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**The complete coding sequence of the human *raf* oncogene and the corresponding structure of the *c-raf-1* gene**

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**ABSTRACT**

The complete 648 amino acid sequence of the human *raf* oncogene was deduced from the 2977 nucleotide sequence of a fetal liver cDNA. The cDNA has been used to obtain clones which extend the human *c-raf-1* locus by an additional 18.9 kb at the 5' end and contain all the remaining coding exons.

**INTRODUCTION**

Oncogenes are genes which have been identified because they cause cellular transformation either when naturally incorporated into a retrovirus or when introduced into tissue culture cells by transfection. Since these genes are evolutionarily well conserved, they are generally thought to have a normal role in the cell which is subverted by their reintroduction into the cell in a form which alters their function or regulation. A normal function has been identified for 3 of the approximately 25 known oncogenes. The *erbB* gene is derived from the receptor gene for epidermal growth factor (EGF) (1), the *sis* oncogene corresponds to a portion of the gene for platelet derived growth factor (PDGF) (2) and the *fms* gene product has been shown to be related to the CSF-1 receptor (3). The *raf* oncogene was originally isolated from a murine transforming retrovirus, 3611-MSV (4). Subsequently the *mil* (or *mht*) oncogene of the avian virus MH2 was identified as being the avian homologue of *raf* (5, 6). The deduced amino acid sequences of these oncogenes are distantly related to a number of oncogenes which encode tyrosine specific kinases, as well as others which apparently do not (6, 7). Although the viral *raf* and *mil* gene products do not have tyrosine specific kinase activity, they have recently been shown to be associated with a serine-threonine-specific kinase (6, 8). We have recently reported that there are two human genes related to *raf* and *mil* (9). The functional gene, *c-raf-1* contains eleven exons which are homologous to *mil*, nine of which are also homologous to *raf*. The second gene, *c-raf-2*, is a processed pseudogene. In order to

