
The complete nucleotide sequence of the *ilvGMEDA* operon of *Escherichia coli* K-12

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ABSTRACT

In this report we present the complete nucleotide sequence of the *ilvGMEDA* operon of *Escherichia coli*. This operon contains five genes encoding four of the five enzymes required for the biosynthesis of isoleucine and valine. We identify and describe the coding regions for these five structural genes and the structural and functional features of the flanking and internal regulatory regions of this operon. This new information contributes to a more complete understanding of the overall control of the biosynthesis of isoleucine and valine.

I. INTRODUCTION

The biosynthesis of isoleucine and valine in *Escherichia coli* and other bacteria, fungi and plants, that have been studied, occurs by the parallel pathway shown in figure 1. The pathway is parallel because the last four steps of isoleucine synthesis are catalyzed by bifunctional enzymes which also function to convert two pyruvate molecules into valine. Additionally, the α -keto acid precursor of valine, α -ketoisovalerate, is a branchpoint intermediate which can be converted to leucine by the four enzymes encoded by the genes of the leucine operon. The first step of the isoleucine biosynthetic pathway, the conversion of threonine to α -ketobutyrate, is not required for the biosynthesis of valine or leucine (for reviews see refs. 1 and 2).

Carbon flow through the isoleucine pathway is metabolically regulated by isoleucine end-product inhibition of the first enzyme of this pathway, threonine deaminase (1). Similarly, excessive carbon flow through the valine pathway is controlled by valine end-product inhibition of two of three acetoxy acid synthase (AHAS) isozymes, AHAS I and AHAS III, that catalyze the first step of the parallel pathway (Fig. 1) (3). The third isozyme, AHAS II is resistant to end-product inhibition by valine. However, unlike other enteric bacteria, and indeed other strains of *E. coli*, AHAS II is not expressed in the *E. coli* K-12 strain because of a frameshift mutation in the *ilvG* gene which encodes the large subunit of this enzyme (4-6). This explains why *E. coli* K-12 is growth inhibited by exogenous valine; that is, the remaining two active AHAS isozymes are both inhibited by valine which blocks the synthesis of isoleucine.

The multiplicity of the AHAS isozymes appears to provide differential regulation of the flow of carbon through the first enzymatic step of the parallel pathway (Fig. 1). Each of the AHAS isozymes are distinguished by the following catalytic and regulatory properties. First, as previously mentioned, AHAS I and III are sensitive to valine feedback inhibition, whereas AHAS II is not regulated by valine. Second, it is thought

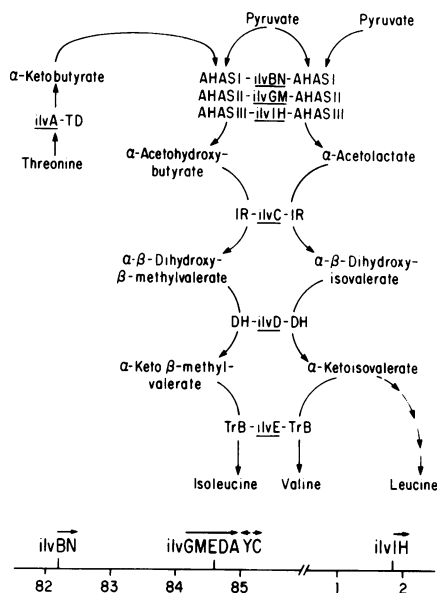


Figure 1. The pathways for the synthesis of isoleucine, leucine and valine in *E. coli*. The enzymes involved in the isoleucine and valine biosynthesis are abbreviated as follows: TD, threonine deaminase; AHAS, acetoxy acid synthase; IR, acetoxy acid isomeroreductase; DH, dihydroxy acid dehydrase; TrB, transaminase B. The genes encoding each of these enzymes are indicated next to the enzyme abbreviation and the chromosomal locations are shown below (7). The *ilvY* gene encodes a positive activator of *ilvC* transcription (8,9). Arrows above genes indicate the direction of transcription of these loci.

that the AHAS isozymes differ in their substrate specificities for α -ketobutyrate and pyruvate (3). Finally, the expression of genes encoding the AHAS isozymes are regulated in different manners. The *ilvBN* operon (AHAS I) is regulated by both catabolite repression (10) and an attenuator mechanism regulated by the intracellular levels of aminoacylated tRNA^{Val} and tRNA^{Leu} (11,12). The *ilvIH* (AHAS III) operon is repressed by leucine by a mechanism not yet fully understood (13) and the *ilvGMEDA* operon, which encodes the subunits of AHAS II (*ilvG* and *ilvM*), is regulated by all three branched chain amino acids via attenuation (14-16). It is not yet fully understood how these differences between the AHAS isozymes function in the regulation of the synthesis of the branched chain amino acids during different physiological conditions.

The *ilvGMEDA* operon of *E. coli* might be thought of as the "backbone" of the *ilv* regulon because this operon encodes four of the five enzymes required for the biosynthesis of isoleucine and valine (Fig. 1). A schematic diagram detailing the structure of this operon is presented in figure 2. The locations of the regulatory regions and structural genes are numbered relative to the *in vivo* transcriptional start site. In the following sections we present the complete nucleotide sequence of the *ilvGMEDA* operon. These sequences are discussed in the context of the regions important for the

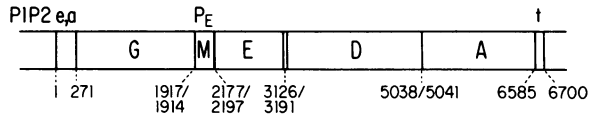


Figure 2. A schematic representation of the *ilvGMEDA* operon of *E. coli*. The tandem *in vitro* promoters P1P2, leader polypeptide (e), attenuator (a), internal promoter, P_E and transcriptional terminator (t) are indicated above the structural genes. Nucleotides are numbered from the *in vivo* transcriptional initiation site (16). The number preceding each gene corresponds to the first nucleotide of the initiation codon and the number following each gene corresponds to the last nucleotide of the termination codon.

regulation of operon expression (Section II) and the five structural genes encoded in the operon (Section III). In the last section (Section IV) we discuss future questions regarding the mechanisms of regulation of operon expression.

II. REGULATORY REGIONS OF THE *ilvGMEDA* OPERON

A. Transcription promoters

The nucleotide sequence of the proximal promoter region of the operon is presented in figure 3. This promoter region consists of tandem *in vitro* promoters P1 and P2, which are separated by 72 base pairs (14,16,18). While *in vitro* RNA polymerase initiates transcription at the P1 promoter at about 50% of the level of the downstream P2 promoter, RNA fingerprint analyses of *in vivo* transcripts from the *ilvGMEDA* attenuator region demonstrates that less than 1% of *in vivo* transcription initiates at the P1 promoter during several growth conditions known to affect operon expression (16). Therefore, it does not appear that differential expression of the tandem promoters can be a mechanism for operon regulation. However, using multicopy *galK* transcriptional fusions, it has been demonstrated that sequences upstream of the P2 promoter are required for maximal *in vivo* expression (18).

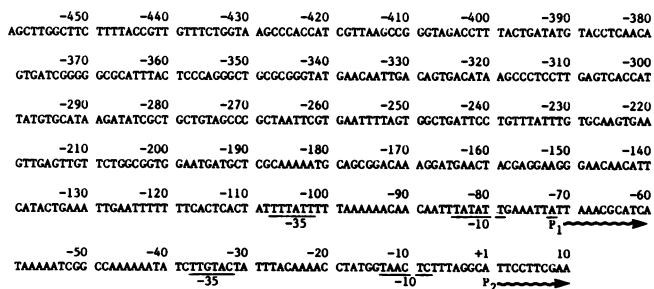


Figure 3. Nucleotide sequence of the promoter regions of the *ilvGMEDA* operon. This sequence begins at an *AluI* restriction site at nucleotide -460. The nucleotides are numbered corresponding to the *in vivo* transcriptional start site (P2 promoter). The transcriptional initiation site of the upstream *in vitro* promoter, P1, is indicated at nucleotide -72. The conserved -10 and -35 hexamer regions (17) of the P1 and P2 promoters are underlined.

