

Published in final edited form as:

*Angew Chem Int Ed Engl.* 2011 March 21; 50(13): 3038–3042. doi:10.1002/anie.201006193.

## Photoregulated Release of Noncovalent Guests from Dendritic Amphiphilic Nanocontainers\*\*

Volkan Yesilyurt, Rajasekharreddy Ramireddy, and Prof. S. Thayumanavan\*

Department of Chemistry, University of Massachusetts Amherst, MA 01003 (USA)

### Keywords

amphiphiles; dendrimers; micelles; supramolecular disassembly

The design of molecular systems with stimuli-sensitive properties is of great interest for applications ranging from drug delivery to gene transfection.<sup>[1]</sup> The use of these molecular systems in drug-delivery applications has been enhanced by imparting amphiphilic character to these designs, as these systems are capable of self-assembling into various supramolecular architectures such as micelles and vesicles, and thus provide interiors that can encapsulate guest molecules noncovalently.<sup>[2]</sup> In this context, significant effort has been devoted to amphiphilic polymers with stimuli-sensitive elements because they are able to 1) form stable micelles, thus providing interiors that can sequester lipophilic guest molecules noncovalently, and 2) release guest molecules in response to both external and internal stimuli such as light,<sup>[3]</sup> pH,<sup>[4]</sup> temperature,<sup>[5]</sup> and reduction/oxidation.<sup>[6]</sup> Since dendrimers can be obtained with a high degree of control over their polydispersity and size,<sup>[7]</sup> it is fundamentally interesting to investigate stimuli-sensitive characteristics in these branched macromolecules. Incorporation of stimuli-sensitive characteristics into dendrimers has been relatively underexplored.<sup>[8]</sup> In particular, amphiphilic dendrimers<sup>[8f,9]</sup> with stimuli-responsive properties would significantly expand the scope of these molecules in a variety of applications. Herein, we describe the design and syntheses of dendritic micelles that can release their guest molecules in response to a light stimulus.

Light-induced release of guest molecules is interesting, because it provides a pathway for releasing a molecule with a remote control, that is, an external physical stimulus.<sup>[3,10]</sup> Light-induced release of lipophilic guest molecules from small-molecule surfactant aggregates has been reported previously.<sup>[3c,11]</sup> While this is interesting, small-molecule-based micelles exhibit rather high critical aggregate concentrations (CACs) and low inherent stabilities. Therefore, light-induced guest release has also been performed with polymer-based micelles.<sup>[3,12]</sup> To the best of our knowledge, there is no prior report on the light-induced disassembly of dendrimer-based micelles, accompanied by the release of a lipophilic guest molecule.

Recently, we reported a unique class of amphiphilic biaryl dendrimers in which every repeating unit in the dendritic backbone contains both lipophilic and hydrophilic functionalities.<sup>[8f,13]</sup> We have shown that these facially amphiphilic biaryl dendrimers form

\*\*We thank the NIGMS of the National Institutes of Health, U.S Army Research Office, and NSF-MRSEC for support.

© 2011 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

\*Fax: (+1)413-545-4490, thai@chem.umass.edu Homepage: <http://www.umass.edu/thaigroup/>.

Dedicated to Professor Peter Beak on the occasion of his 75th birthday.

Supporting information for this article is available on the WWW under <http://dx.doi.org/10.1002/anie.201006193>.

micelle-type aggregates in water and inverted micelle-type aggregates in apolar solvents such as toluene. The micellar aggregates from our dendrimers are formed through aggregation of several dendrimer molecules and can sequester lipophilic guest molecules. The ability of these molecules to assemble and bind guest molecules is dependent on their hydrophilic–lipophilic balance (HLB).<sup>[14]</sup> We hypothesized that a change in the HLB in response to light would result in alterations in the amphiphilic assembly, which should concurrently effect release of guest molecules (Figure 1).

To test this hypothesis we have designed and synthesized amphiphilic dendrimers with a photolabile 2-nitrobenzyl ester moiety as the lipophilic unit (Scheme 1). 2-Nitrobenzyl esters have been widely used as photolabile groups.<sup>[15]</sup> The hydrophilic part of these facially amphiphilic dendrons is based on oligoethylene glycol units. Our molecule is designed in such a fashion that the light-induced cleavage of the nitrobenzyl ester disconnects a significant part of the lipophilic chain from the dendrimer. Moreover, the functionality—a carboxylic acid—in the product, generated on the dendron side of the molecule, is significantly hydrophilic (Figure 1). We envisaged that this transformation should result in a significant change in the HLB and thus cause the supramolecular assembly to release its guest molecules.

