

NIH Public Access

Author Manuscript

J Org Chem. Author manuscript; available in PMC 2013 August 03

Published in final edited form as:

J Org Chem. 2012 August 3; 77(15): 6480-6494. doi:10.1021/jo300963m.

meso-Arylporpholactones and their Reduction Products

Christian Brückner^{†,*}, Junichi Ogikubo[†], Jason R. McCarthy^{†,‡}, Joshua Akhigbe[†], Michael A. Hyland[†], Pedro Daddario[†], Jill L. Worlinsky[†], Matthias Zeller[§], James T. Engle[%], Christopher J. Ziegler[%], Matthew J. Ranaghan[&], Megan N. Sandberg[†], and Robert R. Birge^{†,&}

Department of Chemistry, University of Connecticut, Storrs, CT 06269-3060, United States. Department of Molecular and Cell Biology, University of Connecticut, Storrs, CT 06269-3125, United States. Department of Chemistry, Youngstown State University, One University Plaza, Youngstown, OH 44555-3663, United States. Department of Chemistry, University of Akron, Akron, OH 44325-3601, United States.

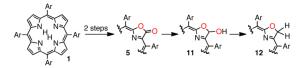
[†]University of Connecticut, Department of Chemistry United States

§Youngstown State University

[%]University of Akron

[&]University of Connecticut, Department of Molecular and Cell Biology

Abstract



The rational syntheses of *meso*-tetraaryl-3-oxo-2-oxaporphyrins 5, known as porpholactones, via MnO_4^- -mediated oxidations of the corresponding meso-tetraaryl-2,3-dihydroxychlorins (7) is detailed. Since chlorin 7 is prepared from the parent porphyrin 1, this amounts to a 2-step replacement of a pyrrolemoiety in 1 by a oxazolone moiety. The step-wise reduction of the porpholactone 5 results in the formation of chlorin analogues, meso-tetraaryl-3-hydroxy-2oxachlorin (11) and meso-tetraaryl-2-oxachlorins (12). The reactivity of 11 with respect to nucleophilic substitution by O-, N-, and S-nucleophiles is described. The profound photophysical consequences of the formal replacement of a pyrrole with an oxazolone (porphyrin-like chromophore) or (substituted) oxazole moiety (chlorin-like chromophore with, for the parent oxazolochlorin 12, red-shifted Q_x band with enhanced oscillator strengths) are detailed and rationalized on the basis of SAC-CI and MNDO-PSDCI molecular orbital theory calculations. The single crystal X-ray structures of the porpholactones point at a minor steric interaction between the carbonyl oxygen and the flanking phenyl group. The essentially planar structures of all chromophores in all oxidation states prove that the observed optical properties originate from the intrinsic electronic properties of the chromophores and are not subject to conformational modulation.

^{*}Author to whom correspondence should be addressed: Fax: (+1) 860 486-2981; Tel: (+1) 860 486-2743; c.bruckner@uconn.edu. Current address: Center for Systems Biology, Massachusetts General Hospital, Boston, MA 02114, United States.

Supporting Information Available. ¹H, ¹³C NMR, and IR spectra of all compounds and experimental details to the crystal structure determination of **5H₂**, **5aZn**, **12H₂**, **13-OMe**, and **15-II-Zn**, including the cif file. This material is available free of charge via the Internet at http://pubs.acs.org.

Introduction

meso-Tetraarylporphyrins **1** and their metal complexes are used in a number of model compounds for naturally occurring porphyrinic cofactors or light-harvesting systems.¹ The great popularity of **1** arises from their straight-forward syntheses and the availability of a wide variety of aryl-functionalized derivatives.²

In a seminal contribution by Crossley and King more than 25 years ago,³ it was recognized that oxidation of β -substituted porphyrins, such as dione **6** (prepared from **1** via **2**, **3**, or **4**), can lead to the loss of one β -carbon and the formal replacement of the porphyrinic β , β '-bond by a lactone moiety, forming porpholactone **5** (Scheme 1).

One other serendipitous finding five years later identified strongly oxidizing reaction conditions (AgNO₃ in refluxing acetic acid containing oxalate) that were suitable for converting porphyrin **1** (with $Ar = C_6F_5$) directly into a porpholactone.^{4,5} However, the latter reaction is not general since it is only applicable toward the synthesis of *meso*-tetrakis(pentafluorophenyl)porpholactone. The method also requires extensive chromatography to separate the porpholactone from the starting material and numerous other non-polar 'over-oxidized' byproducts.^{4,5} Additional specialized reaction pathways toward porpholactones have been discovered since, such as the singlet oxygen oxidation of β -aminoporphyrin **4** (Scheme 1).⁶ Select oxidations of dione **6** also lead to porpholactones, perhaps shedding light on a possible reaction mechanism toward the formation of porpholactones **5**.⁷ Porpholactones also appeared as adventitious products in a variety of oxidation reactions of β -derivatized porphyrins.⁷⁻⁹

Porpholactones have been demonstrated to be of practical value: The Fe(III) and Fe(IV)=O⁺ complexes of *meso*-tetrakis(2,6-dichlorophenyl)-substituted porpholactone were used as model compounds for naturally occurring chlorin-type prosthetic groups.⁶ The catalytic activity of the Fe(III)Cl and Mn(III)Cl complexes of *meso*-tetrakis(phenyl)porpholactone with respect to olefin epoxidation and sulfide oxidation reactions were tested.^{10,11} [*meso*-Tetrakis(pentafluorophenyl)porpholactonato]Pt(II) is a promising component in pressure sensitive paints,¹² allowing the imaging of air flow around objects.¹³ This complex can also be utilized as a high pH sensor in the range of pH 11.5–13.¹⁴ Despite their increasing utility and the emergence of their unique reactivity, no full account on the rational, general, and high-yielding synthesis of *meso*-tetraarylporpholactones has been published to date. In the first part of this report, we will fill this gap by following on our communication.¹⁵

We previously detailed the OsO₄-mediated β , β '-dihydroxylation of *meso*-tetraarylporphyrin **1** and closely related porphyrin derivatives to produce the corresponding *meso*-tetrakis-2,3-dihydroxy-2,3-chlorin **7**.^{14,16-19,20} This reaction was adopted by other groups, providing a range of dihydroxychlorins.^{10,11,21-23} We further demonstrated the versatility of the chlorin diols as starting material for the preparation of porphyrinoids containing non-pyrrolic building blocks using our versatile 'breaking and mending' strategy.^{9,14-18,24,25} The synthesis of porpholactones via the MnO₄⁻-mediated oxidation of the known *meso*-tetraaryl-2,3-dihydroxychlorins also follows this strategy. We will also present the structural description of free base and Zn(II) porpholactones using single crystal X-ray diffraction structure elucidation.

In the second part of this report we present a comprehensive account of the hydridemediated reduction reactions of porpholactones **5** to chlorin-like 2-oxachlorins (also known as oxazolochlorins) containing hemiacetal, acetal, and ether functionalities. Owing to their intense absorbance at wavelengths > 650 nm, the optical window of tissue,²⁶ chlorins (β , β 'dihydroporphyrins) are superior to porphyrins in all applications in which light needs to

penetrate tissue, such as the photodynamic therapy (PDT) of tumors.²⁷ In addition, many naturally occurring light-harvesting chromophores are chlorins.²⁸ In part owing to these applications, the generation of chlorins by total synthesis or by conversion of porphyrins has become a central topic in current porphyrin chemistry.²⁹

We previously presented an isolated example of a reduced porpholactone derivative, hemiacetal 3-hydroxy-3-oxachlorin **11a**, in a comparative study of chlorins,³⁰ and we demonstrated that this compound possesses high photodynamic efficacy *in vivo* in a murine tumor model.³¹ We also identified an adventitious side-product in the oxidative transformations of the silver(II) complex of dihydroxychlorin **7** as an [2-ethoxy-2-oxachlorinato]Ag(II) complex (**11aAg**).^{8,32} Lastly, an oxazolochlorin monomer and dimer formed as products of an intramolecular Cannizzaro reaction of free base secochlorin bisaldehyde **8aH**₂.⁹ However, in spite of their use or occurrence on several occasions, no full synthetic paper describing their rational syntheses has been reported to date. This will be provided herein, and we will also provide chemical evidence that might rationalize the high biological activity of the hemiacetal.³¹

In the final section of this manuscript, we will contrast the UV-visible absorption and fluorescence emission properties of the free base and Zn(II) complexes of the 2-oxaporphyrin and 2-oxachlorin against each other and against the properties of the parent porphyrin **1a** and 2,3-dihydroxychlorin **7aH**₂. The observed trends in the optical spectra are rationalized on the basis of SAC-CI and MNDO-PSDCI molecular orbital theory calculations. Thus, this contribution reveals the synthesis and detailed physical and chemical description of a chlorin-like stable family of chromophores that is readily accessible and for which a number of applications can be foreseen.

Results and Discussion

Synthesis of Porpholactones

Oxidation of dihydroxychlorins 7 with MnO_4^- produced porpholactone 5 in a single step (Scheme 2). The source of MnO_4^- can either be an excess of powdered KMnO₄ suspended at ambient temperature in an organic solvent (toluene, CH₂Cl₂, CHCl₃, THF) in the presence of the phasetransfer agent 18-crown-6, KMnO₄ heterogenized on silica gel,³³ or, most conveniently, the use of one to five equivalents of cetyltrimethylammonium permanganate (CTAP)³⁴ in CH₂Cl₂.

The work-up of the reaction is simple. Filtration of the crude mixture through a plug of diatomaceous earth removes the brown, flocculent to pasty manganese oxides. Short column chromatography and crystallization of the porpholactone isolates the product as crystalline materials in up to 90% yields. The reaction times vary, depending on the concentration (and solubility) of the dihydroxychlorin and the number of equivalents of CTAP (or phase transfer agent) used, and range from 30 min to 12 h. The addition of several smaller portions of oxidant under TLC control to monitor the consumption of the starting material proved to be beneficial. Large excesses of oxidant, elevated temperatures or prolonged reaction times hasten the degradation of the macrocycles, as indicated by the loss of the Soret band in the UV-visible spectra of the reaction mixtures, and low isolated yields of the desired porpholactones.

We recently described the isolation of the dihydroxychlorin osmate esters **8**, the primary products from the reaction of porphyrin **1** with OsO_4 .¹⁹ The osmate esters are also susceptible to CTAP oxidation to the corresponding porpholactones but the reaction is slower than the reaction of the corresponding alcohols, and the fate of the osmium is not clear. Given the high toxicity of OsO_4 that potentially forms as a side product,³⁵ the

oxidation of the diol is given preference over the oxidation of the corresponding osmate ester. We reported the CTAP oxidation of the osmate ester of *meso*-tetrakis(pentafluorophenyl)-2,3-dihydroxychlorin $7gH_2$.¹⁴ The reason for this was that the corresponding diol could at the time not be prepared. Since then, we have reported a method for the preparation of the $7gH_2$,³⁶ and we found that the oxidation of this diol is faster and cleaner than the oxidation of its osmate ester. Also, we will report here that the use of permanganate heterogenized on silica gel offers a very convenient and clean way of generating this *meso*-tetrakis(pentafluorophenyl)porpholactone $5gH_2$ (and it allows facile recovery of starting material). In general, however, the yields of this reaction are very low (25–30% are typical) compared to the yields obtained from oxidation of other tetraaryldihydroxychlorins.

The isolation of the porpholactones by silica gel column chromatography is much facilitated by their lower polarity compared to the corresponding dihydroxychlorins. Their identification is simplified by their characteristic mass and NMR spectra, indicative of the loss of one carbon from the porphyrin framework, the loss of axial symmetry, and the presence of a carbonyl group. As noted previously,^{3,4} the UV-visible spectra of free base porpholactones are surprisingly porphyrin-like, while those of their metal complexes are metallochlorin-like (to be detailed below).

We consider this methodology toward porpholactones to be fairly general. It is applicable to the oxidation of free base dihydroxychlorin **7**, its Ni(II), Zn(II), Ag(II), and Pt(II) complexes, to diol chlorins carrying a variety of electron-donating and -withdrawing *meso*-phenyl substituents,¹⁴ and 5,15-diphenyl-2,3-dihydroxychlorin **14** (see below). We previously also demonstrated the applicability of this method to the synthesis of dithiaporphyrin- and porphyrin *N*-oxide-based porpholactones.^{17,18} As also shown previously,^{6,10,11,12} additional metalloporpholactones are available through metal insertions into the free bases using standard methods (using conventional and microwave heating).

The exact mechanism of formation of the porpholactones remains unclear. Permanganate oxidation of dihydroxychlorin **7** suggests the formation of secochlorin biscarboxylate **10**. In fact, Crossley already surmised that the as yet unobserved secochlorin **10** is the immediate precursor to porpholactone **5**.³ Other reactions that reasonably can be expected to produce the biscarboxylate species also generate porpholactones. For instance, an attempt to insert silver ion into free base bisaldehyde **9** (using excess Ag(I) in pyridine, heat) formed [porpholactonato]Ag(II) **5Ag** in mediocre yield (~20%). Independent evidence points toward 2,3-dioxochlorins of type **6** to be the key intermediates in the conversion of porpholactones.^{7,18} Since porpholactones frequently also appear as adventitious (by)products in a number reactions that treat (β -derivatized) porphyrins under a variety of oxidizing conditions,^{3,4,7,8} one may regard porpholactones as the thermodynamic sink in the β , β '-oxidative degradation pathway of porphyrins. As such, it can be reasonably assumed that multiple pathways lead to this product.

Structural Aspects of Porpholactones

The crystal structures of [porpholactonato]Mn(III)Cl,¹⁰ Fe(III)Cl,⁶ Cu(II),⁷ and Ni(II)⁷ complexes are known. Thus, the connectivity of the porpholactones is not in question. However, the crystal structures of free base porpholactone **5aH**₂ and its Zn complex **5aZn** are interesting with respect to a number of other aspects (Figure 1A). The structural relationship of **5aH**₂ to the corresponding porphyrin **1aH**₂³⁷ is highlighted by the fact that both structures possess essentially isostructural unit cell parameters: same space group P1, with the cell dimension varying by only ~1% when comparing the room temperature structure of **1aH**₂ to the structure of **5aH**₂ determined at 100 K. The chromophore of

porpholactone $5aH_2$ is, analogous to that of porphyrin $1aH_2$, in effect planar with only a minor waving deformation (wav(x)) of the two opposing pyrrole moieties that is likely caused by the repulsion of the two inner hydrogens.³⁸ Like the metalloporpholactone crystal structures described today (see below), the free base porpholactone structure is highly disordered with respect to the relative position of the lactone moiety. In $5aH_2$ all 8 possible positions/orientations are occupied, albeit in a nonstatistical manner (for details, see ESI). While this highlights the structural equivalency of the replacement of a β , β '-bond by a lactone moiety, it also diminishes the significance of small bond lengths and angles differences. We therefore will consider herein mostly general structural trends.

