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Beyond the monoaminergic hypothesis: neuroplasticity and epigenetic changes in a transgenic mouse model of depression

Renaud Massart^{1,2,3}, Raymond Mongeau^{1,2}

and Laurence Lanfumev^{1,2,*}

¹Inserm, U894, Centre de Psychiatrie et Neurosciences, 75013 Paris, France ²UPLC, University Paris 06, UMR 5894, 75013 Paris, France ³Department of Pharmacology and Therapeutics, McGill University, Montreal, Quebec, Canada H3G 1Y6

The monoamine hypothesis of depression has dominated our understanding of both the pathophysiology of depression and the action of pharmacological treatments for the last decades, and it has led to the production of several generations of antidepressant agents. However, there are serious limitations to the current monoamine theory, and additional mechanisms, including hypothalamic-pituitaryadrenal (HPA) axis dysfunctions, as well as neurodegenerative and inflammatory alterations, are potentially associated with the pathogenesis of mood disorders. Moreover, new data have recently indicated that epigenetic mechanisms such as histone modifications and DNA methylation could affect diverse pathways leading to depression-like behaviours in animal models. In a transgenic mouse model of depression, in which a downregulation of glucocorticoid receptors (GR) causes a deficit in the HPA axis feedback control, besides alterations in monoamine neurotransmission and neuroplasticity, we found modifications in the expression of many proteins involved in epigenetic regulation, as well as clock genes, in the hippocampus and the frontal cortex, that might be central in the genesis of depressive-like behaviours.

Keywords: depression; glucocorticoid receptors; mice; epigenetics; stress; monoamines

1. INTRODUCTION

Although we are repeatedly exposed to adversity and stressful situations throughout life, inter-related hormonal, neurobiological and circadian system networks maintain physiological and behavioural homeostasis in most individuals. This functional equilibrium is determined by genetic- and environmentally driven epigenetic elements that interact in complex and still poorly understood ways. Disruptions in the interactions or imbalances between these systems could cause various neurobehavioural disorders such as depression. Depression is a chronic, recurring, life-threatening illness that affects up to 14 per cent of the population and causes a significant burden at both the individual and the society level [1]. Available treatments of depression are suboptimal and still unsatisfactory; only 50 per cent of treated depressed patients achieve full remission. Improvement of therapeutic strategies depends on the identification of underlying pathological processes and mechanisms of action of current treatments. Despite significant advances, the causes of depression and the molecular basis of treatments are still poorly understood. Various theories have been proposed to account for the overall pathophysiological state or particular symptoms of depression based on dysfunction of monoamine neurotransmission [2], the hypothalamic-pituitary-adrenal (HPA) axis [3], circadian rhythms [4] or neuroimmune processes [5]. Besides these major theories, there has recently been considerable interest in the ideas that epigenetic modifications could be biological markers of both depression and antidepressant (AD) efficacies, or that they could be directly involved in the pathophysiology of depression and in the action of AD compounds [6]. This review briefly summarizes the most prominent theories and focuses more specifically on a genetic model of the HPA axis dysregulation in depression, glucocorticoid receptor-impaired (GR-i) mice. We will see that these various hypotheses are far from being mutually exclusive.

2. OVERVIEW OF THE VARIOUS HYPOTHESES (a) The monoamine hypothesis is central but remains insufficient

Our views on the pathophysiology of depression have been dominated by the monoamine hypothesis, based principally on the efficacy of both first- and secondgeneration ADs suggesting that an imbalance, mainly in serotonergic and noradrenergic neurotransmission, is at the core of the pathophysiology of depression. Several classes of ADs are currently available [7], and

^{*} Author and address for correspondance: Centre de Psychiatrie et Neuroscience, Université Pierre et Marie Curie (UPMC), Inserm U 894, Site Pitié Salpêtrière, 91 Bd de l'Hôpital, 75013 Paris, France (laurence.lanfumey@upmc.fr).

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most of them affect monoaminergic transmissions in one way or another. Monoamine oxidase inhibitors (MAOIs), originally derived from drugs developed to treat tuberculosis (e.g. izoniazid) were observed to improve mood in patients. However, the first MAOIs had serious side effects, and their use was problematic because of the strict diet people needed to follow in order to prevent hypertensive reactions induced by food rich in tyramine. Tricyclic AD compounds (TCAs) such as imipramine, derived from structurally related molecules used to treat psychosis, also acted on the monoaminergic system and were very effective in relieving depressive symptoms. However, TCAs also induce aversive effects, presumably because of their action on other transmission systems (mainly histamine and acetylcholine). Drugs selectively blocking monoamine transporters, and thus leading to a specific increase in extracellular monoamine levels, were then developed. Compounds displaying selective serotonin (5-hydroxytryptamine, 5-HT) reuptake inhibition (SSRIs) were first produced in view of clinical findings, such as the strong relationship between suicide completion and low levels of the 5-HT metabolite (5-HIAA) in the cerebrospinal fluid of depressed patients [8]. The majority of ADs prescribed since the 1990s belong to this SSRI family, but more recently, drugs acting selectively on both the norepinephrine and the serotonine transporters (mixed 5-HT/ norepinephrine reuptake inhibitors (SNRIs)) have also shown clinical benefits. In addition, atypical ADs having no direct effects on monoamine reuptake and primarily acting as antagonists at monoaminergic receptors are currently used. Overall, it remains reasonable to hypothesize that depression is caused by inadequate monoamine neurotransmission, as ADs act by increasing monoamine availability and by producing long-term adaptive changes in monoaminergic receptor sensitivity $(5-HT_{1A} \text{ autoreceptors}, 5-HT_2)$ β -adrenergic and α_2 -adrenergic heteroreceptors, etc.) [9]. However, although positive AD responses are transiently reversed in patients under low tryptophan diet leading to 5-HT depletion, this depletion does not worsen symptoms in unmedicated depressed patients [10]. In addition, 5-HT depletion by itself does not cause depression in healthy volunteers, undermining our view about the crucial role of a decrease in the serotonergic tone to trigger depressive episodes. Therefore, factors beyond monoamine deficiency or imbalance are most probably implicated in the development of major depression. Moreover, all available ADs exert their effects only after prolonged administration (several weeks to months), which suggests that their short-term effects on monoaminergic transmission are not directly responsible for the clinical efficacy of these drugs. Rather, long-term adaptations to AD treatment would appear to mediate their therapeutic action. Finally, it is necessary to search beyond the monoaminergic hypothesis because, albeit generally safe, monoamine-based ADs are far from being ideal drugs. The side effects of TCAs and SSRIs may be of sufficient severity to cause discontinuation of treatment in 19 per cent and 15 per cent of patients, respectively [11]. Moreover, in 30 per cent of the cases, there is little or no response to the medication, and almost 50 per cent

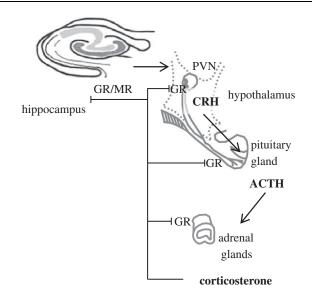


Figure 1. Regulation of the hypothalamic-pituitary-adrenal (HPA) axis. GRs, in particular in the hippocampus and the paraventricular nucleus of the hypothalamus (PVN) exert a negative feedback regulation on the secretion of ACTH and corticosterone. GR-i mice are deficient in GR expression. CRH, corticotropin-releasing hormone; ACTH, adrenocorticotropic hormone; GR, low-affinity glucocorticoid receptor; MR, high affinity mineralocorticoid receptors.

of patients treated with a TCA do not show significant clinical improvements [12].