The structures of the targeted light-sensitive **G1** and **G2** dendrons are shown in Scheme 1. The dendrons were constructed from a biaryl monomer (**3** in Scheme 2), which was synthesized from the arylstannane **1** and bromoarylester **2** by using Stille coupling as the key step (Scheme 2). Reaction between the peripheral unit **4** and the biaryl building block unit **3** in the presence of potassium carbonate afforded the dendron **5** in 80% yield. Similarly, the corresponding G2 acetylene dendron was synthesized from **3** and the brominated version of the G1 dendron **5**. Attachment of the photolabile nitrobenzyl moiety **8** by a Huisgen 1,3-dipolar cycloaddition reaction (click chemistry) led to the targeted **G1** and **G2** dendrons.<sup>[16]</sup> All dendrons were characterized by <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopy as well as MALDI-TOF mass spectrometry; details of the synthesis and characterization data are outlined in the Supporting Information.

First, we investigated the micellar behavior of these dendrons in the aqueous phase by encapsulating a hydrophobic dye, Nile red. Nile red is not soluble in water, unless it is accommodated in a hydrophobic pocket of micellar aggregates. Emission spectra of Nile red in the presence of various concentrations of **G1** and **G2** dendrons were used to calculate the CACs of these dendrons,<sup>[16]</sup> as about 18 and 20 μM, respectively. Dynamic light-scattering (DLS) experiments further verified the formation of micellar aggregates from the **G1** and **G2** dendrons. The micellar aggregates formed by **G1** and **G2** dendrons are about 80 and 85 nm in diameter, respectively, thus indicating that these dendrons are able to form micelle-like aggregates in water. The light-triggered disassembly of dendritic micellar aggregates was first investigated by monitoring the change in the emission spectrum of Nile red. When a 55 μM solution of Nile red encapsulated **G1** dendron was irradiated at a wavelength of 365 nm we observed a systematic decrease in the emission intensity of Nile red over time, thus indicating disassembly of the micelle and the concomitant release of Nile red from the interiors of the dendritic micelles (Figure 2a). The total amount of dye released, after 200 seconds, was about 88%. When a similar experiment was carried out with the **G2** dendron, a 72% release of the guest molecules was observed (Figure 2b). The smaller amount of Nile red released from the **G2** dendron compared to that from the **G1** dendron is likely due to the more tightly packed nature of the assembly generated from the cleaved form of the higher generation **G2** dendron. The difference in the slopes of the lines in Figure 2b also indicates that a generation-dependant controlled release of the guest molecules can be obtained with these dendrons. The cleavage of photolabile ester groups was further verified by UV/Vis spectroscopy. It is known that cleavage of 2-nitrobenzyl esters leads to the formation of a

by-product, 2-nitrosobenzaldehyde, which weakly absorbs at 360 nm and hence can be detected with absorption spectroscopy. Irradiation of the **G1** and **G2** dendrons with UV light at a wavelength of 365 nm resulted in a decrease in the intensities of the absorption at 320 nm and a concomitant increase at 360 nm over time, thus indicating cleavage of the photolabile ester bond and the formation of the by-product (Figure 2c and 2d).

Next, we were interested in evaluating the size evolution of the dendritic micellar aggregates by using DLS (Figure 3a). The size of the aggregates was found to decrease from about 80 to 37 nm upon irradiation. This result indicates that there is some residual nanoscale assembly in the aqueous phase, even after the photochemical reaction. The change in size, however, shows that there is certainly a change in the nature of the supramolecular assembly. The DLS data, combined with the fact that we have effected a significant release of lipophilic guest molecules, suggest that the dendrimer has been converted from an amphiphilic into a significantly hydrophilic structure. Double hydrophilic macromolecules have been observed previously to assemble into core-shell structures and vesicles.<sup>[17]</sup> The 37 nm diameter of the residual hydrophilic dendrimer can be rationalized on the basis of similar arguments. However, the precise nature of the assembly could not be readily discerned at this time.