A comparison of the Ni(II), Cu(II), Fe(III), and Mn(III) complexes of porphyrins³⁹ and porpholactones^{6,7,10} show also their overall similarity with respect to the respective overall macrocycle conformation. The C=O bond length in the zinc porpholactone **5aZn** is 1.26 Å and the C-O bond length is 1.40 Å. The corresponding bond lengths in the Mn(III)Cl complex of **5a** are 1.20 and 1.40 Å.¹⁰

Figure 1B shows a side view of the macrocycles of [porpholactonato]Zn(II) **5aZn** and its porphyrin analogue **1pyZn** ([*meso*-tetrakis(4-pyridyl)porphyrinato]Zn(pyridine))⁴⁰ (and the diphenylporpholactone **15-II-Zn**, described below). The zinc ion sits in all complexes slightly out of plane, drawn toward the side of the axially coordinated pyridine. This is normally observed for square pyramidal zinc(II) porphyrin complexes.³⁹ Most notably when comparing the conformations of the macrocycles in **5aZn** and **1pyZn**, the oxazolone moiety is not, as expected, perfectly co-planar with the macrocycle. This deviation may originate from a steric interaction between the carbonyl oxygen and the flanking phenyl group. A comparable observation was made for the Cu(II) complex of tetrakisphenylporpholactone.⁷

To test whether other experimental evidence can be found for a significant steric interaction between the lactone carbonyl and the flanking phenyl group, we subjected the known diphenyl-2,3-dihydroxychlorin 14^{23} to the CTAP oxidation conditions (Scheme 3). Two isomers of the corresponding diphenylporpholactone 15 can be formed: 2-oxa-3-oxo structure 15-I-H₂, and 3-oxa-2-oxo structure 15-II-H₂.

The oxidation of diol **14** proceeded smoothly and rapidly, generating a porpholactone product mixture. The ¹H NMR of the porpholactone fraction indicated the presence of two isomers in a ~1:5 ratio (see ESI). Repeated preparative plate chromatography and recrystallizations (from MeOH/CHCl₃) allowed the isolation of a pure fraction of the majority product but the minority product could not be isolated in pure and high enough yields to perform a full analysis. 1D and 2D-NMR spectroscopy (see ESI) allowed the unambiguous assignment of the majority product as the 2-oxo-3-oxa isomer **15-II-H**₂, that is unaffected by any steric interaction between the lactone carbonyl and a phenyl group. This was confirmed by single X-ray crystal structure elucidation of **15-II-Zn**, as its pyridine adduct, formed by zinc insertion into the free base (Figure 1).

The fact that the seemingly less sterically inhibited isomer forms in preference over the other isomer serves as an indication for the existence of a small but noticeable steric interaction between the carbonyl and the phenyl group. Most significantly, the macrocycle conformation (Figure 1B) shows that the lactone moiety is near-perfectly co-planar with the oxazole moiety, and that the macrocycle of **15-II-Zn** is overall significantly more planar than that of the tetraaryl analogue **5aZn**. This provides the most convincing proof for the rationalization of the non-planar arrangements of the lactone moiety with the porphyrins in **5aH₂** and **5aZn** on steric grounds.

In addition, we observed reduced disorder in the structure of **15-II-Zn** versus that observed in **5aZn**, and thus can more reliably determine the bond lengths in the oxazolone ring. The C=O lengths in **15-II-Zn** range from 1.106(3) to 1.228(3) Å, while the C-O lengths vary between 1.357(3) to 1.486(8) Å (there are two independent molecules per unit cell). Compared to the free base porpholactones, these bond length changes compared to the porpholactone free base support a chlorin rather than a porphyrin-like electronic structure.

Step-wise reduction of porpholactone 5aZn

Functional group conversions of the lactone moiety offer the opportunity to synthesize oxazolochlorins.^{9,25,30,31} Thus, reduction of the non-polar purple porpholactone **5aZn** with DIBAL-H in dry THF at -78 °C generates in near-quantitative yields a bright blue-green pigment of much higher polarity (isolated yields ~80%, 1 gram scale; Scheme 4). The ¹H NMR spectroscopic signature of this product is lactone-like with two additional signals: One signal of variable chemical shift assigned to the hydroxy functionality (exchangeable with D₂O), and a signal at 3.62 ppm attributed to the proton of the oxazole-moiety (cf. to the pyrrolidine proton of **7aZn** at 6.12 ppm)¹⁶. The corresponding sp³-carbon signal is observed at 112.1 ppm in the ¹³C NMR spectrum. All other analytical and spectroscopic data confirm the structure of the reduction product to be [3-hydroxy-2-oxachlorinato]Zn **11Zn**.

The reduction of free base porpholactones $(5H_2)$ is complicated by the formation of aluminumcontaining side products. Low valent aluminum species are known to metalate free base porphyrins.⁴¹ Therefore, the free base porpholactol $11H_2$ is best prepared by reduction of the zinc complex 5aZn, whereby the work-up procedure includes a wash with 6M aqueous HCl that demetalates the product.

The hemiacetal functionality of $11H_2$ is susceptible to acetal formation under mild conditions.^{8,9} Hence, alcohols must be excluded from any isolation and/or manipulation procedure of $11H_2/Zn$, lest the formation of the corresponding acetals are desired (see also below).

An acid-catalyzed (BF₃·OEt₂ or Amberlyst 15, H⁺ form) silane-induced deoxygenation of the lactol hydroxy group of **11H**₂ (or **11Zn** concomitant with a demetallation reaction) at room temperature provides a green compound that can be identified as the oxazolochlorin **12H**₂. The loss of the hydroxyl group from **11H**₂ is marked by a decrease in polarity of the product, the appearance of the signals in the ¹H NMR at 6.54 ppm for the two oxazolochlorin protons, accompanied by the signal in the ¹³C NMR for a secondary carbon (DEPT) at 76.4 ppm (see ESI).

The connectivity of 2-oxachlorin $12H_2$ could be determined by single crystal X-ray diffraction (Figure 1A). As expected, the chromophore is planar with none of the non-planarity observed in the porpholactones 5. However, the extraordinary degree of disorder of the molecule in the crystal precludes any detailed bond length and angle analysis.

2-Oxachlorin $12H_2$ is generally sensitive toward (photo-sensitized) oxidation back to porpholactol $11H_2$. Thus, the compound must be shielded from exposure to light or oxidizing conditions. The oxazole methylene group is located in a benzylic position with respect to the porphyrinoid aromatic system and α to an oxygen atom in the oxazole moiety. This group thus is highly activated with respect to, for instance, carbocation formation, a fact highlighted by an oxidative dimerization reaction of an oxazolochlorin that is proposed to be an intermediate in the Cannizzaro reaction of *meso*-phenylsecochlorin bisaldehyde.⁹ As a result of this high reactivity, insertion of zinc(II) into $12H_2$ requires anaerobic conditions. Warming $12H_2$ with Zn(OAc)₂·2H₂O in degassed warm DMF under N₂ leads

cleanly to the formation of **12Zn**. The optical properties of the oxazolochlorin derivatives are discussed below.

Attempts at one-step reductions from 5aZn to 12aZn using LiAlH₄ or DIBAL-H under more forcing conditions (large stoichiometric excess, room temperature) failed. These reactions merely lead to the destruction of the porphyrinic macrocycle without the formation of any major product.

Acetalization of Porpholactol 11H₂

The lactol hydroxy group of $11H_2$ is susceptible to facile acidcatalyzed nucleophilic substitution by a range of O-, N-, and S-nucleophiles, providing access to a number of stable chlorin-like derivatives of graded lipophilicity (Scheme 4). A marked nucleophiledependent reactivity difference is noted. Exposure of $11H_2$ to primary, secondary, and tertiary alcohols results in a rapid reaction that is essentially quantitative after 30 min to 1 h at ambient temperature. The resulting acetals **13-OR** showed all the expected spectroscopic data. Diagnostic for the successful formation of acetals containing an α -methylene group, this group shows a diastereotopic splitting. This can be rationalized by its relative position with respect to the macrocycle plane,³² exposing one of the methylene protons to a much larger degree to the diatropic ring current than the other. Alkoxy-substituted morpholinochlorins show a very similar effect.^{15,16} Bulky alcohols like cholesterol or pregnenolone can also be attached to the chromophore with ease.

The crystal structure of **13-OMe**, as its Ag(II) complex³² and a phorpholacol acetal dimer⁹ were previously characterized by single crystal X-ray diffraction. We present here the structure of **13-OMe** as a proof of connectivity and to complete the series of 2-oxachlorins in three different oxidation states (together with the 3-oxo- and the 3-hydroxy-2-oxachlorins), but the structure is disordered to a point that a conformational analysis is meaningless. The conformation of the oxazolochlorin chromophore in the dimer, however, suggested a certain degree of conformational flexibility, though no major distortions from planarity were observed.⁹

Secondary amines required a longer reaction time and azeotropic removal of the water formed (reflux in benzene with 3 Å mol sieves placed in a Soxhlet apparatus over several days) to push the reaction to completion. The hemiaminals **13-NR**₂ showed all the expected spectroscopic and analytical properties, including the diastereotopic split of the α -methylene protons. We did not succeed in reacting primary amines with the porpholactols under these or other conditions tested.

The reactivity of primary thiols was very much similar to the reactivity of the corresponding alcohols. Over time, however, the formation of side products with spectroscopic data suggestive of being the corresponding sulfoxides appeared (m/z = +16 compared to the expected compound; essentially identical ¹H and ¹³C NMR spectra).

This high reactivity of lactol $11H_2$ with respect to acetal, aminal, and thioacetal formation is of immediate interest for the potential application of the 2-oxachlorins as photochemotherapeutics. We previously demonstrated the efficacy of $11H_2$ when incorporated into a biodegradable nanoparticle for the photodynamic treatment of a tumor in a mouse model.³¹ The facile acetalization demonstrated here suggests that the biodistribution of the hemiacetal may also be modulated by its ability to undergo derivatization with biomolecules containing alcohol and thiol groups. Such promiscuity may allow achievement of a biodistribition of the drug it would not have as a single and stable compound. Inversely, the facile derivatization of the 3-hydroxy-2-oxachlorins suggests the

preparation of amphiphilic (pro)drugs using PEGs, carbohydrates, or similarly suitable moieties. These aspects of the oxazolochlorins are currently being studied in detail.

Comparison of Porpholactones and Oxazolochlorins to their Carbaporphyrin Analogues

Carbaporphyrins are porphyrin analogues containing a carbon atom in place of an inner nitrogen. Pawlicki and Latos-Grazynski reported the total synthesis of **17**, the carbaporphyrin analogue to 3-methoxy-2-oxachlorin **13-OEt**.^{44,45} This reaction is in contrast to our 'breaking and mending of porphyrin' strategy toward porpholactones **5**. Like **13-OEt** (see below), 2-oxa-21-carbachlorin **17** possesses a chlorin-like spectrum ($\lambda_{\text{Soret}} = 437$ nm and four Q-bands with $\lambda_{\text{max}} = 672$ nm). The synthesis of the free base nonmacrocycle-aromatic 2-oxa-21-carbaporphyrin could not be achieved. Multi-step oxidation of **17** led to the formation of porphyrnoid **18**, the carbaporphyrin analogue to porpholactone **5**. The UV-visible spectrum of **18** is, in similar fashion to its aza-analogue, porphyrin-like with a sharp Soret band and four Q-bands ($\lambda_{\text{max}} = 690$ nm). Unlike the azaporphyrins, however, the metallocarbaporphyrin chemistry is dominated by reactions on the inner carbon.⁴⁶

Optical Properties of Porpholactones and Oxazolochlorins

Free base porpholactones possess UV-visible and fluorescence emission spectra that are almost indistinguishable from those of the corresponding porphyrins (Figure 2). The similarity of the porphyrin and porpholactone spectra, noted already upon their discovery,^{3,4} is surprising as the modified pyrrolic moiety has lost its cross-conjugated β , β '-double bond. Therefore, porpholactones could have been expected to possess chlorin-like spectra.⁴² Evidently, however, the electronic effects of the carbonyl double bond mimic the presence of a β , β '-double bond. On the other hand, porpholactone Zn(II) complexes exhibit metallochlorin-like UV-visible spectra, though they are slightly hypsochromically shifted when compared to the spectra for metallochlorin **7aZn**. On the basis of iterative extended Hückel calculations, Gouterman and co-workers categorized porpholactones to lie between porphyrins and chlorins.⁴

The reduced porpholactones, 3-hydroxy-2-oxachlorins $11H_2$ ($\lambda_{max} = 646$ nm) and 11Zn, both possess chlorin-like optical spectra (cf. to the spectra for $7aH_2$, $\lambda_{max} = 648$ nm, and 7aZn). The influence of the 3-hydroxyl functionality on the 2-oxachlorin chromophore is profound. The removal of this group in the free base chromophore $12H_2$ results in a 22 nm red-shift ($\lambda_{max} = 668$ nm) in the UV-visible spectrum. We have demonstrated before the distinct blue-shifts caused by OH-substitution of chlorin pyrrolidines.¹⁹ Most surprisingly, the removal of the hydroxy group also results in a significant enhancement of the extinction coefficient of the Q_x band relative to its Soret band. Thus, replacement of the CH₂CH₂ group in chlorins by a CH₂O group has an auxochromic effect, adding to the collection of groups that are known to substantially modify the chlorin chromophore.⁴³ The computations presented below will offer a rationalization for this observation.

Theoretical Analysis of the Free Base Optical Spectra Energies and Intensities

The theoretical studies presented here seek to refine the understanding of the porphyrin-like optical spectra of porpholactones and the photophysical origins of the unusual intensity and red-shifted transition energy of the Q_x band in 2-oxachlorin **12aH2**. Because this compound readily oxidizes, we doubted the reliability of the molar extinction coefficient measurements. Thus, the spectra shown in Fig. 2 are all relative to a normalized Soret band. This approach is logical, but presents a problem because the SAC-CI and MNDO-PSDCI calculations predict that the intensity of the Soret band, which is made up of two or more transitions, will also change with the modifications. To provide a consistent analysis, we

integrated the experimental absorption bands in energy space to provide oscillator strength ratios. The solvent-averaged results allow a consistent comparison of the calculated and observed values (Table 1).

