%), SSSIs (28%), and respiratory tract infections (21%) and were submitted to a central monitoring laboratory (JMI Laboratories, North Liberty, IA) according to established protocols as part of the SENTRY Antimicrobial Surveillance Program. Each participating medical center provided species identifications, which were confirmed by the monitoring laboratory using standard algorithms and the automated Vitek 2 system (bioMérieux, Hazelwood, MO), as required. The species included were as follows: *S. aureus* (7,653 isolates; 49.4%), coagulase-negative staphylococci (CoNS; 1,278 isolates; 8.3%), *Enterococcus faecalis* (1,459 isolates; 9.4%), *Enterococcus faecalis* (2,150 isolates; 13.9%), beta-hemolytic streptococcu (BHS; 1,472 isolates; 9.5%), viridans group streptococci (VGS; 551 isolates; 3.6%), and *Streptococcus anginosus* (97 isolates; 0.6%) and *Streptococcus bovis* (32 isolates; 0.2%) groups.

Isolates were tested for susceptibility by the broth microdilution method according to the Clinical and Laboratory Standards Institute (CLSI; M07-A8, 2009) recommendations (1). Susceptibility testing was performed using customized and validated dryform panels (Thermo Fisher Scientific, Cleveland, OH). Quality assurance was performed by concurrent testing of the CLSI-recommended (M100-S21, 2011) strains: *E. faecalis* ATCC 29212, *S. aureus* ATCC 29213, and *S. pneumoniae* ATCC 49619 (2). The Food and Drug Administration (FDA)-approved breakpoints for telavancin for *S. aureus* ($\leq 1 \mu g/ml$), vancomycin-susceptible *E. faecalis* ($\leq 1 \mu g/ml$), VGS ($\leq 0.12 \mu g/ml$), and BHS ($\leq 0.12 \mu g/$ ml) were applied (16). Interpretation of comparator MIC results was in accordance with published CLSI and European Committee on Antimicrobial Susceptibility Testing (EUCAST) criteria (2, 5).

Telavancin (MIC_{50/90}, 0.12/0.25 μ g/ml) exhibited potent activity when tested against all *S. aureus* and CoNS clinical isolates as well as across different resistance subsets (methicillin susceptibility; Table 1). Some variation in the MIC₅₀ values was noted for telavancin when tested against MRSA strains from different geo-

Received 9 January 2012 Returned for modification 13 March 2012 Accepted 6 April 2012

Published ahead of print 16 April 2012

Address correspondence to Rodrigo E. Mendes, rodrigo-mendes@jmilabs.com. Copyright © 2012, American Society for Microbiology. All Rights Reserved. doi:10.1128/AAC.00011-12