Ultimately, we were interested in testing whether the decrease in the emission intensity of Nile red is solely due to the release of the dye molecules from the micellar interior upon exposure to light. For this purpose, we synthesized a first-generation control dendrimer, **G1-control**, which lacks the photocleavable functionalities (Figure 3c). We hypothesized that there should not be any change in the fluorescence of Nile red upon irradiation with UV light at a wavelength of 365 nm if the light does not have any effect on the electronic properties of the dye molecule that cause a change in the emission spectrum. We were gratified to find that exposure of a 55  $\mu\text{M}$  solution of Nile red containing **G1-control** dendron to UV light of a wavelength of 365 nm caused less than 5% guest release, compared to 88% release with the photolabile **G1** dendron (Figure 3b). This finding supports our hypothesis that the decrease in fluorescence obtained with the photo-labile **G1** and **G2** dendrons is indeed due to release of Nile red. Moreover, we carried out DLS studies with the **G1-control** dendron to determine whether UV light has any effect on the size of the aggregate, which may result in leakage of the guest molecule. We indeed found that exposure of the **G1-control** dendron to UV light did not cause any change in the size of the aggregate (Figure 3d), which indicates that UV light does not cause our biaryl dendrimers to undergo any structural change.

In summary, we have designed and synthesized light-sensitive facially amphiphilic dendrimers that can form micellar aggregates in water. The hydrophobic part of these dendrons consists of photolabile ester groups, which are susceptible to cleavage by UV light. We have shown that light-induced cleavage of the hydrophobic ester groups caused the dendrimers to lose their HLB, thereby resulting in dissociation of the micellar aggregates. Since the facially amphiphilic dendrimers provide the opportunity to sequester lipophilic guest molecules noncovalently, the light-induced supramolecular disassembly provides an opportunity to demonstrate photosensitive release of noncovalently sequestered guest molecules from supramolecular aggregates.

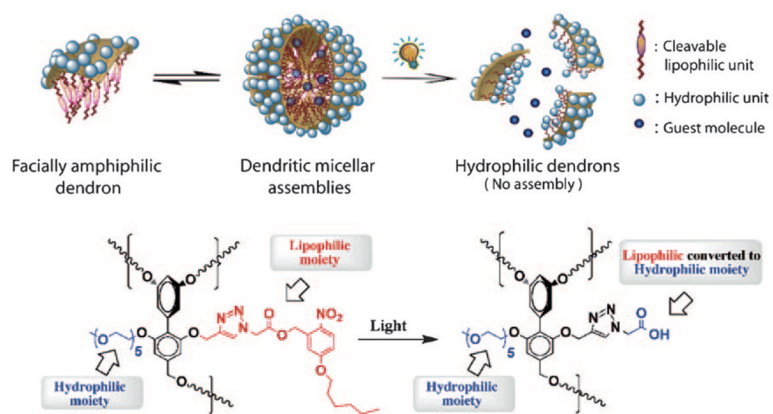
## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