The results of the high-accuracy (level two) SAC-CI calculations are shown in Figure 3 (left panel). These calculations do not include the phenyl groups, and thus we are observing purely those effects associated with substitutions within the macrocycle. The SAC-CI calculations predict that $12aH_2$ will have the most intense Q_x band (Table 1). However, the observed intensity ordering ($5aH_2 < 1aH_2 < 7aH_2 < 11H_2 < 12H_2$) differs from the SAC-CI predicted intensity ordering ($1aH_2 < 7aH_2 < 5aH_2 < 11H_2 < 12H_2$) in that $5aH_2$ is calculated to have significantly more Q_x intensity than is observed. An examination of the MNDO-PSDCI calculations, which include the phenyl groups, indicates that the origin of the failure of the SAC-CI calculations on $5aH_2$ may be associated with the neglect of the phenyl groups. However, the semiempirical calculations (Fig. 3, right panel) underestimate all of the Q_x band intensities, transferring too much oscillator strength into the Q_y bands.

The first and most important observation is that, perhaps contrary to intuition, the dipole moments of the ground state species do not have a significant impact on either the intensities or the red-shifted character of the Q_x bands. The calculated dipole moments, shown in Table 1, correlate with neither the oscillator strengths nor the excitation energies of the bands. Rather, the most red shifted and most intense Q_x band observed in 2-oxachlorin **12H**₂ is observed in a molecule with an intermediate dipole moment of the group studied here.

We examined the configurational characteristics of the Q_x bands and concluded that the key mechanism responsible for both the red shift and the enhanced oscillator strength is mixing of the Q_x band with the low-lying transitions in the Soret region. In the C_{2v} symmetry of chlorin, the Q_x band and the lowest Soret band at ~3.4 eV share the same symmetry (B₂). However, the charge shift upon excitation into the Q_x band decreases the dipole moment of the molecule, a property characteristic of a covalent state (see ESI). In contrast, the lowest Soret band (S3) is an "ionic" state, and despite the highly polar environment of the chlorin, these two states do not share configurational space. The insertion of an oxygen atom in place of one of the methylene moieties in the chlorin macrocycle breaks up the symmetry, and has a significant impact on the excitation induced charge shifts (see ESI). In particular, all of the excited states are now ionic, and this mixing increases the ground state dipole moment of the molecule. The lowest Q_x band mixes with the S3 Soret transition with an approximately 28% configurational overlap, which transfers oscillator strength from S3 into Q_x . This mixing is visually apparent by reference to Figure 2. Note that the oscillator strength of S3 decreases by an amount comparable to the increase in the oscillator strength of Q_x. Furthermore, the configurational mixing "pushes" these two states apart energetically. Thus, electrostatically induced configurational mixing between $S1(Q_x)$ and S3(Soret) is responsible for both the increased oscillator strength and the increased red shift of the Q_x band.

The MNDO-PSDCI calculations provide insight into the impact of the phenyl groups on the relative properties of the Q_x bands. It is noted that while these semiempirical calculations underestimate the intensities of all the Q_x bands of the chlorins, these calculations do indicate that the Q_x band of oxazolochlorin **12H**₂ is significantly more intense than the others, as is observed. The phenyl group near the oxygen atom in this compound rotates slightly to form a weak hydrogen bond between the oxygen atom and the nearby phenyl hydrogen atom (shown in the ESI). Upon excitation, this interaction promotes a charge shift that is attributed to an enhanced mixing of Q_x and the S3 state. Thus in the case of **12H**₂, the phenyl groups increase both the red-shift and oscillator strength of Q_x by enhancing the mechanisms responsible for the analogous effects in the compound without phenyl rings.

Conclusions

We detailed an efficient synthesis of *meso*-tetraarylporpholactones from *meso*-tetraarylporphyrins in two steps using a number of aryl substituents and free base and metallochlorins. We demonstrated this reaction previously for the synthesis of a 21,23-dithiaporpholactone¹⁷ and two porpholactone *N*-oxide isomers.¹⁸ Thus, this methodology can be regarded to be general. In so doing, we have further expanded the synthetic methodologies of converting a porphyrin to pyrrole-modified porphyrins along our 'breaking and mending of porphyrins' strategy.

A number of porpholactones (tetraphenylporpholactones $5aH_2$ and 5aZn, diphenylporpholactone 15-II-Zn) were structurally characterized, displaying their structural relationship to the corresponding porphyrins but also suggest the presence of a steric interaction between the oxazolidone moiety with the flanking phenyl group. The similarity of the optical properties of porphyrins and porpholactones highlight their close electronic relationship.

Replacing a porphyrinoid β , β '-bond by a lactone results in a porphyrin-like optical spectrum for the free base chromophores but more metallochlorin-like properties for the metalloporpholactones. A stepwise reduction of the carbonyl group of **5aH** to an alcohol in porpholactol **11H**₂ establishes a chlorin-like spectrum in the free base and zinc(II) complexes. Further reduction of the hemiacetal moiety forms the novel oxazolochlorin chromophore **12H**₂. Compared to porpholactol **11H**₂ or 2,3-dihydroxychlorin **7aH**₂, it features a significantly red-shift chlorin-like spectrum with an increased extinction coefficient of the Q_x band. This effect could be traced back by computational analysis to an electrostatically induced configurational mixing between S1(Q_x) and S3(Soret) that is made possible by the desymmetrization of the chromophore upon replacement of the pyrrolidine moiety in a regular chlorin by an oxazole moiety. X-ray crystal structure analyses of chromophores in all oxidation states showed their planarity, thus providing evidence for the absence of conformational effects in the modulation of their optical spectra. The presented examples highlight the subtleties that control the structure-electronic properties relationships of porphyrinoids.

The simple syntheses of porpholactones and 2-oxachlorins, their facile derivatization to stable chlorin derivatives potentially bearing many kinds of side chains, and their optical properties are likely to encourage the further study and application of these intriguing chromophores.

Experimental Section

Theoretical

Calculations were carried out by using semiempirical MNDO-PSDCI molecular orbital theory⁴⁷³⁰ as well as SAC-CI theory.⁴⁸ The MNDO-PSDCI calculations used the AM1 Hamiltonian. The configuration interaction included all 64 singles and 2,080 doubles generated from the eight highest energy filled and eight lowest energy un-filled π -orbitals. Calculations were carried out with and without phenyl groups for comparative purposes. The SAC-CI calculations were carried out using a full double zeta D95 basis set,⁴⁹ which has yielded excellent results for large polyatomic chromophores.⁵⁰ Level two integral selection generated a CI basis sets of roughly ~9,500 singles and ~600,000 doubles. The SAC-CI calculations were carried out on modified structures where hydrogens replaced all of the phenyl groups. All calculations were carried out on structures minimized by using B3LYP/ 6-31G(d) density functional theory.

X-Ray Single Crystal Diffractometry

X-ray crystallographic analysis: Single crystals of $5aH_2$, 5aZn, $12H_2$, 13-OMe, and 15-II-Zn were coated in either Paratone®-N (Exxon) or Fomblin® oil, mounted on a pin and placed a goniometer head under a stream of nitrogen cooled to 100 K. The data were collected on a either a Bruker APEX ($5aH_2$, 5aZn, $12H_2$) or APEX2 (13-OMe, and 15-II-Zn) CCD diffractometer with Mo source K_{α} radiation ($\lambda = 0.71073$). The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. Data were corrected for absorption effects using the multi-scan method (SADABS) and the structure was solved and refined using the Bruker SHELXTL Software Package until the final anisotropic full-matrix, least squares refinement of F² converged. Data collection and structural parameters for the structure elucidations of $5aH_2$, 5aZn, $12H_2$, 13-OMe, and 15-II-Zn can be found in the ESI.

Materials and Instrumentation

meso-Tetraarylporphyrins $1H_2$ were synthesized according to the method of Adler.⁵¹ The metal complexes 1aZn, 1aNi, 1aPd, and 1aPt were prepared from free base *meso*-tetraphenylporphyrin $1aH_2$ as described in the literature.⁵²

Flash column chromatography was performed manually in glass columns or on an automated flash chromatography system, on normal-phase silica (solvents used are indicated; isocratic eluation modes).

meso-Tetrakis(4-*i*-propyl)-cis-2,3-dihydroxychlorin 7bH₂—Prepared in 31% overall yield (270 mg, 2.93×10^{-4} mol) from *meso*-tetrakis(4-*i*-propyl)phenylporphyrin (1bH₂) (1.00 g, 1.12×10^{-3} mol) according to a published general procedure.¹⁶ R_f (silica-CH₂Cl₂) = 0.22, (silica-CH₂Cl₂/1% MeOH) 0.35; ¹H NMR (400 MHz, CDCl₃, δ): 8.67 (d, ³*J*= 4.8 Hz, 2H), 8.50 (s, 2H), 8.33 (d, ³*J* = 4.8 Hz, 2H), 8.07–8.02 (m, 6H), 7.85 (d, 7.36 Hz, 2H), 7.61–7.54 (m, 8H), 6.39 (s, 2H), 3.24–3.17 (m, 6H), 1.52–1.48 (m, 24H), –1.74 (s, 2H, exchangeable with D₂O) ppm; UV-visible (CHCl₂) λ_{max} (log ε) 418 (5.18), 522 (4.20), 548 (4.06) 594 (3.75), 644 (4.40) nm; HR-MS (ESI+, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₅₆H₅₇N₄O₂ (MH⁺), 817.4482, found 817.4502.

meso-Tetrakis(4-trifluoromethyl)-cis-2,3-dihydroxychlorin 7fH₂—Prepared in 26% overall yield (270 mg, 2.93×10^{-4} mol) from *meso*-tetrakis(4-trifluoromethyl)phenylporphyrin (1fH₂) (1.00 g, 1.12×10^{-3} mol) according to a published general procedure.¹⁶ R_f (silica-CH₂Cl₂) = 0.28; ¹H NMR (400 MHz, CDCl₃, δ): 8.62 (d, ³J = 4.0 Hz, 2H), 8.43 (s, 2H), 8.30 (d, ³J = 4.0 Hz, 2H), 8.27–8.20 (m, 6H), 8.03–7.93 (m, 10H), 6.27 (s, 2H), 3.10 (s, 1H, exchangeable with D₂O), -1.90 (s, 2H, exchangeable with D₂O) ppm; UV-visible (CHCl₂) λ_{max} (log ε) 408 (5.18), 515 (4.12), 542 (4.07) 593 (3.75), 645 (4.39) nm; HR-MS (ESI+, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₄₈H₂₈F₁₂N₄O₂ (MH⁺) 921.2089, found 921.2128.

meso-Tetrakis(4-methoxy)-cis-2,3-dihydroxychlorin 7eH₂.²²—Prepared in 21% overall yield (220 mg, 2.86×10^{-4} mol) from *meso*-tetrakis(4-methoxy)phenylporphyrin (1eH₂) (1.00 g, 1.36×10^{-3} mol) according to a general procedure.¹⁶ R_f (silica-CH₂Cl₂/5% MeOH) = 0.49; ¹H NMR (400 MHz, CDCl₃, δ): 8.66 (d, ³*J* = 4.0 Hz, 1H), 8.50 (s, 1H), 8.33 (d, ³*J* = 4.0 Hz, 1H), 8.06 (br d, 8.0 Hz 3H), 7.84 (d, 8.0 Hz, 1H), 7.22 (d, 8.0 Hz, 4H), 6.37 (s, 1H), 4.05 (two overlapping s, 6H), 3.21 (s, 1H, exchangeable with D₂O), -1.76 (s, 1H, exchangeable with D₂O) ppm; UV-visible (CHCl₂) λ_{max} (log ε) 418 (5.27), 522 (4.16), 5.51 (4.24), 594 (3.92), 645 (4.38) nm; HR-MS (ESI+, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₄₈H₄₀N₄O₆ (MH⁺) 769.3021, found 769.2991.

General procedure for the preparation of porpholactones 5 by oxidative diol cleavage of 2,3-dihydroxychlorins 7—To a stirring solution of 7 (0.32×10^{-4} mol, for 7aH₂ 206 mg) in 25 mL THF was added 18-C-6 (28 mg, 10^{-4} mol, 0.33 eq). KMnO₄ (251 mg, 1.58 mmol, ~5 equiv) was added to the solution, and the mixture was allowed to react for 12 h at ambient temperature, and the reaction monitored by TLC. If needed, additional oxidant was added after 12 h until the starting material was exhausted. The solution was then filtered through a short plug of silica gel or Celite, and the filter cake washed with CH₂Cl₂ until the filtrate was colorless. The combined filtrates were evaporated to dryness by rotary evaporation. The product was purified by column chromatography (silica/CHCl₃), followed by crystallization.

Alternatively, dihydroxy chlorins 7 can be, under the same reaction conditions, reacted with 2-5 equiv of cetyltrimethylammonium permanganate in CH_2Cl_2 , followed by the workup described above, also providing excellent yields of 5.

meso-Tetrakisphenyl-3-oxo-2-oxaporphyrin (5aH₂)—Prepared as crystalline material in 75% isolated yield (150 mg) according to the general procedure from 7aH₂. Recrystallization from CHCl₃/EtOH. Our IR, UV-vis, LR-MS, ¹H NMR, and analytical data are essentially identical with those reported by Crossley.^{3,53} Supplemented by ¹³C, optical, and HR MS data, they are included here for the purpose of better comparison with the data reported for the reduction products below. R_f (silica-CCl₄/CH₂Cl₂ 1:1) = 0.50; ¹H NMR (400 MHz, CDCl₃, δ): 8.80 (dd, ³*J* = 5.2, ⁴*J* = 1.5 Hz, 1H), 8.77 (dd, ³*J* = 4.8, ⁴*J* = 1.5 Hz, 1H), 8.70 (dd, ${}^{3}J = 4.8$, ${}^{4}J = 1.5$ Hz, 1H), 8.60 (d, ${}^{3}J = 4.6$ Hz, 1H), 8.58 (dd, ${}^{3}J = 5.0$, ${}^{4}J =$ 1.7 Hz, 1H) 8.53 (d, ³*J* = 4.6 Hz, 1H), 8.10-8.16 (m, 6H), 7.97 (dd, *J* = 7.0, 1.7 Hz, 2H), 7.66-7.78 (m, 12), -1.82 (s, 1H, exchangeable with D₂O), -2.05 (s, 1H, exchangeable with D₂O); ¹³C NMR (100 MHz, CDCl₃, δ): 167.7, 157.1, 154.6, 154.2, 141.6, 141.5, 141.4, 139.4, 138.7, 138.4, 137.6, 137.0, 135.3, 134.5, 134.4, 134.3, 133.8, 132.7, 130.2, 130.1, 128.6, 128.4, 128.3, 128.2, 128.0, 127.9, 127.8, 127.7, 127.2, 127.1, 126.3, 125.7, 121.7, 119.4, 102.9 ppm; UV-vis (CHCl₃) λ_{max} (log ε) 420 (5.56), 522 (4.15), 558 (4.16), 588 (3.95), 640 (3.66); UV-visible (TFA/CHCl₃) λ_{max} 430, 588 (sh), 614; HR-MS (FAB, 3-NBA, quadrupole) *m/e* calc'd for C₄₃H₂₈N₄O₂: 632.2212, found 632.2148; Anal. calc'd for C₄₃H₂₈N₄O₂: C, 81.63: H, 4.46: N, 8.85%. Found: C, 81.44; H, 4.44; N, 8.80%.

