One-electron oxidation of an oxoiron(IV) complex to form an [O=Fe^V=NR]⁺ center

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Oxoiron(V) species are postulated to be involved in the mechanisms of the arene cis-dihydroxylating Rieske dioxygenases and of bioinspired nonheme iron catalysts for alkane hydroxylation, olefin cis-dihydroxylation, and water oxidation. In an effort to obtain a synthetic oxoiron(V) complex, we report herein the oneelectron oxidation of the S = 1 complex $[Fe^{IV}(O)(TMC)(NCCH_3)]^{2+}$ (1, where TMC is tetramethylcyclam) by treatment with tert -butyl hydroperoxide and strong base in acetonitrile to generate a metastable $S = \frac{1}{2}$ complex 2 at -44 °C, which has been characterized by UV-visible, resonance Raman, Mössbauer, and EPR methods. The defining spectroscopic characteristic of 2 is the unusual x/y anisotropy observed for the ⁵⁷Fe and ¹⁷O A tensors associated with the high-valent Fe=O unit and for the ¹⁴N A tensor of a ligand derived from acetonitrile. As shown by detailed density functional theory (DFT) calculations, the unusual x/y anisotropy observed can only arise from an iron center with substantially different spin populations in the d_{xz} and d_{yz} orbitals, which cannot correspond to an Fe^{IV}=O unit but is fully consistent with an $S = \frac{1}{2} Fe^{V}$ center, like that found for $[Fe^{V}(O)(TAML)]^{-}$ (where TAML is tetraamido macrocyclic ligand), the only well-characterized oxoiron(V) complex reported. Mass spectral analysis shows that the generation of 2 entails the addition of an oxygen atom to 1 and the loss of one positive charge. Taken together, the spectroscopic data and DFT calculations support the formulation of 2 as an iron(V) complex having axial oxo and acetylimido ligands, namely $[Fe^{V}(O)(TMC)(NC(O)CH_{3})]^{+}$.

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ormally oxoiron(V) oxidants are postulated in the catalytic cycles of several iron enzymes that carry out difficult oxidations. Most prominent of these are cytochromes P450, which can hydroxylate strong C–H bonds (1-3), even those of methane (1, 2). Recent evidence has demonstrated that the active oxidant can be best described as having an oxoiron(IV) unit supported by a porphyrin cation radical (4). On the other hand, the Rieske dioxygenases activate O₂ at an active site with a 2-His-1-carboxylate facial triad motif to effect the cis-dihydroxylation of C=C bonds in the biodegradation of aromatic complexes (5). For these nonheme iron enzymes, the proposed $Fe^{\hat{V}}=O$ oxidant is as yet unobserved. However, unlike the porphyrin ligand in cytochrome P450, none of the ligands in the nonheme iron active site appears likely to undergo one-electron oxidation to stabilize the high-valent state. Fe^V=O oxidants are also implicated in alkane hydroxylation (6, 7), olefin epoxidation and *cis*-dihydroxylation (8–10), and water oxidation (11) by bioinspired nonheme iron catalysts supported by neutral tetradentate ligands, and direct evidence for the formation of oxoiron(V) oxidants has been obtained in two cases by mass spectrometry (10, 12). Despite the wealth of synthetic oxoiron(IV) complexes identified during the last decade (13), to date there is only one spectroscopically well-characterized example of an oxoiron(V) complex, $[Fe^{V}(O)(TAML)]^{-1}$ (where TAML is tetraamido macrocyclic ligand), which is stabilized by the tetraanionic nature of the TAML ligand (14). Given the ligand environments of the Rieske dioxygenases and the synthetic iron catalysts, we have been investigating the feasibility of generating an oxoiron(V) complex supported by a polydentate ligand of lower charge. Toward this end, we have focused on the one-electron oxidation of the oxoiron(IV) complex $[Fe^{IV}(O) (TMC)(NCCH_3)]^{2+}$ (1, see Fig. 1), where the oxoiron(IV) moiety is supported by the neutral macrocyclic tetramethylcyclam (TMC) ligand (15). We chose complex 1 as a starting point for this effort because it is structurally well-characterized and exhibits good thermal stability. One-electron oxidation of 1 could be effected by the addition of *tert*-butyl hydroperoxide and strong base, and our observations are reported herein.

Results and Discussion

The reaction of 1 in CH₃CN at -44 °C with at least three equivalents ^tBuOOH followed by the addition of 1-2 equivalents strong base (KO^tBu or NBu₄OH) generates an orange complex (2) that exhibits distinct UV-visible features at 410 nm ($\varepsilon =$ 4,000 M⁻¹ cm⁻¹) and 780 nm ($\varepsilon = 430$ M⁻¹ cm⁻¹) (Fig. 24). Subsequent addition of five equivalents strong acid, e.g., HBF₄ or HClO₄, produces a green species (2-H⁺) associated with spectral features at 425 nm ($\varepsilon = 4,100 \text{ M}^{-1} \text{ cm}^{-1}$), 600 nm ($\varepsilon =$ 680 M⁻¹ cm⁻¹), and 750 nm ($\varepsilon = 530$ M⁻¹ cm⁻¹) (Fig. 2A). (The molar extinction coefficients shown are based on samples for which Mössbauer quantification of the relevant chromophores has been carried out.) Reaction of $2-H^+$ with an excess of strong base readily regenerates 2, indicating that 2 and $2-H^+$ comprise a conjugate acid/base pair. These complexes are generated in approximately 50% yield relative to 1, as determined by Mössbauer spectroscopy (vide infra). Unlike 1, which is stable in MeCN at -44 °C, 2 and 2-H⁺ decay with $t_{1/2} = 60$ and 30 min, respectively. Notably, both 2 and 2-H⁺ are $S = \frac{1}{2}$ species that exhibit very similar rhombic EPR spectra (Fig. 2A, Inset) with $g_{\text{ave}} \sim 2.0$, suggesting that the S = 1 species 1 has undergone oneelectron oxidation to form 2.