| Ourseiner and moistent arbest | MIC (µg/m | (J | No. (cumulative | %) of isolates inhib | ited at each telavan | cin MIC (µg/ml): | | | | |
|---|--|--------------|-----------------|----------------------|----------------------|------------------|--------------|----------------|------------|------------|
| Organism/ group and resistant subset (no. of isolates tested) | 50% | %06 | ≤0.015 | 0.03 | 0.06 | 0.12 | 0.25 | 0.5 | 1 | 2 |
| Staphylococcus aureus (7,653) | 0.12 | 0.25 | 1(0.0) | 7 (0.1) | 232 (3.1) | 4,155 (57.4) | 2,962 (96.1) | 296 (100.0) | | |
| Oxacillin susceptible (4,565) | 0.12 | 0.25 | 0(0.0) | 7(0.2) | 172 (3.9) | 2,572(60.3) | 1,660(96.6) | $154\ (100.0)$ | | |
| Oxacillin resistant (3,088) | 0.12 | 0.25 | 1(0.0) | 0(0.0) | 60 (2.0) | 1,583~(53.2) | 1,302~(95.4) | 142(100.0) | | |
| Coagulase-negative staphylococci (1,278) | 0.12 | 0.25 | 6(0.5) | 14(1.6) | 88 (8.5) | 659~(60.0) | 473 (97.0) | 37 (99.9) | 1(100.0) | |
| Oxacillin susceptible (332) | 0.12 | 0.25 | 0(0.0) | 9 (2.7) | 36(13.6) | 166(63.6) | 115 (98.2) | 6(100.0) | | |
| Oxacillin resistant (946) | 0.12 | 0.25 | 6 (0.6) | 5(1.2) | 52 (6.7) | 493(58.8) | 358 (96.6) | 31 (99.9) | 1(100.0) | |
| Enterococcus faecalis $(1,459)^a$ | 0.5 | 0.5 | (0.0) | 2(0.1) | 8 (0.7) | 157(11.4) | 461 (43.0) | 761 (95.2) | 38 (97.8) | 2 (97.9) |
| Vancomycin susceptible (1,421) | 0.5 | 0.5 | 0(0.0) | 2(0.1) | 8 (0.7) | 157 (11.8) | 460(44.1) | 758 (97.5) | 36 (100.0) | r. |
| VanA type $(32)^a$ | >2 | >2 | 0(0.0) | 0(0.0) | 0(0.0) | 0(0.0) | 0(0.0) | 0(0.0) | 0(0.0) | 2 (6.3) |
| VanB type (6) | 0.5 | | 0(0.0) | 0(0.0) | 0(0.0) | 0(0.0) | 1 (16.7) | 3 (66.7) | 2(100.0) | |
| Enterococcus faecium $(805)^b$ | 0.25 | >2 | 10 (1.2) | 86 (11.9) | 117 (26.5) | 164(46.8) | 33 (50.9) | 11 (52.3) | 33 (56.4) | 156 (75.8) |
| Vancomycin susceptible (386) | 0.06 | 0.12 | 9 (2.3) | 82 (23.6) | 114(53.1) | 152(92.5) | 29(100.0) | | | |
| VanA type $(392)^b$ | 2 | >2 | (0.0) | 0(0.0) | 0(0.0) | 0(0.0) | 1(0.3) | 9 (2.6) | 31(10.5) | 156(50.3) |
| VanB type (27) | 0.12 | 0.5 | 1 (3.7) | 4(18.5) | 3 (29.6) | 12 (74.1) | 3 (85.2) | 2 (92.6) | 2(100.0) | |
| Streptococcus pneumoniae (2,150) | ≤ 0.015 | 0.03 | 1,811(84.2) | 329 (99.5) | 8 (99.9) | 2(100.0) | | | | |
| Penicillin susceptible (1,330) | $\leq\!0.015$ | 0.03 | 1,103(82.9) | 222 (99.6) | 3 (99.8) | 2(100.0) | | | | |
| Penicillin nonsusceptible (820) | ≤0.015 | 0.03 | 708 (86.3) | 107 (99.4) | 5(100.0) | | | | | |
| Beta-hemolytic streptococci (1,472) | 0.06 | 0.12 | 101 (6.9) | 564 (45.2) | 541(81.9) | 263 (99.8) | 3 (100.0) | | | |
| Group A (521) | 0.03 | 0.06 | 95 (18.2) | 346(84.6) | 60 (96.2) | 20(100.0) | | | | |
| Group B (669) | 0.06 | 0.12 | 0(0.0) | 51 (7.6) | 399 (67.3) | 216(99.6) | 3(100.0) | | | |
| Group C (91) | 0.03 | 0.06 | 2 (2.2) | 54(61.5) | 27 (91.2) | 8(100.0) | | | | |
| Group F (8) | 0.03 | | 0(0.0) | 6(75.0) | 2(100.0) | | | | | |
| Group G (154) | 0.03 | 0.12 | 3(1.9) | 87 (58.4) | 47 (89.0) | 17(100.0) | | | | |
| Viridans group streptococci (551) | 0.03 | 0.06 | 78 (14.2) | 333 (74.6) | 112 (94.9) | 28 (100.0) | | | | |
| S. anginosus group (97) | 0.03 | 0.06 | 7 (7.2) | 68 (77.3) | 19(96.9) | 3(100.0) | | | | |
| S. bovis group (32) | 0.03 | 0.06 | 4(12.5) | 19 (71.9) | 6(90.6) | 3(100.0) | | | | |
| Penicillin susceptible (416) | 0.03 | 0.06 | 66(15.9) | 254 (76.9) | 80 (96.2) | 16(100.0) | | | | |
| Penicillin nonsusceptible (135) | 0.03 | 0.06 | 12 (8.9) | 79 (67.4) | 32 (91.1) | 12(100.0) | | | | |
| ^a A total of 30 VanA-type <i>E. faecalis</i> strains div ^b A total of 195 VanA-type <i>F. faecium</i> strains of | splayed telavanci disnlaved telavan | n MICs of >2 | μg/ml. 2 | | | | | | | |