99  
 CCG AAT GTG ACC GCG TCC CCG CTC ACC CCG CCG GGG GAG GAG CCG GCG AGA AGC TGC CCG CBA ACS ACA GGA CST TGG GCG GCG CTC CCC  
 exon 2  
 TCA GBT TTA ADA AAT GTT TAA GCT GCA TCA ATG GAG CAG ATA CAG GGA GCT TGG AAG ACS ATC ABC AAT GBT TTT BGA TTC AAA GAT GCC GTB TTT  
 MET Glu His Ile Glu Gly Ala Trp Lys Thr Ile Ser Asn Gly Phe Val Phe Lys Asp Ala Val Phe  
 297  
 GGC TCC AGC TGC ATC TTC CCT ACA ATA GTT CAG CAG TTT GGC TAT CAG CCG GCA TCA BAT GAT GGC AAA CTC ACA GAT CCT TCT AAG ACA AGC AAC  
 Gly Ser Ser Cys Ile Ile Ser Pro Thr Ile Val Glu Glu Phe Gly Tyr  
 exon 3  
 ACT ATC CGT GTT TTT TGC CCG AAG CAG CAG ABA ACA GTG GTC AAT GTG CBA AAT GGA ATG AGC TTG CAT GAC TGC CTT ATG AAA CCA CTC AAG GTG AGG  
 Thr Ile Arg Val Phe Leu Pro Asn Lys Glu Arg Thr Val Val Asn Val Arg Asn Gly MET Ser Leu His Asp Cys Leu MET Lys Ala Leu Lys Val Arg  
 exon 4  
 GGC CTG CAA CCA GAG TGC TGT BGA GTT TTA CAG CTT CTC CAC GAA CAG AAA GGT AAA AAA GCA CCG TTA BAT TGG AAT ACT BAT GCT GCS TCT TTS ATT  
 Gly Leu Glu Pro Glu Cys Cys Ala Val Phe Arg Leu Leu His Glu His Lys Gly Lys Lys Ala Arg Leu Asp Trp Asn Thr Asp Ala Ala Ser Leu Ile  
 exon 5  
 GGA GAA GAA CTT CAA GTA GAT TTC CTG GAT CAT GTT CCC CTC ACA CAC AAC TTT GCT CCG AAG ACS TTC CTG AAG CTT GCC TTC TGT GAC ATC TGT  
 Gly Glu Glu Leu Glu Val Asp Phe Leu Asn His Val Pro Leu Thr Thr His Asn Phe Ala Arg Lys Thr Phe Leu Lys Leu Ala Phe Cys Asp Ile Cys  
 493  
 CAG AAA TTC CTG CTC AAT GGA TTT CCA TGT CAG TGT GGC TAC AAA TTT CAT GAG CAC TGT AGC ACC AAA GTA CTT ACT ATG TGT GTG GAC TGG AGT  
 Glu Lys Phe Leu Leu Asn Gly Phe Arg Cys Ile Thr Cys Gly Tyr Lys Phe His Glu His Cys Ser Thr Thr Ile Glu Asp Ser Gly Val Pro Ala Leu Asp Trp Asn Thr Asp Ala Ala Ser Leu Ile  
 exon 6  
 AAC ATC ABA CAA CTC TTA TTT CCA AAT TCC ACT ATT GBT GAT AGT GGA GTC CCA GCA CTA CCT TCT TTG ACT ATG CST GAT ATG CBA GAG TCT GTT  
 Asn Ile Arg Glu Leu Leu Leu Phe Pro Asn Ser Thr Ile Gly Asp Ser Gly Val Pro Ala Leu Pro Ser Leu Thr MET Arg Arg MET Arg Glu Ser Val  
 exon 7  
 TCC AGG ATG CCT GTT AAT TCT CAG CAG ABA TAT TCT ACA CCT CAC GCC TTC ACC TTT ACC ACC TCC AGT CCC TCA TCT GAA GBT TCC CTC CCG AAG  
 Ser Arg MET Pro Val Ser Ser Ser Glu His Arg Tyr Ser Thr Pro His Ala Phe Thr Phe Asn Thr Ser Ser Pro Ser Glu Gly Ser Leu Ser Leu Ser Glu Arg  
 exon 8  
 CAG AAG TCG ACA TCC ACA CCT AAT GTC CAC ATG GTC AGC ACC ACS CTS GCT GTG GAC ABC AAG ATG ATT GAG BAT GCA ATT CBA AGT CAG AAG GAA TCA  
 Glu Arg Ser Thr Ser Pro Asn Val His MET Val Ser Thr Thr Leu Pro Val Asp Ser Arg Arg MET Ile Glu Asp Ala Val Lys Ile Leu Lys Val Val Asp Pro Thr Pro Glu Ser  
 exon 9  
 GCT TCA CCT TCA GCC CTG TCC AGT AGC CCC AAC AAT CTG AAG CCA ACA GGC TGG TCA CAG CCG AAA ACC CCC GTG CCA GCA CAA ABA GAG CCG GCA CCA  
 Ala Ser Pro Ser Ala Leu Ser Ser Ser Pro Asn Asn Leu Ser Pro Thr Gly Trp Ser Glu Pro Lys Thr Pro Val Pro Ala Glu Arg Glu Arg Ala Pro  
 exon 10  
 GTA TCT GGG ACC CAG GAG AAA AAC AAA ATT AAG CCT CBT GGA CAG ABA GAT TCA AGC TAT TAT TGG BAA ATA BAA GGC AGT BAA GTG ATG CTG TCC ACT  
 Val Ser Gly Thr Glu Glu Lys Asn Lys Ile Arg Pro Arg Gly Glu Arg Asp Ser Tyr Tyr Trp Glu Ile Glu Ala Ser Glu Val MET Leu Ser Thr  
 exon 11  
 CCG ATT GGG TCA GGC TCT TTT GGA ACT GTT TAT AAG GBT AAA TGG CAG GBA GAT GTT GCA ATA AAG ATC CTA AAG GTT GTC GAC CCA ACC CCA GAG CAA  
 Arg Ile Glu Ser Gly Ser Phe Gly Thr Val Tyr Lys Gly Lys Trp His Gly Asp Val Ala Val Lys Ile Leu Lys Val Val Asp Pro Thr Pro Glu Ser  