[*meso*-Tetrakisphenyl-3-oxo-2-oxaporphyrinato]Zn(II) (5aZn)—Prepared either according to the general procedure by oxidation of 7Zn in 85% isolated yield or, in quantitative yields, by Zn(II)-insertion into 5H₂ using 2 h reflux in CHCl₃/EtOH containing 3 eq Zn(II)(OAc)₂·2H₂O, followed by slow solvent exchange CHCl₃ to EtOH on the rotary evaporator. The purple course crystals were filtered and air-dried. R_f (silica-CH₂Cl₂) = 0.19 or (silica-ethyl acetate/hexane 3:1) = 0.82; ¹H NMR (400 MHz, CDCl₃, δ): 8.75 (d, ³*J* = 4.5 Hz, 1H), 8.68, 8.67 (two overlapping d, ³*J* = 4.5 Hz, 2H), 8.63 (d, ³*J* = 4.5 Hz, 1H), 8.56 (d, ³*J* = 4.5 Hz, 1H), 8.51 (d, ³*J* = 4.5 Hz, 1H), 8.11 (d, *J* = 7.8, 2.0 Hz, 4H), 8.06 (dd, *J* = 7.8, 2.0 Hz, 2H), 7.93 (dd, *J* = 7.8, 2.0 Hz, 2H), 7.657.77 (m, 12H); ¹³C NMR (75 MHz, CDCl₃, δ): 173.5, 154.1, 143.4, 142.1, 142.0, 139.0, 138.0, 134.3, 134.0, 133.8, 132.4, 132.3, 132.2, 132.0, 130.9, 130.7, 130.6, 129.5, 128.8, 127.9, 127.8, 127.7, 127.6, 127.5, 126.7, 126.6; UV-visible (CH₂Cl₂) λ_{max} (log ε) 402 (sh), 422 (5.53), 520 (3.54), 558 (4.07), 602 (4.44); LR-MS (EI, 250°C) *m/e* 694 (30.9, M+), 638 (7.3), 561 (17.4), 483 (3.5), 28 (100); HR-MS (DART⁺, orifice voltage = 30 V, 100% CH₃CN, TOF) *m/e* calc'd for C₄₃H₂₇N₄O₂ ⁶⁴Zn (MH⁺): 695.1425, found 695.1429.

[*meso*-Tetraphenyl-3-oxo-2-oxaporphyrinato]Ni(II) (5aNi)—Prepared in 55% yield (10^{-4} mol scale) from 7aNi according to the general procedure. R^f (silica-CH₂Cl₂) = 0.74; ¹H NMR (400 MHz, CDCl₃/10% MeOD, δ): 8.55 (d, ³*J* = 4.0 Hz, 1H), 8.51–8.47 (two

overlapping d, ${}^{3}J$ = 4.0 Hz, 2H), 8.45–8.41 (two overlapping d, ${}^{3}J$ = 4.0 Hz, 2H), 8.33 (d, ${}^{3}J$ = 4.0 Hz, 1H), 7.90–7.85 (m, 6H), 7.74 (m, 2H), 7.66–7.60 (m, 12H). 13 C NMR (100 MHz, CDCl₃/10% MeOH-d₄, δ): 149.3, 144.9, 144.5, 141.4, 140.9, 139.9, 137.1, 136.0, 133.5, 133.3, 133.0, 131.8, 128.1, 127.7, 127.4, 127.0, 126.9, 124.9, 121.2, 120.3, 100.7; UV-visible (CHCl₂) λ_{max} (log ϵ) 415 (5.24), 543 (3.96), 586 (4.49) nm; HR-MS (ESI+, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₄₃H₂₆N₄⁵⁸NiO₂ (M⁺): 688.1409, found: 688.1426.

meso-Tetrakis(4-*i*-propylphenyl)-3-oxo-2-oxaporphyrin (5bH₂)—Prepared in 77% yield (13 mg) from 7bH₂ (2.0×10^{-5} mol, 17 mg) according to the general procedure. R_f (silica-CH₂Cl₂) = 0.90; ¹H NMR (400 MHz, CDCl₃, δ): 8.83, 8.81 (two overlapping dd, ³J = 4.5, ⁴J = 1.5 Hz, 2H), 8.74 (dd, ³J = 4.6, ⁴J = 1.3 Hz 1H), 8.63 (d, ³J = 4.5 Hz, 1H) 8.61, 8.59 (dd, ³J = 4.5, ⁴J = 1.5 Hz, 1H), 8.56 (d, ³J = 4.6 Hz, 1H), 8.07–8.02 (m, 6H), 7.90 (d, 8.0 Hz, 2H), 7.60–7.58 (m, 8H), 3.28–3.19 (m, 4H), 1.54–1.50 (m, 24H), -1.62 (s, 1H, exchangeable with D₂O) pcm; ¹³C NMR (100 MHz, CDCl₃, δ): 167.6, 156.9, 154.5, 154.1, 148.7, 148.59, 148.57, 148.54, 141.2, 139.3, 138.8, 138.7, 138.6, 136.9, 135.5, 135.0, 134.8, 134.7, 134.4, 134.29, 134.26, 133.5, 132.6, 129.9, 129.8, 127.7, 127.5, 126.0, 125.9, 125.6, 125.5, 125.0, 124.9, 121.5, 119.3, 102.7, 34.11, 34.06, 24.25, 24.19 ppm; UV-visible (CHCl₂) λ_{max} (log ε) 421 (5.46), 523 (4.01), 562 (4.09), 590 (3.81), 642 (3.10) nm; HR-MS (ESI+, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₅₅H₅₂N₄O₂, (MH⁺) 801.4169, found 801.4149.

meso-Tetrakis(4-*t*-butylphenyl)-3-oxo-2-oxaporphyrin (5cH₂)—Prepared in 73% yield (32 mg) from 7cH₂²² (5.0×10^{-5} mol, 43 mg) according to the general procedure. R_f (silica-CH₂Cl₂) = 0.89; ¹H NMR (400 MHz, CDCl₃, δ): 8.83 (s, 2H), 8.74 (d, ³*J* = 4.0 Hz, 1H), 8.63–8.59 (two overlapping d, ³*J* = 4.0 Hz, 2H), 8.56 (d, ³*J* = 4.0 Hz, 1H), 8.08–8.04 (m, 6H), 7.92–7.90 (d, 2H), 7.77–7.74 (m, 8H), 3.28–3.19 (m, 4H), 1.60, 1.59, 1.57 (3 s, 36H), -1.62 (s, 1H, exchangeable with D₂O), -1.99 (s, 1H, exchangeable with D₂O) ppm. ¹³C NMR (100 MHz, CDCl₃, δ): 167.6, 156.9, 154.3, 154.0, 151.0, 150.9, 150.8, 150.7, 141.2, 139.3, 138.5, 138.4, 138.3, 136.9, 135.1, 135.0, 134.4, 134.19, 134.09, 134.04, 133.4, 132.3, 129.8, 129.7, 127.7, 127.5, 126.0, 125.4, 124.7, 124.4, 123.9, 123.7, 121.4, 119.3, 102.7, 34.92, 34.91, 34.88, 31.66, 31.62 ppm; UV-visible (CHCl₂) λ_{max} (log ε) 421 (5.44), 523 (4.04), 562 (4.12), 590 (3.87), 642 (3.38) nm; HR-MS (ESI+, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₅₉H₆₁N₄O₂ (MH⁺) 857.4795, found 857.4787.

meso-Tetrakis(3,4,5-trimethoxyphenyl)-3-oxo-2-oxaporphyrin (5dH₂)—Prepared in 95% yield (114 mg) from dihydroxychlorin 7dH₂²² (121 mg, 1.2×10^{-4} mol) according to the general procedure. R^f (silica-CH₂Cl₂/5% MeOH) = 0.63; ¹H NMR (400 MHz, CDCl₃, δ): 8.92 (d, ³*J* = 4.0 Hz, 1H), 8.87-8.83 (m, 2H),), 8.72 (d ³*J* = 4.0 Hz, 2H),), 8.65 (d ³*J* = 4.0 Hz, 1H), 7.39 (d ³*J* = 8.0 Hz, 2H), 7.30 (s, 1H), 7.18 (s, 1H), 4.16-4.13 (m, 12H), 3.99-3.94 (m 24H), -1.69 (s, 1H, exchangeable with D₂O), -2.06 (s, 1H, exchangeable with D₂O) ppm; UV-visible (CHCl₂) λ_{max} (log Σ) 425 (5.63), 523 (4.32), 561 (4.33), 589 (4.15), 642 (3.82) nm; HR-MS (ESI+, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₅₅H₅₂N₄O₁₄ (MH⁺) 993.3558, found 993.3528.

[meso-Tetrakis(3,4,5-trimethoxyphenyl)-3-oxo-2-oxaporphyrinato]Ni(II) (5dNi)

—Prepared in 42% yield (17 mg) from **7dNi** (43 mg, 4.0×10^{-5} mol) according to the general procedure. Rf (silica-CH₂Cl₂/10% MeOH) = 0.73; ¹H NMR (400 MHz, CDCl₃, δ): 8.72 (d, ³*J* = 4.0 Hz, 1H), 8.69 (d, ³*J* = 4.0 Hz, 1H), 8.67 (d, ³*J* = 4.0 Hz, 1H), 8.61 (two overlapping d, ³*J* = 4.0 Hz, 2H), 8.51 (d, ³*J* = 4.0 Hz, 1H), 7.20 (s, 4H), 7.13 (s, 2H), 7.01(s, 2H), 4.11–4.08 (t, 12H), 3.96–3.90 (m, 24H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 164.2, 153.2, 152.5, 152.4, 151.9, 151.8, 149.5, 147.4, 145.3, 144.6, 142.1, 141.6, 140.9, 138.2,

138.1, 135.4, 135.3, 134.0, 133.9, 132.2, 131.9, 131.4, 130.9, 130.2, 124.9, 121.2, 120.3, 118.3, 117.8, 111.6, 111.5, 111.4, 110.0, 106.8, 100.8, 61.3, 61.2, 61.00, 56.41, 56.37, 56.28 ppm; UV-visible (CHCl₂) λ_{max} (log ϵ) 421 (5.18), 545 (3.90), 588 (4.46) nm; HR-MS (ESI +, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₅₅H₅₀N₄⁵⁸NiO₁₄ (MH⁺) 1048.2677, found 1048.2667.

meso-Tetrakis(4-methoxyphenyl)-3-oxo-2-oxaporphyrin (5eH₂)—Prepared in 95% yield (7.8 × 10⁻⁵ mol, 56 mg) from dihydroxychlorin 7eH₂²² (7.8 × 10⁻⁵ mol, 60 mg) according to the general procedure. R_f (silica–CH₂Cl₂) = 0.12; ¹H NMR (400 MHz, CDCl₃, δ): 8.80 (dd, ³*J* = 5.1, ⁴*J* = 1.4 Hz, 1H), 8.76 (dd, ³*J* = 5.2, ⁴*J* = 1.6 Hz, 1H), 8.72 (dd, ³*J* = 4.8, ⁴*J* = 1.7 Hz, 1H), 8.61 (d ³*J* = 4.7 Hz, 1H), 8.58 (dd, ³*J* = 4.8 Hz, ⁴*J* = 1.8 Hz, 1H), 8.53 (d, ³*J* = 4.6 Hz, 1H), 8.05-8.00 (m, 6H), 7.88 (m, 2H), 7.32–7.24 (m 8H), 4.04 (s, 12H) – 1.59 (s, 1H, exchangeable with D₂O), –1.95 (s, 1H, exchangeable with D₂O) ppm; UV-visible (CHCl₂) λ_{max} (log ε) 425 (5.66), 527 (4.27), 566 (4.38), 590 (4.18), 644 (3.60) nm; HR-MS (ESI+, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₄₇H₃₆N₄O₆ (MH⁺) 753.2696, found 753.2713.

[meso-Tetrakis(4-methoxyphenyl)-3-oxo-2-oxaporphyrinato]Pd(II) (5ePd)—

meso-Tetrakis (4-methoxyphenyl)-2-oxa-3-oxoporphyrinato **5eH**₂ (65.0 mg, 8.63×10^{-5} mol) was dissolved in PhCN (5 mL) and added to a refluxing solution of PhCN (20 mL) and PdCl₂ (61 mg, 3.44×10^{-4} mol, 4 equiv) in a round-bottom flask equipped with a magnetic stirring bar and N₂ gas inlet. The mixture was heated to reflux for 3 h. When the starting material was consumed (reaction control by UV-vis and TLC), the reaction mixture was allowed to cool and was evaporated to dryness by rotary evaporation. The resulting mixture was separated by column chromatography (silica-CH₂Cl₂). The Pd(II) complex was isolated in 71% (50 mg) yield as a magenta powder: R_f (silica-CH₂Cl₂) = 0.18; ¹H NMR (400 MHz, CDCl₃, δ): 8.67 (d, ³*J* = 5.2 Hz, 1H), 8.65 (d, ³*J* = 4.9 Hz, 1H), 8.64, 8.62, 8.60 (three overlapping d, ³*J* = 5.0 Hz, 3H), 8.50 (d, ³*J* = 4.7 Hz, 1H), 7.99–7.94 (m, 6H), 7.83 (dd, ³*J* = 8.7, ⁴*J* = 2.2 Hz, 2H), 7.27–7.23 (m, 8H), 4.07 (s 6H), 4.04 (d, 6H) ppm; UV-vis (CH₂Cl₂) λ_{max} (log ε) 425 (5.34), 500 (3.74), 538 (4.15), 580 (4.60) nm; HR-MS (DART⁺, orifice voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₄₇H₃₅N₄O₆Pd(II) ([MH]⁺) 857.1608, found 857.1646.