| | | No. of | MIC (µg/ml) | | |
|---------------------|--------------------------|----------|-------------------|-------------------|---------------------|
| Region ^a | Organism | isolates | MIC ₅₀ | MIC ₉₀ | Range |
| North America | S. aureus | 3,488 | 0.12 | 0.25 | 0.03-0.5 |
| | MRSA | 1,768 | 0.12 | 0.25 | 0.06-0.5 |
| | E. faecalis ^b | 571 | 0.5 | 0.5 | 0.06 - 1 |
| | S. pneumoniae | 926 | ≤0.015 | 0.03 | $\leq 0.015 - 0.12$ |
| | BHS | 648 | 0.06 | 0.12 | ≤0.015-0.12 |
| | VGS | 229 | 0.03 | 0.06 | ≤0.015-0.12 |
| Europe | S. aureus | 2,136 | 0.12 | 0.25 | ≤0.015-0.5 |
| * | MRSA | 497 | 0.12 | 0.25 | ≤0.015-0.5 |
| | E. faecalis ^b | 426 | 0.5 | 0.5 | 0.03-1 |
| | S. pneumoniae | 629 | ≤0.015 | 0.03 | ≤0.015-0.06 |
| | BHS | 503 | 0.06 | 0.12 | ≤0.015-0.25 |
| | VGS | 186 | 0.03 | 0.06 | ≤0.015-0.12 |
| Latin America | S. aureus | 914 | 0.12 | 0.25 | 0.03-0.5 |
| | MRSA | 400 | 0.25 | 0.25 | 0.06-0.5 |
| | E. faecalis ^b | 239 | 0.25 | 0.5 | 0.03-1 |
| | S. pneumoniae | 140 | ≤0.015 | 0.03 | ≤0.015-0.03 |
| | BHS | 101 | 0.06 | 0.12 | ≤0.015-0.25 |
| | VGS | 32 | 0.03 | 0.06 | ≤0.015-0.12 |
| Asia-Pacific | S. aureus | 1,115 | 0.25 | 0.5 | 0.06-0.5 |
| | MRSA | 423 | 0.25 | 0.25 | 0.06-0.5 |
| | E. faecalis ^b | 185 | 0.5 | 0.5 | 0.12-1 |
| | S. pneumoniae | 455 | ≤0.015 | 0.03 | ≤0.015-0.12 |
| | BĤS | 220 | 0.06 | 0.12 | ≤0.015-0.25 |
| | VGS | 72 | 0.03 | 0.06 | ≤0.015-0.12 |

 TABLE 2 Antimicrobial activity of telavancin tested against indicated

 Gram-positive bacteria listed by geographic region of origin

^{*a*} Regions and countries surveyed were as follows: North America, United States; Europe, Belgium, France, Germany, Ireland, Italy, Poland, Spain, Sweden, United Kingdom, Israel, and Turkey; Latin America, Argentina, Brazil, Chile, and Mexico; Asia-Pacific, Australia, Hong Kong, Japan, Korea, New Zealand, China, Singapore, and Taiwan.

^b All vancomycin susceptible.

graphic regions. The telavancin MIC_{50} values for strains (0.12 µg/ml) obtained from the United States (56.7% of strains inhibited at 0.12 µg/ml) and Europe (66.2% of strains inhibited at 0.12 µg/ml) were 2-fold lower than the MIC_{50} values (0.25 µg/ml) obtained for Asia-Pacific (40.0% of strains inhibited at 0.12 µg/ml) and Latin American (36.0% of strains inhibited at 0.12 µg/ml) clinical isolates (Table 2). However, similar MIC_{90} and MIC_{100} values (0.25 and 0.5 µg/ml, respectively) were observed for telavancin when tested against each of these subsets (Tables 1 and 2). Overall, telavancin ($\text{MIC}_{50/90}$, 0.12/0.25 µg/ml) and 4- to 8-fold more active than vancomycin and linezolid ($\text{MIC}_{50/90}$ for both, 1/1 µg/ml) against MRSA (Table 3).

When tested against vancomycin-susceptible (MIC_{50/90}, 0.5/ 0.5 µg/ml) and -nonsusceptible VanB (MIC₅₀, 0.5 µg/ml) *E. faecalis* isolates, telavancin inhibited these strains at the FDA breakpoint for susceptibility (≤ 1 µg/ml; Tables 1 and 3). In addition, no variations in the telavancin MIC₅₀ and MIC₉₀ values (0.5 µg/ml for both) were observed among *E. faecalis* strains from different regions, except for Latin American strains that exhibited lower MIC₅₀ results (0.25 µg/ml; Table 2). All *E. faecalis* strains exhibiting higher telavancin MIC results (≥ 2 µg/ml) displayed a VanA phenotype (Tables 1 and 3). Based on the MIC₉₀ values, telavancin (MIC_{50/90}, 0.5/0.5 µg/ml) was 2- to 4-fold more potent than daptomycin (MIC_{50/90}, 1/1 µg/ml), ampicillin (MIC_{50/90}, $\leq 1/2$ µg/ml), vancomycin (MIC_{50/90}, 1/2 µg/ml), and linezolid (MIC_{50/90}, 1/2 µg/ml) when tested against vancomycin-susceptible *E. faecalis* (Table 3). Ampicillin (MIC_{50/90}, $\leq 1/2 \mu g/ml$), daptomycin (MIC_{50/90}, $1/1 \mu g/ml$), and linezolid (MIC_{50/90}, $1/2 \mu g/ml$) ml) showed similar coverage (all 100.0% susceptible) against VanA-type E. faecalis, while telavancin was less active (MIC_{50/90}, >2/>2 µg/ml). Moreover, telavancin (MIC₅₀, 0.5 µg/ml), daptomycin (MIC₅₀, 0.5 µg/ml), and linezolid (MIC₅₀, 1 µg/ml) showed comparable MIC₅₀ results when tested against VanB-type E. faecalis (Table 3). Vancomycin-susceptible E. faecium isolates were highly susceptible to telavancin (MIC_{50/90}, 0.06/0.12 μ g/ml; highest MIC value, 0.25 µg/ml; Table 1), which was 8- to 16-fold more potent than vancomycin (MIC_{50/90}, 1/1 µg/ml) and linezolid (MIC_{50/90}, $1/2 \mu g/ml$) and 16- to 32-fold more active than daptomycin (MIC_{50/90}, 2/2 µg/ml) against vancomycin-susceptible E. faecium (Table 3). VanB- and VanA-type E. faecium demonstrated telavancin MIC₅₀ values 2- and 32-fold greater than those for the wild-type strains, respectively (Tables 1 and 3).