(b) The corticotrope hypothesis: an opening to the neuroimmune and the neuroplasticity theories of depression

Epidemiological data have provided strong support to the idea that stressful life events play a role in the aetiology of depression [13], and this would occur in interaction with genetic factors [14]. The involvement of stress in the development of depressive symptoms may involve several systems, including the monoamines and inflammation factors interacting with the HPA axis, which exerts a central role in the regulation of the stress response. By mediating the perceived stress response, the HPA axis is an essential component of an individual's ability to cope with stress. The activity of the HPA axis is governed by the secretion of corticotropin releasing hormone (CRH) and arginine-vasopressin (AVP) from the hypothalamus, which, in turn, activate the secretion of the adrenocorticotropic hormone (ACTH) from the pituitary. ACTH then stimulates the secretion of corticoids (cortisol in humans and corticosterone in rodents) from the adrenal cortex, and these hormones interact with their receptors (low-affinity glucocorticoid recep-(GRs) and high-affinity mineralocorticoid tors receptors (MRs)) in multiple target tissues throughout the body including the brain. Glucocorticoids exert a negative feedback control on CRH and AVP secretion through the activation of GRs expressed, among others, by hippocampal and paraventricular nucleus neurons (figure 1). Besides their involvement in the negative feedback regulation, these receptors also enhance hippocampal function and thereby promote certain cognitive abilities [15].

Figure 2. Imbalance in tryptophan metabolism caused by pro-inflammatory cytokines and glucocorticoids. The induction by pro-inflammatory cytokines of the enzyme indolamine 2,3-dioxygenase (IDO) and by glucocorticoids of the tryptophan 2,3-dioxygenase (TDO), which both convert tryptophan into kynurenine, leads to decreased 5-HT synthesis via tryptophan hydroxylase 2 (TPH2).

The corticosteroid receptor hypothesis of depression is based on dysfunctions of the HPA axis observed in the majority of depressed patients [3,16]. HPA hyperactivity results from a deficit in the negative feedback regulation of the axis, as evidenced by the failure of GR activation to decrease plasma levels of cortisol in the 'dexamethasone suppression test' [17]. In this test, a low dose of dexamethasone, an exogenous steroid that activates GRs, normally suppresses cortisol secretion in healthy individuals. A high proportion of depressed patients are 'non-suppressor' in the dexamethasone test, in particular individuals with severe psychotic features and suicidal ideation [18]. In addition, dexamethasone administration suppresses mitogen-induced lymphocyte proliferation and the production of the pro-inflammatory cytokine interleukin- 1β in healthy controls, but not in depressed patients [19]. Dexamethasone administration also induces an increase in the number of neutrophils and a decrease in the number of lymphocytes in healthy controls, but these effects are not observed in non-suppressors, suggesting that depression is also associated with changes in immune function which confer a resistance to the effects of glucocorticoids on immunity [20]. Other data indicate that cytokine-mediated inflammatory processes might play an important role in neurochemical changes associated with depression. In particular, both glucocorticoids and pro-inflammatory cytokines lead to a decrease in the synthesis of 5-HT and increase neurotoxic metabolites by enhancing the conversion of its precursor tryptophan into kynurenine [21] (figure 2). Depressive disorders are highly prevalent in infectious, autoimmune and neurodegenerative disorders [22] and, in turn, depressed patients show high levels of pro-inflammatory cytokines and other proteins involved in the immune response [23]. Interestingly, half of the patients treated with the cytokine interferon- α develop depressive symptoms that can be treated with classical ADs [24], further illustrating how inflammation factors may trigger depression. On the other hand, AD drugs were shown to reduce inflammation, possibly through inhibition of the release of pro-inflammatory cytokines from activated macrophages, via monoaminergic receptors located on immune cells [25].

At the anatomo-functional level, the hippocampus seems to be especially sensitive to the deleterious

effects of stress. Neuronal loss has been reported in the hippocampus of stressed or corticosterone-treated animals. Moreover, human brain imaging studies illustrate how stress and depression may cause reductions of the hippocampal volume, probably because of dendritic arborization atrophy and loss of neurons, as observed in post-mortem hippocampi of patients suffering from major depressive disorder [26]. The fact that these morphometric alterations are most often attenuated, or even reversed, by AD drugs further suggests that they are related to depression. They are probably under the regulation of both GRs and MRs since recent data also showed that prenatal stress in rodents impairs the morphological and functional maturation of hippocampal granule cells in adult offspring via MR downregulation [27]. Furthermore, convergent data obtained in several animal species showed that stress and glucocorticoids exert a drastic negative effect on the rate of cell proliferation, leading to a rapid and prolonged decrease in neurogenesis in the hippocampus of adult rodents [28]. Whether this effect is also observed in depressed patients is still unclear [29]. Regardless of the nature of hippocampal damages, these changes are likely to reduce the inhibitory control that the hippocampus exerts on the HPA axis, which would increase circulating glucocorticoid levels and lead to subsequent hippocampal dysfunction. Although ablation of neurogenesis has no effect on basal HPA axis activity, it prevents monoaminergic AD drugs from restoring the negative inhibitory function of the hippocampus when glucocorticoid levels become elevated in chronic stress conditions [30]. Hippocampal neurogenesis was also proposed to contribute to memory formation [31]. A reduction in neurogenesis can theoretically contribute to the cognitive symptoms of depression, even though by itself it is unlikely to produce mood disorders [32].

Neurogenesis is mainly under the control of factors controlling neuroplasticity. Extensive studies in both humans and validated animal models of depression have indicated that monoamines produce their effects through signalling pathways that regulate neuroplasticity and cell survival, such as the calcium/cyclic-AMP responsive-element-binding protein (CREB) and the brain-derived neurotrophic factor (BDNF) pathways. In rodents, both stress and glucocorticoids are known to reduce brain levels of neurotrophic factors, including BDNF mRNA and protein [33-35]. In contrast, different classes of AD drugs increase BDNF expression in the hippocampus [36]. In humans, post-mortem studies have shown that BDNF expression is reduced in the cerebral cortex of depressed patients compared with healthy controls, but increased in patients receiving an AD at the time of death [37]. All these data suggest that neurotrophic factors are involved in AD therapy. How much these neuroplasticity changes are important for mood regulation and adaptive behavioural responses to stress is still an open question. Interestingly, recent data support the hypothesis that hyperactivity of the HPA axis is not a simple consequence or an epiphenomenon of depression, but a risk factor predisposing the patient to the development of depression [38], brought about by genetic liability as well as by early life experiences

which programme molecular changes including epigenetic modifications (see below) [14,39].

