

## Fruiting Body Formation of *Cordyceps militaris* from Multi-Ascospore Isolates and Their Single Ascospore Progeny Strains

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Interest in commercial cultivation and product development of *Cordyceps* species has shown a recent increase. Due to its biochemical and pharmacological effects, *Cordyceps militaris*, commonly known as orange caterpillar fungus, is being investigated with great interest. Cultivation of *C. militaris* has been practiced on a large scale in order to fulfill a demand for scientific investigation and product development. Isolates of *C. militaris* can be easily established from both spores and tissue. For isolation of spores, ascospores released from mature stromata are trapped in sterile medium. Multi-ascospore isolates, as well as combinations of single ascospore strains, are used for production of fruiting bodies. Progeny ascospore strains can be isolated from artificial fruiting bodies, thus, the cycle of fruiting body production can be continued for a long period of time. In this study, we examined fruiting body production from multi-ascospore isolates and their progeny strains for three generations. F<sub>1</sub> progeny strains generally produced a larger number of fruiting bodies, compared with their mother multi-ascospore isolates; however, F<sub>2</sub> and F<sub>3</sub> progeny strains produced fewer fruiting bodies. Optimum preservation conditions could help to increase the vitality of the progeny strains. In order to retain the fruiting ability of the strains, further testing of various methods of preservation and different methods for isolation should be performed.

**KEYWORDS** : Biological efficiency, *Cordyceps militaris*, Multi-ascospore isolate, Progeny strains, Single ascospore isolation

### Introduction

*Cordyceps* species, which are distributed worldwide, are regarded as medicinal herbs in oriental society of Asia, including Korea. *Ophiocordyceps sinensis* (syn. *Cordyceps sinensis*) is a highly regarded medicinal herb, not only among *Cordyceps* species, but also among mushrooms in general [1]. It grows in alpine grasslands of Asia and has significant socio-economic impact in rural areas [2-5]. Due to their similar biochemical compositions and pharmacological properties, *Cordyceps militaris* has recently been viewed as a substitute for *O. sinensis* [6-9]. Many experimental studies for *in vitro* stroma production of *C. militaris* had been conducted [10-14]. Similarly, different insect larvae and pupae have been used for study of the infection process and stromata formation of *C. militaris* [15-18]. Alternatively, brown rice has been used for mass cultivation of *C. militaris* [19, 20].

Isolates derived from spores or tissues are used for establishment of culture and production of fruiting bodies. Among spore isolates, both ascospores and conidia are

frequently used. However, for genetic analysis, single ascospore or single conidial isolates are prerequisites. Comparative studies of fruiting body production from multi-ascospore isolates and progeny strains are lacking in *C. militaris*. Therefore, in this study, we attempted to compare fruiting bodies produced from multi-ascospore isolates, and their progeny strains, as well as from single conidial strains. Findings of the study demonstrated that single ascospore progeny strains isolated from *in vitro* stromata are equally capable of fruiting body production. Single conidial strains isolated from multi-ascospore isolates were also proven as an alternative to single ascospore strains.

### Materials and Methods

**Fungal isolates of *C. militaris* and their fruiting body formation.** Multi-ascospore isolates were derived from all 113 wild specimens of *C. militaris* collected on different mountains in Korea, following the method of Sung *et al.* [21] (Table 1). The isolates were grown on

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**Table 1.** *Cordyceps militaris* specimens used in the experiment

Collection site	No. of specimens	Date of collection
Odae Mt., Gangwon-do	3	Jul 2, 2004
Galchun, Gangwon-do	3	Jul 3, 2004
Chilgab Mt., Chungcheong-do	34	Jul 16, 2004
Songni Mt., Chungcheong-do	4	Jul 17, 2004
Sirang Mt., Gangwon-do	34	Jul 21, 2004
Halla Mt., Jeju-do	24	Jul 27, 2004
Undu Mt., Gangwon-do	1	Aug 12, 2004
Bongmyeong-ri, Gangwon-do	10	Aug 23, 2004
Total	113	