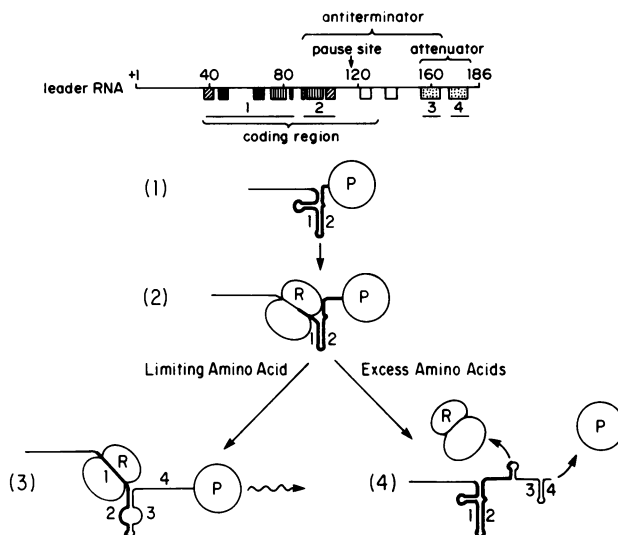


Figure 5. Attenuator control of the *ilvGMEDA* operon. Areas of pattern identity along the leader RNA represent regions of complementary sequences. The bold portion of the leader transcript depicts the leader polypeptide coding region. P represents an RNA polymerase and R represents a ribosome. The following events of attenuator regulation are illustrated. 1) RNA polymerase initiates transcription of leader RNA, stem-loop 1:2 forms and RNA polymerase pauses. 2) A translating ribosome enters stem 1, disrupts stem-loop 1:2 and releases the paused RNA polymerase. 3) During growth conditions limiting for leucine, valine or isoleucine, the translating ribosome stalls on a regulatory codon in stem 1:2, thereby facilitating the formation of the antiterminator, stem-loop 2:3. As a result, the terminator stem-loop 3:4 cannot form, the termination signal is ignored and RNA polymerase continues transcription into the structural genes of the operon (deattenuation). 4) During growth conditions not limiting for leucine, valine or isoleucine, the translating ribosome and pause released RNA polymerase remain coupled. The ribosome clears region 2 and releases at the end of the leader polypeptide coding region before stem 4 is synthesized. Stem-loop 1:2 reforms, stem 4 is synthesized and the terminator, stem-loop 3:4 forms resulting in transcription termination at the attenuator (attenuation).

polypeptide containing five isoleucine, five leucine and six valine residues (Fig. 6). Based on secondary structure predictions of the nucleotide sequence of the leader RNA, it is proposed that the leader RNA can form alternative structures which control the level of transcription into the *ilvGMEDA* structural genes (14,15) (Fig. 6B). In order to accommodate multivalent attenuation by three amino acids, tandem codons for isoleucine, valine and leucine must be located in the stem 1 region of the leader transcript. It is possible that the requirement for this much coding sequence in the stem 1 region necessitates the rather complicated bifurcated 1:2 stem-loop structure shown in figure 6B.

The regulatory codons for leucine are at positions four and five in the polypeptide coding region of the leader RNA (Fig. 6). Assuming that a ribosome covers

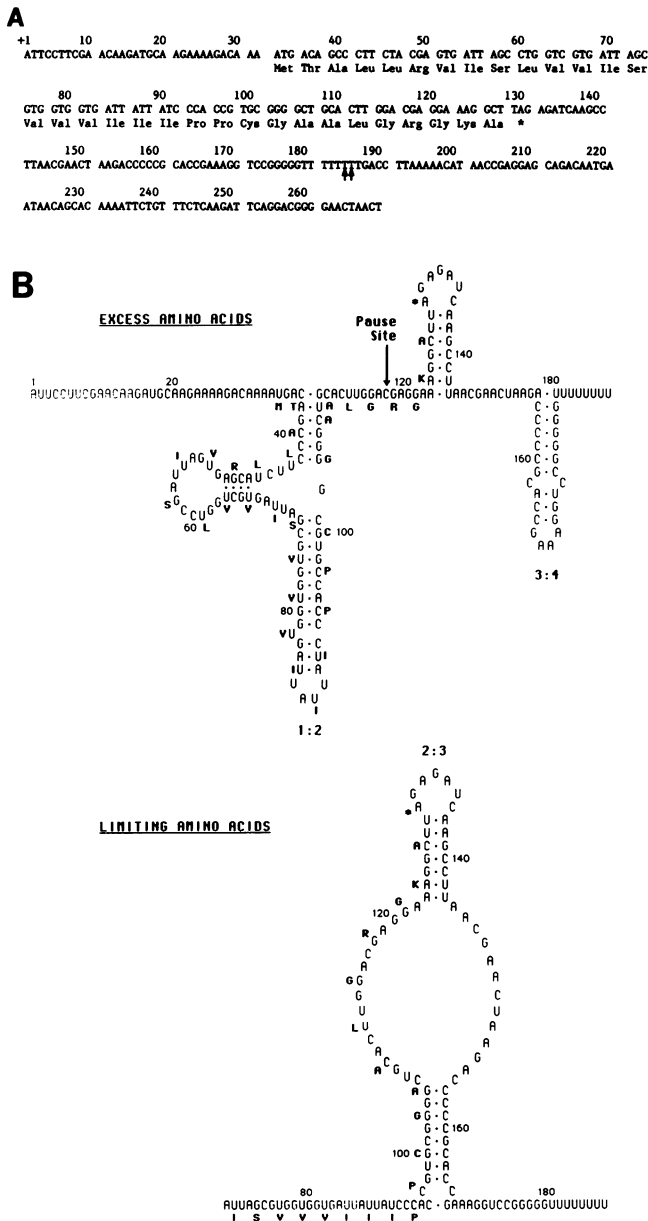


Figure 6. The leader region of the *ilvGMEDA* operon. (A) Nucleotide sequence of the nontranscribed DNA strand of the leader region and the deduced sequence of the leader polypeptide. The arrows indicate the nucleotides at which the leader transcript terminates. (B) The alternative RNA secondary structures that are believed to form during excess and limiting branched chain amino acids. The sequence of the encoded leader polypeptide is shown using the one letter amino acid symbols.

approximately 9 nucleotides on either side of a translated codon (28,29), a ribosome stalled on one of these leucine codons would cover a region of the leader transcript from approximately nucleotides 33 to 56. The position of this stalled ribosome precludes this covered nucleotide sequence from base pairing with its complementary sequence in stem 2, thus facilitating pairing of the stem 2 sequence with complementary sequences in stem 3 to form the antiterminator (Fig. 6B).

The role of the valine, isoleucine and leucine codons in the bifurcated stem-loop (nucleotides 43-72, Fig. 6) is not clear. Regulatory codons in attenuators are usually observed in tandem. The only exception is the single leucine regulatory codon in the leader transcript of the *ilvGMEDA* operon of *Serratia marcescens* (30). This suggests that the single valine, isoleucine and leucine codons specifying amino acids at positions 7,8 and 10 in the leader polypeptide, respectively, might be unimportant for regulation of attenuation. Consistent with this point, these codon positions within the bifurcated stem-loop of stem 1 are located 9 nucleotides or more away from RNA sequences in stem 1 which are involved in base pairing with stem 2. The tandem valine codons at positions 11 and 12, however, are close enough to disrupt the 1:2 stem-loop base pairing if a ribosome stalled on them. A ribosome stalled at one of the three tandem valine codons specifying amino acids at positions 15-17 of the leader polypeptide should also facilitate deattenuation. However, if it is accepted that a ribosome covers only 21 nucleotides of the RNA transcript, the second and third tandem isoleucine codons (specifying amino acids at positions 19-20 of the leader polypeptide) at the bottom of the 1:2 stem-loop are not situated to effect deattenuation (Fig. 6). In order to account for the fact that the *ilvGMEDA* operon is deattenuated by isoleucine limitation, it has been postulated that a second ribosome could queue behind a ribosome stalled at either of these isoleucine codons at the bottom of stem-loop 1:2 (14). This second ribosome would preclude 1:2 stem-loop formation and thus facilitate deattenuation. However, the recent documentation of a transcriptional pause site at nucleotide 117 near the base of stem 1:2 casts doubt on the validity of this model (31). If the first ribosome entering stem 1 releases an RNA polymerase molecule paused at nucleotide 117 then it is not likely that there would be sufficient time to load and position a second ribosome before the released RNA polymerase completed the remainder of the leader transcript resulting in termination of transcription (27). A more plausible explanation is that a ribosome stalled at an isoleucine codon at the bottom of the 1:2 stem-loop has a longer range effect on destabilization of RNA base pairing than the 21 or so nucleotides that are directly masked by the ribosome. The denaturing effect of the physical interaction of the ribosome with the 21 or so bases around the Ile codon would disrupt the bottom half of stem-loop 1:2 and spatially separate the remaining complementary base pairing regions of the stems 1 and 2. Therefore, a ribosome at this position would favor the base pairing of the unmasked downstream sequences leading to the formation of the antiterminator stem-loop 2:3.