Overall, telavancin (MIC_{50/90}, $\leq 0.015/0.03 \ \mu g/ml$) demonstrated similar highly potent activities when tested against penicillin-susceptible and -nonsusceptible S. pneumoniae isolates collected during 2010, and equivalent results were noted for telavancin (MIC_{50/90}, $\leq 0.015/0.03 \ \mu g/ml$) across all sampled regions (Tables 1 and 2). Telavancin (MIC_{50/90}, $\leq 0.015/0.03 \,\mu$ g/ml) showed MIC₅₀ and MIC₉₀ results at least 16-fold more potent than those of vancomycin (MIC_{50/90}, 0.25/0.5 µg/ml; 100.0% susceptible), levofloxacin (MIC_{50/90}, 1/1 µg/ml; 98.8% susceptible), and linezolid (MIC_{50/90}, $1/1 \mu g/ml$; >99.9% susceptible) when tested against S. pneumoniae (Table 3). While the penicillin and telavancin MIC₅₀ results (≤ 0.03 and $\leq 0.015 \mu g/ml$, respectively) obtained against S. pneumoniae were comparable, the telavancin MIC₉₀ result (0.03 µg/ml) was 128-fold lower than that for penicillin (MIC_{50/90}, $\leq 0.03/4 \,\mu$ g/ml; Table 3). Telavancin MIC₅₀ results when tested against BHS serogroup B (MIC₅₀, 0.06 µg/ml) were slightly higher (2-fold) than those noted against all other serogroups tested (MIC₅₀, 0.03 µg/ml; Table 1). Consistent MIC_{50/90} results (0.06/0.12 µg/ml) were noted for telavancin against BHS from all sampled geographic regions (Table 2). Furthermore, telavancin (MIC_{50/90}, 0.06/0.12 µg/ml) and penicillin (MIC_{50/90}, \leq 0.03/0.06 µg/ml) showed the lowest MIC₉₀ values among tested agents against BHS (Table 3).

VGS isolates were highly susceptible to telavancin ($MIC_{50/90}$, 0.03/0.06 µg/ml), as were the *S. anginosus* ($MIC_{50/90}$, 0.03/0.06 µg/ml) and *S. bovis* ($MIC_{50/90}$, 0.03/0.06 µg/ml; Table 1) groups. In addition, penicillin susceptibility did not affect the telavancin MIC_{50} and MIC_{90} results ($MIC_{50/90}$, 0.03/0.06 µg/ml), which were 2- and 16-fold more potent than those noted for penicillin ($MIC_{50/90}$, 0.06/1 µg/ml) against VGS, respectively. Furthermore, telavancin was 8- to 32-fold more active than vancomycin ($MIC_{50/90}$, 0.5/0.5 µg/ml), daptomycin ($MIC_{50/90}$, 0.25/0.5 µg/ml), linezolid ($MIC_{50/90}$, 1/1 µg/ml), and levofloxacin ($MIC_{50/90}$, 1/2 µg/ml) against VGS (Table 3).

The telavancin spectrum of activity has been monitored against clinical organisms since 2007 via the SENTRY Program (13-15) and elsewhere (3, 4, 6, 9, 11), from which consistent and potent *in vitro* activity against important Gram-positive isolates has been documented. This study reports the activity of telavancin tested against a worldwide contemporary (2010) collection of clinical pathogens. The results described here demonstrate continued activity of telavancin when tested against indicated Grampositive isolates (16), regardless of susceptibility phenotype or geo-

| TABLE 3 Antimicrobial activities of telavancin and comparator antimicrobia | vial agents tested against a worldwide collection of Gram-po | sitive clinical |
|--|--|-----------------|
| isolates (2010) | | |