Sabouraud dextrose agar plus yeast extract (SDAY; dextrose 20 g, yeast extract 5 g, peptone 5 g, and agar 15 g per 1,000 mL; pH 5.6) medium at  $24 \pm 1^\circ\text{C}$  for 3 wk for use in the experiment. The specimens have been preserved in the Cordyceps Research Institute (CRI), Mushtech, Korea. For induction of fruiting body formation, all 113 isolates were inoculated in brown rice medium, following the method of Shrestha *et al.* [22]. Brown rice medium was prepared by mixing 50 g of brown rice and 10 g of silkworm pupae in 70 mL of distilled water in 1,000 mL polypropylene (PP) bottles and was then sterilized. After 60 days of culture in brown rice medium, stroma length (SL) of fresh fruiting bodies was measured in mm and biological efficiency (BE) of fruiting bodies was calculated as a percentage (%). For calculation of BE, measurement of dry wt. (DW) of fruiting bodies from each PP bottle was performed by drying them at  $60^\circ\text{C}$  for 24 hr. BE (%) was calculated according to the formula: [DW of fruiting body (g)/substrate wt. (60 g)]  $\times$  100. Ten specimens, C-11408, C-11445, C-11821, C-11876, C-11894, C-11913, C-12086, C-12167, C-12434, and C-12448, were selected for calculation of correlation co-efficient ( $r$ ) between herbarium size of *C. militaris* and artificial fruiting bodies produced from their isolates, following <http://easycalculation.com/statistics/correlation.php>.

#### Fruiting body formation from single ascospore strains.

Fruiting bodies produced by the isolates CRI C-12448, CRI C-11913, CRI C-12434, CRI C-11894, and CRI C-12086 were used for isolation of progeny strains. Following the method of Shrestha *et al.* [23], six single ascospores were isolated from fruiting bodies produced by each isolate and were crossed among themselves in brown rice medium. Single ascospore strains derived from each isolate were numbered from 1 to 6. For example, CRI C-12448 1, CRI C-12448 2, CRI C-12448 3, CRI C-12448 4, CRI C-12448 5, and CRI C-12448 6 were derived from the isolate CRI C-12448. The single ascospore strains were designated as  $F_1$  progeny strains. Calculation of BE of the fruiting bodies of each crossing was performed as described above.

Among the crosses, fruiting bodies produced by the crossing CRI C-12448 4  $\times$  CRI C-12448 6 (denoted in short as CRI C-12448 4  $\times$  6) were utilized again for further isolation of progeny strains. Six single ascospore strains isolated from that crossing were numbered as CRI C-12448 (4  $\times$  6) 1, CRI C-12448 (4  $\times$  6) 2, CRI C-12448 (4  $\times$  6) 3, CRI C-12448 (4  $\times$  6) 4, CRI C-12448 (4  $\times$  6) 5, and CRI C-12448 (4  $\times$  6) 6. They were designated as  $F_2$  progeny strains and crossed among themselves for fruiting body formation. BE was calculated, as described above. Six single ascospores were again isolated from the crossing CRI C-12448 (4  $\times$  6) 3  $\times$  4 and numbered from CRI C-12448 (4  $\times$  6) (3  $\times$  4) 1 to CRI C-12448 (4  $\times$  6) (3  $\times$  4) 6. The strains were designated as  $F_3$  progeny strains and were crossed among themselves; BE was then calculated. In all of the crosses, perithecial stroma formation was indicated as (+) and non-perithecial stroma or no stroma formation as (-).

#### Fruiting body production from single-conidial strains.

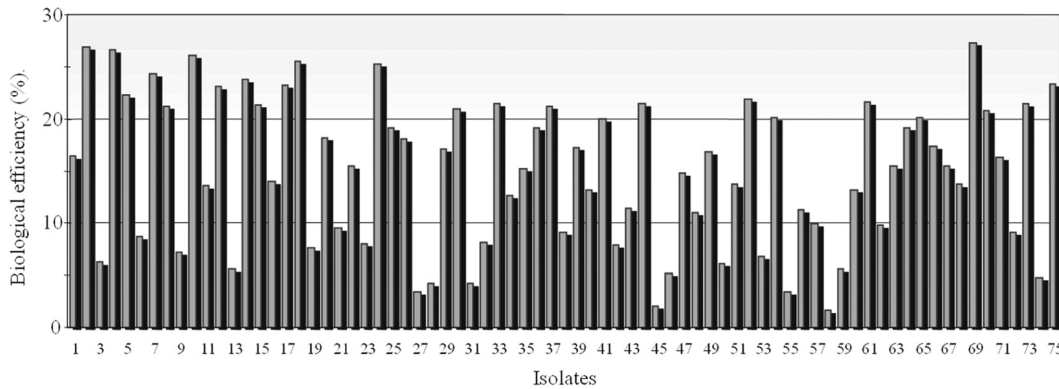
The dilution method was used for isolation of single conidial strains from the isolates CRI C-12448 and CRI C-12086. The strains were crossed among themselves in brown rice medium and calculation for BE was performed. Six single conidial strains were also isolated from each single ascospore strain, CRI C-12448 4 and CRI C-12448 6, and crossed among themselves. In addition, crossings between single conidial strains isolated from strains CRI C-12448-4 and CRI C-12448-6 were also made, and calculation for BE was performed.