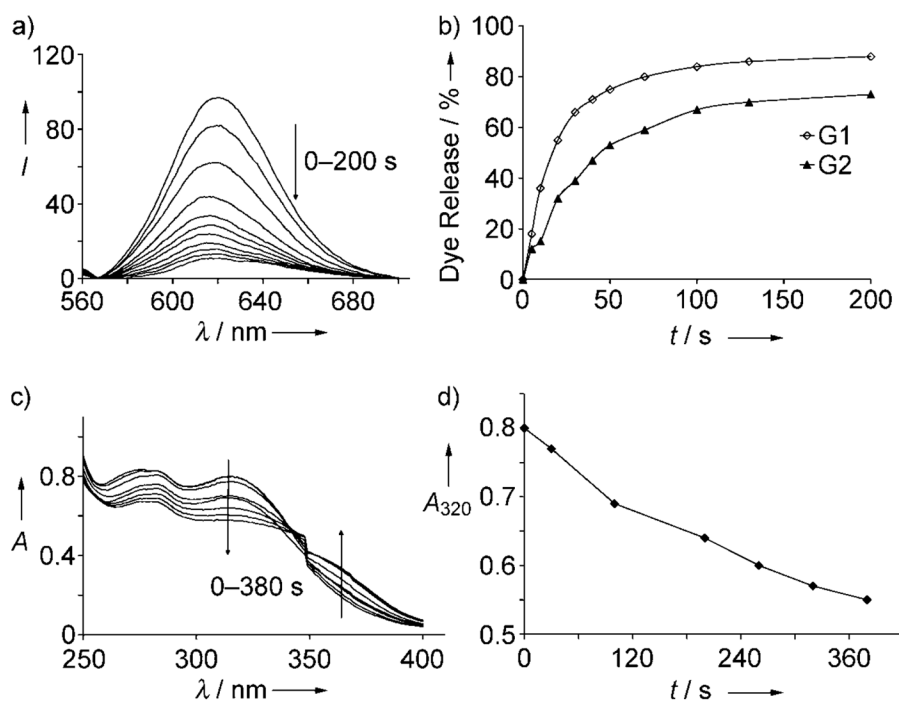
## References

1. a) Han G, Mokari T, Franklin CA, Cohen BE. *J Am Chem Soc.* 2008; 130:15811–15813. [PubMed: 18983148] b) Han G, You CC, Kim BJ, Turingan RS, Forbes NS, Martin CT, Rotello VM. *Angew Chem.* 2006; 118:3237–3271. *Angew Chem Int Ed.* 2006; 45:3165–3169. c) Zhao YL, Li Z, Kabehie S, Botros YY, Stoddart JF, Zink JI. *J Am Chem Soc.* 2010; 132:13016–13025. [PubMed: 20799689] d) Climent E, Bernardos A, Manez RM, Maquieira A, Marcos MD, Navarro NP, Puchades R, Sancenon F, Soto J, Amoros P. *J Am Chem Soc.* 2009; 131:14075–14080. [PubMed: 19739626]
2. a) Brunsveld L, Folmer JB, Meijer EW, Sijbesma RP. *Chem Rev.* 2001; 101:4071–4097. [PubMed: 11740927] b) Andresen TL, Jensen SS, Kent J. *Prog Lipid Res.* 2005; 44:68–97. [PubMed: 15748655] c) Eliaz RE, Nir S, Marty C, Szoka FC. *Cancer Res.* 2004; 64:711–718. [PubMed: 14744789] d) Torchilin VP. *Nat Rev Drug Discovery.* 2005; 4:145–160. e) Duncan R. *Nat Rev Drug Discovery.* 2003; 2:347–360. f) Davis ME, Chen Z, Shin DM. *Nat Rev Drug Discovery.* 2008; 7:771–782. g) Savic R, Laibin L, Eisenberg A, Maysinger D. *Science.* 2003; 300:615–618. [PubMed: 12714738] h) Discher DE, Eisenberg A. *Science.* 2002; 297:967–973. [PubMed: 12169723]
3. a) Jiang J, Tong X, Morris D, Zhao Y. *Macromolecules.* 2006; 39:4633–4640. b) Jiang J, Tong X, Zhao Y. *J Am Chem Soc.* 2005; 127:8290–8291. [PubMed: 15941255] c) Goodwin AP, Mynar JL, Ma Y, Feleming GR, Fréchet JMJ. *J Am Chem Soc.* 2005; 127:9952–9953. [PubMed: 16011330] d) Babin J, Pelletier M, Lepage M, Allard JF, Morris D, Zhao Y. *Angew Chem.* 2009; 121:3379–3382. *Angew Chem Int Ed.* 2009; 48:3329–3332.
4. a) Klaiherd A, Nagamani C, Thayumanavan S. *J Am Chem Soc.* 2009; 131:4830–4838. [PubMed: 19290632] b) Gillies ER, Jonsson TB, Fréchet JMJ. *J Am Chem Soc.* 2004; 126:11936–11943. [PubMed: 15382929] c) Gillies ER, Fréchet JMJ. *Chem Commun.* 2003:1640–1641. d) Jung J, Lee IH, Lee E, Park J, Jon S. *Biomacromolecules.* 2008; 8:3401–3407. [PubMed: 17939711] e) Gohy JF, Willet N, Varshney S, Zhang JX, Jerome R. *Angew Chem.* 2001; 113:3314. *Angew Chem Int Ed.* 2001; 40:3214–3216.