[meso-Tetrakis (4-methoxyphenyl)-3-oxo-2-oxaporphyrinato] Pt(II) (5ePt)-

Prepared in good yields 72% (58 mg) as a red powder as described for the Pd(II) complex from **5eH**₂ (65 mg, 8.63 × 10⁻⁵ mol) and PtCl₂ (46 mg, 1.73 × 10⁻⁴ mol, 2.0 equiv). R_f (silica-CH₂Cl₂) = 0.17; ¹H NMR (400 MHz, CD₂Cl₂, δ): 8.68–8.62 (m, 5H), 8.50 (d, ³*J* = 5.1 Hz, 1H), 7.97–7.93 (m, 6H), 7.82 (d, *J* = 6.9, 1.9 Hz, 2H), 7.29–7.22 (m, 8H), 4.03 (d, 7.8 Hz, 12H) ppm; UV-vis (CH₂Cl₂) λ_{max} (log ε) 421 (5.14), 527 (4.07), 568 (4.49) nm; HR-MS (DART⁺, orifice voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₄₇H₃₅N₄O₆Pt(II) ([MH]⁺) 946.2208, found 946.2217.

meso-Tetrakis(4-trifluoromethylphenyl)-3-oxo-2-oxaporphyrin (5fH₂)—Prepared in 73% yield (8.0×10^{-5} mol, 73 mg) from dihydroxychlorin 7fH₂ (1.1×10^{-4} mol, 100 mg) according to the general procedure. R_f (silica-CH₂Cl₂) = 0.91; ¹H NMR (400 MHz, CDCl₃, δ): 8.78, 8.76 (two overlapping dd, ³*J* = 5.0, ⁴*J* = 1.8 Hz 2H), 8.67 (dd, ³*J* = 4.9, ⁴*J* = 1.9 Hz, 1H), 8.54 (two overlapping d and dd, ³*J* = 4.6, ⁴*J* = 2.6 Hz, 2H), 8.47 (d, ³*J* = 4.7 Hz, 1H), 8.26–8.21 (m, 6H), 8.09–7.99 (m, 10H), –1.70 (s, 1H, exchangeable with D₂O), –2.06 (s, 1H, exchangeable with D₂O) ppm; UV-visible (CH₂Cl₂) λ_{max} (log ε) 416 (5.67), 518 (4.63), 555 (4.35), 588 (4.11), 641 (4.01) nm; HR-MS (ESI⁺, cone voltage = 30 V, 100% CH₃CN, TOF) *m/e* calcd for C₄₇H₂₄F₁₂N₄O₂²³Na (MNa⁺) 927.1605, found 927.1575.

[meso-Tetrakis(4-trifluoromethylphenyl)-3-oxo-2-oxaporphyrinato]Zn(II) (5fZn) —Prepared in near-quantitative yield by zinc(II) insertion into **5fH**₂ according to the method described for **5aZn**. MW= 968.06 g/mol; R_f (silica–CH₂Cl₂) = 0.72; ¹H NMR (300 MHz, CDCl₃, δ): 8.76 (d, ³*J* = 4.8 Hz, 1H), 8.71 (d, ³*J* = 4.8 Hz, 1H), 8.68 (m, 2H), 8.61 (d, ³*J* = 4.6 Hz, 1H), 8.49 (d, ³*J* = 4.6 Hz, 1H), 8.27 (d, ³*J* = 7.8 Hz, 4H), 8.22 (d, ³*J* = 7.9, 2H), 8.04 (m, 8H), 7.97 (d, ³*J* = 8.1 Hz, 2H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 166.2, 154.3, 152.4, 151.9, 151.2, 149.9, 148.1, 147.9, 145.5, 142.4, 141.5, 134.7, 134.3, 134.1, 132.9, 132.8, 132.6, 131.3, 131.1, 131.0, 130.9, 130.6, 130.5, 130.3, 130.1, 126.3, 126.1, 126.0, 125.2, 125.00, 124.97, 124.69, 124.67, 124.20, 124.16, 123.3, 121.0, 119.6, 101.3 ppm; UVvisible (CHCl₃) λ_{max} (log ε): 424 (5.54), 522 (3.64), 560 (4.13), 604 (4.49) nm; Fl λ_{max} (CHCl₃, $\lambda_{exc} = 424$ nm) 607, 660 nm, $\phi = 0.078$; HR–MS (ESI+ of MH⁺, 100% CH₃CN, TOF): *m/z* calc'd for C₄₇H₂₂F₁₂N₄O₂ ⁶⁴Zn: 966.0843, found 966.0857.

meso-Tetrakis(pentafluorophenyl)-3-oxo-2-oxaporphyrin (5gH₂)—This optimized procedure for the synthesis of this known porpholactone^{4,14,22} is based on the recent availability of the corresponding dihydroxychlorin $7gH_2^{36}$, and it the most efficient method of making this porpholactone reported to date: 5,10,15,20-Tetrakis(pentafluorophenyl)-2,3dihydroxychlorin **7gH**₂ (400 mg, 3.97×10^{-4} mol) was dissolved in CH₂Cl₂ (150 mL) in a 250 mL round-bottom flask equipped with a stir bar. 1 Equiv of cetyltrimethylammonium permanganate was added every 20 min over a course of 100 min (total addition 800 mg, 1.99×10^{-3} mol, 5 equiv). In between additions, the flask was stoppered, shielded from light with aluminum foil, and magnetically stirred at ambient temperature. The disappearance of the starting material/appearance of the product was monitored by TLC. The crude reaction mixture was absorbed onto silica by addition of ~10 g silica gel and evaporation of the solvent by rotary evaporation. The crude material loaded onto silica was purified via column chromatography (24 g silica-CH₂Cl₂/30% hexanes). The red low polarity product was collected and the solvent was removed by rotary evaporation. The product was re-dissolved in the minimal amount of CHCl₃ and crystallized by slow solvent exchange with EtOH on a rotary evaporator. The bright red product was isolated by filtration and air dried. Yield: 25-30% (125 mg 1.26×10^{-4} mol). Spectroscopic data as reported previously.^{4,14,22}

Alternative procedure using KMnO₄ heterogenized onto silica gel:³³ Either *meso*-Tetrakis(pentafluorophenyl)-2,3-dihydroxychlorin osmate ester (0.300 g, 2.16×10^{-4} mol) or *meso*-Tetrakis(pentafluorophenyl)-2,3-dihydroxychlorin **7gH**₂ (0.300 g, 2.98×10^{-4} mol) were dissolved in a 250 mL round-bottom flask equipped with a stir bar in CHCl₃ (100 mL). To the stirred porphyrinoid solution was added KMnO₄ heterogenized onto silica (5.19 g; corresponding to a ~15-fold stoichiometric excess of oxidant). The flask was stoppered, shielded from light with aluminum foil, and stirred at ambient temperature for 24 h. The crude mixture was filtered through Celite® to remove the KMnO₄-silica. The resulting solution was evaporated to dryness using to rotary evaporation and purified by column chromatography (silica, CH₂Cl₂/70% hexanes) to provide *meso*-tetrakis(pentafluorophenyl) porpholactone in 10-20% yield. *meso*-Tetrakis(pentafluorophenyl)-2,3-dihydroxychlorin osmate ester was recovered in 85% yield, the chlorin **7gH**₂ in 65% yield.

5,15-Diphenylporpholactones 15-I-H₂ and 15-II-H₂—In a 50 mLmL round bottom flask shielded from light with aluminum foil, diphenyldihydroxychlorin 14^{23} (50 mg, 5.7×10^{-5} mol) was dissolved in CHCl₃ (20 mL). The solution was stirred magnetically and cetytrimethylammonium permanganate (CTAP) was added (69 mg, 0.17 mmol, ~3 equiv) at ambient temperature. TLC was used to monitor the formation of a bright pink, non-polar spot. UV-visible spectroscopy was used to monitor the disappearance of the chlorin peak (at ~650 nm) and the formation of porphyrin-like peaks. The reaction was stirred until the full consumption of 14 was observed (~2 h). The products were then isolated by flash

chromatography (silica, CH₂Cl₂), providing an isomeric mixture (approximate ratio 5:1 by ¹H NMR; see ESI) of the diphenylporpholactones in high (~80%) yield. The isomers were separated by preparative TLC (silica-50% petroleum ether 30-60/CHCl₃) but compound **15-I-H**₂ could not be isolated in high purity in large enough quantity for its full characterization. For an alternative reaction using the osmate ester of **14**, see SI. **15-II-H**₂. R_f (silica-CH₂Cl₂) = 0.84; ¹H NMR (300 MHz, CDCl₃, δ): 10.10 (s, 1H), 10.03 (s, 1H), 9.34 (dd, ³J = 4.9 Hz, ⁴J = 1.8 Hz, 1H), 9.21 (d, J = 4.6 Hz, 1H), 9.09 (d, J = 4.5 Hz, 1H), 9.03 (dd, ³J = 4.8 Hz, ⁴J = 1.8 Hz, 1H), 8.96 (d, J = 8.0, 4.7 Hz 1H), 8.85 (d, J = 8.0, 4.5 Hz, 1H), 8.18-8.22 (m, 4 H), 7.77-7.85 (m, 6H), -1.99 (s, 1H), -2.58 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 170.2, 155.2, 154.9, 154.7, 141.4, 140.8, 137.6, 137.4, 137.0, 136.7, 135.4, 134.7, 134.6, 132.9, 131.8, 131.1, 129.9, 129.0, 128.4, 128.3, 128.25, 127.5, 127.3, 126.35, 124.3, 107.0, 102.6, 101.1 ppm; UV-visible (CHCl₃) λ_{max} (log ϵ): 409 (5.23), 511 (3.92), 549 (4.00), 581 (3.76), 633 (3.81) nm; HR-MS (ESI+, 100% CH₃CN, TOF): *m/z* calc'd for C₃₁H₂₁N₄O₂ (MH⁺): 481.1665, found 481.1629.

[5,15-Diphenylporpholactonato]Zn(II) (15-II-Zn)—To a stirring solution of **15-II-H**₂ (30 mg, 6.3×10^{-5} mol) in CHCl₃ (~5 mL), was added a solution of Zn(OAc)₂·4 H₂O in MeOH (27 mg, 1.3×10^{-4} mol, ~2 equiv). The mixture was heated to reflux for ~1 h. TLC was used to monitor the formation of a more polar green product. Upon completion, the solvents were evaporated using rotary evaporation and the product was isolated by column chromatography (silica, 1% MeOH/CH₂Cl₂) to provide **15-II-Zn** in near quantitative yield (33 mg). R_f (silica, 1% MeOH/CH₂Cl₂) = 0.46; ¹H NMR (300 MHz, CDCl₃, δ): 9.84 (s, 1H), 9.69 (s, 1H), 9.16 (d, *J* = 4.7 Hz, 1H), 9.08 (d, *J* = 4.5 Hz, 1H), 8.95 (d, *J* = 4.3 Hz, 1H), 8.83-8.87 (m, 3H), 8.08-8.15 (m, 4 H), 7.76-7.79 (m, 6H) ppm; UV-visible (CH₂Cl₂) λ_{max} (log ϵ): 416 (5.81), 514 (3.76), 553 (4.33), 597 (4.79) nm; MS (ESI+, 100% CH₃CN, 30 V cone voltage): *m/z* = 543.1 (MH⁺); HRMS (ESI+ of MH⁺, 100% CH₃CN, TOF): *m/z* calc'd for C₃₁H₁₉N₄O₂Zn: 543.0799, found 543.0833.

General procedure for the preparation of hemiacetals 11Zn by DIBAI-H

reduction of lactones 5Zn—Lactone 5Zn $(1.4 \times 10^{-4} \text{ mol}, ~95 \text{ mg for 5aZn})$ were dissolved under an atmosphere of N₂ in dry THF (20-25 mL) and cooled to -78° C. To it was added 20% DIBAl-H (1.0 mL of a 20 weight % solution in hexane, ~7.0 equiv). The reaction mixture stirred for 60 min at this temperature and then allowed to warm to ambient temperature. A noticeable color change from green to blue took place during this time. Once warm, the reaction was quenched by addition of a few drops of water. The solution was then transferred to a separatory funnel, diluted with CH₂Cl₂ (~25 mL), washed twice with 0.1 M aq. HCl, and once with H₂O. The organic layer was collected and dried over anhyd. MgSO₄. The solution was then evaporated to dryness by rotary evaporation and used as is or purified by preparative plate or column chromatography (Note: Care should be taken not to expose the hemiacetal **9Zn** to any alcohols or the corresponding acetal **10Zn** will be isolated).

[*meso*-Tetrakisphenyl-3-hydroxy-2-oxachlorinato]Zn(II) (11Zn)—Prepared as a purple powder in 80-90% isolated yields (180 mg) from porpholactol zinc complex 5aZn (200 mg, 0.287 mmol) according to the general procedure. R_f (silica– CH_2Cl_2) = 0.06; ¹H NMR (400 MHz, CDCl₃, δ): 8.61 (d, ³*J* = 4.4 Hz, 1H) 8.52 (d, ³*J* = 4.2 Hz, 1H), 8.42 (d, ³*J* = 4.0 Hz, 1H), 8.38 (d, ³*J* = 4.4 Hz, 1H), 8.34 (d, ³*J* = 4.0 Hz, 1H), 8.20 (brs, 1H), 8.12 (m, 2H), 8.04 (m, 2H), 7.73 (m, 8H), 7.50 (m, 9H), 7.26 (s, 1H), 3.02 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 190.7, 162.7, 187.3, 157.7, 153.9, 149.6, 145.3, 143.2, 140.6, 140.3, 139.3, 138.0, 137.2, 136.7, 136.6, 136.2, 135.0, 133.8, 133.2, 131.9, 129.2, 129.1, 128.5, 128.4, 128.3, 127.6, 127.4, 127.3, 126.2, 125.9, 125.7, 124.2, 121.6, 112.1 ppm; UV-visible (CH₂Cl₂) λ_{max} (log ϵ): 415 (5.37), 453 (3.87), 513 (3.75), 538 (3.70), 580 (3.97), 615

(4.60), 795 (3.37) nm; Fl (CH₂Cl₂, $\lambda_{excitation} = 420$ nm) λ_{max} : 621, 672 nm; HR-MS (FAB+ of M⁺, PEG, quadrupole): *m/z* calc'd for C₄₃H₂₈O₂N₄ ⁶⁴Zn: 696.1504, found 696.1519.

meso-Tetrakisphenyl-3-hydroxy-2-oxachlorins (11H₂)—Prepared in > 90% yields (up to 500 mg scale) by demetallation of **11Zn**. The dilute HCl wash in the general procedure for the preparation of 11Zn was replaced with a wash of half-concentrated aqueous HCl, followed by several washes with H₂O saturated aq. Na₂CO₃ solution, and again H₂O. The organic layer was dried over anhyd. MgSO₄, evaporated to dryness and purified by preparative plate or column chromatography (Note: Care should be taken not to expose the hemiacetal 11H₂ to any alcohols or the corresponding acetals 13H₂ will be isolated). R_f (silica–CH₂Cl₂) = 0.54; ¹H NMR (400 MHz, CDCl₃, δ): 8.58 (d, ³J = 4.5 Hz, 1H), 8.50 (d, ${}^{3}J$ = 4.5 Hz, 1H), 8.42 (t, ${}^{3}J$ = 4.5 Hz, 3.51, 2H), 8.34 (d, ${}^{3}J$ = 4.0 Hz, 1H), 8.17 (d, ${}^{3}J$ = 4.5 Hz, 1H), 8.10 (m, 5H), 7.87 (t, ${}^{3}J$ = 5.8 Hz, 7.77, 2H), 7.71 (brs, 13H), 3.83 (d, ${}^{3}J$ = 7.2 Hz, 1H), -0.76 (s, 1H), -1.13 (s, 1H) ppm; ${}^{13}C$ NMR (100 MHz, CDCl₃, δ): 164.3, 155.1, 152.0, 151.6, 142.0, 136.9, 135.1, 134.1, 133.9, 133.7, 131.9, 131.5, 129.9, 128.2, 128.2, 128.1, 128.0, 127.9, 127.7, 127.1, 127.0, 125.3, 122.1, 112.2, 100.2 ppm; UVvis $(CH_2Cl_2) \lambda_{max}$ (log ϵ): 416 (5.26), 515 (4.11), 550 (4.18), 592 (3.87), 646 (4.51) nm; Fl $(CH_2Cl_2, \lambda_{excitation} = 420 \text{ nm}) \lambda_{max}$: 651, 704 nm; +ESI-MS (cone voltage 70 V, 100%) CH₃CN): m/z = 635 (MH⁺); HR-MS (FAB+ of MH⁺, PEG, qudrupole): m/z calc'd for C₄₃H₃₁O₂N₄: 635.2447, found 635.2435; Anal. calc'd for C₄₃H₃₀N₄O₂: C, 81.37: H, 4.76: N, 8.83%. Found: C, 80.72; H, 4.94; N, 8.70%.