The resonance Raman (rR) spectra of 2 and 2-H⁺ obtained with 413-nm excitation show features associated with an Fe=O unit. Complex 2 exhibits an intense vibrational feature at 798 cm⁻¹ with an associated overtone at 1,587 cm⁻¹ (Fig. 2*B*); corresponding features for 2-H⁺ are observed at 811 and 1,613 cm⁻¹. All four features downshift when ¹⁸O-labeled 1 is used as the precursor; the observed downshifts are in accord with values calculated for an Fe=O diatomic oscillator using Hooke's law (e.g., $\Delta \nu = -35$ and -36 cm⁻¹, respectively). The

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Fig. 1. Proposed steps in the one-electron oxidation of 1.

observed vibrational frequencies of 798 and 811 cm⁻¹ fall at the low end of the range of $\nu_{Fe=O}$ values previously observed for **1** (839 cm⁻¹) and its axial-ligand-substituted derivatives $[Fe^{IV}(O)$ $(TMC)(N_3)]^+$ (814 cm⁻¹) and $[Fe^{IV}(O)(TMC)(CF_3CO_2)]^+$ (854 cm⁻¹) (16). Thus the lower $\nu_{Fe=O}$'s of **2** and **2-H**⁺ suggest the presence of a highly basic *trans* anionic ligand that weakens the Fe=O bond.

To obtain further insight into the nature of the $S = \frac{1}{2}$ species, we have studied samples of **2** and **2-H**⁺ with Mössbauer spectroscopy. In a typical sample, **2** and **2-H**⁺ represented 40–55% of the total Fe in a sample. Fig. 3*A* shows a 4.2 K Mössbauer spectrum of **2** recorded in a 50 mT field applied parallel to the observed γ rays. In addition to **2**, which accounts for approximately 55% of Fe, the sample contained several S = 1 Fe^{IV} species yielding quadrupole doublets for **1** (green, $\Delta E_Q = 1.23$ mm/s, $\delta =$ 0.17 mm/s, approximately 8%), [Fe^{IV}(O)(TMC)(OH)]⁺ (**1-OH**: blue, $\Delta E_Q = 0.16$ mm/s, $\delta = 0.15$ mm/s, 16%), and an unidentified Fe^{IV} species ($\Delta E_Q \sim 0.6$ mm/s, $\delta \sim 0.15$ mm/s, $\approx 10\%$). There is also a high-spin Fe^{III} species ($g \sim 6.9, 5.1, \approx 10\%$), which,



Fig. 2. Spectroscopic data for 2 and 2-H⁺. (*A*) UV-visible spectra of 1 (black dashed line), 2 (thick red line), and 2-H⁺ (blue solid line) collected at -44 °C. (*Inset*) EPR spectra of 2 in CD₃CN and 2-H⁺ in 1:9 CH₃CN: CH₂Cl₂. (*B*) Resonance Raman spectra of 2 (*Left*) and 2-H⁺ (*Right*) collected at 77 K using 413-nm laser excitation for frozen solution samples prepared with ¹⁶O (black line) and ¹⁸O (red line). Acetonitrile solvent peaks are marked as "s".

fortunately, absorbs mostly outside the spectral region of 2. Similarly, the sample of 2-H⁺ studied here contained the following species: $S = \frac{1}{2}$ complex 2-H⁺ (41%), two S = 1 complexes 1 (27%) and 1-OH (2%), and S = 5/2 Fe^{III} (30%, see *SI Appendix*).

The presence of multiple contaminants in the sample for **2** complicates the analysis of the $S = \frac{1}{2}$ species of interest. However, two Mössbauer attributes allow us to uniquely associate a spectral component with the $S = \frac{1}{2}$ species, and to eliminate the S = 1 and $S = \frac{5}{2}$ contaminants. First, we can record spectra by applying a weak (e.g., 50 mT) magnetic field either parallel or perpendicular to the observed γ rays. If the intensities of the absorption lines are different between parallel and perpendicular field, the species must be associated with an EPR active species; in this way a unique association between an $S = \frac{1}{2}$ EPR signal and a Mössbauer spectral component is established. Second, for a difference spectrum thus taken, all contributions from species yielding quadrupole doublets cancel, leaving us with the analysis of a pure $S = \frac{1}{2}$ paramagnetic species. This procedure yielded the difference spectrum of Fig. 3*B*, which represents **2**.

To simulate the Mössbauer and EPR spectra, we used the $S = \frac{1}{2}$ spin Hamiltonian, as follows:

$$\mathcal{H} = \mathbf{g}\beta \mathbf{B} \bullet \mathbf{S} + \mathbf{S} \bullet \mathbf{A}({}^{\mathsf{S}}{}^{\mathsf{T}}\mathbf{F}\mathbf{e}) \bullet \mathbf{I} - g_{n}\beta_{n}\mathbf{B} \bullet \mathbf{I} + \mathcal{H}_{Q} + \Sigma_{n}\mathbf{S} \bullet \mathbf{A}(n) \bullet \mathbf{I}(n),$$
^[1]



Fig. 3. A 4.2 K Mössbauer spectra of **2** and **2**-H⁺. Mössbauer spectra of samples containing **2** (*A*) and **2**-H⁺ (*C*) recorded in a 50 mT field applied parallel to the observed γ rays. (*B*) Difference spectrum parallel minus perpendicular, representing **2**, obtained with 50 mT applied fields. Red lines are simulations for **2** and **2**-H⁺ based on Eq. 1, using the parameters listed below and in Table 1. The major Fe^{TV}=O contaminants are shown by the green (1) and blue (1-OH) lines. The black solid line in *A* is a spectral simulation for the sum of **2** (55% of Fe), **1** (8%), and 1-OH (16%). The doublets in C represent 27% of **1** and 2% of 1-OH. The arrows in C point to absorption due to a high-spin Fe^{III} contaminant (30%). The δ , ΔE_Q , and η values used in the simulations are +0.10(4) mm/s, -0.5 mm/s, and -3, respectively, for **2** and +0.10(4) mm/s, -0.2 mm/s, and -3, respectively.