## Results and Discussion

#### Fruiting body formation from multi-ascospore isolates.

Of the 113 multi-ascospore isolates of *C. militaris*, only 75 isolates produced fruiting bodies (Fig. 1). The remaining isolates did not produce fruiting bodies on brown rice medium. Multi-ascospore isolates of *C. militaris* produce unstable fruiting bodies [22]. Multi-ascospore isolates are not favorable for stable and uniform fruiting body formation; however, they can be used for testing of fruiting ability and selection for superior isolates. In this study, we observed variation in fruiting body formation by different isolates of *C. militaris*. Observation of the positive correlation between herbarium specimens and their respective artificial fruiting bodies was of particular interest (Table 2, Figs. 2~4). Correlation co-efficient between SL of herbarium specimens and SL of artificial fruiting bodies, and between SL of herbarium specimens and BE of fruiting bodies was 0.6711 and 0.6253, respectively (Table 2).

**Fruiting body formation from crossing of  $F_1$  progeny strains.** Among the 113 isolates, CRI C-12448 produced the highest BE, followed by CRI C-11408 and CRI C-



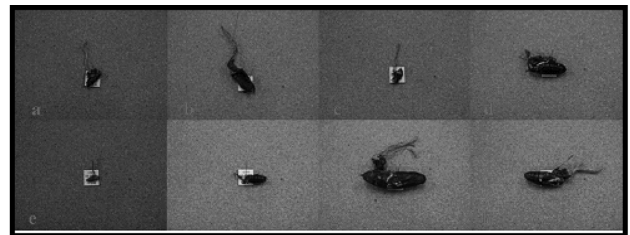
**Fig. 1.** Biological efficiency of fruiting bodies produced by 75 isolates of *Cordyceps militaris*. 1, CRI C-11255; 2, CRI C-11408; 3, CRI C-11444; 4, CRI C-11445; 5, CRI C-11559; 6, CRI C-11732; 7, CRI C-11734; 8, CRI C-11738; 9, CRI C-11740; 10, CRI C-11741; 11, CRI C-11743; 12, CRI C-11744; 13, CRI C-11745; 14, CRI C-11748; 15, CRI C-11818; 16, CRI C-11819; 17, CRI C-11820; 18, CRI C-11821; 19, CRI C-11822; 20, CRI C-11823; 21, CRI C-11825; 22, CRI C-11826; 23, CRI C-11828; 24, CRI C-11829; 25, CRI C-11831; 26, CRI C-11832; 27, CRI C-11834; 28, CRI C-11841; 29, CRI C-11845; 30, CRI C-11876; 31, CRI C-11892; 32, CRI C-11893; 33, CRI C-11894; 34, CRI C-11896; 35, CRI C-11899; 36, CRI C-11901; 37, CRI C-11902; 38, CRI C-11904; 39, CRI C-11905; 40, CRI C-11906; 41, CRI C-11907; 42, CRI C-11908; 43, CRI C-11909; 44, CRI C-11913; 45, CRI C-12019; 46, CRI C-12023; 47, CRI C-12025; 48, CRI C-12027; 49, CRI C-12028; 50, CRI C-12084; 51, CRI C-12086; 52, CRI C-12167; 53, CRI C-12169; 54, CRI C-12170; 55, CRI C-12171; 56, CRI C-12171; 57, CRI C-12176; 58, CRI C-12292; 59, CRI C-12293; 60, CRI C-12294; 61, CRI C-12295; 62, CRI C-12296; 63, CRI C-12297; 64, CRI C-12304; 65, CRI C-12434; 66, CRI C-12443; 67, CRI C-12445; 68, CRI C-12446; 69, CRI C-12448; 70, CRI C-12449; 71, CRI C-12450; 72, CRI C-12451; 73, CRI C-12452; 74, CRI C-12453; 75, CRI C-12454. CRI, Cordyceps Research Institute.