| | MIC (µg/ml) | | | % susceptible/resista | nt ^a |
|--|-------------------------------------|-------------|------------------|------------------------|-----------------|
| Organism (no. tested) and antimic robial agent^f | Range | 50% | 90% | CLSI | EUCAST |
| MSSA (4,565) | | | | | |
| Telavancin ^b | 0.03-0.5 | 0.12 | 0.25 | 100.0/-c | 100.0/- |
| Vancomycin | 0.25-2 | 1 | 1 | 100.0/0.0 | 100.0/0.0 |
| Teicoplanin | $\leq 1-4$ | ≤1 | ≤ 1 | 100.0/0.0 | >99.9/<0.1 |
| Daptomycin | $\leq 0.06 - 1$ | 0.25 | 0.5 | 100.0/- | 100.0/0.0 |
| Linezolid | ≤0.12-2 | 1 | 2 | 100.0/0.0 | 100.0/0.0 |
| Levofloxacin | $\leq 0.5 -> 4$ | ≤0.5 | ≤0.5 | 91.7/7.6 | 91.7/7.6 |
| Erythromycin | ≤0.25->4 | ≤0.25 | >4 | 76.4/21.7 | 76.4/22.7 |
| Clindamycin | ≤0.25->2 | ≤0.25 | ≤0.25 | 95.1/4.6 | 94.5/4.9 |
| Quinupristin-dalfopristin | $\leq 0.5 - 4$ | ≤0.5 | ≤0.5 | 99.9/0.1 | 99.9/0.1 |
| Gentamicin | ≤1->8 | ≤1 | ≤ 1 | 97.5/2.1 | 96.6/3.4 |
| Tetracycline | ≤0.25->8 | ≤0.25 | 0.5 | 94.3/5.0 | 93.6/6.1 |
| Trimethoprim-sulfamethoxazole | ≤0.5->4 | ≤0.5 | ≤0.5 | 99.1/0.9 | 99.1/0.7 |
| MRSA (3,088) | | | | | |
| Telavancin | ≤0.015-0.5 | 0.12 | 0.25 | 100.0/- | |
| Vancomycin | 0.25-2 | 1 | 1 | 100.0/0.0 | 100.0/0.0 |
| Teicoplanin | ≤1-4 | ≤ 1 | ≤ 1 | 100.0/0.0 | 99.5/0.5 |
| Daptomycin | ≤0.06-2 | 0.25 | 0.5 | 99.9/- | 99.9/0.1 |
| Linezolid | ≤0.12-8 | 1 | 1 | >99.9/<0.1 | >99.9/<0.1 |
| Levofloxacin | $\leq 0.5 -> 4$ | >4 | >4 | 24.1/74.1 | 24.1/74.1 |
| Erythromycin | $\leq 0.25 ->4$ | >4 | >4 | 16.4/82.9 | 16.4/83.2 |
| Clindamycin | ≤0.25->2 | ≤0.25 | >2 | 59.2/40.7 | 58.9/40.8 |
| Quinupristin-dalfopristin | ≤0.5->4 | ≤0.5 | ≤0.5 | 99.7/0.1 | 99.7/0.1 |
| Gentamicin | ≤1->8 | ≤1 | >8 | 83.0/16.5 | 82.3/17.7 |
| Tetracycline | ≤0.25->8 | ≤0.25 | >8 | 87.6/12.1 | 84.3/12.7 |
| Trimethoprim-sulfamethoxazole | ≤0.5->4 | ≤0.5 | ≤0.5 | 95.0/5.0 | 95.0/4.7 |
| CoNS (1,278) | | | | | |
| Telavancin | ≤0.015-1 | 0.12 | 0.25 | -/- | -/- |
| Vancomycin | 0.25-4 | 1 | 2 | 100.0/0.0 | 99.3/0.7 |
| Teicoplanin | ≤1->8 | 2 | 4 | 99.1/0.0 | 90.8/9.2 |
| Daptomycin | ≤0.06-2 | 0.25 | 0.5 | 99.8/- | 99.8/0.2 |
| | ≤0.12->8 | 0.5 | 1 | 99.4/0.6 | 99.4/0.6 |
| | $\leq 0.25 - > 2$ | >2 | >2 | 26.0/74.0 | 26.0/74.0 |
| Levonoxacin | $\leq 0.5 - > 4$ | 2 | >4 | 46.2/48.8 | 46.2/48.8 |
| Clindomusin | $\leq 0.25 - 24$ | 24 20.25 | >4 | 50.1/02.0 65.5/22.0 | 50.1/05.2 |
| Quinunristin delfonristin | $\leq 0.25 - 2$ | ≤ 0.25 | <pre>>2</pre> | 05.5/55.0 | 04.2/34.3 |
| Centamicin | $\leq 0.5 = 24$ | ≤0.5 ≤1 | 0.5 _8 | 67 5/23 0 | 57 8/42 2 |
| Tetracycline | $\leq 1 = > 0$ $\leq 0.25 = > 8$ | 1 | >8 | 85 2/13 4 | 72 0/16 1 |
| Trimethoprim-sulfamethoxazole | $\leq 0.5 > 4$ | ≤0.5 | >4 | 63.7/36.3 | 63.7/20.8 |
| E. faecalis (1.459) | | | | | |
| Vancomvcin-susceptible (1.421) | | | | | |
| Telavancin | 0.03-1 | 0.5 | 0.5 | 100.0/- | |
| Ampicillin | ≤1-8 | ≤1 | 2 | 100.0/0.0 | 99.6/0.0 |
| Vancomycin | 0.25-4 | 1 | 2 | 100.0/0.0 | 100.0/0.0 |
| Teicoplanin | ≤1-2 | ≤1 | ≤ 1 | 100.0/0.0 | 100.0/0.0 |
| Daptomycin | ≤0.06-4 | 1 | 1 | 100.0/- | -/- |
| Linezolid | 0.25->8 | 1 | 2 | 99.9/0.1 | 99.9/0.1 |
| Levofloxacin | $\leq 0.5 -> 4$ | 1 | >4 | 69.2/30.2 | -/- |
| VanA-type E. faecalis (32) | | | | | |
| Telavancin | 2->2 | >2 | >2 | _/_ | -/- |
| Ampicillin | ≤1-2 | ≤ 1 | 2 | 100.0/0.0 | 100.0/0.0 |
| Teicoplanin | >8 | >8 | >8 | 0.0/100.0 | 0.0/100.0 |
| Daptomycin | 0.5-2 | 1 | 1 | 100.0/- | -/- |
| Linezolid | 0.5–2 | 1 | 1 | 100.0/0.0 | 100.0/0.0 |
| Levofloxacin | 1 -> 4 | $>\!\!4$ | >4 | 6.3/93.8 | -/- |