Table 1. g- and A-tensors (in megahertz) experimentally observed for 2, 2-H⁺, and [Fe^V(O)(TAML)]¹⁻ and calculated for 2 and 2-H⁺

Compound	g			A (⁵⁷ Fe)			A (¹⁷ O)			A (¹⁴ N)		
	x	у	z	x	У	z	x	У	z	x	у	z
HRP Cpd I	_	_	-	-26*	-26*	-8*	35 [†]	36 [†]	ND	_	_	-
1	-	_	-	-28 [‡]	-28 [‡]	-4	–27§	–27§	+61 [§]	-	-	_
2 ¹	2.053	2.010	1.971	-47(2)	-17(2)	0(5)	25(15)	128(3)	≈20	29(2)	11(3)	11(3)
2 (DFT)	2.03	2.00	1.97	-43.5	-18.5	3.3	-1.0	-70.8	75.7	-39.3	10.8	6.1
2-H ⁺¹	2.054	2.011	1.975	-47(2)	-17(2)	0(5)	25(15)**	133(3)	23(10)	30(2)**	11(3)	10(3)
2-H ⁺ (DFT) [∥]	2.03	2.00	1.97	-45.1	-14.0	-6.0	18.4	-91.4	84.7	-35.5 [∥]	-1.2	3.9
$[Fe^V(O)(TAML)]^{1-tt}$	1.99	1.97	1.74	-49.3	-1.5	-16.3**	ND	ND	ND	-	-	-

ND, not determined.

*From Mössbauer data analyzed with an S = 1 Hamiltonian (34).

[†]From ENDOR data, S = 1 (19).

[†]Refit of Mössbauer data reported in ref. 15 using g values and zero-field splitting parameters derived from more recent high-field EPR data reported in ref. 35.

[§]This work; B3LYP calculations yield a good match for $A(^{57}Fe)$.

¹This work; data analyzed with Eq. **1**; 4.2 K Mössbauer spectra recorded at 2.0, 4.0, and 8.0 T show that $A_x(^{57}Fe)$ and $A_y(^{57}Fe)$ of **2** and **2**-H⁺ are negative, see *SI Appendix*, Fig. S4. The *z* axis is along the Fe–O bond in all systems. Results for **2** apply to the majority species.

Values for the $2(Fe^V)$ model obtained from DFT calculations using BP86 functionals (see below).

**Relative to g, $A(^{17}O)$ and $A(^{14}N)$ are rotated 8° around the z axis of the g-tensor.

^{††}From ref. 14.

^{**}Correcting A_z for the orbital contribution, which is proportional to $(g_z - 2)$, would yield $A_z \sim 0$.

where $A({}^{57}Fe)$ is the ${}^{57}Fe$ magnetic hyperfine tensor, I is the ${}^{57}Fe$ nuclear spin operator, and \mathscr{H}_Q describes the ${}^{57}Fe$ quadrupole interaction. As needed, the last term will be used to describe ligand hyperfine interactions observed by EPR; $n = {}^{14}N$, ${}^{15}N$, or ${}^{17}O$. From extensive simulations of the spectra of 2 and 2-H⁺, we obtained the parameters listed in Table 1 and *SI Appendix*, Fig. S6. Both 2 and 2-H⁺ exhibit the same isomer shifts within the uncertainties, namely $\delta = +0.10(4)$ mm/s. The δ values are smaller than those reported for S = 1 Fe^{IV}(O)(TMC) complexes (range 0.15–0.19 mm/s) (16), suggesting that the oxidation state of the iron site in 2 and 2-H⁺ is above Fe^{IV}. More importantly, 2 and 2-H⁺ exhibit unusual ${}^{57}Fe$ A tensors with large x/y anisotropy, suggesting an electronic structure that differs significantly from that of 1.

We have conducted extensive EPR studies of 2 and 2-H⁺. The use of glassing solvent mixtures (1:3 CH₃CN:CH₂Cl₂ or 1:3 CH₃CN:butyronitrile) sharpened the spectra considerably (4 G linewidth). The higher resolution provided decisive clues about the ligand structure of 2 and $2-H^+$, as well as important magnetic hyperfine structure data. Selected spectra are displayed in Fig. 4. We found that **2** reproducibly displayed two $S = \frac{1}{2}$ species in essentially constant proportions (ca. 2:1 in all preparations studied); the minority species is recognized by a shoulder on the high-field feature in Fig. 4A. Our studies revealed that these two species did not differ in any significant way. The minor species, 2^m , has g-values at 2.05, 1.99, and 1.96. As $2-H^+$ was found to be free of EPR-active contaminants in the g = 2.0region, we focus here on this species. (EPR analysis of 2, summarized in Table 1 and detailed in the SI Appendix, afforded similar conclusions.) For **2-H**⁺ (Fig. 4*B*), the signal at $g_x = 2.054$ exhibits a 1:1:1 hyperfine triplet, corresponding to a single ¹⁴N nucleus with $|A_x({}^{14}N)| = 28.5$ MHz and $|A_{y,z}({}^{14}N)| \approx 11$ MHz (the signs of $A_{x,y,z}$ cannot be determined by EPR). A nearly identical pattern can be discerned in the spectrum of 2 shown in Fig. 4A. This splitting pattern changed to a 1:1 doublet with $|A_x({}^{15}N)| =$ 40 MHz (Fig. 4C) when $2-H^+$ was prepared using 1:3 ¹⁵NCCH₃: CH₂Cl₂ as solvent, unequivocally demonstrating that the observed hyperfine structure arises from a nitrogen atom derived from acetonitrile. This procedure yielded the parameters listed in Table 1.