 exon 12  
 TTC CAG GCC TTC AAG AAT GAG GTG GCT GTT CTG CCG AAA ACA CCG CAT GTG AAG AAC ATT CTG CTT TTC ATG GGG TAC ATG ACA AAG GAC AAC CTG CCA ATT  
 Phe Glu Ala Phe Arg Asn Glu Val Ala Val Leu Arg Lys Thr Arg His Val Asn Ile Leu Leu Phe MET Gly Tyr MET Thr Lys Asp Asn Leu Ala Ile  
 1485  
 GTG ACC CAG TGG TGC GAG GGC AGC AGC CTC TAC AAA CAG CTG CAT GTC CAG GAG ACC AAG TTT CAG ATG TTC CAG CTA ATT GAC ATT ACC CCG CAG ACS  
 Val Thr Glu Trp Cys Glu Gly Ser Phe His Ala Val His Arg Asp MET Lvs Ser Asn Asn Gly Val Glu Glu Thr Lys Phe Glu MET Phe Glu Ile Asp Ile Ala Ser Glu Thr  
 exon 13  
 GCT CAG GGA ATG BAC TAT TTG CAT GCA AAG AAC ATC ATC CAT ABA BAC ATG AAA TCC AAC AAT ATA TTT CTC CAT GAA GGC TTA ACA GTG AAA ATT GGA  
 Ala Glu Glu MET Asp Thr Leu His Ala Val His Arg Asp MET Lvs Ser Asn Asn Gly Val Glu Glu Thr Lys Phe Glu MET Phe Glu Ile Asp Ile Ala Ser Glu Thr  
 exon 14  
 GAT TTT GBT TTB GCA ACA GTA AAG TCA CCG TGG AGT GBT TCT CAG CAG GTT GAA CAA CCT ACT GGC TCT GTC CTC TGG ATG GCG CCA BAG GTG ATC CBA  
 Asp Phe Gly Leu Ala Thr Val Lys Ser Arg Trp Ser Gly Ser Glu Glu Val Glu Glu Pro Thr Gly Ser Val Leu Trp MET Ala Pro Glu Val Ile Arg  
 1782  
 ATG CAG GAT AAC AAC CCA TTC AAT TCT CAG TCG BAT GTC TAC TCC TAT GGC ATC GTA TTS TAT GAA CTG ATG ACS GGG GAG CTT CCT TAT TCT CAC ATC  
 MET Glu Asp Asn Asn Pro Phe Ser Phe Glu Ser Asp Val Tyr Ser Tyr Gly Ile Val Leu Tyr Glu Leu MET Thr Gly Glu Leu Pro Tyr Ser Ser His Ile  
 exon 16  
 AAC AAC CBA GAT CAG ATC ATC TTC ATG GTG GGC CBA GBA TAT GCC TCC CCA GAT CTT AGT AAG CTA TAT AAG AAC TGC CCC AAA GCA ATG AAG AAG CTG  
 Asn Asn Arg Asp Glu Ile Ile Phe MET Val Gly Arg Gly Tyr Ala Ser Pro Asp Lys Ser Lys Leu Tyr Lys Asn Cys Pro Lys Ala MET Lys Arg Leu  
 exon 17  
 GTA GCT GAC TGT GTG AAG AAA GTA AAG GAG GAG AAG CCT CTT TTT CCC CAG ATC CTG TCT TCC ATT GAG CTG CTC CAA CAC TCT CTA CCG AAG ATC  
 Val Ala Asp Cys Val Lys Lys Val Lys Glu Glu Arg Pro Leu Phe Pro Glu Ile Leu Ser Ser Ile Glu Leu Leu Glu His Ser Leu Pro Lys Ile Asn  
 2879  
 CCG AAG GCT TCC BAG CCA TCC TTB CAT CCG GCA GCG CAG ACT GAG BAT ATC AAT GCT TGC ACC CCG ACC CCG CCG AAG CTG GCT CTC TTC TAG TTB  
 Arg Ser Ala Ser Glu Pro Ser Leu His Arg Ala Ala His Thr Glu Asp Ile Asn Ala Cys Thr Leu Thr Ser Pro Arg Leu Pro Val Phe  
 2199  
 ACTTTCACG TBTCTCAGG CTCCAGGGG AGGAGGAGAA GCCACGACCC ACCACTTTTC TBTCCCTTT CTCCAGAGCC AGACACATG TTTTCABAGA AGCTCTCCTA ABACCTTCT  
 2319  
 AGACTGCTG CAGGGCTTA ACTGATGTT GCLTCTTTT CTATCCCTTT GGGCCCTGG AGAGGAGAGC CATTTCAGT GCTGTGTGT CBTGCTCCT CCCACATTC CCAATGCTG  
 2439  
 AGCCCAACG TCTGTAGT GCGCAGTGG ATTTTATGAG TATACAAA AGCAGGGCC CAGCCCAAC TBTTCCTAC ATGATATT AGAGAGATA AGTATCAGG CAGTCCAGC  
 2597  
 CTATGTGA CAGACATGG ATTTTBAAA TACGCTTCS BABNATGCA TGTACAGCC GGCACCTTCT TCAAGATG GTCCAGGCC AGACATTTT CACATAGCC ACCAGAAC  
 CAGACTG CAGACTCTG GCGCCGAAA GAGGCTGCT TBTACTAT GBACTTTTC TTAGGAGCA CBTCTCCTT TENGACATC TAGGTGTC AGTCATG BATTGTTTC  
 CAGCAGCC ACTCGCAA TCCCATCT AGCCCTCTA GAGCAGTCT TCCATATG TBAATTTT CTCCAGAGG CTGCCCTAT GGGCCGGCC GAGGGCCAG CTTGTTTTC  
 2799  
 TACAGAAA ACAAGAAC AGCTTBTTT CTCTATGAC ATCATGTGA TACAGAAA CAGAGATAC AGTTTTCTT BATTGTTTB GTTTTAMTT TTTTITTTT GACTGTACA  
 2919  
 AAATAGTT ATCTATBT CCTCATTA TBTATTTTA ATAGATAGA TAAATTTAA AAGAAA  
 AAATAGTT ATCTATBT CCTCATTA TBTATTTTA ATAGATAGA TAAATTTAA GATATATG TBTCTATG TBTCTTTC CTCTTTAA ATATATCTG AGCTCAGAC TTATGCCC  
 2944  
 CATTGTCA CCTCTCAG GATCAGATT C  
 CGAT