**Table 2.** Correlation co-efficient between herbarium specimens of *Cordyceps militaris* and their artificial fruiting bodies

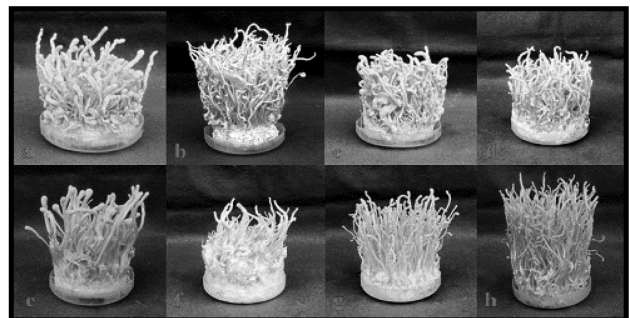
Herbarium specimen		Artificial fruiting bodies	
No.	SL	SL	BE
C-12448	39	106	27.25
C-11408	31	60	26.90
C-11445	51	103	26.57
C-11821	30	87	25.48
C-12167	25	85	21.81
C-11913	22	93	21.47
C-11876	29	77	20.84
C-12434	38	94	20.07
C-11894	35	81	19.41
C-12086	16	49	13.68
<i>r</i>		0.6711 <sup>a</sup>	0.6253 <sup>b</sup>

SL, stroma length; BE, biological efficiency; *r*, correlation co-efficient. <sup>a</sup>*r* between stroma lengths of herbarium specimens and artificial fruiting bodies. <sup>b</sup>*r* between stroma lengths of herbarium specimens and BE of artificial fruiting bodies.

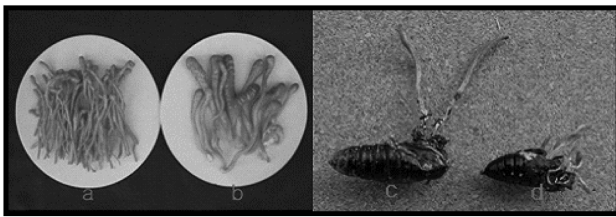
11445 (Table 2, Fig. 1). In this study, F<sub>1</sub> progeny strains were isolated from isolates that differed in fruiting ability based on BE (Table 2). Among F<sub>1</sub> progeny strains of the isolate CRI C-12448, strains 1, 2, 4, and 5 and strains 3 and 6 were opposite in mating type (Table 3, Fig. 5). Similarly, strains 1, 3, 5, and 6 of the isolate CRI C-11913 were opposite in mating type to strains 2 and 4 (Table 4, Fig. 6). In the case of the isolate CRI C-12434, strains 1,



**Fig. 2.** Herbarium specimens of *Cordyceps militaris*. a, CRI C-11408; b, CRI C-11445; c, CRI C-11821; d, CRI C-11876; e, CRI C-11913; f, CRI C-12167; g, CRI C-12434; h, CRI C-12448. CRI, Cordyceps Research Institute.



**Fig. 3.** Fruiting bodies produced by *Cordyceps militaris* isolates. a, CRI C-11408; b, CRI C-11445; c, CRI C-11821; d, CRI C-11876; e, CRI C-11913; f, CRI C-12167; g, CRI C-12434; h, CRI C-12448. CRI, Cordyceps Research Institute.



**Fig. 4.** Fruiting bodies and herbarium specimens of *Cordyceps militaris*. a, c, CRI C-11894; b, d, CRI C-12086. CRI, Cordyceps Research Institute.

**Table 3.** Stromata formation from crossing of F<sub>1</sub> progeny strains of *Cordyceps militaris* CRI C-12448

	1	2	3	4	5	6
1	-	-	+	-	-	+
2		-	+	-	-	+
3			-	+	+	-
4				-	-	+
5					-	+
6						-

CRI, Cordyceps Research Institute.