Species 2-H⁺ obtained from ¹⁷O-labeled 1 gave rise to a spectrum for which the (middle) $g = g_y = 2.01$ feature was split to afford satellite features at both low and high field (Fig. 4*E*); these features arise from ¹⁷O superhyperfine splitting with $|A_y(^{17}O)| =$ 133 MHz. We have studied two preparations, with ¹⁷O enrichment of 10% and 30%; a spectrum of the latter shown in Fig. 4*E*. To analyze this spectrum we have taken the results of the fit of Fig. 4*B* and added an ¹⁷O hyperfine tensor. After obtaining a sufficient understanding of the splitting pattern, we have fitted the spectrum, visually focusing on the resolved two high-field peaks and the low-field feature. The simulation of Fig. 4*F* (plotted for 100% ¹⁷O enrichment) indicates the complexity of the spectrum, which stems to a major extent from the fact that the ¹⁷O splitting along *y* does not belong to a unique direction and that the splitting covers the entire Δg range. The final fit, obtained after exploring tensor rotations, gives a good value for A_y (¹⁷O) and an upper limit for A_z (¹⁷O). Although the calculated intensity for the low field feature is a bit high, the fit could be improved by increasing A_x (¹⁷O) together with a distribution of that parameter.

Lastly, the ⁵⁷Fe-enriched sample (Fig. 4*D*) exhibits significant broadening of the g_x feature corresponding to $|A_x({}^{57}Fe)| =$ 40 MHz, a value in good agreement with the largest A-tensor component of -47 MHz observed in the Mössbauer spectra of **2** and **2-H**⁺ (EPR can only give the magnitude of the A-values); our EPR spectral simulations show that A_y and A_z are at least a factor of two smaller than A_x . Most importantly, the EPR spectra



of the ⁵⁷Fe-enriched sample establish that the largest ⁵⁷Fe A-tensor component is along *x*, the same direction for which the largest ¹⁴N splitting is observed. It should be noted that the EPR spectra establish a spatial correlation between the dominant components of the ⁵⁷Fe, ¹⁷O, and ¹⁴N hyperfine tensors that provides insight into the electronic structure of **2** and **2-H**⁺.

Our accumulated spectroscopic data suggest that 2 and 2-H⁺ are one-electron oxidized derivatives of 1 and consist of a (TMC)Fe=O unit with an axial ligand derived from acetonitrile. In Fig. 1, we propose that 2 is formed by the attack of the *t*-butylperoxide anion on the bound acetonitrile to form a peroxyimidic ester intermediate (17) that then undergoes homolytic cleavage of the O-O bond. To verify that homolysis occurs in this instance, we replaced 'BuOOH with 2-methyl-1-phenylpropan-2vl hydroperoxide (MPPH), a useful mechanistic probe to distinguish between homolytic and heterolytic O-O bond cleavage (18). Although substitution of MPPH for ^tBuOOH did not diminish the yield of 2, product analysis by GC showed quantitative conversion of MPPH to benzaldehyde. This result demonstrates that O-O bond homolysis occurs to generate 2. The formulation of 2 as shown in Fig. 1 is supported by low temperature electrospray ionization (ESI)-MS studies of 2 that reveal a feature at m/z 385.2, consistent with the formulation of 2 as [Fe(O)] $(TMC)(NCCH_3) + O]^+$ (SI Appendix, Figs. S8 and S9). This feature shifts to m/z 388.2 upon preparation of 2 in CD₃CN and disappears after the sample is allowed to decay. We postulate in Fig. 1 two possible limiting electronic descriptions for 2: an oxoiron(IV) unit with an axial acetaminyl radical ligand $(2(Fe^{IV}N\bullet))$ at one end, and an iron(V) center with *trans* oxo and acetylimido ligands $(2(Fe^V))$ at the other end. Our efforts to refine electronic descriptions for 2 and 2-H⁺ more precisely are discussed below.

The unusual A-tensor anisotropies in the *xy* plane listed in Table 1 for **2** and **2**-**H**⁺ indicate that their electronic structures must be significantly different from that of **1**. The ⁵⁷Fe and ¹⁷O tensors of **1** as well as those of horseradish peroxidase compound I, an S = 1 Fe^{IV}=O species antiferromagnetically coupled to a porphyrin radical (19), are axial with respect to the Fe=O bond, the *z* axis in our coordinate frames. The axial anisotropy observed reflects the $(d_{xz})^1(d_{yz})^1$ electronic configuration associated with the S = 1 Fe^{IV}=O unit. In contrast, **2** and **2**-**H**⁺ have drastically different A-tensor components along *x* and *y*, indicating substantially different unpaired spin densities on the d_{xz} and d_{yz} orbitals. This distribution can be achieved by transferring the electron from d_{xz} to the p_x orbital of the aminyl ligand to create an Fe^V=O center with a *trans* imido ligand, a notion supported by the similar ⁵⁷Fe A-tensor anisotropy observed for the bona fide Fe^V=O complex [Fe^V(O)(TAML)]⁻ (Table 1) (14).