further characterize the raf gene and its protein product, we have cloned a human cDNA which contains all the coding sequence of the raf gene and have used this DNA as a probe to identify the remaining 5 coding exons of the c-raf-1 gene.

#### MATERIALS AND METHODS

**Library Screening:** Two human cDNA libraries, one derived from fetal liver mRNA (provided by E.F. Fritsch) and one from placental mRNA, in the  $\lambda$ gt10 vector were screened using a raf-specific portion of the cloned 3611-MSV virus (the 0.75 kb XhoI-SstII fragment) as probe (4). Filters were hybridized in 3X SSC at 60°C and washed at 1X SSC at 60°C. Genomic clones were obtained by screening partial EcoRI and HaeIII-AluI libraries in Charon 4A (10) using the 0.9 kb EcoRI-SalI fragment from the 5' end of the 2.9 kb liver cDNA clone.

#### RESULTS

##### Characterization of cDNA Clones

Screening the fetal liver cDNA library with raf specific probe yielded two large clones (2.98 and 2.89 kb). These clones are only slightly smaller than the 3.4 kb raf message which has been observed in numerous tissues (J. Cleveland, personal communication) and contain the complete coding sequence. Similar but shorter clones were also isolated from the placental library. Upon analysis, two of the placental clones were found to contain intron as well as exon sequences.

The sequence of the long liver cDNA clones, as shown in Figure 1, contains a single large open reading frame from nucleotides 121 to 2073 followed by an untranslated region of 905 nucleotides which ends in a poly(A) stretch of 9 nucleotides. This poly(A) stretch is preceded by two AATAAA sequences which could serve as polyadenylation signals (11). To confirm that the apparent poly(A) sequence is not due to priming of the cDNA from an A-rich sequence of the gene, we have compared the cDNA sequence with the sequence of the corresponding region of the human c-raf-1 gene.