5. a) Yotaro M. *Angew Chem.* 2007; 119:1392–1394. *Angew Chem Int Ed.* 2007; 46:1370–1372. b) You YZ, Oupicky D. *Biomacromolecules.* 2007; 8:98–105. [PubMed: 17206794] c) Zhang Q, Clark CG, Wang M, Remsen EE, Wooley KL. *Nano Lett.* 2002; 2:1051–1054.
6. a) Ghosh S, Irvin K, Thayumanavan S. *Langmuir.* 2007; 23:7916–7919. [PubMed: 17567057] b) Ryu J, Roy R, Ventura J, Thayumanavan S. *Langmuir.* 2010; 26:7086–7092. [PubMed: 20073533] c) Takae S, Miyata K, Oba M, Ishii T, Nishiyama N, Itaka K, Yamasaki Y, Koyama H, Kataoka K. *J Am Chem Soc.* 2008; 130:6001–6009. [PubMed: 18396871] d) Li Y, Lokitz BS, Armes SP, McCormick CL. *Macromolecules.* 2006; 39:2726–2728.
7. a) Bosman AW, Janssen HM, Meijer EW. *Chem Rev.* 1999; 99:1665–1688. [PubMed: 11849007] b) Grayson SM, Fréchet JMJ. *Chem Rev.* 2001; 101:3819–3867. [PubMed: 11740922] c) Fischer M, Vögtle F. *Angew Chem.* 1999; 111:934–955. *Angew Chem Int Ed.* 1999; 38:884–905. d) Tomalia DA, Svenson S. *Adv Drug Delivery Rev.* 2005; 57:2106–2129. e) Tomalia DA, Fréchet JMJ. *J Polym Sci Part A.* 2002; 40:2719. f) Menjoge AR, Kannan RM, Tomalia DA. *Drug Discovery Today.* 2010; 15:171–185. [PubMed: 20116448] g) Rosen BM, Wilson CJ, Wilson DA, Peterca M, Imam MR, Percec V. *Chem Rev.* 2009; 109:6275–6540. [PubMed: 19877614]
8. a) Jiang DL, Aida T. *Nature.* 1997; 388:454–456. b) Amir RJ, Pessah N, Shamis M, Shabat D. *Angew Chem.* 2003; 115:4632–4637. *Angew Chem Int Ed.* 2003; 42:4494–4499. c) Kojima C, Haba Y, Fukui T, Kono K, Takagishi T. *Macromolecules.* 2003; 36:2183–2186. d) Avital-Shmilovici M, Shabat D. *Soft Matter.* 2010; 6:1073–1080. e) Kostianen MA, Kasyutich O, Cornelissen JJLM, Nolte RJM. *Nat Chem.* 2010; 2:394–399. [PubMed: 20414241] f) Aathimanikandan SV, Savariar EN, Thayumanavan S. *J Am Chem Soc.* 2005; 127:14922–14929. [PubMed: 16231948] g) Nishiyama N, Iriyama A, Jang WD, Miyata K, Itaka K, Inoue Y, Takahashi H, Yanagi Y, Tamaki Y, Koyama H, Kataoka K. *Nat Mater.* 2005; 4:934–941. [PubMed: 16299510] h) Kostianen MA, Smith DK, Ikkala O. *Angew Chem.* 2007; 119:7744–7748. *Angew Chem Int Ed.* 2007; 46:7600–7604.
9. For some examples of amphiphilic dendrimers see: Percec V, Wilson DA, Leowanawat P, Wilson CJ, Hughes AD, Kaucher MS, Hammer DA, Levine DH, Kim AJ, Bates FS, Davis KP, Lodge TP, Klein ML, DeVane RH, Aqad E, Rosen BM, Argintaru AO, Sienkowska MJ, Rissanen K,