(c) Circadian system dysregulations: old observations and new concepts

Circadian rhythm abnormalities have been considered for years to play a role in mood disorders, but the exact nature and the mechanisms of these abnormalities are still poorly understood [40]. Biological clocks and the so-called clock genes regulate many behaviour and physiological functions, such as sleep or feeding. These functions are greatly altered in depression and the severity of depressive symptoms varies with a daily pattern. Circadian rhythms of many biological variables such as the daily profiles of body temperacortisol, thyrotropin, prolactin, growth ture, hormone, melatonin and excretion of various metabolites are disrupted in depressed patients [41]. Although no clock gene polymorphism is apparently associated with major depression per se, a polymorphism has been linked to the recurrence of bipolar depression [42]. Most AD drugs and mood stabilizers influence endogenous rhythms. For instance, the AD agomelatine, a melatonin receptor agonist and a 5-HT_{2C} receptor antagonist, improves disturbed sleep-wake rhythms in depressed patients and adjusts the circadian system by resynchronizing the body temperature and cortisol levels [43,44]. Moreover, the SSRI fluoxetine appears to modulate clock gene expression [45]. Finally, 'chronotherapeutics' such as sleep deprivation have been found to be efficient for clinical treatment of depressed patients [46]. Their rapid and transient effects can be stabilized with common AD treatments [47].

(d) Current issues about the role of epigenetics in depression

Aetiological studies of depression have traditionally focused primarily on genetic factors. However, epigenetic factors, independent of DNA sequence variations, appear to be of considerable interest in understanding the effect of early life stress in depression [48-50]. The epigenome that programmes the genome consists of covalent modifications of DNA by methylation of cytosines and a panel of different histone modifications and chromatin remodelling factors. Non-coding RNAs, including siRNA, antisense RNA and microRNAs are emerging as important regulators of stable gene expression and are integrated with both chromatin and DNA methylation epigenetic functions. Although the epigenetic programme is ruled by a highly organized developmental process that is tightly controlled to maintain tissue-specific patterns of gene expression and is generally similar in different individuals, recent data suggest that it might still be sensitive to input from the environment, especially in the early life period. Environmental conditions can modulate the pattern of DNA methylation, while maintaining the overall cell-type specificity. It has been hypothesized [51] that cues from social and physical environments early in life can cause variations in epigenetic programming that serve as an 'adaptive' response of the genome to the anticipated lifelong

environment. A misfit between the 'adaptive' response and the actual environment later in life would result in maladaptation and an increased risk of developing diseases. In this context, certain environmental exposures early in life might result in persistent epigenetic reprogramming that contributes to the risk of later developing psychiatric diseases. For instance, the role of DNA methylation variations in sustaining the effects of early environmental experience in moodrelated disorders, notably the dysregulation of the HPA axis activity, has been first demonstrated by Meaney and Szyf's laboratories in the context of postnatal mother-infant interactions [52]. Interestingly, this involves 5-HT-dependent synaptic transmission and binding of the NGFIA transcription factor to the GR promoter site [53]. Since then, other epigenetic modifications, notably of BDNF or arginine/vasopressin, were found to be linked to depression [54,55]. Epigenetic variations, in contrast to genetic variations, are potentially reversible by subsequent modulation of epigenetic enzymes [56]. Interestingly, various classical AD compounds display epigenetic effects, while histone deacetylases or DNA methylation inhibitors exhibit ADlike effects [57]. Therefore, the study of epigenetics in mood disorders bears the potential optimism to reshape our comprehension of the molecular aetiology of depression and the possibility of pharmacological as well as therapeutic interventions through the environment.

3. THE GLUCOCORTICOID RECEPTOR-IMPAIRED MOUSE MODEL OF MOOD DISORDERS

Investigations on depression have mainly focused on animal models of 'mood-related behavioural disorders' generated by exposure to chronic stress or gene manipulation in mice that can drive strong dysfunctions in HPA axis feedback regulation. Nevertheless, basic understanding of both molecular- and system-level perturbations resulting in the HPA axis dysregulations in depression is still lacking. A better comprehension of these mechanisms might possibly lead to the identification of novel therapeutic targets.

To investigate dysfunctions brought about by dysregulations of HPA axis, GR-i mice were developed as a genetic model of depressive disorders [58]. As a result of the expression of GR antisense mRNA under the control of a human neurofilament promoter element, GR-i mice exhibited a decrease in both GR-specific binding (approx. 50% in the hippocampus and the cerebral cortex) and GR mRNA levels (approx. 25% in the hippocampus and in the dorsal raphe nucleus, DRN) compared with paired wild-type (WT) mice [59]. Neuroendocrine regulation in these mice is heavily disturbed, as exemplified by a reduction in both glucocorticoid feedback efficiency and stress-induced increase in plasma ACTH, but not corticosterone [59-62]. Furthermore, it has been reported that GR-i mice have altered behavioural responses in models of anxiety and depression [62-64]. Most of these alterations are reduced by various AD treatments, such as TCAs, SSRIs or new generation ADs, such as agomelatine. Normalization by AD treatments of the hyperactive HPA system in GR-i mice is relevant to what occurs during successful AD therapy

gene	full name	family/action	role	GR-i vs WT
Arc	activity-regulated cytoskeleton-associated protein	member of the immediate-early gene (IEG) family	plasticity	+
NeuroD1	neurogenic differentiation 1	member of the NeuroD family of basic helix-loop-helix (bHLH) transcription factors	plasticity	-
Nocturnin	CCR4 carbon catabolite repression 4-like	circadian deadenylase	post-transcriptional control of circadian gene expression	-
rev-erbB	nuclear receptor subfamily 1, group D, member 2	member of the Rev-ErbB family of nuclear receptors	circadian transcriptional repression	-
Per 1	period circadian protein homologue 1	member of the Period family of genes	circadian transcriptional repression	+
Per 3	period circadian protein homologue 3	member of the Period family of genes	circadian transcriptional repression	+
NPAS2	neuronal PAS domain- containing protein 2 i	member of the bHLH-PAS family of transcription factors	circadian transcriptional regulation	_
H2AZ	histone H2A variant histone family, member Z	variant of H2A localized to promoters	gene expression regulation	_
HDAC2	histone deacetylase 2 HDAC Class I	removes acetyl groups from an N-acetyl lysine on a histone	transcriptional repression	_
HDAC5	histone deacetylase 5 HDAC Class II A	removes acetyl groups from an N-acetyl lysine on a histone	transcriptional repression	_
MYST2	MYST histone acetyltransferase 2	acetylates lysine by transferring an acetyl group from acetyl CoA to form N-acetyl lysine	steroid receptor-mediated transcription regulation	_
NCor1	nuclear receptor co-repressor 1	recruits histone deacetylases to DNA promoter regions	transcriptional repression	_
MeCP2	methyl CpG-binding protein 2	binds methylated DNA	transcriptional regulation	+