Figure 1. The nucleotide sequence of raf cDNA and the deduced amino acid sequence of the raf protein. The bottom two lines show the sequence of the 3' end of the human c-raf-1 gene (9) aligned with the 3' end of the cDNA. The EcoRI site in c-raf-1 is at 40.5 kb in Fig. 2. The presumptive ATP binding site is located near nucleotide 1200 and the majority of the amino acid homology with other kinases lies between nucleotides 1480 and 1773. The first nucleotide of each exon is indicated by an asterisk above the sequence.

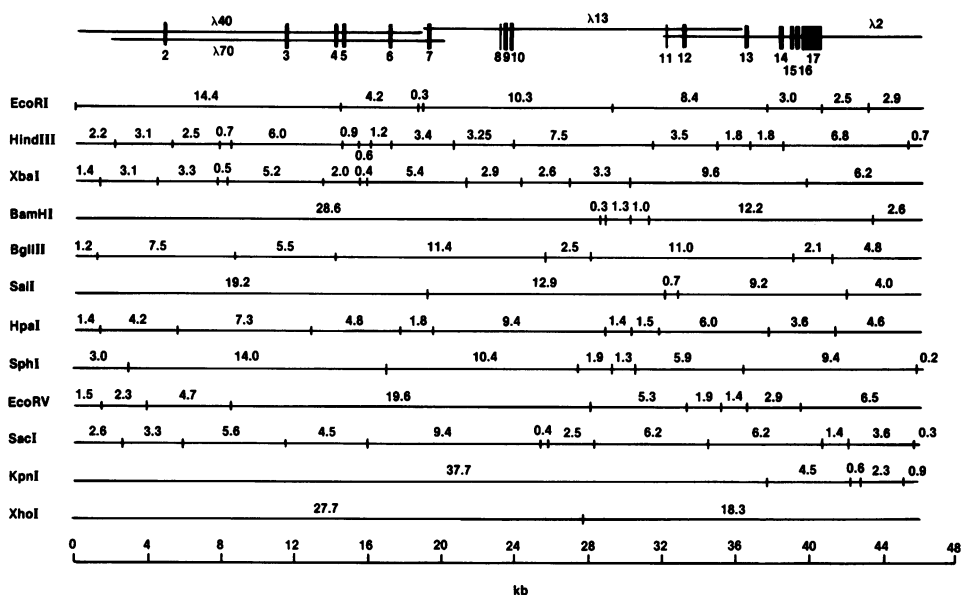


Figure 2. Restriction map of the human *c-raf-1* locus derived from overlapping  $\lambda$  phage clones. Clones  $\lambda 13$  and  $\lambda 2$  have been previously described (9). The positions of four phage clones and the exons which they contain are shown above the map. The 14.4, 10.3, and 8.4 kb *EcoRI* fragments but not the 3.0 and 2.5 kb fragments hybridize to Alu family repeated sequences [BLUR8 (17)].

The sequence alignment shown in Figure 1 demonstrates that the gene has the sequence AGGTGTAAT in place of the poly(A) and thus the poly(A) stretch of the clone represents the true poly(A) of the message. The 1953 nucleotide open reading frame is the only open reading frame of more than 280 nucleotides and coincides with the reading frame of the viral *raf* and *mil* proteins. Its sequence has been confirmed by sequencing the corresponding 16 exons of the human *c-raf-1* gene (see below). The cDNA thus encodes the complete 648 amino acid *raf* protein with a predicted molecular weight of 73023 daltons. This result extends the amino terminal of the sequence known from *v-mil* by 267 amino acids. However, this additional sequence has no significant homology to protein sequences in the National Biomedical Research Foundation Protein Data Bank. As judged by the criterion of Kyte and Doolittle (12), the complete protein contains no extensive hydrophobic regions which would be candidates for transmembrane regions. This characteristic is consistent with the observation of a 75000 molecular weight *raf* protein primarily in the cytosolic fraction of normal rat cells (13).