(Continued on following page)

TABLE 3 (Continued)

| | MIC (µg/ml) | | | % susceptible/resistant ^a | |
|--|-----------------|----------|----------|---|----------------------|
| Organism (no. tested) and antimic robial agent^f | Range | 50% | 90% | CLSI | EUCAST |
| VanB-type E. faecalis (6) | | | | | |
| Telavancin | 0.25-1 | 0.5 | _ | _/_ | -/- |
| Ampicillin | ≤1-2 | ≤ 1 | _ | 100.0/0.0 | 100.0/0.0 |
| Teicoplanin | ≤1-2 | ≤ 1 | — | 100.0/0.0 | 100.0/0.0 |
| Daptomycin | 0.5-2 | 0.5 | _ | 100.0/- | -/- |
| Linezolid | 0.5-1 | 1 | _ | 100.0/0.0 | 100.0/0.0 |
| Levofloxacin | >4 | >4 | - | 0.0/100.0 | -/- |
| E. faecium | | | | | |
| Vancomycin-susceptible (386) | | | | | |
| Telavancin | ≤0.015-0.25 | 0.06 | 0.12 | _/_ | -/- |
| Ampicillin | ≤1->8 | >8 | $>\!\!8$ | 14.2/85.8 | 13.7/85.8 |
| Vancomycin | 0.25-4 | 1 | 1 | 100.0/0.0 | 100.0/0.0 |
| Teicoplanin | $\leq 1 - 4$ | ≤ 1 | ≤ 1 | 100.0/0.0 | 99.7/0.3 |
| Daptomycin | ≤0.06-4 | 2 | 2 | 100.0/- | -/- |
| Linezolid | 0.5-8 | 1 | 2 | 99.2/0.8 | 99.2/0.8 |
| Levofloxacin | ≤0.5->4 | >4 | >4 | 12.7/82.9 | -/- |
| Quinupristin-dalfopristin | ≤0.5->4 | ≤0.5 | 4 | 71.8/11.9 | 71.8/1.0 |
| VanA-type E. faecium (392) | | | | | |
| Telavancin | 0.25->2 | 2 | >2 | -/- | -/- |
| Ampicillin | ≤1->8 | > 8 | > 8 | 0.3/99.7 | 0.3/99.7 |
| Teicoplanin | > 8 | > 8 | $>\!\!8$ | 0.0/100.0 | 0.0/100.0 |
| Daptomycin | 0.12-8 | 2 | 2 | 99.5/- | -/- |
| Linezolid | 0.5->8 | 1 | 1 | 98.7/0.3 | 99.7/0.3 |
| Levofloxacin | >4 | >4 | >4 | 0.0/100.0 | -/- |
| Quinupristin-dalfopristin | $\leq 0.5 -> 4$ | ≤0.5 | 1 | 97.2/1.0 | 97.2/1.0 |
| VanB-type E. faecium (27) | | | | | |
| Telavancin | ≤0.015-1 | 0.12 | 0.5 | -/- | -/- |
| Ampicillin | > 8 | > 8 | $>\!\!8$ | 0.0/100.0 | 0.0/100.0 |
| Teicoplanin | $\leq 1 - 8$ | ≤1 | 8 | 100.0/0.0 | 77.8/22.2 |
| Daptomycin | 0.12-2 | 2 | 2 | 100.0/- | -/- |
| Linezolid | 0.5-4 | 1 | 2 | 96.3/0.0 | 100.0/0.0 |
| Levofloxacin | 4->4 | >4 | >4 | 0.0/100.0 | -/- |
| Quinupristin-dalfopristin | ≤0.5-4 | ≤0.5 | 1 | 88.9/7.4 | 88.9/0.0 |
| S. pneumoniae (2,150) | | | | | |
| Telavancin | ≤0.015-0.12 | ≤0.015 | 0.03 | -/- | -/- |
| Vancomycin | ≤0.12-1 | 0.25 | 0.5 | 100.0/- | 100.0/0.0 |
| Teicoplanin | ≤ 1 | ≤1 | ≤ 1 | -/- | 100.0/0.0 |
| Linezolid | ≤0.12-4 | 1 | 1 | >99.9/- | 100.0/0.0 |
| Penicillin | ≤0.03->4 | ≤0.03 | 4 | 87.6/0.6 ^d /61.9/22.4 ^e | 61.9/12.4 |
| Levofloxacin | $\leq 0.5 -> 4$ | 1 | 1 | 98.8/1.1 | 98.8/1.2 |
| Erythromycin | ≤0.25->4 | ≤0.25 | >4 | 59.2/40.0 | 59.2/40.0 |
| Clindamycin | ≤0.25->1 | ≤0.25 | >1 | 72.3/27.3 | 72.7/27.3 |
| Tetracycline | ≤0.25->8 | 0.5 | >8 | 67.7/32.0 | 67.3/32.3 |
| Beta-hemolytic streptococci (1,472) | | | | | |
| Telavancin | ≤0.015-0.25 | 0.06 | 0.12 | 99.8/- | |
| Penicillin | ≤0.03-0.12 | ≤0.03 | 0.06 | 100.0/- | 100.0/0.0 |
| Vancomycin | ≤0.12-1 | 0.5 | 0.5 | 100.0/- | 100.0/0.0 |
| Teicoplanin | ≤ 1 | ≤ 1 | ≤ 1 | -/- | 100.0/0.0 |
| Daptomycin | ≤0.06-0.5 | ≤0.06 | 0.25 | 100.0/- | 100.0/0.0 |
| Linezolid | ≤0.12-2 | 1 | 1 | 100.0/- | 100.0/0.0 |
| Levofloxacin | $\leq 0.5 -> 4$ | ≤0.5 | 1 | 98.4/1.3 | 94.7/1.6 |
| Erythromycin | ≤0.25->4 | ≤0.25 | >4 | 75.3/23.6 | 75.3/23.6 |
| Clindamycin | ≤0.25->2 | ≤0.25 | >2 | 87.5/12.0 | 88.0/12.0 |
| Viridans group streptococci (519) All (519) | | | | | |
| Telavancin | ≤0.015-0.12 | 0.03 | 0.06 | 100.0/- | 100.0/- |
| | | | | (Continue | d on following page) |