General procedure for the conversion of hemiacetals 11 to MeO-based acetals 13—Excess MeOH was added to a stirring solution of $11H_2$ or 11Zn in CH₂Cl₂. Traces of TFA vapors (from a TFA bottle head space, delivered via pipette) were added, and the reaction was monitored by TLC for completion. The acid was then neutralized with Et₃N (1 drop), the solution washed, dried over anhyd. MgSO₄, evaporated to dryness, and purified by column or preparative plate chromatography.

[*meso*-Tetrakisphenyl-3-methoxy-2-oxachlorinato]Zn(II) (13-OMeZn)—Prepared according to the general procedure from 11aZn and MeOH in 85% yields: R_f (silica–CH₂Cl₂) = 0.54; ¹H NMR (400 MHz, CDCl₃) δ 3.32 (s, 3H), 7.56 (s, 1H), 7.66 (m, 12 H), 7.82 (m, 1H), 7.87 (brs, 1H), 8.00 (m, 3H), 8.10 (m, 3H), 8.12 (d, ${}^{3}J$ = 4.6 Hz, 1H), 8.31 (d, ${}^{3}J$ = 4.4 Hz, 1H), 8.34 (d, ${}^{3}J$ = 4.6 Hz, 1H), 8.38 (d, ${}^{3}J$ = 4.4 Hz, 1H), 8.48 (d, ${}^{3}J$ = 4.4 Hz, 1H), 8.52 (d, ${}^{3}J$ = 4.6 Hz, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃) δ 162.8, 156.2, 154.5, 146.8, 142.6, 139.6, 134.7, 134.3, 134.0, 134.0, 133.8, 131.5, 130.6, 128.7, 128.0, 128.0, 127.9, 127.6, 127.6, 127.2, 127.1, 126.2, 123.4, 113.6, 106.5, 99.4, 55.2 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ε): 419 (5.23), 519 (3.60), 572 (shoulder), 618 (4.53) nm; Fl (CH₂Cl₂, λ_{excitation} = 420 nm) λ_{max}: 621, 675 nm; HR-MS (FAB+ of M⁺, PEG, quadrupole): *m/z* calc'd for C₄₄H₃₀O₂N₄ ⁶⁴Zn: 710.1660, found 710.1673.

meso-Tetrakisphenyl-2-oxachlorin (12H₂)—To 11H₂ (30 mg, 4.8×10^{-2} mmol) dissolved in dry CH₂Cl₂ under N₂ and stirred at room temperature was added excess Et₃SiH (125 µL, 16 equiv) and excess BF₃·OEt₂ (250 µL, 30 equiv). The reaction proceeded for 10 min. The reaction can also be catalyzed by Amberlyst 15 (600 mg) but then takes 12 h for completion. The solution was then washed twice with a concd. aq. NaHCO₃ solution, dried over anhyd. MgSO₄, and evaporated to dryness by rotary evaporation. Flash chromatography (silica/CH₂Cl₂) was used to isolate and purify the product. It was precipitated by slow solvent exchange with cyclohexane to produce 12H₂ as a purple powder in near-quantitative yields. The product undergoes spontaneous (photo)oxidations over time. R_f (silica–CH₂Cl₂) = 0.98; ¹H NMR (400 MHz, CDCl₃, δ): 8.43 (dd, ³*J* = 5.0, ⁴*J* = 1.6 Hz, 1H), 8.38 (dd, ³*J* = 4.8, ⁴*J* = 1.8 Hz, 1H), 8.28 (d, ³*J* = 4.4 Hz, 1H), 8.22 (dd, ³*J* =

5.0, ${}^{4}J$ = 1.8 Hz, 1H), 8.20 (d, ${}^{3}J$ = 4.4 Hz, 1H), 8.07 (m, 4H), 7.98 (m, 3H), 7.83 (dd, ${}^{3}J$ = 7.8, ${}^{4}J$ = 1.6 Hz, 2H), 7.69 (m, 12H), 6.54 (s, 2H), 0.15 (s, 1H), -0.25 (s, 1H) ppm; ${}^{13}C$ NMR (100 MHz, CDCl₃, δ): 170.1, 156.0, 154.5, 151.6, 144.0, 142.0, 133.9, 133.8, 133.7, 132.8, 131.9, 131.2, 130.0, 129.0, 128.3, 128.2, 128.1, 128.0, 127.9, 127.7, 127.1, 127.0, 124.2, 107.9, 99.0, 76.4 ppm; UV-vis (CH₂Cl₂) λ_{max} (rel. intensity): 373 (shoulder), 422 (1.00), 519 (0.07), 554 (0.06), 612 (0.04), 668 (0.26) nm; Fl (CH₂Cl₂, $\lambda_{excitation}$ 420 nm) λ_{max} : 674, 732 (sh) nm; HR-MS (ESI+, 100% CH₃CN, TOF) expected for C₄₃H₃₁N₄O (MH⁺): 619.2492, found 619.2477.

[*meso*-Tetrakisphenyl-2-oxachlorinato]Zn(ll) (12Zn)—To 12H₂ (30 mg, 4.8×10^{-2} mmol) dissolved in dry CH₂Cl₂ in DMF was added Zn(OAc)₂·2H₂O (2 equiv). The solution was warmed to 80 °C and metalation was monitored by TLC. Upon completion, the solution was evaporated to dryness under high vacuum. The residue was dissolved in CH₂Cl₂, and flash chromatography (silica-CH₂Cl₂) was used to separate the product. The product was evaporated to dryness to yield a green-purple product in 90% yield. The product undergoes spontaneous (photo)oxidations. ¹H NMR (400 MHz, CDCl₃, δ): 8.31 (d, ³*J* = 4.8 Hz, 1H), 8.29 (d, ³*J* = 4.6 Hz, 1H), 8.17 (d, *J* = 4.5 Hz, 1H), 8.09 (d, ³*J* = 4.5 Hz, 1H), 8.07 (d, ³*J* = 4.8 Hz, 1H), 7.97–8.01 (m, 3H), 7.89 (m, 2H), 7.87 (m, 1H), 7.86 (d, ³*J* = 4.5 Hz, 1H), 7.77 (dd, *J* = 7.8, 1.6 Hz, 2H), 7.62-7.67 (m, 12H), 6.54 (s, 2H) ppm; UV-vis (CH₂Cl₂) λ_{max} (rel. intensity): 420 (1.00), 520 (0.02), 591 (0.04), 640 (0.19); Fl (CH₂Cl₂, $\lambda_{excitation}$ 420 nm) λ_{max} : 651, 670, 699 nm; HR-MS (ESI+, 100% CH₃CN, TOF) expected for C₄₃H₂₉N₄O⁶⁵Zn (MH⁺): 681.1627, found 681.1619.

meso-Tetrakisphenyl-3-isopropoxy-2-oxachlorin (13-OⁱPr)—General procedure for the conversion of hemiacetals 11 to acetals 13. Isopropanol (1 mL) was added to a stirring solution of 11H₂ (11.5 mg, 1.9×10^{-5} mol) in CHCl₃ (3 mL) at room temperature. Traces of TFA vapors (from a TFA bottle head space, delivered via pipette) were added, and the reaction was monitored by TLC. The reaction was complete within in 3 h. Upon completion, the acid was neutralized with Et₃N (1 drop), the solution washed, dried over anhydrous MgSO₄, evaporated to dryness using rotary evaporation, and purified by flash column chromatography (DCM) or preparative plate. Yield >95% (12 mg). R_f (silica–CH₂Cl₂) = 0.96; ¹H NMR (300 MHz, CDCl₃, δ): 8.60 (d, ³*J* = 4.2 Hz, 1H), 8.51 (d, ³*J* = 4.2 Hz, 1H), 8.47 (d, ³*J* = 4.2 Hz, 1H), 8.43 (d, ³*J* = 4.2 Hz, 1H), 8.35 (d, ³*J* = 4.2 Hz, 1H), 8.04–8.19 (m, 7H), 7.89 (br, 1H), 7.66–7.73 (m, 14H), 3.96 (m, 1 H), 1.27 (d, ³*J* = 6.0 Hz, 3H), 0.91 (d, ³*J* = 6.0 Hz, 3H), -0.73 (s, 1H), -1.09 (s, 1H) ppm; UV-vis (CH₂Cl₂) λ_{max} (log ϵ): 418 (5.38), 516 (4.21), 550 (4.26), 593 (3.97), 647 (4.63) nm; LR-MS (ESI+, 100% CH₃CN, 30 V cone voltage, TOF): *m/z* 677.1 (MH⁺), 634.7 (MH⁺ –C₃H₇); HR-MS (ESI+ of M⁺, 100% CH₃CN): *m/z* calc'd for C₄₆H₃₆N₄O₂: 677.2917, found 677.3018.

meso-Tetrakisphenyl-3-methoxy-2-oxachlorin (13-OMe)—Prepared according to the general procedure from $11H_2$ (10 mg, 0.016 mmol) and methanol (1 mL) in nearquantitative yield. Spectroscopic properties as described previously.⁸

meso-Tetrakisphenyl-3-cyclohexanoxy-2-oxachlorin (13-O^cHex)—Prepared according to the general procedure from $11H_2$ (10 mg, 0.016 mmol) and cyclohexanol (1 mL) in >95% isolated yields 11 mg): R_f (silica–CH₂Cl₂) = 0.98; ¹H NMR (300 MHz, CDCl₃, δ): 8.59 (dd, ³*J* = 4.9, ⁴*J* = 1.5Hz, 1H) 8.51 (dd, ³*J* = 3.8, ⁴*J* = 1.7Hz, 1H), 8.46 (dd, ³*J* = 4.9, ⁴*J* = 1.7Hz, 1H), 8.43 (d, ³*J* = 4.5 Hz, 1H), 8.35 (d, ³*J* = 4.5 Hz, 1H), 7.88–8.19 (m, 8H), 7.69–7.72 (m, 13H), 3.59–3.66 (m, 1H), 0.89–1.72 (m, 10H), -0.71 (s, 1H), -1.08 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 165.5, 154.9, 151.8, 151.4, 142.9, 142.1, 142.0, 141.1, 140.1, 139.1, 136.7, 135.3, 134.5, 134.0, 133.9, 131.7, 131.1, 129.7, 127.9, 127.85, 127.8, 127.7, 127.6, 127.5, 127.0, 126.9, 125.9, 125.0, 121.9, 121.3, 112.2, 105.1, 100.4,

79.4, 33.7, 31.9, 25.8, 24.1 ppm; UV-visible (CH₂Cl₂) λ_{max} (log ϵ): 418 (5.31), 516 (4.14), 550 (4.19), 594 (3.88), 647 (4.55) nm; HR-MS (ESI+ of M⁺, 100% CH₃CN, TOF): m/z calc'd for C₄₉H₄₁N₄O₂: 717.3230, found 717.3212.

meso-Tetrakisphenyl-3-tert-butoxy-2-oxachlorin (13-O^tBu)—Prepared according to the general procedure from 11H₂ (12 mg, 0.019 mmol) and *tert*-butanol (1 mL) in 80% isolated yields (9.8 mg): R_f (silica–CH₂Cl₂) = 0.92; ¹H NMR (300 MHz, CDCl₃, δ): 8.58 (d, ³*J* = 4.7 Hz, 1H), 8.44–8.49 (m, 2H), 8.41 (d, ³*J* = 4.5 Hz, 1H), 8.34 (d, ³*J* = 4.5 Hz, 1H), 8.03–8.19 (m, 8H), 7.84–7.89 (m, 2H), 7.63–7.72 (m, 12H), 1.09 (s, 9H), –0.71 (s, 1H), – 1.07 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 165.3, 154.8, 152.1, 151.7, 142.9, 142.2, 142.1, 142.0, 141.2, 140.4, 139.1, 136.7, 135.5, 134.4, 134.3, 134.1, 134.0, 133.9, 133.3, 131.6, 131.4, 129.6, 128.0, 127.96, 127.92, 127.8, 127.7, 127.6, 127.54, 127.51, 127.0, 126.9, 125.9, 125.0, 121.7, 121.2, 112.0, 105.8, 100.6, 100.4, 64.8, 28.5 ppm; UV–visible (CH₂Cl₂) λ_{max} (log ε): 418 (5.20), 514 (4.05), 550 (4.08), 594 (3.79), 647 (4.46) nm; HR-MS (DART⁺, 20 V orifice voltage, 100% CH₃CN, TOF): *m*/*z* calc'd for C₄₇H₃₉N₄O₂ (MH⁺): 691.3037, found 691.3056.