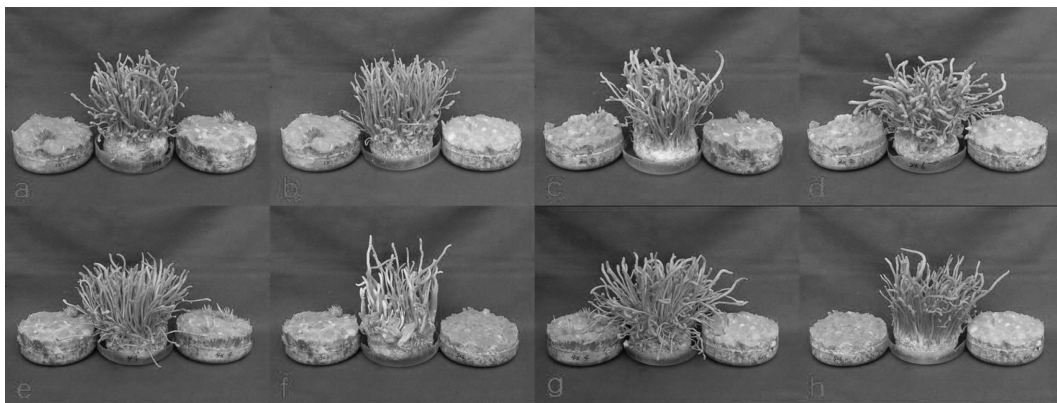
**Table 4.** Stromata formation from crossing of F<sub>1</sub> progeny strains of *Cordyceps militaris* CRI C-11913

	1	2	3	4	5	6
1	-	+	-	+	-	-
2		-	+	-	+	+
3			-	+	-	-
4				-	+	+
5					-	-
6						-

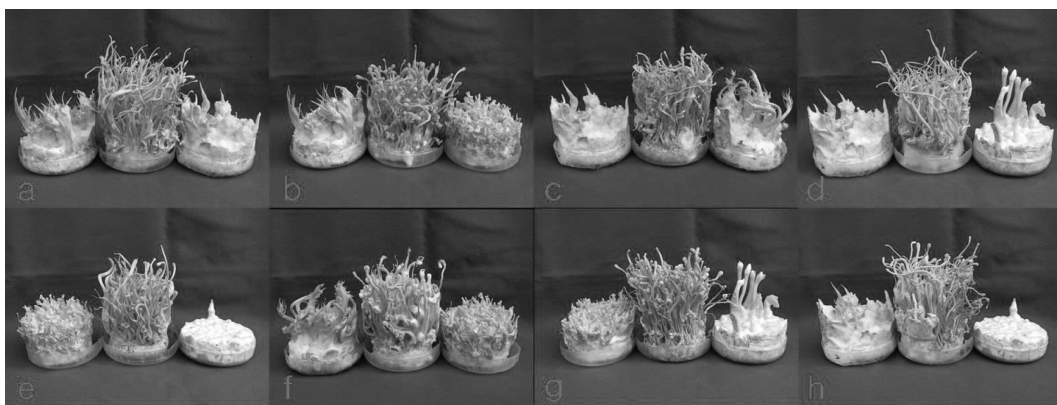
CRI, Cordyceps Research Institute.

4, and 6 and strains 2, 3, and 5 were opposite in mating type (Table 5). In isolate CRI C-11894, strains 1, 2, and 3 were of opposite type to strains 4, 5, and 6 (Table 6). All crossings of CRI C-12086 produced perithecial stromata (Table 7). This was probably due to error in isolation of single ascospore strains or cytological abnormality in the strains. Except for the isolate C-12448, BE of crossings of F<sub>1</sub> progeny strains were better, compared to their respective multi-ascospore isolates (Fig. 7).

*Cordyceps militaris* is a heterothallic fungus and



**Fig. 5.** Fruiting bodies produced from crossings of F<sub>1</sub> progeny strains of *Cordyceps militaris* CRI C-12448. a, 1 × 3; b, 1 × 6; c, 2 × 3; d, 2 × 6; e, 3 × 4; f, 3 × 5; g, 4 × 6; h, 5 × 6. CRI, Cordyceps Research Institute.



**Fig. 6.** Fruiting bodies produced from crossings of F<sub>1</sub> progeny strains of *Cordyceps militaris* CRI C-11913. a, 1 × 2; b, 1 × 4; c, 2 × 3; d, 2 × 5; e, 2 × 6; f, 3 × 4; g, 4 × 5; h, 4 × 6. CRI, Cordyceps Research Institute.

**Table 5.** Stromata formation from crossing of F<sub>1</sub> progeny strains of *Cordyceps militaris* CRI C-12434

	1	2	3	4	5	6
1	-	+	+	-	+	-
2		-	-	+	-	+
3			-	+	-	+
4				-	+	-
5					-	+
6						-

CRI, Cordyceps Research Institute.