Table 1. Changes in gene expression in the hippocampus of GR-i versus control WT paired mice. Microarray experiments were conducted using the Illumina MouseRef-8 Expression BeadChip [68].

of depressive illness [63]. These observations fit well with the hypothesis that GR-i mice model a subgroup of depressed patients who do show a similar hyperactivity of the HPA axis. In addition, impaired GR functioning affects normal adaptive responses to chronic mild stress. Under non-stressful conditions, GR-i mutants present no alterations in 5-HT_{1A} autoreceptor function in the dorsal DRN. However, whereas in WT mice, a chronic mild stress session produces a functional desensitization of DRN 5-HT_{1A} autoreceptors, this is not observed in GR-i mice [59]. In addition, chronic mild stress was found to have a pro-cognitive effect in WT but not GR-i mice in a decision-making task [59]. When subjected to a psychological stressor, GR-i mice displayed hyperresponsiveness of the serotonergic system as shown by a profound rise in hippocampal 5-HT during a rat exposure, but no increase of free corticosterone and behavioural impairments when compared with WT mice [65]. These results demonstrate that impaired GR functioning affects normal adaptive responses to stress at both the HPA and the serotonergic levels and alters stress-related consequences on cognition.

Presumably, owing to a decrease in BDNF mRNA expression in the hippocampus of GR-i compared with WT mice, a reduction in cell proliferation and survival together with reduced neurogenesis have also been observed within the dentate gyrus of GR-i compared with WT mice [64]. These alterations were reversed by chronic AD treatments with either the MT1/MT2 receptor agonist and $5\text{-HT}_{2\text{C}}$ receptor antagonist agomelatine or the SSRI fluoxetine [64]. Interestingly, GR-i mice have been shown to display numerous abnormalities regarding spatial learning and short-term memory [66], anxiety [62] and responses to stress [59,65]. All these alterations could be linked to the deficits observed in hippocampal cell proliferation and survival.

(a) Genetic and epigenetic regulations in glucocorticoid receptor-impaired mice

We have recently investigated the genome-wide expression in GR-i mice [67]. Genes whose expression was modified in these mutants versus paired WT mice were deciphered using oligonucleotide-based microarrays in three brain areas relevant to mood disorders (anterior raphe area, hippocampus and frontal cortex). Important gene expression differences were observed in the hippocampus and in the frontal cortex, which are key sites for the negative feedback regulation of HPA axis activity. Among the genes differentially expressed in GR-i versus WT mice, those encoding Arc, NeuroD1, nocturnin, rev-erbB, Per 1-3 and NPAS2 are of particular interest because of their implications in synaptic plasticity, neurotransmission, neurogenesis and circadian rhythms (table 1). Some of the differences were confirmed using RT-PCR (figure 3 and see [68,69]). Interestingly, numerous mRNA-encoding proteins involved in epigenetic

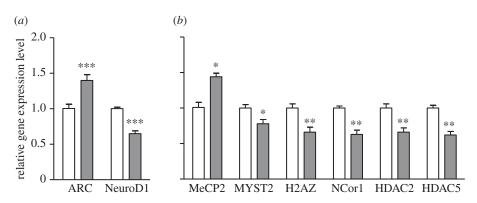


Figure 3. Gene expression levels in GR-i mice compared with paired WT controls. (a) mRNA expression of various genes implicated in synaptic plasticity (a) and related to epigenetic regulations (b). Brains were removed between 9.00 and 11.00. Semiquantitative determinations were made with reference to reporter gene encoding hypoxanthine guanine phosphoribosyl transferase (HPRT). Data are mean \pm s.e.m. (n = 6). *p < 0.05, **p < 0.01, ***p < 0.001 GR-i (grey bars) versus WT (empty bars) mice, paired Student's *t*-test.

regulations such as the variant H2AZ of the histone 2A, the histone deacetylases HDAC2 and HDAC5, the MYST histone acetyltransferase 2 MYST2 and the nuclear receptor co-repressor NCor1 were downregulated in GR-i mutants (table 1 and figure 3). In contrast, methyl CpG-binding protein 2 (MeCP2) was upregulated. Finally, genes encoding various enzymes or complexes involved in cellular metabolism (cytochrome oxidase, NADH dehydrogenase, pyruvate dehydrogenase kinase, etc.) were also affected in the transgenic mice [67–69]. All these gene expression modulations might be involved in the depressive-like behaviours of the GR-i mice.

Emerging data underline that circadian rhythms, regulated at organ and cellular levels via the clock machinery, govern physiological and behavioural functions in most organisms [70]. Notably, the clock transcription-translation feedback loop system possesses intimate links with cellular metabolism. These connections are maintained by epigenetic mechanisms [71]. For instance, HDAC3 activation by Ncor1, the expression of which is decreased in GR-i mice, is critical for the epigenetic regulation of circadian and metabolic physiology [72]. Interestingly, data suggest that clock genes expression is regulated via direct interactions between NCor1 and rev-erbB [73], whose expression is also decreased in GR-i compared with WT mice. Moreover, the circadian CLOCK system and the HPA axis strongly interact. In particular, clock proteins directly regulate GR transcriptional activity [74] and glucocorticoids modulate energy homeostasis by entraining the clock machinery [75] via stimulation of rev-erbB and NPAS2 expressions that are also affected in GR-i mice. The dysregulation of NPAS2 in GR-i mice is of particular interest, since this homologue of the protein CLOCK operating in the mammalian forebrain [76] is involved in the acquisition of specific memory [77] and in the maintenance of circadian behaviours [78].

The observed downregulations of HDAC2 and HDAC5 expression in GR-i mice are consistent with results obtained by other groups. For example, Nestler's group reported that after AD treatment, HDAC5 expression is decreased in the hippocampus of stressed mice and might participate in the efficacy

of the treatment [55]. The same group suggested that HDAC5 may regulate adaptation to chronic stress [79]. Therefore, the decrease in HDAC5 expression in GR-i mice may be a compensatory mechanism induced by the decrease in GR function. The downregulation in HDAC2 expression observed in the hippocampus of GR-i mice [69] may also be an adaptive mechanism, since it was found that, in the hippocampus, HDAC2 negatively regulates synaptic plasticity and morphological changes necessary for learning [80]. Furthermore, a decrease in HDAC2 protein expression in the nucleus accumbens was shown to be associated with both depression in humans and chronic social-stress-induced behavioural deficits in mice [81]. Increased expression of MeCP2 has also been observed in GR-i mice. MeCP2 was identified as a protein that binds specifically to methylated DNA and is primarily considered a transcriptional repressor through associated HDACs. However, MeCP2 appears to be a multi-functional protein. For example, an association with the transcriptional activator CREB1 was reported [82] and it is now believed that MeCP2 can function as both an activator and repressor of transcription. Moreover, MeCP2 has a widespread effect on gene expression [83] and could prevent aberrant transcriptional events [84]. A role of MeCP2 in regulating chromatin architecture has also been proposed [85]. It has recently been reported that a mouse line overexpressing MeCP2 in neurons only exhibits heightened anxiety, impairments in learning and memory, as well as hippocampal synaptic plasticity alterations that may be attributable to transcriptional repression via HDAC recruitment [86]. These results suggest that MeCP2 overexpression may directly cause some of the deficits observed in GR-i mice and strengthen the hypothesis that HDAC downregulation in GR-i mice could be a compensatory mechanism counteracting MeCP2 overexpression. However, detrimental or beneficial impacts of hippocampal MeCP2 overexpression in GR-i mice may be difficult to evaluate since repeated injections of fluoxetine in rats upregulate MeCP2 in the dentate gyrus of hippocampus [87].