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### Structure of the c-raf-1 Gene

The availability of a nearly full length cDNA clone allowed us to extend the characterization of the 5' end of the c-raf-1 gene beyond the region of homology to v-mil. Using the 5' portion of the cDNA as probe we have isolated two additional genomic clones,  $\lambda$ HR40 and  $\lambda$ HR70, from partial EcoRI and partial HaeIII-AluI libraries, respectively. These clones extend the 5' end of the restriction map of the locus by 18.9 kb (Fig. 2). The exons in these clones were located approximately by hybridization of cDNA probes to restriction digests of the clones and the appropriate regions were sequenced to precisely identify the exons. Five additional exons were identified which account for all the coding sequences. The intron boundaries all contain characteristic splice acceptor and splice donor sequences (13). Their positions have been indicated on the map of the genomic clones (Fig. 2) and the sequence of the cDNA (Fig. 1). The exon which contains the first 103 nucleotides of the cDNA sequence is not contained within these clones. Although there are about 500 nucleotides of mRNA unaccounted for in the genomic clones, we have assumed for the purpose of numbering the exons that there is only one additional exon which contains all but 26 nucleotides of the 5' untranslated sequence. The structure of the gene has been confirmed by the detection of the expected bands in genomic blots of human DNA using the cDNA as probe (not shown).

### DISCUSSION

We have characterized a nearly fully length cDNA which contains the entire coding sequence of the human raf oncogene. Taking advantage of the HgiAI site which spans the second and third codons of the coding sequence we have begun an attempt to express the complete protein in E. coli. If substantial amounts of protein can be produced it should be very useful in characterizing the function of the gene and in verifying that the raf protein is a serine-threonine specific kinase. The cDNA has also allowed us to identify all the coding exons of the human c-raf-1 gene. This information should be useful in characterizing rearrangements of the gene in human tumors. Five transforming variants of the raf gene have been described, all apparently being truncated at the amino end. The viral raf and mil sequences in the transforming viruses 3611-MSV and MH2 begin within exons 9 and 7 of the mouse and chicken genes, respectively, and extend beyond the termination codon in exon 17. A promoter insertion activated form of the mouse gene has been described (14) in which a retroviral long terminal

repeat sequence was inserted between exons 5 and 6. In addition, two transforming raf DNAs have been identified by transfection of DNA derived from human tumors (15, 16). In the first of these, obtained from a stomach tumor, the transforming DNA has been cloned and partially mapped with EcoRI and Bam HI. Comparison of the map of this DNA with Figure 2 indicates that the EcoRI site at 29.2 kb and the cluster of Bam sites at 28-32 kb (Fig. 2) is present. The 10.3 and 8.4 kb EcoRI fragments on either side of this site in our map agree well with the 10.0 and 8.0 kb fragments of the transforming DNA suggesting that exons 7-13 are present. However, the EcoRI site at 40.6 kb (Fig. 2) is absent in the transforming DNA suggesting a rearrangement near the poly(A) site. The EcoRI fragment immediately 5' of the 10.0 kb fragment in the transforming DNA is a 13.0 kb fragment containing two Bam sites while our clones have 0.3, 4.2, and 14.4 kb EcoRI fragments with no Bam sites. Thus, the transforming DNA is also rearranged at the 5' end. If the agreement between the 10.0 and 10.3 kb EcoRI fragments is not fortuitous, this would place the rearrangement near exons 6 or 7. The second transforming DNA, obtained from a glioblastoma, has only been characterized by hybridization to secondary and tertiary transformants. These transformants all contain human EcoRI fragments of 10, 8, 5, and 4.2 kb detected with a human Alu-family repeated sequence probe. Three of these fragments agree well with the 10.3, 8.4 and 4.2 kb fragment of our clones while the 5 kb fragment does not. Presumably the 5 kb fragment represents a rearrangement in the vicinity of exon 3 or 4. In addition a v-raf probe (containing exons 10-17) detected HindIII fragments of 3.5 and 1.9 as expected from Figure 2 but a 4.0 kb band instead of the 6.8 kb band of normal human DNA. The detection of a Pst 5.65 kb band which maps at 36.3 - 41.9 on Figure 2 (not shown), indicates that the rearrangement of the 6.8 HindIII band occurs 1.4 - 4.8 kb 3' of the poly(A) site (40.5 kb on the map). Thus both transforming DNAs appear to be rearranged at both the 5' and 3' ends. More precise characterization of the rearrangements should now be possible. Nevertheless, since all the transforming variants of the raf gene appear to be truncated at the amino terminus, it is tempting to speculate that this region (exons 2-9) represents a regulatory domain, the loss of which allows inappropriate activity of the apparent kinase domain (exons 10-16).

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