C. Termination of transcription at the end of the *ilvGMEDA* operon

The nucleotide sequence following the *ilvA* gene is presented in figure 7. This sequence contains a possible rho-independent terminator (32) centered at nucleotide 6686. There is a G+C rich inverted repeat of seven nucleotides followed by 5 T residues in the *ilvA* nontranscribed DNA strand. In vitro transcription experiments using a

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        6590      6600      6610      6620      6630      6640      6650      6660
TAGGGAAAAA TGCCTGATAG CGCTTCGCTT ATCAGGCCTA CCCCGCGCAG AACGTCATTT GTGGTTCCGC AAMAATCTCC
*
        6670      6680      6690      6700      6710      6720      6730      6740
AAGATCGCTC AATTAGCGGC TCATGTAGCC GCITTTTCTG CGCACACAGG CCCAGCTCAA ACCGGCTTTT CTCATCGCTC
        44
    
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Figure 7. Transcriptional termination at the end of the *ilvGMEDA* operon. Nucleotide sequence of the nontranscribed DNA strand of the region distal to the *ilvA* gene. The sequence begins with the *ilvA* translational stop codon. The *ilvY* gene is transcribed in a direction opposite to the *ilvGMEDA* operon. The stop codon of the *ilvY* product is centered at nucleotide 6638 (designated by asterisk) on the DNA strand complementary to the sequence presented. Centered at nucleotide 6686 is an inverted repeat followed by 5 T residues. The arrows indicate the site of the rho-independent transcription termination at the end of the *ilvGMEDA* operon.

A

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ilvC
280      290      300      310      320      330      340      350      360
ATG AAT GGC GCA CAG TGG GTG GTA CAT GCG TTC CGG GCA CAG GGT GTG AAC ACC GTT TTC GGT TAT CCC GGT GGC GCA ATT ATG CCG GTT
Met An Gly Ala Gln Trp Val Val His Ala Leu Arg Ala Gln Gly Val Asn Thr Val Phe Gly Tyr Pro Gly Gly Ala Ile Met Pro Val
1
370      380      390      400      410      420      430      440      450
TAC GAT GCA TTC TAT GAC GGC GGC GTG GAG CAC TTG CTA TGG CGA CAT GAG CAG GGT GCG GGA ATG GCG GCT ATC GGT TAT GCT CGT GCT
Tyr Asp Ala Leu Tyr Asp Gly Gly Val Glu His Leu Leu Cys Arg His Glu Gln Gly Ala Ala Met Ala Ala Ile Gly Tyr Ala Arg Ala
40
460      470      480      490      500      510      520      530      540
ACC GGC AAA ACT GGC GTA TGT ATC GCC ACG TCT GGT CCG GCG GCA ACC AAC CTG ATA ACC GGG CTT GCG GAC GCA CTG TTA GAT TCC ATC
Thr Gly Lys Thr Gly Val Cys Ile Ala Thr Ser Gly Pro Gly Ala Thr Asn Leu Ile Thr Thr Gly Leu Ala Asp Ala Leu Leu Asp Ser Ile
70
550      560      570      580      590      600      610      620      630
CCT GTT GTT GCC ATC ACC GGT CAA GTG TCC GCA CCG TTT ATC GGC ACT GAC GCA TTT CAG GAA GTG GAT GTG CTG GGA TTG TCG TTA GCC
CCT Val Val Ala Ile Thr Gly Gln Val Ser Ala Pro Phe Ile Gly Thr Asp Ala Phe Gln Glu Val Asp Val Leu Gly Leu Ser Leu Ala
110
640      650      660      670      680      690      700      710      720
TCT ACC AAG CAC AGC TTT CTG CTG CAG TCG CTG GAA CAG TTG CCG CGC ATC ATG GCT GAA GGA TTC CAG GCT GGC TGC TCA GGT CGT GCT
Cys Thr Lys His Ser Phe Leu Val Gln Ser Leu Glu Glu Leu Pro Arg Ile Met Ala Glu Ala Phe Asp Val Ala Cys Ser Gly Arg Pro
130
730      740      750      760      770      780      790      800      810
GGT CCG GTT CTG GTC GAT ATC CCA AAA CAT ATC CAG TTA GCC AGC GGT GAC CTG GAA CCG TGG TTC ACC ACC GTT GAA AAC GAA GTG ACT
Gly Pro Val Leu Val Asp Ile Pro Lys Asp Ile Gln Leu Ala Ser Gly Asp Leu Glu Pro Trp Phe Thr Thr Val Glu Asn Glu Val Thr
160
820      830      840      850      860      870      880      890      900
TTC CCA CAT GGC GAA GTT GAG CAA GCG CCG CAG ATG CTG GCA AAA GCG CAA AAA CCG ATC CTG TAC GTT GCG GGT GGC GTG GGT ATG GCG
Phe Pro His Ala Glu Val Glu Gln Ala Arg Gln Met Leu Ala Lys Ala Gln Lys Pro Met Leu Tyr Val Gly Gly Gly Val Gly Met Ala
200
910      920      930      940      950      960      970      980      990
CAG GGA GTT CCG GCT TTT CGT GAA TTT CTC GCT GCC ACA AAA ATG GCT GCC ACC TGT ACC CTG AAA GGG CTG GCG CCA GTA GAA GCA GAT
Gln Ala Val Pro Ala Leu Arg Glu Phe Leu Ala Ala Thr Lys Met Pro Ala Thr Cys Thr Leu Lys Gly Leu Gly Ala Val Glu Ala Asp
220
1000     1010     1020     1030     1040     1050     1060     1070     1080
TAT CCG TAC TAT CTG GGC ATG CTG GGG ATG CAC GGC ACC AAA GCG GCA AAC TTC GCG GTG CAG GAG TGT GAC CTG ATC GCC GTG GCG
Tyr Pro Tyr Tyr Leu Gly Met Leu Gly Met His Gly Thr Lys Ala Ala Asn Phe Ala Val Gln Glu Cys Asp Leu Leu Ile Ala Val Gly
250
1090     1100     1110     1120     1130     1140     1150     1160     1170
GCA CGT TTT GAT CAC GCG GTG ACC GGC AAA CTG AAC ACC TCC GCG CCA CAC GCC AGT GTT ATC CAT ATG GAT ATC GAC CCG GCA GAA ATG
Ala Arg Phe Asp Asp Arg Val Thr Gly Lys Leu Asn Thr Ser Ala Pro His Ala Ser Val Ile His Met Asp Ile Asp Pro Ala Glu Met
280
1180     1190     1200     1210     1220     1230     1240     1250     1260
AAC AAG CTU CGT CAG GCA CAT CTG GCA TTA CAA CGT GAT TTA AAT GCT CTG TTA CCA GCA TTA CAG CAG CCG TTA AAT CAA TAT GAC TGG
Asn Lys Leu Arg Gln Ala His Val Ala Leu Gln Gly Asp Leu Asn Ala Leu Leu Pro Ala Leu Gln Gln Pro Leu Asn Gln Tyr Asp Trp
310
1270     1280     1290     1300     1310     1320     1330     1340     1350
CAG CAA CAC TGC GCG CAG CTG GAT GAA CAT TCC TGC CTT TAC GAC CAT CCC GGT GAC GGT ATC TAC GCG CCG TTG TTG TTA AAA CAA
Gln Gln His Cys Ala Gln Leu Arg Asp Glu His Ser Trp Arg Tyr Asp His Pro Gly Asp Ala Ile Tyr Ala Pro Leu Leu Leu Lys Gln
340
1360     1370     1380     1390     1400     1410     1420     1430     1440
CTG TCC GAT CGT AAT CCT GCG GAT TCG GTC ACC ACA GAT GTG GCG CAG CAC CAG ATG TGG GCT GCG CAG CAC ATC CCG ACT CCG
Leu Ser Asp Arg Lys Pro Ala Asp Cys Val Val Thr Thr Asp Val Gly Gln His Gln Met Trp Ala Ala Gln His Ile Ala His Thr Arg
370
1450     1460     1470     1480     1490     1500     1510     1520     1530
CCG GAA AAT TTC ATC ACC TCC AGC GGT TTA GGT ACC ATG GGT TTT GGT TTA CCG GCG GCG GTT GCG GCA CAA GTC GCG GCA CCG AAC GAT
Pro Glu Asn Phe Ile Thr Ser Ser Gly Leu Gly Thr Met Gly Phe Gly Leu Pro Ala Ala Val Gly Ala Gln Val Ala Arg Pro Asn Asp
400
1540     1550     1560     1570     1580     1590     1600     1610     1620
ACC GTT GTC TGT ATC TCC GGT GAC GGC TCT TTC ATG ATG AAT GTG CAA GAG CTG GCG ACC GTA AAA GCG AAG CAG TTA CCG TTG AAA ATC
Thr Val Val Cys Ile Ser Gly Ser Phe Met Met Met Met Met Met Met Met Met Met Met Met Met Met Met Met Met Met Met Met Met
430
1630     1640     1650     1660     1670     1680     1690     1700     1710
GTC TTA CTC GAT AAC CAA CCG TTA GGG ATG GTT CGA CAA TGG CAG CAA CTG TTT TTT CAG GAA CGA TAC ACC GAA ACC ACC CTT ACT GAT
Val Leu Leu Asp Asn Gln Arg Leu Gly Met Val Arg Gln Trp Gln Gln Leu Phe Phe Gln Glu Arg Tyr Ser Glu Thr Thr Leu Thr Asp
460
    