The H2AZ variant of the histone 2A is involved in diverse biological functions, such as gene transcription [88,89]. H2AZ may prepare chromatin structure for recruitment of the transcriptional machinery and

regulates promoter capacity to trigger transcription. Interestingly, this variant is enriched at GR promoter sites and this enrichment is modulated by GR receptor activity [90]. The decrease in H2AZ expression in GR-i mice could be a direct consequence of HPA axis activity dysregulation. However, it remains to be determined whether these H2AZ variations contribute to the regulation of the global expression pattern modulated by glucocorticoids, and whether it is of any relevance to depression or represents an adaptation mechanism similar to the changes observed in HDAC5, HDAC2 or MeCP2 expression levels.

Finally, GR-i mice also manifest immuneendocrine abnormalities. Indeed, in addition to HPA axis dysfunctions, GR-i mice show alterations in the trafficking and the responsiveness of T lymphocytes [91]. Hence, further work on these transgenic mice should be particularly informative regarding immune regulationrelated to depressive disorders. Moreover, the study of these mice in the context of gene \times environment interactions appears particularly promising. It is indeed well demonstrated that adverse conditions early in life have a major influence on the normal development of GR-HPA interactions, and that events during development have long-term impact on the capacity of the glucocorticoid system to cope with stress. We should investigate whether early manipulations, such as handling and maternal separation in G-R-i mice, might alter the relationships between immune, neuroendocrine and epigenetic changes in the context of mood-related disorders.

4. CONCLUSION

Adaptation to environmental stresses is one of the most fundamental biological regulatory processes. Chronic stress entails long-term dysregulations at neurotransmitter, neurohormone and cellular levels leading to behavioural changes. Homeostasis is maintained by complex physiological and behavioural systems coordinated by the brain. The effectors are diverse and comprise endocrine and immune regulations, neural plasticity, circadian rhythms and chromatin modulations. Recent studies suggested that all these systems are inter-related and may underlie brain diseases, especially mood-related disorders. Perturbations of neurotransmitter metabolism, circadian clock genes and chromatin modifications interact and certainly determine, at least in part, behavioural disorders. Chromatin regulation, clock genes expression and neurotransmitter metabolism processes thought to be involved in mood disorders and HPA axis regulation are also found to be altered in GR-i mice. It is therefore of importance to determine the exact nature of such perturbations, as well as their relationships, in order to design new strategies for the treatment of depression.

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REFERENCES

 Sobocki, P., Jonsson, B., Angst, J. & Rehnberg, C. 2006 Cost of depression in Europe. *J. Ment. Health Policy Econ.* 9, 87–98.

- 2 Prins, J., Olivier, B. & Korte, S. M. 2011 Triple reuptake inhibitors for treating subtypes of major depressive disorder: the monoamine hypothesis revisited. *Expert. Opin. Investig. Drugs.* 20, 1107–1130. (doi:10.1517/ 13543784.2011.594039)
- 3 Anacker, C., Zunszain, P. A., Carvalho, L. A. & Pariante, C. M. 2011 The glucocorticoid receptor: pivot of depression and of antidepressant treatment? *Psychoneuroendocrinology* 36, 415–425. (doi:10.1016/j.psyneuen. 2010.03.007)
- 4 McClung, C. A. 2007 Circadian genes, rhythms and the biology of mood disorders. *Pharmacol. Ther.* **114**, 222–232. (doi:10.1016/j.pharmthera.2007.02.003)
- 5 Miller, A. H. 2010 Depression and immunity: a role for T cells? *Brain Behav. Immun.* 24, 1–8. (doi:10.1016/j. bbi.2009.09.009)
- 6 Krishnan, V. & Nestler, E. J. 2008 The molecular neurobiology of depression. *Nature* 455, 894–902. (doi:10. 1038/nature07455)
- 7 Slattery, D. A., Hudson, A. L. & Nutt, D. J. 2004 Invited review: the evolution of antidepressant mechanisms. *Fundam. Clin. Pharmacol.* 18, 1–21. (doi:10.1111/j. 1472-8206.2004.00195.x)
- 8 Asberg, M., Traskman, L. & Thoren, P. 1976 5-HIAA in the cerebrospinal fluid: a biochemical suicide predictor? *Arch. Gen. Psychiatry* **33**, 1193–1197. (doi:10.1001/archpsyc.1976.01770100055005)
- 9 Mongeau, R., Blier, P. & de Montigny, C. 1997 The serotonergic and noradrenergic systems of the hippocampus: their interactions and the effects of antidepressant treatments. *Brain Res. Brain Res. Rev.* 23, 145–195. (doi:10.1016/S0165-0173(96)00017-3)
- 10 Delgado, P. L. 2006 Monoamine depletion studies: implications for antidepressant discontinuation syndrome. J. Clin. Psychiatry 67(Suppl. 4), 22-26.
- Montgomery, S. A., Henry, J., McDonald, G., Dinan, T., Lader, M., Hindmarch, I., Clare, A. & Nutt, D. 1994 Selective serotonin reuptake inhibitors: meta-analysis of discontinuation rates. *Int. Clin. Psychopharmacol.* 9, 47–53. (doi:10.1097/00004850-199400910-00008)
- 12 Nemeroff, C. B. 1998 Psychopharmacology of affective disorders in the 21st century. *Biol. Psychiatry* 44, 517–525. (doi:10.1016/S0006-3223(98)00068-7)
- 13 Kendler, K. S., Kessler, R. C., Walters, E. E., MacLean, C., Neale, M. C., Heath, A. C. & Eaves, L. J. 1995 Stressful life events, genetic liability, and onset of an episode of major depression in women. *Am. J. Psychiatry* 152, 833–842.
- 14 Caspi, A. & Moffitt, T. E. 2006 Gene-environment interactions in psychiatry: joining forces with neuroscience. *Nat. Rev. Neurosci.* 7, 583–590. (doi:10.1038/ nrn1925)
- 15 Oitzl, M. S. & de Kloet, E. R. 1992 Selective corticosteroid antagonists modulate specific aspects of spatial orientation learning. *Behav. Neurosci.* 106, 62–71. (doi:10.1037/0735-7044.106.1.62)
- 16 Pariante, C. M. 2003 Depression, stress and the adrenal axis. *J. Neuroendocrinol.* **15**, 811–812. (doi:10.1046/j. 1365-2826.2003.01058.x)
- 17 Dam, H. 1988 Dexamethasone suppression test. Acta Psychiatr. Scand. Suppl. 345, 38–44.
- 18 Fountoulakis, K. N., Gonda, X., Rihmer, Z., Fokas, C. & Iacovides, A. 2008 Revisiting the dexamethasone suppression test in unipolar major depression: an exploratory study. Ann. Gen. Psychiatry 7, 22. (doi:10.1186/1744-859X-7-22)
- 19 Maes, M., Bosmans, E., Suy, E., Vandervorst, C., DeJonckheere, C. & Raus, J. 1991 Depression-related disturbances in mitogen-induced lymphocyte responses and interleukin-1 beta and soluble interleukin-2 receptor

production. *Acta Psychiatr. Scand.* **84**, 379–386. (doi:10. 1111/j.1600-0447.1991.tb03163.x)