- Nummelin S, Ropponen J. *Science*. 2010; 328:1009–1014. [PubMed: 20489021] Joester D, Losson M, Pugin R, Heinzlmann H, Walter E, Merkle HP, Diederich F. *Angew Chem*. 2003; 115:1524–1528. *Angew Chem Int Ed*. 2003; 42:1486–1490. Cooper AI, Londono JD, Wignall G, McClain JB, Samulski ET, Lin JS, Dobrynin A, Rubinstein M, Burke ALC, Fréchet JMJ, DeSimone JM. *Nature*. 1997; 389:368–371. Newkome GR, Moorefield CN, Baker GR, Johnson AL, Behera RK. *Angew Chem*. 1991; 103:1205–1207. *Angew Chem Int Ed Engl*. 1991; 30:1176–1178. Hawker CJ, Wooley KL, Fréchet JMJ. *J Chem Soc Perkin Trans 1*. 1993:1287–1297.
10. Wang Y, Han P, Xu H, Wang Z, Zhang X, Kabanov AV. *Langmuir*. 2010; 26:709–715. [PubMed: 19627165]
11. a) Wang Y, Ma N, Wang Z, Zhang X. *Angew Chem*. 2007; 119:2881–2884. *Angew Chem Int Ed*. 2007; 46:2823–2826. b) Orihara Y, Matsumura A, Saito Y, Ogawa N, Saji T, Yamaguchi A, Sakai H, Abe M. *Langmuir*. 2001; 17:6072–6076.
12. Liu X, Jiang M. *Angew Chem*. 2006; 118:3930–3934. *Angew Chem Int Ed*. 2006; 45:3846–3850.
13. a) Vutukuri DR, Basu S, Thayumanavan S. *J Am Chem Soc*. 2004; 126:15636–15637. [PubMed: 15571373] b) Gomez-Escudero A, Azagarsamy MA, Theddu N, Vachet RW, Thayumanavan S. *J Am Chem Soc*. 2008; 130:11156–11163. [PubMed: 18661986]
14. a) Azagarsamy MA, Sokkalingam P, Thayumanavan S. *J Am Chem Soc*. 2009; 131:14184–14185. [PubMed: 19757790] b) Azagarsamy MA, Yesilyurt V, Thayumanavan S. *J Am Chem Soc*. 2010; 132:4550–4551. [PubMed: 20232866]
15. a) Mayer G, Heckel A. *Angew Chem*. 2006; 118:5020–5042. *Angew Chem Int Ed*. 2006; 45:4900–4921. b) Fomina N, McFearin C, Sermsakdi M, Edigin O, Almutairi A. *J Am Chem Soc*. 2010; 132:9540–9542. [PubMed: 20568765] c) Kloxin AM, Kasko AM, Salinas CN, Anseth KS. *Science*. 2009; 324:59–63. [PubMed: 19342581]
16. See the Supporting Information for details.
17. a) An Z, Shi Q, Tang W, Tsung CK, Hawker CJ, Stucky GD. *J Am Chem Soc*. 2007; 129:14493–14499. [PubMed: 17967023] b) Pasparkis G, Alexander C. *Angew Chem*. 2008; 120:4925–4928. *Angew Chem Int Ed*. 2008; 47:4847–4850. c) Savariar EN, Aathimanikandan SV, Thayumanavan S. *J Am Chem Soc*. 2006; 128:16224–16230. [PubMed: 17165775]