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1720 1730 1740 1750 1760 1770 1780 1790 1800
AAC CCC GAT TTC CTC ATG TTA GCC AGC GGC TTC GGC ATC CAT GGC CAA CAC ATC ACC CGG AAA GAC CAG GTT GAA GCG GCA CTC GAC ACC
Asn Pro Asp Phe Leu Met Leu Ala Ser Ala Phe Gly Ile His Gly Gln His Ile Thr Arg Lys Asp Gln Val Glu Ala Ala Leu Asp Thr
510
1810 1820 1830 1840 1850 1860 1870 1880 1890
ATG CTG AAC AAT GAT GCG CGA TAC CTG CTT CAT GTC TCA ATC GAC GAA CTT GAG AAC GTC TGG CCG CTG GTG CCG CCT GGC GCC AAT AAT
Met Leu Asn Ser Asp Gly Pro Tyr Leu His Val Ser Ile Asp Glu Leu Glu Asn Val Trp Pro Leu Val Pro Pro Gly Ala Ser
520 530 540
1900 1910
TCA GAA ATG TTG GAG AAA TTA TCA TGA
Ser Glu Met Leu Glu Lys Leu Ser *

B

livM
1920 1930 1940 1950 1960 1970 1980 1990 2000
ATG ATG CAA CAT CAG GTC AAT GTA TGG CCG TTC AAT CCA GAA ACC TTA GAA CGT GTT TTA GCG GTG GTG CGT CAT CGT GGT TTC CAC
Met Met Gln His Gln Val Asn Val Ser Ala Arg Phe Asn Pro Glu Thr Leu Glu Arg Val Leu Arg Val Val Arg His Arg Gly Phe His
1 10 20 30
2010 2020 2030 2040 2050 2060 2070 2080 2090
GTC TGC TCA ATG AAT ATG GCC GCG GCC AGC GAT GCA CAA AAT ATA AAT ATC GAA TTG ACC GTT GCG ACC CCA CCG TCG GTC GAC TTA CTG
Val Cys Ser Met Asn Met Ala Ala Ala Ser Asp Ala Gln Asn Ile Asn Ile Glu Leu Thr Val Ala Ser Pro Arg Ser Val Asp Leu Leu
50 60
2100 2120 2130 2140 2150 2160 2170
TTT AGT CAG TTA AAT AAA CTG GTG GAC GTC GCA CAC GTT GGC ATC TGC CAG AGC ACA ACC ACA TCA GAA CAA ATC CCG GCC TGA
Phe Ser Gln Leu Asn Lys Leu Val Asp Val Ala His Val Ala Ile Cys Gln Ser Thr Thr Ser Gln Gln Ile Arg Ala *
70 80

C

2180 2190
SPACER CGCCAAAAGG AATATAAAA
livE
2200 2210 2220 2230 2240 2250 2260 2270 2280
ATG ACC ACC AAG AAA GCT GAT TAC ATT TGG TTC AAT GGC GAG ATG GTT GCG TCG GAA GAC GCG AAG GTC CAT GTG ATG TCG CAC GCG CTG
Met Thr Thr Lys Lys Ala Asp Tyr Ile Trp Phe Asn Gly Glu Met Ala Lys Val Arg Trp Glu Asp Ala Lys Val His Val Met Ser His Ala Leu
1 10 20 30
2290 2300 2310 2320 2330 2340 2350 2360 2370
CAC TAT GCG ACT TCG GTT TTT GAA GGC ATC GGT TCG TAC GAC TCG CAC AAA GGA CCG GTT GTA TTC CCG CAT CGT GAG CAT ATG CAG CGT
His Tyr Gly Thr Ser Val Phe Glu Gly Ile Arg Cys Tyr Asp Ser His Lys Gly Pro Val Val Phe Arg His Arg Glu His Met Gln Arg
40 50 60
2380 2390 2400 2410 2420 2430 2440 2450 2460
CTG CAT GAC TCC GCC AAA ATC TAT CCG TTC CCG GTT TCG CAG AGC ATT GAT GAG CTG ATG GAA GCT TGT CGT GAC GTC ATC CCG AAA AAC
Leu His Asp Ser Ala Lys Ile Tyr Arg Phe Pro Val Ser Gln Ser Ile Asp Glu Leu Met Glu Ala Cys Arg Asp Val Ile Arg Lys Asn
70 80 90
2470 2480 2490 2500 2510 2520 2530 2540 2550
AAT CTC ACC AGC GCC TAT ATC CGT CCG CTG ATC TTC GTC GGT GAT GTT GCG ATG GGA GTA AAC CCG CCA GCG GGA TAC TCA ACC GAC GTG
Asn Leu Thr Ser Ala Tyr Ile Arg Pro Leu Ile Phe Val Gly Asp Val Gly Met Gly Val Asn Pro Pro Ala Gly Tyr Ser Thr Asp Val
100 110 120
2560 2570 2580 2590 2600 2610 2620 2630 2640
ATT ATC GCT GCT TTC CCG TCG GCA GCG TAT CTC GCG GCA GAA GCG CTG GAC GAG GGG ATC GAT GCG ATG GTT TCC TCG TCG AAC CCG CCA
Ile Ile Ala Ala Phe Pro Trp Gly Tyr Leu Gly Ala Glu Ala Leu Glu Gln Gly Ile Asp Ala Met Val Ser Ser Trp Asn Arg Ala
130 140 150
2650 2660 2670 2680 2690 2700 2710 2720 2730
GCA CCA AAC ACC ATC CCG ACC GCG GCA AAA GCG GGT GGT AAC TAC CTC TCT TCC CTG CTG GTG GGT AGC GAA GCG CCG GCC CAC GGT TAT
Ala Pro Asn Thr Ile Pro Thr Ala Ala Lys Ala Gly Gly Asn Tyr Leu Ser Ser Leu Ala Arg His Gly Ser His Gly Tyr
160 170 180
2740 2750 2760 2770 2780 2790 2800 2810 2810
CAG GAA GGT ATC GCG CTG GAT GTC AAC GGT TAT ATC TCT GAA GCG GCA GCG GAA AAC CTG TTT GAA GTG AAA GAT GGT GTG CTC TTC ACC
Gln Glu Gly Ile Ala Leu Asp Val Asn Gly Tyr Ile Ser Glu Gly Ala Gly Glu Asn Leu Phe Glu Val Lys Asp Gly Val Lys Phe Thr
190 200 210
2830 2840 2850 2860 2870 2880 2890 2900 2910
CCA CCG TTC ACC TCC TCC GCG CTG CCG GGT ATT ACC GGT GAT GCC ATC ATC AAA CTC CCG AAA GAG CTG GGA ATT GAA GTA CTT GAG CAG
Pro Pro Phe Thr Ser Ser Ala Leu Pro Gly Ile Thr Arg Asp Ala Ile Ile Lys Leu Ala Lys Glu Leu Gly Ile Glu Val Arg Glu Gln
220 230 240
2920 2930 2940 2950 2960 2970 2980 2990 3000
GTG CTG TCG CCG GAA TCC CTC TAC CTG GCG GAT GAA GTG TTT ATG TCC GGT ACC GCG GCA GAA ATC ACG CCA GTG CCG ACC GTA GAC GGT
Val Leu Ser Arg Glu Ser Leu Tyr Leu Ala Asp Glu Val Phe Met Ser Gly Thr Ala Ala Glu Ile Thr Pro Val Arg Ser Val Asp Gly
250 260 270
3010 3020 3030 3040 3050 3060 3070 3080 3090
ATT GAT CTT GGC GAA GGC CGT TGT GCG CCG GTT ACC AAA CCG ATT CAG CAA GCG TTC TTC GGC CTC TTC ACT GCG GAA ACC GAA GAT AAA
Ile Gln Val Gly Glu Gly Arg Cys Gly Pro Val Thr Lys Arg Ile Gln Gln Ala Phe Phe Gly Leu Phe Thr Gly Glu Thr Glu Asp Lys
280 290
3100
TGG GCG TGG TTA GAT CAA GTT AAT CAA TAA
Trp Gly Trp Leu Asp Gln Val Asn Gln *
310