Density functional theory (DFT) studies of 2 and 2-H⁺ lend substantial support to the above conclusion (see SI Appendix, Tables S1–S7 for full computational details). For comparison, DFT calculations were also carried out for the experimentally characterized complexes 1 and $[Fe^{V}(O)(TAML)]^{-1}$ as well as the hypothetical iron(V) species 1_{ox} (a computational model obtained by removing one electron from 1; italics denote purely computational models). The use of the hybrid functional B3LYP led to the $2(Fe^{IV}N\bullet)$ description, which is similar to that reported for HRP Cpd I, but with the radical trans rather than cis to the oxo group. This Fe^{IV}-aminyl radical description with an iron $(d_{xz})^{1}(d_{vz})^{1}$ electron configuration predicts 2 and 2-H⁺ to have essentially axial 57 Fe and 17 O A-tensors, inconsistent with the experimental data. In stark contrast, the use of the pure functional BP86 generated the $2(Fe^V)$ limiting description with an iron $(d_{yz})^1(d_{xz})^0$ electron configuration and yielded solutions that reproduce the observed anisotropies of the ⁵⁷Fe A-tensor and rationalize the essential features of the entire dataset. In this model, the acetaminyl radical becomes an acetylimido ligand, and 2 and 2-H⁺ are best formulated as complexes having $[O=Fe^{V}=NR]^{+}$ units (see Fig. 1). The dependence of the calculated ground state on whether a hybrid (B3LYP) or pure (BP86) functional is used may be related to the propensity of Hartree-Fock exchange to stabilize high spin states in the case of the former functional. A similar dependence on the type of functionals used has been reported for the description of electronic structures of {FeNO}⁷ and {FeO₂}⁸ complexes, systems involving a noninnocent ligand radical (see for example refs. 20 and 21). In all cases, the functional chosen for further investigation most closely reproduced the experimentally observed spectroscopic parameters to generate an experimentally validated computational model.

Table 2 compares the unpaired spin populations calculated for key orbitals of several complexes. Interestingly, although the d_{vz} spin population remains more or less constant at about 0.58 across the series, the d_{xz} spin population is equal to the d_{vz} spin population for 1 and $2(Fe^{IV}N\bullet)$ but decreases progressively for $2(Fe^V)$, 1_{ox} , and $[Fe^V(O)(TAML)]^-$. As viewed from the progression of d_{xz} spin density listed in Table 2, the electronic configuration of 2 more closely resembles that of 1_{ox} and $[Fe^V(O)(TAML)]^-$ than of 1 and $2(Fe^{IV}N\bullet)$. Because the transfer of electron density from the metal d_{xz} orbital to the singly occupied p_r orbital of the acetaminyl radical is not complete, the axial nitrogen retains a net β spin population in p_r (n = -0.30), which is much smaller than the value (n = -0.85) calculated for $2(Fe^{IV}N\bullet)$. Consistent with these ideas, the Mulliken Fe spin populations for $2(Fe^{V})$ and $2(Fe^{V})-H^{+}$ are calculated to be 0.86 and 0.77, respectively, which is lower than the 1.30 value calculated for 1. The expectation value of the operator S^2 provides quantitative insight into the oxidation state predicted by DFT. The $2(Fe^{V})$ model was found to have $\langle \Psi_{BP86} | \mathbf{S}^{2} | \Psi_{BP86} \rangle = 0.94$, which is close to the ideal value of S(S + 1) = 0.75 for $S = \frac{1}{2} \text{ Fe}^{V}$ but considerably smaller than the value of 1.75 anticipated for the broken symmetry state (BS) of $2(Fe^{IV}N\bullet)$. Using the expansion $|\Psi_{BP86} > = c|Fe^V$, $S = \frac{1}{2} > + (1-c^2)^{1/2}|Fe^{IV}N\bullet$, BS > and the expectation values $\langle \mathbf{S} = 1/2 | \mathbf{S}^2 | \mathbf{S} = 1/2 \rangle = 0.75$ and $\langle \mathbf{BS} | \mathbf{S}^2 | \mathbf{BS} \rangle = 1.75$, we obtain $c^2 = 0.81$ from $\langle \Psi_{\text{BP86}} | \mathbf{S}^2 | \Psi_{\text{BP86}} \rangle =$ $0.75 c^2 + 1.75(1-c^2) = 0.94$. As spin unrestricted calculations show always some degree of spin contamination, the Fe^V character of 2 (and $2-H^+$) is greater than 80%.

If we view 2 and 2-H⁺ as Fe^V complexes, the spin densities at the axial ligand atoms must originate from covalent spin polarization (22). Interaction of the oxo $O(p_y)$ electron pair with the Fe(d_{yz}) orbital results in partial transfer of a β oxo electron to pair off some of the α spin density associated with the unpaired Fe(d_{yz}) electron, leaving a net α spin population in $O(p_y)$ (blue in Fig. 5). The imido N(p_x) electron pair transfers net α spin density to the empty Fe(d_{xz}) orbital, guided by Hund's rule, leaving a net β spin population in N(p_x) (red in Fig. 5). The electron pair in the out-of-plane N(p_y) orbital of the imido ligand is calculated by DFT to be 10,000 cm⁻¹ lower in energy than the redox-active in-plane N(p_x) orbital and thus is much less involved in covalent delocalization to the metal.

Table 2. Calculated unpaired spin populations $n = n_{\alpha} - n_{\beta}$ of Fe-d_{xz}, Fe-d_{vzr}, and N_{axial}-p_x orbitals

	$Fe^{V}(O)(TAML) \textbf{*}$	$1_{ox}(\mathit{Fe^{V}})^{\dagger}$	2(<i>Fe^V</i>)*	1	2 (Fe ^{IV} N•)§
Fe-d _{xz}	0.07	0.15	0.23	0.58	0.58
Fe-d _{vz}	0.57	0.63	0.53	0.58	0.58
N _{axial} -p _x	_	~0	-0.30	0	-0.85

*Tiago de Oliveira et al. (14).