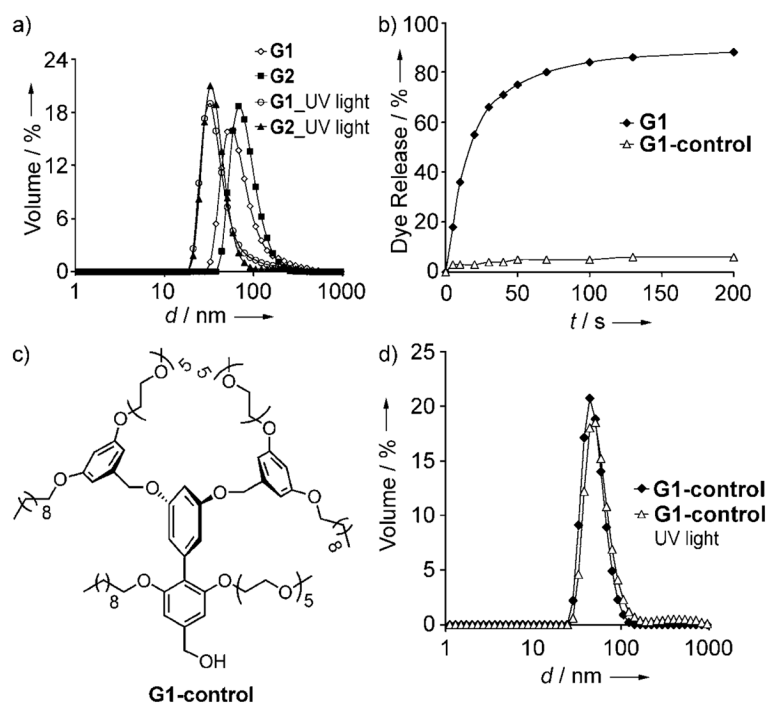


**Figure 1.** Schematic representation of the light-induced disassembly of dendritic micellar assemblies.



**Figure 2.**

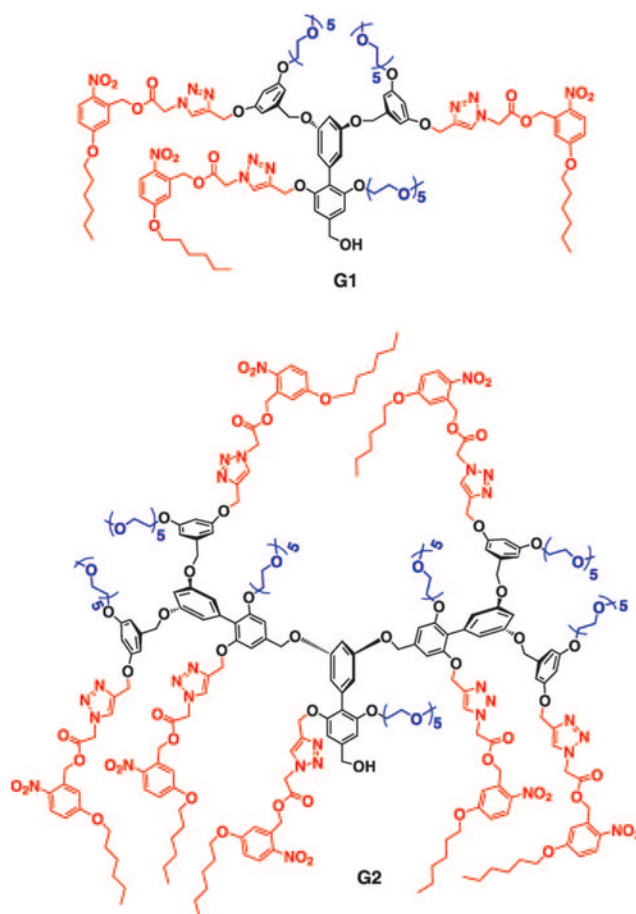
a) Release of Nile red from a 55  $\mu\text{M}$  micellar solution of the **G1** dendron upon irradiation with UV light for different time intervals (0–200 s), b) release of Nile red from the **G1** and **G2** dendrons upon irradiation with UV light, c) UV/Vis spectra of the **G1** dendron upon irradiation with UV light for different time intervals (0–380 s), d) plot of the absorbance at 320 nm, which illustrates cleavage of the photolabile ester bond.



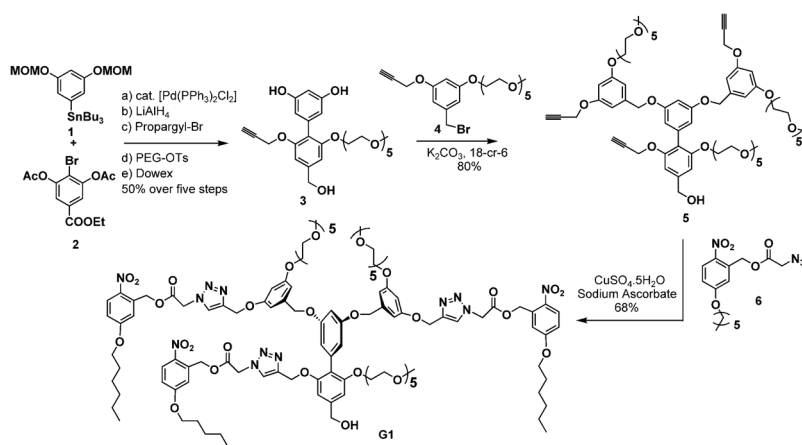
**Figure 3.**

a) Size evolution of 55  $\mu\text{M}$  solutions of the **G1** and **G2** dendrons upon irradiation with UV light, b) comparison of dye release with the photolabile **G1** and **G1-control** dendrons, c) structure of the **G1-control** dendron, d) sizes of the **G1-control** dendron before and after UV irradiation.





**Scheme 1.**  
Structures of the photo-cleavable **G1** and **G2** dendrons.

**Scheme 2.**

Synthetic scheme for the photolabile **G1** dendron (18-cr-6 = [18]crown-6; MOM = methoxymethyl; PEG-OTs = tosylate of pentaethylene-glycol monomethyl ether).