D

3130 3140 3150 3160 3170 3180 3190
SPACER ATACAAAAAT GGCAGCCAC GCACCGTCCC ATTTACGACA CAGACACTGG GAGTAAATAA AGT
livD
3200 3210 3220 3230 3240 3250 3260 3270 3280
ATG CTT AAG TAC GGT TCC GCG ACC ACC ACT CAT GGT GGT AAT ATG CCG GGT GCT CTT GCG CTC TCG GCG ACC ACC GGA ATG ACC GAC CCG
Met Pro Lys Tyr Arg Ser Ala Thr Thr His Gly Arg Asn Met Ala Gly Ala Arg Ala Leu Trp Arg Ala Thr Gly Met Thr Asp Ala
1 10 20 30
3290 3300 3310 3320 3330 3340 3350 3360 3370
GAT TTC GGT AAG CCG ATT ATC GCG GTT GTG AAC TCG TTC ACC CAA TTT GTA CCG GGT CAC GTC CAT CTG CCG GAT CTC GGT AAA CTC GTC
Asp Phe Gly Lys Pro Ile Ile Ala Val Val Asn Ser Phe Thr Gln Phe Val Pro Gly His Val His Leu Arg Asp Leu Gly Lys Leu Val
30 40 50 60
3380 3390 3400 3410 3420 3430 3440 3450 3460
GCC GAA CAA ATT GAA GCG GGT GCG GCG GTT GCC AAA GAG TTC AAC ACC ATT GCG GTG GAT GAT GCG ATT GCG ATG GCG CAC GCG GCG ATG
Ala Glu Gln Ile Glu Ala Ala Gly Val Ala Lys Glu Phe Asn Thr Ile Ala Val Asp Asp Gly Ile Ala Met Gly His Gly Met
70 80 90

Nucleic Acids Research

3470 3480 3490 3500 3510 3520 3530 3540 3550
 CTT TAT TCA GCT CCA TCT CGC GAA CTC ATG GCT GAT TCC GTT GAG TAT ATG GTC AAC GCC CAC TGC GCC GAC GCC ATG GTC TGC ATC TCT
 Leu Tyr Ser Leu Pro Ser Arg Glu Leu Ile Ala Asp Ser Val Glu Tyr Met Val Aen Ala His Cys Ala Asp Ala Met Val Cys Ile Ser
 100
 3560 3570 3580 3590 3600 3610 3620 3630 3640
 AAC TGC GAC AAA ATC ACC CGC GGG ATG CTG ATC GTC GGC CTG AAT ATT CCG GTC ATT TTT GTT TCC GGC GGC CGC ATG GAG GCC
 Aen Cys Asp Lys Ile Thr Pro Gly Met Leu Met Ala Ser Leu Arg Leu Aen Ile Pro Val Ile Phe Val Ser Gly Gly Pro Met Glu Ala
 150
 3650 3660 3670 3680 3690 3700 3710 3720 3730
 GGC AAA ACC AAA CTT TCC GAT CAC ATC ATC CAG CTT GAT CCG GGT GAT CCG GGC GGC GGC GGC AAA GTA TCT GAC TCC CAG
 Gly Lys Thr Lys Leu Ser Asp Glu Ile Ile Lys Leu Asp Pro Val Asp Ala Met Ile Glu Gly Ala Asp PCC Pro Lys Val Ser Asp Ser
 180
 3740 3750 3760 3770 3780 3790 3800 3810 3820
 AGC GAT CAC GTT GAA CGT TCC GCG TGT CCG ACC TCC GGT TCC TCC TCC GGC ATG TTT ACC GCT AAC TCA ATG AAC TGC CTG ACC GAA GGC
 Ser Asp Glu Val Glu Arg Ser Ala Cys Pro Thr Cys Gly Ser Cys Ser Gly Met Phe Thr Ala Aen Ser Met Aen Cys Leu Thr Glu Ala
 210
 3830 3840 3850 3860 3870 3880 3890 3900 3910
 CTG GGC CTG TGC GAG CCG GGC AAC GGC TGC CTG CCA ACC CAC GGC CAC GGT AAG CAG CTC TTC CTT ATT GGT GGT AAA GGC ATT GTT
 Leu Gly Leu Ser Glu Pro Gly Aen Gly Ser Leu Leu Ala Thr His Ala Asp Arg Lys Glu Leu Phe Leu Aen Ala Gly Lys Arg Ile Val
 240
 3920 3930 3940 3950 3960 3970 3980 3990 4000
 GAA TTG ACC AAA CGT TAT TAC GAG CAA AAC GAC GAA AGT GCA CTG CCG GGT AAT ATC CCG ACC AAG GGC GGC TTT GAA AAC GCC ATC AGC
 Glu Leu Thr Lys Arg Tyr Tyr Glu Glu Aen Asp Glu Ser Ala Leu Pro Arg Aen Ile Ala Ser Lys Ala Ala Phe Glu Aen Ala Met Thr
 270
 4010 4020 4030 4040 4050 4060 4070 4080 4090
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 Leu Asp Ile Ala Met Gly Gly Ser Thr Aen Thr Val Leu His Leu Ala Ala Glu Ile Asp Phe Glu Thr Met Ser Asp
 300
 4100 4110 4120 4130 4140 4150 4160 4170 4180
 ATC GAT AAC CTT TCC CCG AAG GTT CCA CAG CTG TGT AAA GTT GCG CCG AGC ACC CAG TAC CAT ATG GAA GAT GTT CCA CGT GCT GGT
 Ile Asp Lys Leu Ser Arg Lys Val Pro Glu Lys Cys Lys Val Ala Pro Ser Thr Glu Lys Tyr His Met Glu Asp Val His Arg Ala
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 4190 4200 4210 4220 4230 4240 4250 4260 4270
 GGT CTT ATC GGT ATT CTC GGC GAA CTG GAT CCG GGC TTA CTG AAC CGT GAT GTG AAA AAC GTA CTT GGC CTG ACC TTG CCG GAA ACC
 Gly Val Ile Gly Ile Leu Gly Glu Leu Asp Arg Ala Gly Leu Leu Aen Arg Asp Val Lys Aen Val Leu Gly Leu Thr Leu Pro Glu Thr
 360
 4280 4290 4300 4310 4320 4330 4340 4350 4360
 CTG GAA CAA TAC GAC CTT ATG CTC ACC CAG CAT GAC CCG GTA AAA AAT ATG TTC CCG GCA GGT CGT GCA GGC ATT GCT ACC ACA CAG GCA
 Leu Glu Glu Tyr Asp Val Met Leu Thr Glu Asp Asp Ala Val Lys Aen Met Phe Arg Ala Gly Pro Ala Gly Ile Arg Thr Thr Glu Ala
 390
 4370 4380 4390 4400 4410 4420 4430 4440 4450
 TTC TCG CAA GAT TCC CCT TCG GAT ACG CTC GAC GAC GAT CCG CCG AAT GGC GTA TCC CCG CCG TCG AAC ACG CCT ACA GCA AAG ACG GCG
 Phe Ser Glu Asp Cys Arg Trp Asp Thr Leu Asp Asp Asp Arg Ala Aen Gly Val Ser Ala Arg Trp Aen Thr Pro Thr Ala Lys Thr Ala
 420
 4460 4470 4480 4490 4500 4510 4520 4530 4540
 GCC TGG CCG TCG TCT ACG GTA ATC TTG CCG AAA CCG CTG CAT GGT GAA ACG GCA GGC CTC GAT GAC AGC ATC CTC AAA TTC ACC GGC GGC
 Ala Trp Arg Cys Ser Thr Val Ile Leu Arg Lys Arg Leu His Arg Glu Thr Ala Gly Val Asp Asp Ser Ile Leu Lys Phe Thr Gly Pro
 450
 4550 4560 4570 4580 4590 4600 4610 4620 4630
 GGC AAA CTG TAC CAA ACG CAG GAC GAT GGC GTA GAA GGC ATT CTC GCG GGT AAA GTT CTC CCG GCA GAT GTG GTA GTA ATT CCG TAT
 Ala Lys Val Tyr Glu Ser Glu Asp Asp Ala Val Glu Ala Ile Leu Gly Gly Lys Val Val Ala Gly Asp Val Val Val Ile Arg Tyr Glu
 480
 4640 4650 4660 4670 4680 4690 4700 4710 4720
 GGC CCG AAA GGC GGT CCG GGC ATG CAG GAA ATC CTC TAC CCA ACC GCG TTC CTG AAA TCA ATT GGT CTC GGC AAA GCC TGT GCG CTG ATC
 Gly Pro Lys Gly Gly Pro Gly Met Glu Gly Met Leu Tyr Pro Thr Ser Phe Leu Lys Ser Met Met Leu Gly Lys Ala Cys Ala Leu Ile
 510
 4730 4740 4750 4760 4770 4780 4790 4800 4810
 ACC GAC GGT CTT TCT TCT GGT GGC ACC TGT GGT CTT TCC ATC GGC CAC CTC TCA CCG GAA GGC GCA ACC GGC ACC GAT ACC GCT ATT
 Thr Asp Gly Arg Phe Ser Gly Gly Thr Ser Gly Leu Ser Ile Gly His Val Ser Pro Glu Ala Ala Ser Gly Gly Ser Ile Gly Leu Ile
 540
 4820 4830 4840 4850 4860 4870 4880 4890 4900
 GAA GAC GGT GAC CTG ATC GCT ATC GAC ATC CCG AAC CGT GGC ATT CAG TTA CAG GTA ACC GAT CCG GAA CTG CCG GCT CGT GAA GGC
 Glu Asp Gly Asp Leu Ile Ala Ile Asp Ile Pro Aen Arg Gly Ile Glu Leu Glu Val Ser Ser Asp Ala Glu Leu Ala Ala Arg Arg Glu Ala
 570
 4910 4920 4930 4940 4950 4960 4970 4980 4990
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 Glu Asp Ala Arg Gly Asp Lys Ala Trp Thr Pro Lys Aen Arg Glu Arg Glu Val Ser Phe Ala Leu Arg Ala Tyr Ala Ser Ala Thr
 600
 5000 5010 5020 5030
 ACC GCC CAG AAA GGC GCG GTG CCG GAT AAA TCG AAA CTG GGC GGT TAA
 Ser Ala Asp Lys Gly Ala Val Arg His Lys Ser Lys Leu Gly Gly *
 610