**Table 6.** Stromata formation from crossing of F<sub>1</sub> progeny strains of *Cordyceps militaris* CRI C-11894

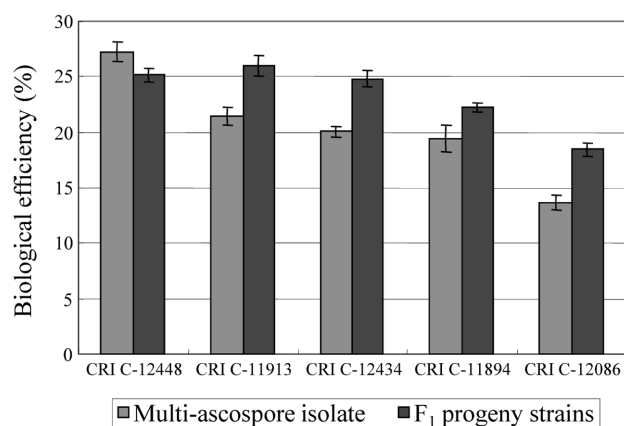
	1	2	3	4	5	6
1	-	-	-	+	+	+
2		-	-	+	+	+
3			-	+	+	+
4				-	-	-
5					-	-
6						-

CRI, Cordyceps Research Institute.

**Table 7.** Stromata formation from crossing of F<sub>1</sub> progeny strains of *Cordyceps militaris* CRI C-12086

	1	2	3	4	5	6
1	+	+	+	+	+	+
2		+	+	+	+	+
3			+	+	+	+
4				+	+	+
5					+	+
6						+

CRI, Cordyceps Research Institute.



**Fig. 7.** Biological efficiency of multi-ascospore isolates and F<sub>1</sub> progeny strains of *Cordyceps militaris*. CRI, Cordyceps Research Institute.

therefore requires a combination of two opposite mating type strains in order to form fruiting bodies [23]. Many studies have followed crossing between single ascospore

**Table 8.** Stromata formation from crossing of F<sub>2</sub> progeny strains of *Cordyceps militaris* CRI C-12448 4 × 6

	1	2	3	4	5	6
1	-	+	-	+	+	+
2		-	+	-	-	-
3			-	+	+	+
4				-	-	-
5					-	-
6						-

CRI, Cordyceps Research Institute.

**Table 9.** Stromata formation from crossing of F<sub>3</sub> progeny strains of *Cordyceps militaris* CRI C-12448 (4 × 6) 3 × 4

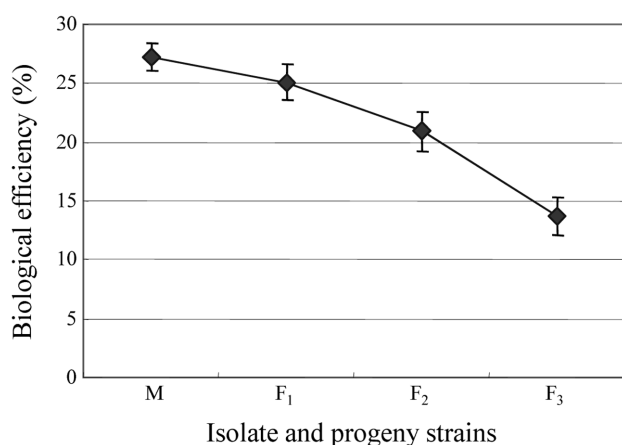
	1	2	3	4	5	6
1	-	+	+	+	+	-
2		-	-	-	-	+
3			-	-	-	+
4				-	-	+
5					-	+
6						-

CRI, Cordyceps Research Institute.

strains in order to produce fruiting bodies of *C. militaris* [24-29]. However, isolation of a single ascospore is a time-consuming process requiring a good laboratory facility and a sterile environment. By contrast, isolation of multiple ascospores is an easy process and can also be performed in the field. Hence, isolation of multiple ascospores is a good option for establishment of culture in the lab. Once the culture has been established, single ascospore progeny strains can be isolated from fruiting bodies produced by multi-ascospore isolates, and, in turn, can be used for production of stable fruiting bodies, as demonstrated by Sung *et al.* [26].