TABLE 3 (Continued)

| | MIC (µg/ml) | | | % susceptible/resist | ant ^a |
|---|-------------|----------|-------|----------------------|------------------|
| Organism (no. tested) and antimicrobial $agent^f$ | Range | 50% | 90% | CLSI | EUCAST |
| Penicillin | ≤0.03->4 | 0.06 | 1 | 74.0/5.4 | 84.0/5.4 |
| Vancomycin | ≤0.12-1 | 0.5 | 0.5 | 100.0/- | 100.0/0.0 |
| Teicoplanin | ≤ 1 | ≤ 1 | ≤1 | -/- | 100.0/0.0 |
| Daptomycin | ≤0.06-2 | 0.25 | 0.5 | 99.6/- | -/- |
| Linezolid | ≤0.12-2 | 1 | 1 | 100.0/- | -/- |
| Levofloxacin | ≤0.5->4 | 1 | 2 | 92.5/6.2 | -/- |
| Erythromycin | ≤0.25->4 | ≤0.25 | >4 | 52.2/45.1 | -/- |
| Clindamycin | ≤0.25->2 | ≤0.25 | >2 | 86.9/12.1 | 87.9/12.1 |
| S. bovis group (32) | | | | | |
| Telavancin | ≤0.015-0.12 | 0.03 | 0.06 | 100.0/- | 100.0/- |
| Penicillin | ≤0.03-0.12 | ≤0.03 | 0.06 | 100.0/0.0 | 100.0/0.0 |
| Vancomycin | 0.25-0.5 | 0.25 | 0.5 | 100.0/- | 100.0/0.0 |
| Teicoplanin | ≤ 1 | ≤ 1 | ≤1 | -/- | 100.0/0.0 |
| Daptomycin | ≤0.06-0.12 | ≤0.06 | ≤0.06 | 100.0/- | -/- |
| Linezolid | 0.5-2 | 1 | 1 | 100.0/- | -/- |
| Levofloxacin | ≤0.5-2 | 1 | 2 | 100.0/0.0 | -/- |
| Erythromycin | ≤0.25->4 | ≤0.25 | >4 | 59.4/37.5 | -/- |
| Clindamycin | ≤0.25->2 | ≤0.25 | >2 | 75.0/25.0 | 75.0/25.0 |

^{*a*} Criteria for susceptibility as published by the Clinical and Laboratory Standards Institute (M100-S21, 2011) and the European Committee on Antimicrobial Susceptibility Testing (2011).

^{*b*} For telavancin, the FDA-approved susceptible breakpoints for *S. aureus* ($\leq 1 \mu g/ml$), vancomycin-susceptible *E. faecalis* ($\leq 1 \mu g/ml$), viridans group streptococci ($\leq 0.12 \mu g/ml$), and beta-hemolytic streptococci ($\leq 0.12 \mu g/ml$) were applied.