[meso-Tetrakisphenyl-3-octoxy-2-oxachlorin] (13-OⁿOct)—Prepared according to the general procedure from **11H**₂ (10 mg, 0.016 mmol) and *n*-octanol (1 mL) in 92% isolated yields (11.4 mg): R_f (silica– CH_2Cl_2) = 0.92; ¹H NMR (300 MHz, CDCl₃, δ): 8.59 (dd, ³*J* = 4.5, ⁴*J* = 1.5 Hz, 1H), 8.52 (dd, ³*J* = 4.6, ⁴*J* = 1.7 Hz, 1H), 8.45 (dd, ³*J* = 5.8, ⁴*J* = 1.7 Hz, 1H), 8.43 (d, ³*J* = 4.6 Hz, 1H), 8.35 (d, ³*J* = 4.5 Hz, 1H), 8.21 (dd, ³*J* = 5.8, ⁴*J* = 1.7Hz, 1H), 7.85–8.18 (m, 8H), 7.64–7.77 (m, 12H), 7.59 (s, 1H), 3.64–3.67 (m, 1H), 3.42–3.45 (m, 1H), 1.24–1.28 (m, 17H), –0.73 (s, 1H), –1.09 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃) δ 165.0, 154.9, 151.9, 151.0, 143.0, 142.1, 142.0, 140.9, 139.9, 139.0, 136.8, 135.0, 134.5, 134.2, 134.1, 134.0, 133.9, 133.5, 131.8, 131.2, 129.7, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 127.65, 127.0, 126.9, 125.9, 125.1, 121.9, 121.4, 112.2, 105.9, 100.3, 69.4, 32.0, 29.9, 29.5, 29.4, 26.3, 22.9, 14.3 ppm; UV-visible (CH₂Cl₂) λ_{max} (log ϵ): 417 (5.23), 515 (4.06), 550 (4.11), 593 (3.79), 646 (4.48) nm; HR-MS (ESI+ of M⁺, 100% CH₃CN, TOF): *m/z* calc'd for C₅₁H₄₇N₄O₂: 747.3699, found 747.3705.

meso-Tetrakisphenyl-3-(+)cholesteroxy-2-oxachlorin (13-O-Chol)—Prepared according to the general procedure from $11H_2$ (33.9 mg, 0.053 mmol) and cholesterol (20.7 mg, 2 eq.) in 88% isolated yield (47 mg): R_f (silica–CH₂Cl₂) = 0.96; ¹H NMR (300 MHz, CDCl₃, δ): 8.63 (d, ³*J* = 4.8 Hz, 1H), 8.55 (m, 1H), 8.51 (d, ³*J* = 1.5 Hz, 1H), 8.47 (d, ³*J* = 4.5 Hz, 1H), 8.39 (d, ${}^{3}J$ = 4.5 Hz, 1H), 8.10–8.25 (m, 8H), 7.9 (d, ${}^{3}J$ = 0.3 Hz, 2H), 7.66– 7.76 (m, 13H), 5.29–5.36 (m, 1H), 3.55–3.63 (m, 1H), 2.32–2.64 (m, 1H), 2.12–1.72 (m, 6H), 1.28-1.62 (m, 15H), 0.99-1.22 (m, 12H), 0.96 (s, 3H), 0.98 (s, 3H), 0.71 (s, 3H), -0.74 (s, 1H), -1.06 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 165.1, 165.0, 155.0, 154.9, 151.9, 151.2, 151.1, 143.0, 142.2, 142.1, 141.2, 141.1, 141.0, 140.2, 140.1, 139.1, 139.0, 136.8, 136.7, 135.4, 135.3, 134.6, 134.2, 134.1, 134.0, 133.5, 131.8, 131.1, 131.0, 129.8, 128.2, 128.1, 128.0, 128.0, 127.9, 127.9, 127.9, 127.7, 127.6, 127.1, 126.9, 126.0, 125.1, 125.1, 122.1, 122.0, 122.0, 121.9, 121.4, 121.4, 112.2, 105.3, 105.2, 100.5, 100.4, 81.1, 81.0, 57.0, 56.4, 50.3, 42.6, 40.8, 40.0, 39.8, 38.6, 37.6, 37.4, 36.9, 36.8 36.5, 36.1, 32.3, 32.2, 32.1, 30.1, 28.5, 28.3, 28.2, 24.6, 24.1, 23.1, 22.9, 21.4, 21.3, 19.6, 19.0, 12.1 ppm; UV-visible (CH₂Cl₂) λ_{max} (log ϵ): 419 (5.24), 515 (4.16), 552 (4.18), 594 (3.95) 655 (3.79) nm; HR-MS (ESI+, 100% CH₃CN, TOF): *m/z* calc'd for C₇₀H₇₅N₄O₂ (MH+): 1003.5890, found 1003.5893.

meso-Tetrakisphenyl-3-pregnenolonoxy-2-oxachlorin (13-O-Preg)—Prepared according to the general procedure from $11H_2$ (33.5 mg, 0.053 mmol) and pregnenolone (33.4 mg, 2 equiv) in 83% isolated yield (41 mg): R_f (silica– CH_2Cl_2) = 0.55; ¹H NMR (300

MHz, CDCl₃, δ): 8.61 (d, ³*J* = 4.5 Hz, 1H), 8.53 (d, ³*J* = 4.5 Hz, 1H), 8.47 (t, ³*J* = 4.5 Hz, 1H), 8.44 (d, ³*J* = 4.5 Hz, 1H), 8.36 (d, ³*J* = 4.5 Hz, 1H), 7.89–8.22 (m, 9H), 7.65–7.74 (m, 13H), 5.30 (dd, ³*J* = 23.3 Hz, 4.5 Hz, 1H), 3.55–3.61 (m, 1H), 2.29–2.62 (m, 2H), 2.20 (m, 1H), 2.14 (s, 3H), 2.02–2.013 (m, 3H), 1.04–1.94 (m, 14H), 0.96 (s, 3H), 0.63 (s, 3H), -0.74 (s, 1H), -1.08 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃) δ 209.8, 151.9, 142.9, 142.1, 142.0, 141.1, 141.1, 141.0, 140.9, 136.8, 136.7, 135.4, 135.3, 134.5, 134.2, 134.1, 134.0, 133.5, 131.8, 131.1, 131.0, 129.8, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 127.1, 126.9, 126.0, 125.1, 125.0, 122.0, 121.9, 121.7, 121.6, 112.2, 105.3, 105.2, 100.4, 80.9, 80.7, 63.9, 57.1, 50.1, 44.2, 40.7, 39.0, 38.5, 37.5, 37.4, 36.9, 36.8, 32.1, 31.8, 30.0, 28.1, 24.7, 23.0, 21.3, 21.2, 19.6, 19.5, 13.4 ppm; UV-visible (CH₂Cl₂) λ_{max} (log ϵ): 419 (5.28), 517 (4.13), 550 (4.17), 593 (3.88) 647 (4.52) nm; HR-MS (ESI+, 100% CH₃CN, TOF): *m/z* calc'd for C₆₄H₆₁N₄O₃ (MH⁺): 933.4744, found 933.4733.

meso-Tetrakisphenyl-3-ethioxy-2-oxachlorin (13-SEt)—General procedure for the conversion of hemiacetals 11H₂ to RS-based thiaacetals 13-SR. Excess ethanethiol (1 to 2 mL) was added to a stirring solution of 11H₂ (10 mg, 0.015 mmol) in CHCl₃ (3-5 mL) at room temperature. Traces of TFA vapors (from a TFA bottle head space, delivered via pipette) were added, and the reaction was monitored by TLC. The reaction was complete within in 3 to 5 h. Upon completion, the acid was neutralized with Et₃N (1 drop), the solution washed, dried over anhydrous MgSO₄, evaporated to dryness using rotary evaporation, and purified by silica gel flash column or preparative plate chromatography (CH_2Cl_2) . Isolated yield 90% (9.6 mg): R_f (silica– CH_2Cl_2) = 0.96; ¹H NMR (300 MHz, CDCl₃, δ): 8.50-8.52 (m, 1H), 8.43-8.44 (m, 1H), 8.33-8.35 (m, 2H), 8.26-8.27 (m, 1H), 7.99 (m, 8H), 7.90–7.91 (m, 1H), 7.67–7.80 (m, 12H), 2.31–2.41 (m, 1H), 2.12–2.19 (m, 1H), 0.89–0.93 (m, 3H), -0.30 (s, 1H), -0.65 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 165.9, 154.9, 153.1, 151.9, 143.4, 141.9, 141.8, 141.4, 139.9, 138.7, 136.8, 135.5, 134.6, 134.1, 133.9, 133.8, 133.3, 131.6, 131.1, 129.9, 128.5, 128.2, 128.0, 127.92, 127.94, 127.97, 127.8, 127.7, 127.1, 126.95, 126.9, 126.3, 124.9, 121.8, 110.6, 100.1, 90.8 ppm; UVvisible $(CH_2Cl_2) \lambda_{max}$ (log ϵ): 420 (5.31), 457 (4.43), 518 (4.13), 555 (4.07), 608 (3.88), 660 (4.51) nm; LR-MS (ESI+, 100% CH₃CN, 30 V cone voltage): m/z 679.1 (MH⁺), upon exposure to ambient light and environment, significant peak of 695.7 (MHO⁺) is observed; HR-MS (ESI +, 100% CH₃CN, TOF): *m/z* calc'd for C₄₅H₃₅N₄OS (MH⁺): 679.2532, found 679.2471.

meso-Tetrakisphenyl-3-hexanethioxy-2-oxachlorin (13-SⁿHex)—Prepared according to the general procedure from $11H_2$ (10 mg 0.014 mmol) and hexanethiol (1 mL) in 50% isolated yields (5.8 mg): R_f (silica – CH₂Cl₂) = 0.92; ¹H NMR (300 MHz, CDCl₃, δ): 8.50–8.51 (m, 1H), 8.42–8.43 (m, 1H) 8.33–8.34 (m, 2H), 8.26–8.27 (m, 1H), 7.98–8.23 (m, 8H), 7.89–7.91 (m, 1H), 7.65–7.75 (m, 13H), 2.35–2.42 (m, 1H), 2.15–2.18 (m, 1H), 0.71–1.28 (m, 13H), -0.30 (s, 1H), -0.65 (s, 1H) ppm; UV-visible (CH₂Cl₂) λ_{max} (log ϵ): 422 (5.17), 518 (3.99), 553 (3.93), 603 (3.65), 659 (4.35) nm; HR-MS (ESI+, 100% CH₃CN, TOF): *m*/*z* calc'd for C₄₉H₄₃N₄OS (MH⁺): 735.3153, found 735.3097.

meso-Tetrakisphenyl-3-N-morpholinyl-2-oxachlorin (13-N^{morph})—General procedure for the conversion of hemiacetals 11 to aminals. A small-scale Soxhlet containing 3 Å molecular sieves was attached to a 50 mL round bottom flask. Excess amine (5 to 10 eq) was then added to a stirring solution of $11H_2$ (30 mg, 0.043 mmol) in benzene (10-15mL). A few drops of TFA were added and the mixture was refluxed for several days (3-5 days). The reaction progress was monitored by TLC and upon completion, the acid was neutralized with Et₃N (1 drop), dried over anhydrous MgSO₄, evaporated to dryness using rotary evaporation, and purified by flash column chromatography (CH₂Cl₂) or preparative plate to give 13-N^{morph} in 30% isolated yield (9 mg): R_f (silica–CH₂Cl₂) = 0.60; ¹H NMR (300 MHz, CDCl₃, δ): 8.52–8.54 (m, 1H), 8.45–8.47 (m, 1H), 8.36–8.37 (m, 2H), 8.28–8.29

(m, ¹H), 7.84–8.15 (m, 10H), 7.64–7.70 (m, 11H), 7.44 (s, 1H), 3.47–3.48 (m, 2H), 3.29–3.30 (m, 2H), 2.58–2.59 (m, 2H), 2.38–2.43 (m, 2H), –0.44 (s, 1H), –0.86 (s, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃, δ): 166.5, 154.9, 151.6, 150.6, 143.4, 142.0, 141.9, 141.2, 140.5, 139.2, 136.8, 135.2, 134.2, 134.0, 133.9, 133.8, 133.3, 131.5, 130.1, 129.7, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 127.2, 127.1, 126.93, 126.97, 126.1, 125.1, 121.6, 121.2, 111.6, 101.2, 99.7 ppm; UV-visible (CH₂Cl₂) λ_{max} (log ε): 420 (5.08), 517 (3.91), 552 (3.91), 598 (3.16) 654 (4.34) nm; HR-MS (ESI+, 100% CH₃CN, TOF): *m/z* calc'd for C₄₇H₃₈N₅O₂ (MH⁺): 704.3026, found 704.3009.

meso-Tetrakisphenyl-3-N-dibenzylamine-2-oxachlorin (13-N(Bn)₂)—Prepared according to the general procedure from $11H_2$ (10.7 mg, 0.017 mmol) and dibenzylamine (1 mL) in 70% isolated yield (9.6 mg): R_f (silica–CH₂Cl₂) = 0.96; ¹H NMR (300 MHz, CDCl₃, δ): 8.54–8.56 (m, 1H), 8.39–8.44 (m, 2H), 8.34–8.35 (m, 1H), 8.26–8.28 (m, 1H), 7.98–8.15 (m, 7H), 7.84–7.89 (m, 1H), 7.71–7.71 (m, 12H), 7.49–7.55 (m, 1H), 7.28 (s,1H), 7.11–7.12 (br, 5H), 6.84–6.86 (m, 4H), 3.47–3.61 (m, 4H), -0.33 (s, 1H), -0.70 (s, 1H) ppm; UV-visible (CH₂Cl₂) λ_{max} (log ε): 420 (5.16), 518 (3.97), 555 (3.97), 600 (3.69) 655 (4.39) nm; HR-MS (ESI+, 100% CH₃CN, TOF): *m/z* calc'd for C₅₇H₄₄N₅O (MH⁺): 814.3546, found 814.3590.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This work was supported by the US National Science Foundation (CHE-0517782 and CHE-1058846 to CB). The 400 MHz NMR used was supported by the NSF (CHE-1048717). The diffractometer at YSU was funded by NSF grant 0087210, by Ohio Board of Regents grant CAP-491, and by YSU. The theoretical studies were supported by grants from the National Institutes of Health (GM-34548) and the National Science Foundation (EMT-08517; both to RRB). CJZ acknowledges the National Science Foundation (CHE-0840446) for funds used to purchase the diffractometer at Akron used in this work. We thank Renee Rubenstein, Eileen Meehan, Alison Rossi (supported by a UConn SURF grant), Douglas Mooney (supported by NSF-REU grant CHE-0754580), John Haskoor (supported by a UConn SURF grant), and Kim Panther (supported by NSF-REU grant CHE-1062946) for experimental assistance.