**Fruiting body formation from crossings of F<sub>2</sub> and F<sub>3</sub> single ascospore strains.** Eight combinations of F<sub>2</sub> progeny strains isolated from a combination of CRI C-12448 4 × 6 produced fruiting bodies (Table 8). Similarly, eight combinations of F<sub>3</sub> progeny strains isolated from CRI C-12448 (4 × 6) 3 × 4 produced fruiting bodies (Table 9). On average, isolate CRI C-12448 produced the highest BE, followed by F<sub>1</sub>, F<sub>2</sub>, and F<sub>3</sub> progeny strains (Fig. 8).

**Fruiting body formation from single conidial strains.** Among the single conidial strains derived from the isolate CRI C-12448, strains 1, 3, 5, and 6 were opposite in mating type to remaining strains 2 and 3 (Table 10). Similarly, strains 1, 2, 4, 5, and 6, isolated from CRI C-12086, were opposite in mating type to strain 3 (Table 11). However, BE of combinations of single conidial strains were almost half or less than half of that of the



**Fig. 8.** Biological efficiency of a multi-ascospore isolate and progeny strains of *Cordyceps militaris*. M, multi-ascospore isolate CRI C-12448; F<sub>1</sub>, F<sub>1</sub> progeny strains of CRI C-12448; F<sub>2</sub>, F<sub>2</sub> progeny strains of C-12448 (4 × 6); F<sub>3</sub>, F<sub>3</sub> progeny strains of C-12448 (4 × 6) (3 × 4). CRI, Cordyceps Research Institute.

**Table 10.** Fruiting bodies produced from crossings of single-conidium isolates of *Cordyceps militaris* isolate CRI C-12448

	1	2	3	4	5	6
1	-	+	-	+	-	-
2		-	+	-	+	+
3			-	+	-	-
4				-	+	+
5					-	-
6						-

CRI, Cordyceps Research Institute.

**Table 11.** Fruiting bodies produced from crossings of single-conidium isolates of *Cordyceps militaris* isolate CRI C-12086

	1	2	3	4	5	6
1		-	+	-	-	-
2			+	-	-	-
3				+	+	+
4					-	-
5						-
6						

CRI, Cordyceps Research Institute.

isolate CRI C-12448. Single conidial strains isolated from each F<sub>1</sub> progeny strain, C-12448-4 and C-12448-6, produced no fruiting bodies (Tables 12 and 13) that demonstrated stability of mating type loci among the conidial strains.

Among *Cordyceps* species, *C. militaris* is an ideal species for the study of *in vitro* stromata formation, as demonstrated by its successful formation [11-13]. *Cordyceps militaris* isolates collected from different mountains in Korea have been used for selection of superior strains [24-27]. Wen *et al.* [30] demonstrated variation in biochemical

**Table 12.** Crossings of single conidium isolates derived from F<sub>1</sub> progeny strain of *Cordyceps militaris* CRI C-12448 4

	1	2	3	4	5	6
1	-	-	-	-	-	-
2		-	-	-	-	-
3			-	-	-	-
4				-	-	-
5					-	-
6						-

CRI, Cordyceps Research Institute.

**Table 13.** Crossings of single conidium isolates derived from F<sub>1</sub> progeny strain of *Cordyceps militaris* CRI C-12448 6

	1	2	3	4	5	6
1	-	-	-	-	-	-
2		-	-	-	-	-
3			-	-	-	-
4				-	-	-
5					-	-
6						-

CRI, Cordyceps Research Institute.

composition among different *C. militaris* isolates collected from different sites. This variation provides us an opportunity to select superior strains among the natural isolates.

In most studies, multispore isolates are used for production of fruiting bodies. Since the revelation of heterothallic mating system in *C. militaris*, many studies have involved isolation of single ascospores and crossing between them [23-29]. Results of genetic analysis have also demonstrated that mating type and mycelium pigmentation are independent of one another [24]. Optimum preservation of the superior strains is a persistent problem for continuous cultivation of *C. militaris*. Results of preservation studies conducted at low temperatures have demonstrated successful fruiting body production from successive subcultures of superior strains in *C. militaris* [26]. In addition, findings of this study have demonstrated that successive isolation of single ascospores from *in vitro* stromata is another option for preserving strains with desirable characteristics. In conclusion, subcultures of the strains and isolation of single ascospores and conidia can be applied for large scale cultivation of *C. militaris* and should be used complementarily.

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