^tComputational model obtained by removing one electron from **1**. ^tBP86 solution.

 $^{s}B3LYP$ solution, S = 1 Fe^{IV}=O coupled to S = 1/2 radical on N of axial ligand.



Fig. 5. Spin density plots of geometry optimized BP86 solutions for $2(Fe^{V})$, 1_{ox} , and $[Fe(O)(TAML)]^-$. The plot for $2(Fe^{V})$, shown in two views, reveals the contours of the orbitals carrying spin density. From top to bottom, p_x (N_{am}), d_{yz} (Fe), and p_y (¹⁷O). For 1_{ox} the TMC ligand has been rotated by approximately 90° around the Fe=O bond relative to the orientation shown for $2(Fe^{V})$. Majority spin α in blue; minority spin β in red.

The resulting spin densities shown in Fig. 5 rationalize the spatial pattern observed in the A-tensors for ¹⁷O, ⁵⁷Fe, and ¹⁴N. Thus, the net β spin population of the N(p_x) gives a magnetic hyperfine tensor with a large component along x and two smaller components along y and z for 2 (and 2-H⁺), with calculated $A_{x,y,z}$ values of (-39, 11, 6) MHz for ¹⁴N in good agreement with the data. The net α spin population of Fe(d_{vz}) produces a spin-dipolar hyperfine contribution with a large negative component along x, which together with the negative Fermi contact term yields a magnetic hyperfine tensor of which the dominant component is along x; the calculated $A_{x,y,z}$ values of (-44, -19, 3) MHz (Table 1) are also in excellent agreement with the results obtained for **2** and **2-H**⁺. Finally, the net α spin population of O(p_v) gives an ¹⁷O A-tensor that has a large component along y, with $A_{x,y,z}$ values of (-1, -71, +76) MHz. The agreement for ¹⁷O between experiment and calculations is not as good as that for ⁵⁷Fe and ¹⁴N; A_v is still too small and A_z comes out too large but, importantly, the calculated tensor lacks axial symmetry around z and exhibits the x/y anisotropy observed experimentally. Finally, the g-values calculated for $2(Fe^{V})$ and reported in Table 1 match both in magnitude and direction the experimental data for 2 and 2-H⁺. Thus the congruence of the experimental EPR results with the $2(Fe^{V})$ model provides a strong rationale for the description of 2 and $2-H^+$ as Fe^V complexes.



Fig. 6. Calculated geometric and spectral parameters for the energyminimized BP86 models $1_{\alpha x_i}$ 2, and 2- H^+ . Bond lengths are reported in angstroms "L" denotes the TMC ligand.

In contrast, the $\nu_{\text{Fe}=0}$ frequencies hint at an Fe^{IV} oxidation state for **2** and **2**-**H**⁺ and therefore may at first glance favor $2(Fe^{IV}N\bullet)$ over $2(Fe^V)$. It should be noted, however, that $\nu_{\text{Fe}=0}$ is a spectroscopic parameter that reflects the length of the Fe-oxo bond (23). The Fe–O bond in the DFT model $2(Fe^V)$ (1.70 Å) is longer than the oxoiron(V) bond in 1_{ax} (1.60 Å) and the oxoiron(IV) bond in **1** (1.66 Å) due to delocalization of electron density over the oxoironimido moiety* (see Fig. 6) and is therefore predicted to exhibit a lower $\nu_{\text{Fe}=0}$ than 1_{ax} . The $2(Fe^V)$ model predicts $\nu_{\text{Fe}=0}$ values of 785 cm⁻¹ for $2(Fe^V)$ ($\nu_{\text{exp}} = 798$ cm⁻¹) and of 829 cm⁻¹ for $2(Fe^V)$ -**H**⁺ ($\nu_{\text{exp}} = 811$ cm⁻¹) in excellent agreement with the trend observed in our experimental data, indicating that the imido ligand exerts a sizable *trans* influence on the Fe–O unit.

Similarly, the weaker Fe=O bonds of 2 and 2-H⁺ are expected to give rise to higher Mössbauer isomer shifts, as this spectroscopic parameter reflects the 3 s electron density at the ⁵⁷Fe nucleus and the extent to which the 3 s electrons are shielded by 3d electrons. Thus, we expect the δ value of 2 to be more positive than that of an Fe^{V} complex with an O=Fe-NCCH₃ unit. As a further test of the *trans* influence of the O=Fe=N-C(O)-R unit on the value of δ , we calculated the properties for the hypothetical 1-electron reduced 2 (denoted 2_{red}) and obtained an Fe–O bond length of 1.767 Å, which is 0.10 Å longer than the Fe–O distance 1.664 Å calculated for 1. The increase in bond length is indeed accompanied by an increase of the δ value by 0.09 mm/s and a 164 cm⁻¹ decrease in $\nu_{Fe=O}$. The significant trans influence of the acetylimido ligand is also manifested in the comparison of the calculated values for $2(Fe^{V})$ ($\mathbf{r}_{Fe-O} =$ 1.701 Å, $\delta = 0.03$ mm/s, $\nu_{Fe=O} = 785$ cm⁻¹) and 1_{ox} ($r_{Fe=O} = 1.603$ Å, $\delta = -0.08$ mm/s, $\nu_{Fe=O} = 944$ cm⁻¹). These calculation tions support our view that $\nu_{Fe=O}$ and δ are poor indicators for the oxidation state of 2 and 2-H⁺, without properly accounting for the trans influence of the axial ligand. Judged from the full data set, 2 and $2-H^+$ are best described as Fe^V complexes.