- 20 Maes, M. 1994 Cytokines in major depression. *Biol. Psychiatry* 36, 498–499. (doi:10.1016/0006-3223(94) 90652-1)
- 21 Maes, M., Leonard, B. E., Myint, A. M., Kubera, M. & Verkerk, R. 2011 The new '5-HT' hypothesis of depression: cell-mediated immune activation induces indoleamine 2,3-dioxygenase, which leads to lower plasma tryptophan and an increased synthesis of detrimental tryptophan catabolites (TRYCATs), both of which contribute to the onset of depression. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **35**, 702–721. (doi:10.1016/j.pnpbp.2010.12.017)
- 22 Pollak, Y. & Yirmiya, R. 2002 Cytokine-induced changes in mood and behaviour: implications for 'depression due to a general medical condition', immunotherapy and antidepressive treatment. *Int. J. Neuropsychopharmacol.* 5, 389–399. (doi:10.1017/S1461145702003152)
- 23 Raison, C. L., Lin, J. M. & Reeves, W. C. 2009 Association of peripheral inflammatory markers with chronic fatigue in a population-based sample. *Brain Behav. Immun.* 23, 327–337. (doi:10.1016/j.bbi.2008.11.005)
- 24 Raison, C. L., Capuron, L. & Miller, A. H. 2006 Cytokines sing the blues: inflammation and the pathogenesis of depression. *Trends Immunol.* 27, 24–31. (doi:10. 1016/j.it.2005.11.006)
- 25 Leonard, B. E. 2010 The concept of depression as a dysfunction of the immune system. *Curr. Immunol. Rev.* 6, 205–212. (doi:10.2174/157339510791823835)
- 26 Stockmeier, C. A., Mahajan, G. J., Konick, L. C., Overholser, J. C., Jurjus, G. J., Meltzer, H. Y., Uylings, H. B., Friedman, L. & Rajkowska, G. 2004 Cellular changes in the postmortem hippocampus in major depression. *Biol. Psychiatry* 56, 640–650. (doi:10.1016/ j.biopsych.2004.08.022)
- 27 Tamura, M., Sajo, M., Kakita, A., Matsuki, N. & Koyama, R. 2011 Prenatal stress inhibits neuronal maturation through downregulation of mineralocorticoid receptors. *J. Neurosci.* 31, 11505–11514. (doi:10.1523/JNEUROSCI.3447-10.2011)
- 28 Warner-Schmidt, J. L. & Duman, R. S. 2006 Hippocampal neurogenesis: opposing effects of stress and antidepressant treatment. *Hippocampus* 16, 239–249. (doi:10.1002/hipo.20156)
- 29 Reif, A., Fritzen, S., Finger, M., Strobel, A., Lauer, M., Schmitt, A. & Lesch, K. P. 2006 Neural stem cell proliferation is decreased in schizophrenia, but not in depression. *Mol. Psychiatry* **11**, 514–522. (doi:10.1038/ sj.mp.4001791)
- 30 Surget, A. *et al.* 2011 Antidepressants recruit new neurons to improve stress response regulation. *Mol. Psychiatry* 16, 1177–1188. (doi:10.1038/mp.2011.48)
- 31 Aasebo, I. E., Blankvoort, S. & Tashiro, A. 2011 Critical maturational period of new neurons in adult dentate gyrus for their involvement in memory formation. *Eur. J. Neurosci.* 33, 1094–1100. (doi:10.1111/j.1460-9568.2011.07608.x)
- 32 Lucassen, P. J., Meerlo, P., Naylor, A. S., van Dam, A. M., Dayer, A. G., Fuchs, E., Oomen, C. A. & Czeh, B. 2010 Regulation of adult neurogenesis by stress, sleep disruption, exercise and inflammation: implications for depression and antidepressant action. *Eur. Neuropsychopharmacol.* 20, 1–17. (doi:10.1016/j. euroneuro.2009.08.003)
- 33 Duman, R. S. & Monteggia, L. M. 2006 A neurotrophic model for stress-related mood disorders. *Biol. Psychiatry* 59, 1116–1127. (doi:10.1016/j.biopsych.2006.02.013)
- 34 Smith, M. A., Makino, S., Kvetnansky, R. & Post, R. M. 1995 Effects of stress on neurotrophic factor expression

in the rat brain. Ann. N. Y. Acad. Sci. 771, 234–239. (doi:10.1111/j.1749-6632.1995.tb44684.x)