E

11vA
 5040
 TA ATG GCT GAC TCG CAA CCC CTG TCC GGT GCT CCG GAA GGT GCC GAA TAT TTA AGA GCA GTG CTG CCG CCG CCG GTT TAC GAG CCG GCG
 Met Ala Asp Ser Glu Pro Leu Ser Gly Ala Pro Glu Gly Leu Ala Glu Tyr Leu Arg Ala Val Leu Arg Ala Pro Val Tyr Glu Ala Ala
 1
 10
 5130 5140 5150 5160 5170 5180 5190 5200 5210
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 Glu Val Thr Pro Leu Lys Met Glu Lys Leu Ser Ser Arg Leu Asp Aen Val Ile Leu Val Lys Arg Glu Asp Arg Glu Asp Arg Glu Val His
 30
 40
 5220 5230 5240 5250 5260 5270 5280 5290 5300
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 60
 70
 5310 5320 5330 5340 5350 5360 5370 5380 5390
 CAC GCG CAG GGC GTC CCG TTT TCT TGT CCG GCG TTA GGC GTG AAG GCC CTG ATC GTT ATG CCA ACC GCG ACC GCG GAC ATC AAA GTC GAC
 His Ala Glu Gly Val Ala Phe Ser Ser Ala Arg Leu Gly Val Lys Ala Leu Ile Val Met Pro Thr Ala Thr Ala Asp Ile Lys Val Asp
 90
 100
 5400 5410 5420 5430 5440 5450 5460 5470 5480
 GCG GTG CCG GGC TTC GGC GAA GTG CTG CTC CAC GCG GGC AAC TTT GAT GAA GCG AAA GCG AAA CCG ATC GAA CTG TCA CAG CAG CAG
 Ala Val Arg Gly Phe Gly Gly Glu Val Leu Leu His Gly Ala Aen Phe Asp Glu Ala Lys Ala Lys Ala Ile Glu Leu Ser Glu Glu Glu
 120
 130

5490 5500 5510 5520 5530 5540 5550 5560 5570
GGG TTC ACC TGG GTG CCG CCG TTC GAC CAT CCG ATG GTG ATT GCC GGC CAA GGC ACG CTG GCG CTG GAA CTG CTC CAG CAG GAC GCC CAT
Gly Phe Thr Trp Val Pro Pro Phe Asp His Pro Met Val Ile Ala Gly Gln Gly Thr Leu Ala Leu Glu Leu Leu Gln Gln Asp Ala His
150
5580 5590 5600 5610 5620 5630 5640 5650 5660
CTC GAC CGC GTA TTT GTG CCA GTC GGC GGC GGC GGT CTG GCT GCT GGC GTG GCG GTG CTG ATC AAA CAA CTG ATG CCG CAA ATC AAA GTG
Leu Asp Arg Val Phe Val Pro Val Gly Gly Gly Leu Ala Ala Gly Val Ala Val Leu Ile Lys Gln Leu Met Pro Gln Ile Lys Val
180
5670 5680 5690 5700 5710 5720 5730 5740 5750
ATC GCC GTA GAA GCG GAA GAC TCC GCC TGC CTG AAA GCA GCG CTG GAT GCG GGT CAT CCG GTT GAT CTG CCG CGC GTA GGG CTA TTT GCT
Ile Ala Val Glu Ala Glu Asp Ser Ala Cys Leu Lys Ala Ala Leu Asp Ala Gly His Pro Val Asp Leu Pro Arg Val Gly Leu Phe Ala
210
5760 5770 5780 5790 5800 5810 5820 5830 5840
GAA GGC GTA GGC GTA AAA CGC ATC GGT GAC GAA ACC TTC CGT TTA TGC CAG GAG TAT CTC GAC GAC ATC ATC ACC GTC GAT AGC GAT GCC
Glu Gly Val Gly Val Lys Arg Ile Gly Asp Glu Thr Phe Arg Leu Cys Gln Gly Tyr Leu Asp Asp Ile Ile Thr Val Asp Ser Asp Ala
240
5850 5860 5870 5880 5890 5900 5910 5920 5930
ATC TGT CCG GCG ATG AAG GAT TTA TTC GAA GAT GTG CCG GCG GTG CCG CCA CCG TCT GCG CGC CTG CCG CTG CCG GGA ATG AAA AAA TAT
Ile Cys Ala Ala Met Lys Asp Leu Phe Glu Asp Val Arg Ala Val Ala Glu Pro Ser Gly Ala Leu Ala Leu Ala Gly Met Lys Lys Tyr
270
5940 5950 5960 5970 5980 5990 6000 6010 6020
ATC GCC CTG CAC AAC ATT CCG GGC GAA GGC CTG CCG CAT ATT CTT TCC GGT GCC AAC GTG AAC TTC CAC GGC CTG CCG TAC GTC TCA GAA
Ile Ala Leu His Asn Ile Arg Gly Glu Arg Leu Ala His Ile Leu Ser Gly Ala Asn Val Phe His Gly Leu Arg Tyr Val Ser Glu
300
6030 6040 6050 6060 6070 6080 6090 6100 6110
CGC TGC GAA CTG GGC GAA CAG CCG GAA GGC TTC TTG CCG GTG ACC ATT CCG GAA GAA AAA GGC AGC TTC CTC AAA TTC TGC CAA CTG CTT
Arg Cys Glu Leu Gly Glu Gln Arg Glu Ala Leu Leu Ala Val Thr Ile Pro Glu Glu Lys Gly Ser Phe Leu Lys Phe Cys Gln Leu Leu
330
6120 6130 6140 6150 6160 6170 6180 6190 6200
GGC GGG CGT TCG GTC ACC GAG TTC AAC TAC CGT TTT GCC GAT GCC AAA AAC GCC TCC ATC TTT GTC GGT GTG CCG CTG AGC CGC GGC CTC
Gly Gly Arg Ser Val Thr Glu Phe Asn Tyr Arg Phe Ala Asp Ala Lys Asn Ala Cys Ile Phe Val Gly Val Arg Leu Ser Arg Gly Leu
360
6210 6220 6230 6240 6250 6260 6270 6280 6290
GAA GAG CGC AAA GAA ATT TTG CAG ATG CTC AAC GAC GGC GGC TAC AGC GTG GTT GAT CTC TCC GAC GAC GAA ATG CCG AAC CTA CAC GTC
Glu Arg Lys Glu Ile Leu Gln Met Leu Asn Asp Gly Gly Tyr Ser Val Val Asp Leu Ser Asp Asp Glu Met Ala Lys Leu His Val
390
6300 6310 6320 6330 6340 6350 6360 6370 6380
CGC TAT ATG GTC GGC GGA CCG CCA TCG CAT CCG TTG CAG GAA CGC CTC TAC AGC TTC GAA TTC CCG GAA TGA CCG GGC GGC CTC GCG
Arg Tyr Met Val Gly Gly Arg Pro Ser His Pro Leu Gln Glu Arg Leu Tyr Ser Phe Ser Phe Glu Phe Pro Glu Ser Pro Gly Ala Leu Leu Arg
420
6390 6400 6410 6420 6430 6440 6450 6460 6470
TTC CTC AAC ACG CTG GGT ACG TAC TGG AAC ATT TCT TTG TTC CAC TAT CCG AGC CAT GGC ACC GAC TAC GGG CGC GTA CTG GCG GCG TTC
Phe Leu Asn Thr Leu Gly Thr Tyr Trp Asn Ile Ser Leu Phe His Tyr Arg Ser His Gly Thr Asp Tyr Gly Arg Val Leu Ala Ala Phe
450
6480 6490 6500 6510 6520 6530 6540 6550 6560
GAA CTT GGC GAC CAT GAA CCG GAT TTC GAA ACC CCG CTG AAT GAG CTG GGC TAC GAT TGC CAC GAC GAA ACC AAT AAC CCG GCG TTC AGG
Glu Leu Gly Asp His Glu Pro Asp Phe Glu Thr Arg Leu Asn Glu Leu Gly Tyr Asp Cys His Asp Glu Thr Asn Asn Pro Ala Phe Arg
480
6570 6580
TTC TTT TTG GCG GGT TAG
Phe Phe Leu Ala Gly *
510