The characterization of 2 and 2-H⁺ reported in this paper increases to three the number of synthetic oxoiron(V) complexes for which detailed spectroscopic and electronic structure information is available. Such complexes demonstrate that it is possible to stabilize such a high-valent state in an iron coordination complex. For $[Fe^{V}(O)(TAML)]^{-}$, the iron(V) oxidation state was achieved by the use of a tetraanionic tetraazamacrocyclic ligand that mitigates the high charge of the metal center (14). In contrast, 2 and 2-H⁺ are supported by a neutral tetraazamacrocycle, so the stabilization of the iron(V) state, it would seem, must derive from the presence of the charged axial oxo and imido ligands. The structure we favor for 2 is closely related to that of $[Mn^{V}(O)_{2}(TPFPP)]^{-}$ [where TPFPP is meso-tetrakis(pentafluorophenyl)porphinate dianion], which is the only known example of a first row transition metal containing an $[M^{V}(O)_{2}]$ moiety (24). Examples of nitridoiron(V) and nitridoiron(VI) complexes have also been reported, demonstrating the ability of the N^{3-} ligand to allow access to high-valent iron complexes, even with supporting polydentate ligands that are neutral or monoanionic (25–29). Lastly, the characterization of 2 and 2-H⁺ demonstrates the possibility of a neutral nonheme ligand to support an Fe(V) center and lends credence to mechanistic proposals for $Fe^{V}(O)(OH)$ oxidants in arene cis-dihydroxylation by Rieske dioxygenases (5) and in alkane, olefin, and water oxidation by a series of bioinspired synthetic iron catalysts (6-9, 11). For the latter, evidence for the fleeting iron(V) intermediates has been obtained from low

^{*}Unfortunately, our Raman experiments have not been able to identify vibrational modes that can be associated with the Fe=NR unit. This result may not be surprising, as vibrational data are also not available for the only previously reported six-coordinate Fe=NR complex (32), although Raman data for four-coordinate Fe=NR complexes have been obtained (33).

Experimental Procedures

Preparation of 1, 2, and 2-H⁺. All reagents and solvents were purchased from commercial sources and used as received unless otherwise noted. As previously reported (15), a solution of $[Fe^{II}(TMC)(O_3SCF_3)]$ in acetonitrile was reacted with 1.2 equivalents of PhIO solubilized in methanol to obtain 1 in quantitative yield, as monitored by the appearance of its distinctive chromophore at 820 nm. Compound 2 was prepared at -44 °C in acetonitrile by the addition of at least three equivalents of tert-butyl hydroperoxide followed by the addition of 1–2 equivalents of NBu₄OH or KO'Bu. Subsequent addition of at least five equivalents of strong acid, HBF₄ or HClO₄, yielded **2-H**⁺. Forty percent ¹⁷O-enriched H₂O was used to prepare the ¹⁷O EPR samples.

Physical Methods. A Hewlett Packard 8453A diode-array spectrometer equipped with a Unisoku Scientific Instruments cryostat to maintain a constant temperature was employed to collect electronic absorption spectrum. Resonance Raman spectra were generated via excitation with Kr^+ and Ar^+ lasers (Spectra Physics BeamLok 2060-RM) with a power of <10 mW at the samples, and the spectra were collected using an ACTON AM-506M3

- Denisov IG, Makris TM, Sligar SG, Schlichting I (2005) Structure and chemistry of cytochrome P450. Chem Rev 105:2253–2278.
- Zilly FE, et al. (2011) Tuning a P450 enzyme for methane oxidation. Angew Chem Int Ed 50:2720–2724.
- Kawakami N, Shoji O, Watanabe Y (2011) Use of perfluorocarboxylic acids to trick cytochrome P450BM3 into initiating the hydroxylation of gaseous alkanes. Angew Chem Int Ed 50:5315–5318.
- Rittle J, Green MT (2010) Cytochrome P450 compound I: Capture, characterization, and C–H bond activation kinetics. *Science* 330:933–937.
- Kovaleva EG, Lipscomb JD (2008) Versatility of biological non-heme Fe(II) centers in oxygen activation reactions. Nat Chem Biol 4:186–193.
- Chen K, Que L, Jr (2001) Stereospecific alkane hydroxylation by nonheme iron catalysts: Mechanistic evidence for an Fe^V=O active species. J Am Chem Soc 123:6327–6337.
- Company A, et al. (2007) Alkane hydroxylation by a nonheme iron catalyst that challenges the heme paradigm for oxygenase action. J Am Chem Soc 129:15766–15767.
- Chen K, Costas M, Kim J, Tipton AK, Que L, Jr (2002) Olefin *cis*-dihydroxylation versus epoxidation by nonheme iron catalysts: Two faces of an Fe^{III}—OOH coin. J Am Chem Soc 124:3026–3035.
- Company A, et al. (2009) Olefin-dependent discrimination between two nonheme HO-Fe^V = tautomeric species in catalytic H₂O₂ epoxidations. *Chem Eur J* 15:3359–3362.
- Chow TW-S, et al. (2010) Cis-dihydroxylation of alkenes with oxone catalyzed by iron complexes of a macrocyclic tetraaza ligand and reaction mechanism by ESI-MS spectrometry and DFT calculations. J Am Chem Soc 132:13229–13239.
- Fillol JL, et al. (2011) Efficient water oxidation catalysts based on readily available iron coordination complexes. *Nat Chem* 3:807–813
- Prat I, et al. (2011) Observation of Fe(V)=0 using variable temperature mass spectrometry and its enzyme-like C–H and C=C oxidation reactions. Nat Chem 3:788–793.
- Que L, Jr (2007) The road to non-heme oxoferryls and beyond. Acc Chem Res 40:493–500.
- Tiago de Oliveira F, et al. (2007) Chemical and spectroscopic evidence for an Fe^V—oxo complex. Science 315:835–838.
- Rohde J-U, et al. (2003) Crystallographic and spectroscopic evidence for a nonheme Fe^{IV}=O complex. Science 299:1037–1039.
- Jackson TA, et al. (2008) Axial ligand effects on the geometric and electronic structures of nonheme oxoiron(IV) complexes. J Am Chem Soc 130:12394–12407.
- Payne GB, Deming PH, Williams PH (1961) Reactions of hydrogen peroxide. VII. Alkalicatalyzed epoxidation and oxidation using a nitrile as co-reactant. J Org Chem 26:659–663.
- MacFaul PA, Arends IWCE, Ingold KI, Wayner DDM (1997) Oxygen activation by metal complexes and alkyl hydroperoxides. Applications of mechanistic probes to explore the role of alkoxyl radicals in alkane functionalization. J Chem Soc Perkin Trans 2:135–146.