- 35 Gronli, J., Bramham, C., Murison, R., Kanhema, T., Fiske, E., Bjorvatn, B., Ursin, R. & Portas, C. M. 2006 Chronic mild stress inhibits BDNF protein expression and CREB activation in the dentate gyrus but not in the hippocampus proper. *Pharmacol. Biochem. Behav.* 85, 842–849. (doi:10.1016/j.pbb.2006.11.021)
- 36 Martinowich, K. & Lu, B. 2008 Interaction between BDNF and serotonin: role in mood disorders. *Neuropsychopharmacology* 33, 73–83. (doi:10.1038/sj.npp.1301571)
- 37 Lee, B. H. & Kim, Y. K. 2010 The roles of BDNF in the pathophysiology of major depression and in antidepressant treatment. *Psychiatry Invest.* 7, 231–235. (doi:10. 4306/pi.2010.7.4.231)
- 38 Pariante, C. M. & Lightman, S. L. 2008 The HPA axis in major depression: classical theories and new developments. *Trends Neurosci.* 31, 464–468. (doi:10.1016/j. tins.2008.06.006)
- 39 Meaney, M. J., Szyf, M. & Seckl, J. R. 2007 Epigenetic mechanisms of perinatal programming of hypothalamicpituitary-adrenal function and health. *Trends Mol. Med.* 13, 269–277. (doi:10.1016/j.molmed.2007.05.003)
- 40 Schulz, P. 2007 Biological clocks and the practice of psychiatry. *Dialogues Clin. Neurosci.* 9, 237–255.
- 41 Marques, A. H., Silverman, M. N. & Sternberg, E. M. 2009 Glucocorticoid dysregulations and their clinical correlates. From receptors to therapeutics. *Ann. N. Y. Acad. Sci.* 1179, 1–18. (doi:10.1111/j.1749-6632.2009.04987.x)
- 42 Benedetti, F., Serretti, A., Colombo, C., Barbini, B., Lorenzi, C., Campori, E. & Smeraldi, E. 2003 Influence of CLOCK gene polymorphism on circadian mood fluctuation and illness recurrence in bipolar depression. *Am. J. Med. Genet. B. Neuropsychiatr. Genet.* 123B, 23–26. (doi:10.1002/ajmg.b.20038)
- 43 Gorwood, P. 2010 Restoring circadian rhythms: a new way to successfully manage depression. *J. Psychopharmacol.* 24, 15–19. (doi:10.1177/1359786810372981)
- 44 Popoli, M. 2009 Agomelatine: innovative pharmacological approach in depression. CNS Drugs 23(Suppl 2), 27–34. (doi:10.2165/11318640-00000000-00000)
- 45 Uz, T., Ahmed, R., Akhisaroglu, M., Kurtuncu, M., Imbesi, M., Dirim Arslan, A. & Manev, H. 2005 Effect of fluoxetine and cocaine on the expression of clock genes in the mouse hippocampus and striatum. *Neuro-science* **134**, 1309–1316. (doi:10.1016/j.neuroscience. 2005.05.003)
- 46 Dallaspezia, S. & Benedetti, F. 2011 Chronobiological therapy for mood disorders. *Expert. Rev. Neurother.* 11, 961–970. (doi:10.1586/ern.11.61)
- 47 Benedetti, F., Barbini, B., Lucca, A., Campori, E., Colombo, C. & Smeraldi, E. 1997 Sleep deprivation hastens the antidepressant action of fluoxetine. *Eur. Arch. Psychiatry Clin. Neurosci.* 247, 100–103. (doi:10.1007/ BF02900200)
- 48 Mill, J. & Petronis, A. 2007 Molecular studies of major depressive disorder: the epigenetic perspective. *Mol. Psychiatry* **12**, 799–814. (doi:10.1038/sj.mp.4001992)
- 49 Schroeder, M., Krebs, M. O., Bleich, S. & Frieling, H. 2010 Epigenetics and depression: current challenges and new therapeutic options. *Curr. Opin. Psychiatry* 23, 588–592. (doi:10.1097/YCO.0b013e32833d16c1)
- 50 Tsankova, N., Renthal, W., Kumar, A. & Nestler, E. J. 2007 Epigenetic regulation in psychiatric disorders. *Nat. Rev. Neurosci.* 8, 355–367. (doi:10.1038/nrn2132)
- 51 Szyf, M. 2009 The early life environment and the epigenome. *Biochim. Biophys. Acta* 1790, 878–885. (doi:10.1016/j.bbagen.2009.01.009)
- 52 Weaver, I. C., Cervoni, N., Champagne, F. A., D'Alessio, A. C., Sharma, S., Seckl, J. R., Dymov, S.,

Szyf, M. & Meaney, M. J. 2004 Epigenetic programming by maternal behavior. *Nat. Neurosci.* 7, 847–854. (doi:10.1038/nn1276)

- 53 Zhang, T. Y. & Meaney, M. J. 2010 Epigenetics and the environmental regulation of the genome and its function. *Annu. Rev. Psychol.* **61**, 439–466. (doi:10.1146/annurev. psych.60.110707.163625)
- 54 Murgatroyd, C. et al. 2009 Dynamic DNA methylation programs persistent adverse effects of early-life stress. Nat. Neurosci. 12, 1559–1566. (doi:10.1038/nn.2436)
- 55 Tsankova, N. M., Berton, O., Renthal, W., Kumar, A., Neve, R. L. & Nestler, E. J. 2006 Sustained hippocampal chromatin regulation in a mouse model of depression and antidepressant action. *Nat. Neurosci.* 9, 519–525. (doi:10.1038/nn1659)
- 56 Weaver, I. C., Champagne, F. A., Brown, S. E., Dymov, S., Sharma, S., Meaney, M. J. & Szyf, M. 2005 Reversal of maternal programming of stress responses in adult offspring through methyl supplementation: altering epigenetic marking later in life. *J. Neurosci.* 25, 11 045– 11 054. (doi:10.1523/JNEUROSCI.3652-05.2005)
- 57 Boks, M. P., de Jong, N. M., Kas, M. J., Vinkers, C. H., Fernandes, C., Kahn, R. S., Mill, J. & Ophoff, R. A. 2012 Current status and future prospects for epigenetic psychopharmacology. *Epigenetics* 7, 20–28.
- 58 Pepin, M. C., Pothier, F. & Barden, N. 1992 Impaired type II glucocorticoid-receptor function in mice bearing antisense RNA transgene. *Nature* 355, 725–728. (doi:10.1038/355725a0)
- 59 Froger, N. et al. 2004 Neurochemical and behavioral alterations in glucocorticoid receptor-impaired transgenic mice after chronic mild stress. *J. Neurosci.* 24, 2787–2796. (doi:10.1523/JNEUROSCI.4132-03.2004)
- 60 Barden, N., Stec, I. S., Montkowski, A., Holsboer, F. & Reul, J. M. 1997 Endocrine profile and neuroendocrine challenge tests in transgenic mice expressing antisense RNA against the glucocorticoid receptor. *Neuroendocrinology* 66, 212–220. (doi:10.1159/000127240)
- 61 Karanth, S., Linthorst, A. C., Stalla, G. K., Barden, N., Holsboer, F. & Reul, J. M. 1997 Hypothalamic– pituitary–adrenocortical axis changes in a transgenic mouse with impaired glucocorticoid receptor function. *Endocrinology* **138**, 3476–3485. (doi:10.1210/en. 138.8.3476)
- Montkowski, A. et al. 1995 Long-term antidepressant treatment reduces behavioural deficits in transgenic mice with impaired glucocorticoid receptor function. J. Neuroendocrinol. 7, 841–845. (doi:10.1111/j.1365-2826.1995.tb00724.x)
- 63 Barden, N., Shink, E., Labbe, M., Vacher, R., Rochford, J. & Mocaer, E. 2005 Antidepressant action of agomelatine (S 20098) in a transgenic mouse model. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 29, 908–916. (doi:10.1016/j.pnpbp.2005.04.032)
- 64 Paizanis, E. *et al.* 2010 Behavioural and neuroplastic effects of the new-generation antidepressant agomelatine compared to fluoxetine in glucocorticoid receptor-impaired mice. *Int. J. Neuropsychopharmacol.* 13, 759–774. (doi:10.1017/S1461145709990514)
- 65 Linthorst, A. C., Flachskamm, C., Barden, N., Holsboer, F. & Reul, J. M. 2000 Glucocorticoid receptor impairment alters CNS responses to a psychological stressor: an in vivo microdialysis study in transgenic mice. *Eur. J. Neurosci.* 12, 283–291. (doi:10.1046/j. 1460-9568.2000.00878.x)
- 66 Rousse, I., Beaulieu, S., Rowe, W., Meaney, M. J., Barden, N. & Rochford, J. 1997 Spatial memory in transgenic mice with impaired glucocorticoid receptor function. *Neuroreport* 8, 841–845. (doi:10.1097/ 00001756-199703030-00007)