Figure 8. The complete nucleotide sequence of the nontranscribed DNA strand of the structural genes of the *ilvGMEDA* operon. The nucleotide sequence is numbered relative to the *in vivo* transcriptional initiation site. The amino acid residues of each encoded polypeptide are numbered from the amino terminal Met. The nucleotide sequence of the intercistronic region preceding each gene is identified as spacer. The nucleotide sequence of each structural gene listed in this figure are presented as follows: A) *ilvG*; B) *ilvM*; C) *ilvE*; D) *ilvD*; E) *ilvA*. It is noted that two nucleotides, a C at 3257 and a G at 3260, have been inserted compared to a previously published sequence (33). These nucleotide insertions are consistent with the fact that a *ApaI* restriction site (GGGCCC), which was present in the previously published sequence, does not exist, while a *BssHII* restriction site (GCGCGC) is conserved at this site in both *E. coli* K-12 and *S. typhimurium*.

restriction fragment containing this sequence fused to the *tac* promoter indicate that this is a rho-independent termination site (unpublished observation). When termination factor rho (32) was added to the system, a rho-dependent termination site was detected about 100 bp downstream of the rho-independent termination site. Therefore, the arrangement of transcriptional terminators at the 3'-terminus of the *ilvGMEDA* operon

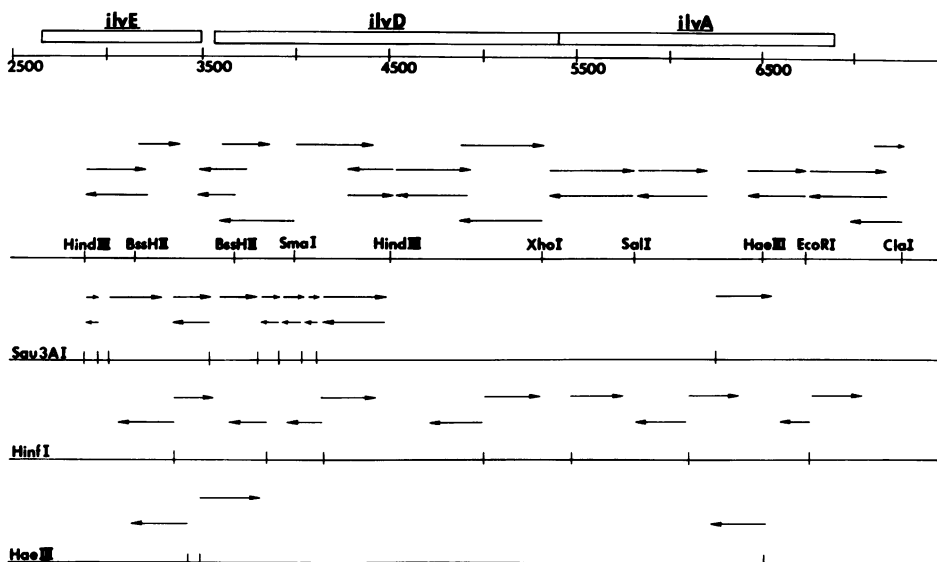


Figure 9. Nucleotide sequence strategy for the *ilvE*, *ilvD* and *ilvA* genes. The lengths of the arrows represent the extent of readable sequence determined by the Maxam and Gilbert chemical cleavage method (34) and dideoxy chain termination method of Sanger et al. (35). It is noted that the nucleotide sequence of the *ilvE* gene and the 5'-portion of the *ilvD* gene have been previously published (33). Nucleotide differences between the sequence determined in this strategy and that of previous work (33) are indicated in the figure 8 legend.

appears to be very similar to the well characterized *trp* *tt'* terminators of the *trp* operon of *E. coli* (32) (unpublished observations). It is interesting to note that both transcriptional termination sites are located in the *ilvY* coding region. This gene is transcribed in an opposite direction to the *ilvA* gene (9). The positions of the *ilvA* and *ilvY* stop codons are indicated by asterisks in figure 7.

III. STRUCTURAL GENES OF THE *ilvGMEDA* OPERON

A schematic diagram of the *ilvGMEDA* operon outlining the location of each structural gene is presented in figure 2. The complete nucleotide sequence of the non-transcribed DNA strand and the corresponding deduced amino acid sequence of the five structural genes are presented in figure 8. The nucleotide sequence of the *ilvG* and *ilvM* genes are from previous publications (5) and the sequence of the *ilvE*, *ilvD* and *ilvA* genes were determined according to the strategy shown in figure 9.

The following criteria were used to predict each polypeptide sequence. First, each structural gene had an open translational reading frame which encodes a product with a molecular weight consistent with the electrophoretic analysis of the gene products synthesized *in vivo*. Second, the initiation codons for the *ilvG*, *ilvM* and *ilvE* deduced

amino acid sequences were selected based on the amino terminal sequences determined from the purified protein products of these genes (36-38). Finally, the nucleotide sequence of the intergenic regions of the structural genes of the ilvGMEDA operon in S. typhimurium were determined and compared to the homologous regions in E. coli (Fig. 10). Because these two enteric bacteria are so closely related, coding sequences are usually conserved while noncoding sequences, unless they perform a regulatory role, are diverged. Based on this coding conservation the probable initiation codons for the ilvD and ilvA genes of E. coli were determined and the deduced carboxy terminal sequences for each structural gene were confirmed. These points will be addressed more fully in the description of the structural genes of the operon below.

The ilvG and ilvM genes encode the large and small subunits, respectively, of the AHAS II isozyme. The AHAS II isozyme is one of three AHAS isozymes in E. coli K-12. As mentioned in the introduction, the ilvG gene in E. coli K-12 contains a frameshift mutation which results in termination of translation approximately in the middle of the gene (4-6). Mutations have been isolated in E. coli K-12 which displace this frameshift site. These ilvG mutations (previously referred to as ilvQ mutations) result in the full length expression of the ilvG gene and, therefore, a functional AHAS II isozyme. The deduced amino acid sequences of the ilvG and ilvM polypeptides are presented in figures 8A and 8B, respectively. The ilvG sequence shown contains the mutation ilvG2096 (5), which differs from the wild type E. coli K-12 sequence by the deletion of a G residue between nucleotides 1252 and 1253 in figure 8A (5,6). Several other mutations have been characterized and each mutation involves a one base pair deletion or a two base pair insertion within a ten base pair region preceding the frameshift site in the ilvG gene of E. coli K-12 (6).

Recently, the AHAS II isozyme from S. typhimurium has been purified and characterized (36). The AHAS II isozyme was shown to be a tetramer composed of two large and two small subunits with molecular weights of 59,300 and 9,700, respectively. The amino terminal sequences of the ilvG and ilvM encoded subunits were determined and these compare almost identically with the deduced E. coli subunit sequences (Fig. 8A and 8B). Of the 35 amino terminal residues determined for the S. typhimurium ilvG subunit there is only a single amino acid difference, a lysyl residue substituted for an asparaginyl residue at position 17 of the E. coli sequence. The amino terminal analysis of the ilvM subunit showed the first 25 residues are identical between S. typhimurium and E. coli. An amino acid composition analysis of the small and large AHAS II subunits of S. typhimurium were also very similar to the deduced polypeptide sequences of E. coli.

The intergenic region between the ilvG and ilvM genes of E. coli and the homologous region of S. typhimurium is presented in figure 10. The ilvG chain termination codon overlaps the ilvM initiation codon by four nucleotides in both enteric bacteria. There are similar examples of overlapping termination and initiation codons in the trpE-trpD and trpB-trpA intergenic regions of the trp operon of E. coli. In both examples the pairs of genes which overlap encode non-identical polypeptides which are associated in multi-subunit enzyme complexes. It has been suggested that this juxtaposition of translational signals is important in the coordinate synthesis of these subunits (39-42). It is possible, therefore, that the overlap of the ilvG and ilvM genes

also functions in an analogous manner to insure the equimolar expression of the large and small AHAS II subunits (36).

The primary branched chain amino acid transaminase, transaminase B, is the product of the *ilvE* gene. Transaminase B from *E. coli* has been purified to apparent homogeneity and the native enzyme was characterized as a hexamer of identical subunits with molecular weights of about 32,000 (37). The amino terminal sequence and amino acid composition was determined for the *ilvE* encoded product and both compare favorably with the deduced amino acid sequence determined from the nucleotide sequence (Fig. 8C) (37,38). One interesting point concerning the amino terminal sequence of transaminase B is that it appears to be post-translationally modified. Two independent determinations of this sequence begin Gly Thr Lys Lys Ala. When compared to the deduced amino acid sequence in figure 8C, it appears that the amino terminal Met residue has been cleaved and the Thr residue modified to a Gly residue. In *S. typhimurium*, the amino terminal sequence of transaminase B is Thr Thr Lys Lys Ala (R. R. Randall, M. H. Wallis, G. J. Young and F. B. Armstrong, Fed. Am. Abstr. 38, 325, 1979). In this example it appears that only the Met residue has been cleaved.