monochromator and a Princeton Instruments ACTON PyLoN LN/CCD-1340 \times 400 detector. X band (9.28 GHz) EPR spectra were recorded on a Bruker ESR 300 spectrometer equipped with an Oxford ESR 910 liquid Helium cryostat and an Oxford temperature controller. Mössbauer spectra were recorded using Janis Research Super-Varitemp dewars that allowed studies in applied magnetic fields up to 8.0 T. Mössbauer spectral simulations were performed using the WMOSS software package (SEE Co), and EPR spectra were simulated with SpinCount, a program developed by Prof. M. P. Hendrich of Carnegie Mellon University. Isomer shifts are quoted relative to Fe metal at 298 K. ESI-MS experiments were conducted using a Bruker BioTOF II mass spectrometer. The spray chamber voltage was set to 4,000 V and the gas carrier temperature was set at 60 °C.

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- Roberts JE, Hoffman BM, Rutter R, Hager LP (1981) ¹⁷O ENDOR of horseradish peroxidase compound I. J Am Chem Soc 103:7654–7656.
- Schenk G, Pau MYM, Solomon EI (2004) Comparison between the geometric and electronic structures and reactivities of {FeNO}⁷ and {FeO₂}⁸ complexes: A density functional theory study. J Am Chem Soc 126:505–515.
- Hopmann KH, Conradie J, Ghosh A (2009) Broken-symmetry DFT spin densities of iron nitrosyls, including Roussin's red and black salts: Striking differences between pure and hybrid functionals. J Phys Chem B 113:10540–10547.
- 22. Drago RS (1992) Physical Methods for Chemists (Saunders, Philadelphia), 2nd Ed.
- Green MT (2006) Application of Badger's rule to heme and non-heme iron-oxygen bonds: An examination of ferryl protonation states. J Am Chem Soc 128:1902–1906.
- Jin N, Ibrahim M, Spiro TG, Groves JT (2007) Trans-dioxo manganese(V) porphyrins. J Am Chem Soc 129:12416–12417.
- Meyer K, Bill E, Mienert B, Weyhermüller T, Wieghardt K (1999) Photolysis of *cis* and *trans*-[Fe^{III}(cyclam)(N₃)₂]⁺ complexes: Spectroscopic characterization of a nitridoiron (V) species. *J Am Chem Soc* 121:4859–4876.
- 26. Grapperhaus CA, Mienert B, Bill E, Weyhermüller T, Wieghardt K (2000) Mononuclear (nitrido)iron(V) and (oxo)iron(IV) complexes via photolysis of [(cyclam-acetato)Fe^{III} (N₃)]⁺ and ozonolysis of [(cyclam-acetato)Fe^{III}(O₃SCF₃)]⁺ in water/acetone mixtures. *Inorg Chem* 39:5306–5317.
- Aliaga-Alcalde N, et al. (2005) The geometric and electronic structure of [(cyclam-acetato)Fe(N)]⁺: A genuine iron(V) species with a ground-state spin S = ¹/₂. Angew Chem Int Ed 44:2908–2912.
- Berry JF, et al. (2006) An octahedral coordination complex of iron(VI). Science 312:1937–1941.
- Scepaniak JJ, et al. (2011) Synthesis, structure, and reactivity of an iron(V) nitride. Science 331:1049–1052.
- Lyakin OY, Bryliakov KP, Britovsek GJP, Talsi EP (2009) EPR spectroscopic trapping of the active species of nonheme iron-catalyzed oxidation. J Am Chem Soc 131:10798–10799.
- Lyakin OY, Bryliakov KP, Talsi EP (2011) EPR, ¹H and ²H NMR, and reactivity studies of the iron-oxygen intermediates in bioinspired catalyst systems. *Inorg Chem* 50:5526–5538.
- Klinker EJ, et al. (2006) A tosylimido analogue of a nonheme oxoiron(IV) complex. Angew Chem Int Ed 45:7394–7397.
- Mehn MP, Brown SD, Jenkins DM, Peters JC, Que L, Jr (2006) Vibrational spectroscopy and analysis of pseudo-tetrahedral complexes with metal imido bonds. *Inorg Chem* 45:7417–7427.
- Schulz CE, Rutter R, Sage JT, Debrunner PG, Hager LP (1984) Mössbauer and electron paramagnetic resonance studies of horseradish peroxidase and its catalytic intermediates. *Biochemistry* 23:4743–4754.
- Krzystek J, et al. (2008) Determination by high-frequency and -field EPR of zero-field splitting in iron(IV) oxo complexes: Implications for intermediates in nonheme iron enzymes. *Inorg Chem* 47:3483–3485.