- 67 Hoyle, D. et al. 2011 Shared changes in gene expression in frontal cortex of four genetically modified mouse models of depression. Eur. Neuropsychopharmacol. 21, 3–10. (doi:10.1016/j.euroneuro.2010.09.011)
- 68 Massart, R., Fabre, V., Païzanis, E., Hamon, M. & Lanfumey, L. 2010 Epigenetic and clock genes' expression perturbations in genetic and environmental models of depressive-like disorders. In 7th World Congress on Stress, 26 August 2010, Leiden, The Netherlands.
- 69 Massart, R., Stragier, E., Boulle, F., Païzanis, E., Gabriel, C., Mocaer, E., Hamon, M. & Lanfumey, L. 2011 Epigenetic modifications in genetic and environmental models of depressive-like disorder: reversal by chronic treatment with Agomelatine. In *Program no. 394.18/WW16 Neuroscience Meeting Planner, 14 November 2011, Washington, DC.* Washington, DC: Society for Neuroscience.
- 70 Turek, F. W. 2008 Circadian clocks: tips from the tip of the iceberg. *Nature* **456**, 881–883. (doi:10.1038/ 456881a)
- 71 Grimaldi, B., Nakahata, Y., Kaluzova, M., Masubuchi, S. & Sassone-Corsi, P. 2009 Chromatin remodeling, metabolism and circadian clocks: the interplay of CLOCK and SIRT1. *Int. J. Biochem. Cell Biol.* 41, 81-86. (doi:10.1016/j.biocel.2008.08.035)
- 72 Alenghat, T. et al. 2008 Nuclear receptor corepressor and histone deacetylase 3 govern circadian metabolic physiology. *Nature* 456, 997–1000. (doi:10.1038/ nature07541)
- 73 Raghuram, S. *et al.* 2007 Identification of heme as the ligand for the orphan nuclear receptors REV-ERBα and REV-ERBβ. *Nat. Struct. Mol. Biol.* 14, 1207– 1213. (doi:10.1038/nsmb1344)
- 74 Nader, N., Chrousos, G. P. & Kino, T. 2010 Interactions of the circadian CLOCK system and the HPA axis. *Trends Endocrinol. Metab.* 21, 277–286. (doi:10.1016/j. tem.2009.12.011)
- 75 Balsalobre, A., Brown, S. A., Marcacci, L., Tronche, F., Kellendonk, C., Reichardt, H. M., Schutz, G. & Schibler, U. 2000 Resetting of circadian time in peripheral tissues by glucocorticoid signaling. *Science* 289, 2344–2347. (doi:10.1126/science.289.5488.2344)
- 76 Reick, M., Garcia, J. A., Dudley, C. & McKnight, S. L. 2001 NPAS2: an analog of clock operative in the mammalian forebrain. *Science* 293, 506–509. (doi:10.1126/ science.1060699)
- 77 Garcia, J. A., Zhang, D., Estill, S. J., Michnoff, C., Rutter, J., Reick, M., Scott, K., Diaz-Arrastia, R. & McKnight, S. L. 2000 Impaired cued and contextual memory in NPAS2-deficient mice. *Science* 288, 2226– 2230. (doi:10.1126/science.288.5474.2226)
- 78 Dudley, C. A., Erbel-Sieler, C., Estill, S. J., Reick, M., Franken, P., Pitts, S. & McKnight, S. L. 2003 Altered patterns of sleep and behavioral adaptability in NPAS2deficient mice. *Science* **301**, 379–383. (doi:10.1126/ science.1082795)
- 79 Renthal, W. et al. 2007 Histone deacetylase 5 epigenetically controls behavioral adaptations to chronic emotional stimuli. *Neuron* 56, 517–529. (doi:10.1016/j. neuron.2007.09.032)
- 80 Guan, J. S. et al. 2009 HDAC2 negatively regulates memory formation and synaptic plasticity. *Nature* **459**, 55–60. (doi:10.1038/nature07925)
- 81 Covington III, H. E. *et al.* 2009 Antidepressant actions of histone deacetylase inhibitors. *J. Neurosci.* 29, 11 451– 11 460. (doi:10.1523/JNEUROSCI.1758-09.2009)
- 82 Chahrour, M., Jung, S. Y., Shaw, C., Zhou, X., Wong, S. T., Qin, J. & Zoghbi, H. Y. 2008 MeCP2, a key contributor to neurological disease, activates and represses transcription. *Science* **320**, 1224–1229. (doi:10.1126/ science.1153252)

- 83 Ben-Shachar, S., Chahrour, M., Thaller, C., Shaw, C. A. & Zoghbi, H. Y. 2009 Mouse models of MeCP2 disorders share gene expression changes in the cerebellum and hypothalamus. *Hum. Mol. Genet.* 18, 2431–2442. (doi:10. 1093/hmg/ddp181)
- 84 Guy, J., Cheval, H., Selfridge, J. & Bird, A. 2011 The role of MeCP2 in the brain. Annu. Rev. Cell Dev. Biol. 27, 631–652. (doi:10.1146/annurev-cellbio-092910-154121)
- 85 Chadwick, L. H. & Wade, P. A. 2007 MeCP2 in Rett syndrome: transcriptional repressor or chromatin architectural protein? *Curr. Opin. Genet. Dev.* 17, 121–125. (doi:10.1016/j.gde.2007.02.003)
- 86 Na, E. S., Nelson, E. D., Adachi, M., Autry, A. E., Mahgoub, M. A., Kavalali, E. T. & Monteggia, L. M. 2012 A mouse model for MeCP2 duplication syndrome: MeCP2 overexpression impairs learning and memory and synaptic transmission. *J. Neurosci.* 32, 3109-3117. (doi:10.1523/JNEUROSCI.6000-11.2012)

- 87 Cassel, S., Carouge, D., Gensburger, C., Anglard, P., Burgun, C., Dietrich, J. B., Aunis, D. & Zwiller, J. 2006 Fluoxetine and cocaine induce the epigenetic factors MeCP2 and MBD1 in adult rat brain. *Mol. Pharmacol.* **70**, 487–492. (doi:10.1124/mol.106.022301)
- 88 Draker, R. & Cheung, P. 2009 Transcriptional and epigenetic functions of histone variant H2A.Z. Biochem. Cell Biol. 87, 19–25. (doi:10.1139/O08-117)
- 89 Zlatanova, J. & Thakar, A. 2008 H2A.Z: view from the top. Structure 16, 166–179. (doi:10.1016/j.str.2007.12.008)
- 90 John, S. et al. 2008 Interaction of the glucocorticoid receptor with the chromatin landscape. Mol. Cell. 29, 611-624. (doi:10.1016/j.molcel.2008.02.010)
- 91 Marchetti, B., Morale, M. C., Testa, N., Tirolo, C., Caniglia, S., Amor, S., Dijkstra, C. D. & Barden, N. 2001 Stress, the immune system and vulnerability to degenerative disorders of the central nervous system in transgenic mice expressing glucocorticoid receptor antisense RNA. *Brain Res. Rev.* **37**, 259–272. (doi:10. 1016/S0165-0173(01)00130-8)