^c –, no breakpoint available.

^d Penicillin, parenteral (nonmeningitis).

^e Penicillin (oral penicillin V).

^f Abbreviations: MSSA, methicillin-susceptible S. aureus; MRSA, methicillin-resistant S. aureus; CoNS, coagulase-negative staphylococci.

graphic region. In addition, telavancin demonstrated potency at least 2-fold greater than that of comparators when tested against staphylococci, including MRSA strains. As previously reported, telavancin was less active against vancomycin-resistant enterococcal species; however, these pathogens are not included in the FDA-approved prescribing information listing of susceptible organisms (16).

ACKNOWLEDGMENTS

Expert technical and information support was kindly provided by the following staff members at JMI Laboratories: R. Flamm, M. Castanheira, P. Rhomberg, G. Moet, D. Biedenbach, S. Benning, and M. Janechek.

The study and publication process were funded by Astellas Pharma Global Development, Inc. Circulation of the draft manuscript for scientific review by Astellas Pharma Global Development, Inc., and Theravance, Inc., and collation of comments was conducted by Emily Hutchinson, an employee of Envision Scientific Solutions funded by Astellas Pharma Global Development, Inc.

REFERENCES

- 1. Clinical and Laboratory Standards Institute. 2009. M07-A8. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard, 8th ed. Clinical and Laboratory Standards Institute, Wayne, PA.
- Clinical and Laboratory Standards Institute. 2011. M100-S21. Performance standards for antimicrobial susceptibility testing: 21st informational supplement. Clinical and Laboratory Standards Institute, Wayne, PA.
- 3. Draghi DC, et al. 2008. Comparative surveillance study of telavancin activity against recently collected Gram-positive clinical isolates from across the United States. Antimicrob. Agents Chemother. 52:2383–2388.
- Draghi DC, et al. 2008. In vitro activity of telavancin against recent Gram-positive clinical isolates: results of the 2004–05 Prospective European Surveillance Initiative. J. Antimicrob. Chemother. 62:116–121.
- 5. European Committee on Antimicrobial Susceptibility Testing (EUCAST). January 2011. Breakpoint tables for interpretation of MICs and zone diame-

ters. Version 1.3, January 2011. European Society of Clinical Microbiology and Infectious Diseases, Basel, Switzerland. http://www.eucast.org/clinical _breakpoints/.

- 6. Farrell DJ, Krause KM, Benton BM. 2011. *In vitro* activity of telavancin and comparator antimicrobial agents against a panel of genetically defined staphylococci. Diagn. Microbiol. Infect. Dis. **69**:275–279.
- Gould IM. 2011. Clinical activity of anti-Gram-positive agents against methicillin-resistant *Staphylococcus aureus*. J. Antimicrob. Chemother. 66(Suppl. 4):iv17–iv21.
- Higgins DL, et al. 2005. Telavancin, a multifunctional lipoglycopeptide, disrupts both cell wall synthesis and cell membrane integrity in methicillin-resistant *Staphylococcus aureus*. Antimicrob. Agents Chemother. 49: 1127–1134.
- Karlowsky JA, Adam HJ, Poutanen SM, Hoban DJ, Zhanel GG. 2011. In vitro activity of dalbavancin and telavancin against staphylococci and streptococci isolated from patients in Canadian hospitals: results of the CANWARD 2007–2009 study. Diagn. Microbiol. Infect. Dis. 69:342–347.
- Kosowska-Shick K, et al. 2009. Activity of telavancin against staphylococci and enterococci determined by MIC and resistance selection studies. Antimicrob. Agents Chemother. 53:4217–4224.
- 11. Krause KM, et al. 2008. *In vitro* activity of telavancin against resistant Gram-positive bacteria. Antimicrob. Agents Chemother. **52**:2647–2652.
- 12. Lunde CS, et al. 2009. Telavancin disrupts the functional integrity of the bacterial membrane through targeted interaction with the cell wall precursor lipid II. Antimicrob. Agents Chemother. 53:3375–3383.
- 13. Mendes RE, Moet GJ, Janechek MJ, Jones RN. 2010. *In vitro* activity of telavancin against a contemporary worldwide collection of *Staphylococcus aureus* isolates. Antimicrob. Agents Chemother. 54:2704–2706.
- 14. Mendes RE, Sader HS, Farrell DJ, Jones RN. 2011. Update on the telavancin activity tested against European staphylococcal clinical isolates (2009–2010). Diagn. Microbiol. Infect. Dis. 71:93–97.
- Putnam SD, Sader HS, Moet GJ, Mendes RE, Jones RN. 2010. Worldwide summary of telavancin spectrum and potency against Gram-positive pathogens: 2007 to 2008 surveillance results. Diagn. Microbiol. Infect. Dis. 67:359–368.
- 16. Theravance. 2009. Vibativ package insert. Theravance, South San Francisco, CA. http://www.vibativ.com.