The *ilvD* gene encodes dihydroxy acid dehydrase, which catalyzes the dehydration of the α,β -dihydroxy acids to yield the corresponding α -keto acids (Fig. 1). The nucleotide sequence of the nontranscribed DNA strand and corresponding deduced amino acid sequence of the *ilvD* product is presented in figure 8D. The encoded polypeptide has a molecular weight of approximately 66,000 which is in agreement with that determined from electrophoresis of labeled maxicell products of the *ilvD* gene (43,44). It is interesting to note that in the 64 bp intercistronic region between the *ilvE* and *ilvD* genes there is an 11 bp inverted repeat (Fig. 8D). Although there is no direct evidence, it is possible that this region, which ends with 3 T residues in row, could function as a weak, operon internal, rho-independent terminator (45). In the *ilvE-ilvD* intercistronic region of *S. typhimurium* this region of dyad symmetry is not present (Fig. 10); however, there is approximately 100 bp of additional sequence in this intercistronic region of *S. typhimurium* which is not represented in *E. coli*. At present we are not certain of the origin or function of this insertion/deletion in the *ilvE-ilvD* intercistronic region of *E. coli* and *S. typhimurium*.

The most distal gene of the operon, *ilvA* encodes threonine deaminase. This enzyme catalyzes the first enzymatic step of the isoleucine pathway (Fig. 1). In 1956, Umbarger made the classical observation that threonine deaminase was inhibited by isoleucine (46). This phenomena whereby the end-product of a pathway inhibits the first enzymatic step of a pathway was termed end-product inhibition. End-product inhibition has subsequently been shown to be a universal type of metabolic regulation.

Threonine deaminase has been purified to near homogeneity in many organisms, including *Bacillus subtilis* (47), *S. typhimurium* (48), *E. coli* (49-51) and *Saccharomyces cerevisiae* (52,53). It is a tetramer of identical subunits, and, in *E. coli*, has a monomer molecular weight of 53,000 as determined by SDS gel electrophoresis (49). The nucleotide sequence of the nontranscribed DNA strand of the *ilvA* gene and the corresponding deduced threonine deaminase sequence is presented in figure 8E. We reported previously the nucleotide sequence of the 3'-portion of the *ilvA* gene (9). The deduced 76 amino acids of this carboxy terminal region of *E. coli* threonine deaminase

shares 51% homology with the deduced carboxy terminus of the S. cerevisiae enzyme (9,54). With the completion of the nucleotide sequence of the ilvA gene from E. coli we compared the entire deduced threonine deaminase sequence of E. coli with the deduced sequence from S. cerevisiae (54). The gene encoding threonine deaminase from S. cerevisiae has an open translational reading frame which could encode a polypeptide of 63,700, but the purified subunit has been characterized to have a molecular weight of 48,000 (52). Kielland-Brandt et al. (54) suggested that since threonine deaminase in S. cerevisiae is located in the mitochondria a larger molecular weight precursor polypeptide is processed by proteolytic cleavage into a 48,000 molecular weight subunit. This processing is consistent with the cleaving of a signal sequence characterized for many proteins that are synthesized in the cytoplasm and transported into the mitochondria (55). As predicted by the processing model there is no detectable homology between the first 62 amino acids of threonine deaminase from S. cerevisiae and the amino terminal sequence of E. coli. Amino acids 16 through 514 from the threonine deaminase sequence of E. coli were aligned with residues 63 through 592 from the S. cerevisiae sequence. A minimum number of gaps were allowed to maximize the alignment between the two sequences. This alignment shows 50% identical amino acid homology between the paired sequences. The homology is interspersed throughout the threonine deaminase sequences with a large gap of non-homology between the E. coli residues 371-390 and S. cerevisiae residues 434-462.

IV. QUESTIONS TO BE ADDRESSED IN THE FUTURE

In the preceding sections we presented the complete nucleotide sequence and described the structural and functional features of the regulatory and coding regions of the ilvGMEDA operon of E. coli. In this section we consider some remaining questions concerning the role of the ilvGMEDA proximal promoter and attenuator in the regulation of this operon.

Attenuation, a mechanism involved in the control of several amino acid biosynthetic operons, was first described in the trp operon of E. coli. In addition to attenuation, the trp operon is also regulated by repression. The trp repressor regulates transcription initiation of the operon in response to changes in the intracellular availability of free tryptophan, while attenuation regulates transcription termination in the leader region of the trp operon in response to the intracellular levels of aminoacylated tRNA^{Trp}. Yanofsky et al. (56) have shown that repression and attenuation respond to different degrees of tryptophan starvation. The trp repressor regulates transcription initiation during growth with excess to moderately limiting levels of tryptophan, whereas, attenuation is not relaxed until tryptophan limitation is in the range of moderate to severe. Unlike the trp operon, however, there is no direct evidence for the involvement of a repressor in the regulation of the ilvGMEDA operon. This implies that this operon is regulated exclusively by attenuation. Such a possibility creates an apparent paradox. During increasing growth rates accompanying a shift from a poor to a good carbon source, protein synthesis rates increase. Concomitantly, the rates of synthesis of the branched chain amino acids increase. If attenuation of the ilvGMEDA operon is responsible for this growth rate regulation, and if attenuation of the ilvGMEDA operon parallels that of the trp operon, then relaxed attenuation at increased

growth rates must be accompanied by a moderate to severe starvation for aminoacylated branched chain tRNAs. However, such limitations for aminoacylated tRNAs are not observed during rapid rates of growth and protein synthesis. In order to accommodate this dilemma it has been noted that amino acid biosynthetic operons which are apparently regulated solely by attenuation (ilvGMEDA, his, thr, leu) contain several tandem regulatory codons, whereas, the attenuator region of the trp operon contains only two tandem trp codons. It has been proposed that there is a translational delay at each regulatory codon which is amplified at each successive regulatory codon (57). This "queuing effect" could make attenuation hypersensitive to relatively small changes in the intracellular levels of the regulatory aminoacylated tRNA. Additionally, the fact that rare codons recognized by minor tRNA isoacceptor species are used as regulatory codons for attenuation may further enhance this sensitivity. One last point, which is surely important, is the coding context of these tandem regulatory codons. For example, the leader polypeptide of the ilvGMEDA operon of S. marcescens encodes a single Leu regulatory codon (30). Since this codon, albeit rare, appears in structural gene coding sequences, its role as a regulatory codon in the leader transcript is probably enhanced by non-leucine flanking codons.

Structural similarities between the promoter region of the ilvGMEDA operon and the tyrT gene, which is regulated by growth rate control, suggests an alternative explanation for the mechanism of growth rate regulation of the ilvGMEDA operon that does not involve attenuation. A deletion analysis of the promoter region of the tyrT gene of E. coli has demonstrated that, like the ilvGMEDA promoter region, nucleotide sequences upstream of this promoter (between positions -40 to -98) are required for maximal expression of the tyrT gene (58). These sequences upstream of the tyrT promoter have affinity for RNA polymerase in vitro as judged by DNAase I protection assays (59). The promoter region of the ilvGMEDA operon consists of tandem in vitro promoters P1P2 (Fig. 5) (14,16,18). In vivo, however, initiation of transcription is detected at only the downstream P2 promoter (16). Travers et al. (59) have proposed a model in which an RNA polymerase bound at an upstream binding site activates transcription from a polymerase bound at the downstream primary promoter. Such an activation could involve protein-protein contact between the adjacent RNA polymerase molecules. If this is so, then tyrT promoter activity would be proportional to the square of polymerase concentration. Since the concentration of RNA polymerase in the cell increases linearly with the growth rate, such a mechanism could explain why the synthesis of stable RNA is proportional to the square of the growth rate. An analogous model can be invoked to explain the role of the dual promoters of the ilvGMEDA operon. The cooperative action of RNA polymerase bound at the upstream P1 binding site could activate transcription, in vivo, by a second RNA polymerase bound at the downstream P2 promoter. This cooperative action could account for the growth rate regulation of the ilvGMEDA operon.

In conclusion, a fundamental problem for the future is to elucidate the physiological roles of the ilvGMEDA promoter and attenuator regions. Most certainly, the attenuation mechanism functions to increase expression of the operon during conditions of severe limitation of aminoacylated branched chain amino acid tRNAs. But this only explains why elevated levels of operon expression are observed during

nutritional shift up or auxotroph starvation experiments. The sites and mechanisms of action of growth rate control and end-product repression remain uncertain.

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