References

- For a representative synthetic light-harvesting systems, see, e.g.: For Tomizaki K, Loewe RS, Kirmaier C, Schwartz JK, Retsek JL, Bocian DF, Holten D, Lindsey JS. J. Org. Chem. 2002; 67:6519–6534. and references therein. [PubMed: 12201776]
- (2). Lindsey, JS. Porphyrin Handbook. Kadish, KM.; Smith, KM.; Guilard, R., editors. Vol. Vol. 1. Academic Press; San Diego: 2000. p. 45-118.
- (3). Crossley MJ, King LG. J. Chem. Soc., Chem. Commun. 1984:920-922.
- (4). Gouterman M, Hall RJ, Khalil GE, Martin PC, Shankland EG, Cerny RL. J. Am. Chem. Soc. 1989; 111:3702–3707.
- (5). Khalil G, Gouterman M, Ching S, Costin C, Coyle L, Gouin S, Green E, Sadilek M, Wan R, Yearyean J, Zelelow B. J. Porphyrins Phthalocyanines. 2002; 6:135–145.
- (6). Jayaraj K, Gold A, Austin RN, Ball LM, Terner J, Mandon D, Weiss R, Fischer J, DeCian A, Bill E, Müther M, Schünemann V, Trautwein AX. Inorg. Chem. 1997; 36:4555–4566. [PubMed: 11670121]
- (7). Köpke T, Pink M, Zaleski JM. Chem. Commun. 2006:4940–4942.
- (8). McCarthy JR, Melfi PJ, Capetta SH, Brückner C. Tetrahedron. 2003; 59:9137-9146.
- (9). Akhigbe J, Ryppa C, Zeller M, Brückner C. J. Org. Chem. 2009; 74:4927–4933. [PubMed: 19489565]
- (10). Cetin A, Ziegler CJ. Dalton Trans. 2005:25–26. [PubMed: 15709252]

- (11). Rahimi R, Tehrani AA, Fard MA, Sadegh BMM, Khavasi HR. Catal. Commun. 2009; 11:232– 235.
- (12) (a). Zelelow B, Khalil GE, Phelan G, Carlson B, Gouterman M, Callis JB, Dalton LR. Sens. Actuators, B. 2003; 96:304–314.(b) Gouterman M, Callis J, Dalton L, Khalil G, Mebarki Y, Cooper KR, Grenier M. Meas. Sci. Technol. 2004; 15:1986–1994.(c) Khalil GE, Costin C, Crafton J, Jones G, Grenoble S, Gouterman M, Callis JB, Dalton LR. Sens. Actuators, B. 2004; 97:13–21.
- (13) (a). Gouterman M. J. Chem. Educ. 1997; 74:697–702.(b) Schäferling M. Angew. Chem. Int. Ed. 2012; 51:3532–3554.
- (14). Khalil GE, Daddario P, Lau KSF, Imtiaz S, King M, Gouterman M, Sidelev A, Puran N, Ghandehari M, Brückner C. Analyst. 2010; 135:2125–2131. [PubMed: 20552107]
- (15). McCarthy JR, Jenkins HA, Brückner C. Org. Lett. 2003; 5:19–22. [PubMed: 12509880]
- (16). Brückner C, Rettig SJ, Dolphin D. J. Org. Chem. 1998; 63:2094–2098.
- (17). Lara KK, Rinaldo CK, Brückner C. Tetrahedron. 2005; 61:2529–2539.
- (18). Banerjee S, Zeller M, Brückner C. J. Org. Chem. 2010; 75:1179–1187. [PubMed: 20067246]
- (19). Samankumara LP, Zeller M, Krause JA, Brückner C. Org. Biomol. Chem. 2010; 8:1951–1965. [PubMed: 20449503]
- (20). The numbering of chlorins as 2,3-dihydroporphyrins is against IUPAC recommendations (a chlorin is a 7,8-dihydroporphyrin; IUPAC-IUB Joint Commission on Biochemical Nomenclature, Pure Appl. Chem. 1987; 59:779–832., but frequently used, intuitive, and convenient.
- (21) (a). Starnes SD, Rudkevich DM, Rebek J Jr. J. Am. Chem. Soc. 2001; 123:4659–4669. [PubMed: 11457274] (b) Sutton JM, Clarke OJ, Fernandez N, Boyle RW. Bioconjugate Chem. 2002; 13:249–263.(c) Rancan F, Wiehe A, Noebel M, Senge MO, Omari SA, Boehm F, John M, Roeder BJ. Photochem. Photobiol., B. 2005; 78:17–28.(d) Choi Y, McCarthy JR, Weissleder R, Tung CH. ChemMedChem. 2006; 1:458–463. [PubMed: 16892381] (e) Al-Omari S. Biomed. Mater. 2007; 2:107–115. [PubMed: 18458443]
- (22). MacAlpine JK, Boch R. Dolphin, D. J. Porphyrins Phthalocyanines. 2002; 6:146-155.
- (23). Wang TY, Chen JR, Ma JS. Dyes Pigm. 2002; 52:199–208.
- (24) (a). Brückner C, Sternberg ED, MacAlpine JK, Rettig SJ, Dolphin D. J. Am. Chem. Soc. 1999; 121:2609–2610.(b) Campbell CJ, Rusling JF, Brückner C. J. Am. Chem. Soc. 2000; 122:6679–6685.(c) Daniell HW, Brückner C. Angew. Chem., Int. Ed. 2004; 43:1688–1691.(d) McCarthy JR, Hyland MA, Brückner C. Org. Biomol. Chem. 2004; 2:1484–1491. [PubMed: 15136804] (e) Brückner C, Hyland MA, Sternberg ED, MacAlpine J, Rettig SJ, Patrick BO, Dolphin D. Inorg. Chim. Acta. 2005; 358:2943–2953.(f) Ryppa C, Niedzwiedzki D, Morozowich NL, Srikanth R, Zeller M, Frank HA, Brückner C. Chem.–Eur. J. 2009; 15:5749–5762. [PubMed: 19388039] (g) Akhigbe J, Haskoor J, Zeller M, Brückner C. Chem. Commun. 2011; 47:8599–8601.(h) Akhigbe J, Peters G, Zeller M, Brückner C, Org. Biomol. Chem. 2011; 9:2306–2313. [PubMed: 21321766] (i) Brückner C, Götz DCG, Fox SP, Ryppa C, McCarthy JR, Bruhn T, Akhigbe J, Banerjee S, Daddario P, Daniell HW, Zeller M, Boyle RW, Bringmann G. J. Am. Chem. Soc. 2011; 133:8740–8752. [PubMed: 21534626]
- (25). Ogikubo J, Brückner C. Org. Lett. 2011; 13:2380-2383. [PubMed: 21456604]
- (26). The range from ~650 to 1300 nm in which tissue has the largest depth of penetration; the wavelength of maximum penetration of breast tissue is ~725 nm: Cerussi AE, Berger AJ, Bevilacqua F, Shah N, Jakubowski D, Butler J, Holcombe RF, Tromberg B. J. Acad. Radiol. 2001; 8:211–218.
- (27) (a). Sternberg ED, Dolphin D, Brückner C. Tetrahedron. 1998; 54:4151–4202.(b) Bonnett, R. Chemical Aspects of Photodynamic Therapy. Gordon & Breach; Langhorne, PA: 2000. (c) Pandey, RK.; Zheng, G. The Porphyrin Handbook. Kadish, KM.; Smith, KM.; Guilard, R., editors. Vol. Vol. 6. Academic Press; San Diego: 2000. p. 157-230.
- (28). Grimm, B.; Porra, RJ.; Rüdinger, W.; Scheer, H., editors. Chlorophylls and Bacteriochlorophylls. Vol. Vol. 25. Springer; Dordrecht, NL: 2006.
- (29) (a). Flitsch W. Adv. Heterocycl. Chem. 1988; 43:73–126.(b) Montforts F-P, Gerlach B, Hoeper F. Chem. Rev. 1994; 94:327–347.(c) Shanmugathasan S, Edwards C, Boyle RW. Tetrahedron.

2000; 56:1025–1046.(d) Galezowski M, Gryko DT. Curr. Org. Chem. 2007; 11:1310–1338.(e) Taniguchi M, Mass O, Boyle PD, Tang Q, Diers JR, Bocian DF, Holten D, Lindsey JS. J. Mol. Struct. 979:27–45.(f) Taniguchi M, Kim H-J, Ra D, Schwartz JK, Kirmaier C, Hindin E, Diers JR, Prathapan S, Bocian DF, Holten D, Lindsey JS. J. Org. Chem. 2002; 67:7329–7342. [PubMed: 12375962] (g) Taniguchi M, Ptaszek M, McDowell BE, Boyle PD, Lindsey JS. Tetrahedron. 2007; 63:3850–3863. [PubMed: 17479169] (h) Taniguchi M, Ptaszek M, McDowell BE, Lindsey JS. Tetrahedron. 2007; 63:3840–3849. [PubMed: 17479170] (i) Kee HL, Kirmaier C, Tang Q, Diers JR, Muthiah C, Taniguchi M, Laha JK, Ptaszek M, Lindsey JS, Bocian DF, Holten D. Photochem. Photobiol. 2007; 83:1125–1143. [PubMed: 17880507] (j) Kee HL, Kirmaier C, Tang Q, Diers JR, Muthiah C, Taniguchi M, Laha JK, Ptaszek M, Lindsey JS, Bocian DF, Holten D. Photochem. Photobiol. 2007; 83:1110–1124. [PubMed: 17880506] (k) Laha JK, Muthiah C, Taniguchi M, McDowell BE, Ptaszek M, Lindsey JS. J. Org. Chem. 2006; 71:4092–4102. [PubMed: 16709048] (l) Ruzie C, Krayer M, Lindsey JS. Org. Lett. 2009; 11:1761–1764. [PubMed: 19296621]

- (30). Brückner C, McCarthy JR, Daniell HW, Pendon ZD, Ilagan RP, Francis TM, Ren L, Birge RR, Frank HA. Chem. Phys. 2003; 294:285–303.
- (31). McCarthy JR, Perez MJ, Brückner C, Weissleder R. Nano Lett. 2005; 5:2552–2556. [PubMed: 16351214]
- (32). Zeller M, Hunter AD, McCarthy JR, Capetta SH, Brückner C. J. Chem. Crystallogr. 2005; 35:935–942.
- (33). Tércio J, Ferreira B, Cruz WO, Vieira PC, Yonashiro M. J. Org. Chem. 1987; 52:3698–3699.
- (34). Vogel, AI.; Tatchell, AR.; Furnis, BS.; Hannaford, AJ.; Smith, PWG. Vogel's Textbook of Practical Organic Chemistry. 5th Ed. Pearson; Edinburgh Gate: 1989.
- (35). National Institute for Occupational Health and Safety Registry of Toxic Effects of Chemical Substances (RTECS): RN1140000; TSCA 8(b) inventory.
- (36). Hyland MA, Morton MD, Brückner C. J. Org. Chem. 2012; 77:3038–3048. [PubMed: 22390228]
- (37). CSD code TPHPOR04
- (38). Jentzen W, Song X-Z, Shelnutt JA. J. Phys. Chem. B. 1997; 101:1684–1699.
- (39). Senge, MO. Porphyrin Handbook. Kadish, KM.; Smith, KM.; Guilard, R., editors. Vol. Vol. 10. Academic Press; San Diego: 2000. p. 1-218.
- (40). CSD code PYZNPO10
- (41) (a). Inoue S, Takeda N. Bull. Chem. Soc. Jpn. 1977; 50:984–986.(b) Buchler, JW. Porphyrins and Metalloporphyrins. 2nd ed. Smith, KM., editor. Elsevier Scientific Publishing Company; Amsterdam: 1975. p. 157-232.
- (42). Gouterman, M. The Porphyrins. Dolphin, D., editor. Vol. Vol. 3. Academic Press; New York: 1978. p. 1-165.
- (43). See, for example, the studies relating to the optical properties of the chlorins synthesized by Lindsey and co-workers, ref. 29e through 29l, and references therein
- (44). Pawlicki M, Latos-Grazynski L. Chem.-Eur. J. 2003; 9:4650-4660. [PubMed: 14566870]
- (45). Pawlicki M, Latos-Grazynski L. J. Org. Chem. 2005; 70:9123-9130. [PubMed: 16268581]
- (46) (a). Grzegorzek N, Pawlicki M, Latos-Grazynski L. J. Org. Chem. 2009; 74:8547–8553.
 [PubMed: 19860402] (b) Grzegorzek N, Pawlicki M, Szterenberg L, Latos-Grazynski L. J. Am. Chem. Soc. 2009; 131:7224–7225. [PubMed: 19422240]
- (47) (a). Martin CH, Birge RR. J. Phys. Chem. A. 1998; 102:852–860.(b) Ren L, Martin CH, Wise KJ, Gillespie NB, Luecke H, Lanyi JK, Spudich JL, Birge RR. Biochemistry. 2001; 40:13906–13914. [PubMed: 11705380] (c) Shima S, Ilagan RP, Gillespie N, Sommer BJ, Hiller RG, Sharples FP, Frank HA, Birge RR. J. Phys. Chem. A. 2003; 107:8052–8066.
- (48) (a). Miyahara T, Nakatsuji H, Hasegawa J, Osuka A, Aratani N, Tsuda A. J. Chem. Phys. 2002; 117:11196–11206.(b) Nakatsuji H, Hirao K. J. Chem. Phys. 1978; 68:2053–2065.(c) Nakatsuji H. Chem. Phys. Lett. 1991; 177:331–337.
- (49). Dunning, TH., Jr.; Hay, PJ. Modern Theoretical Chemistry. Schaefer, HF., editor. Vol. Vol. 3. Plenum, New York; New York: 1976. p. 1-28.
- (50). Premvardhan L, Sandberg DJ, Fey H, Birge RR, Buchel C, Grondelle R. v. J. Phys. Chem. B. 2008; 112:11838–11853. [PubMed: 18722413]

- (51). Adler AD, Longo FR, Finarelli JD, Goldmacher J, Assour J, Korsakoff L. J. Org. Chem. 1967; 32:476.
- (52) (a). Adler AD, Longo FR, Kampas F, Kim J. J. Inorg. Nucl. Chem. 1970; 32:2443–2445.(b) Dean ML, Schmink JR, Leadbeater NE, Brückner C. Dalton Trans. 2008:1341–1345. [PubMed: 18305846]
- (53). Crossley MJ, Hambley TW, King LG. Bull. Soc. Chim. Fr. 1996; 133:735-742.

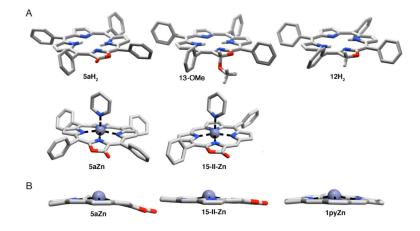


Figure 1.

Single crystal X-ray structures of porpholactone **5aH**₂, [porpholactonato]Zn(py) **5aZn**, 2oxachlorin **12H**₂, 3-methoxy-2-oxachlorin **13-OMe**, and [diphenylporpholactonato]Zn(py) **15-II-Zn**. A. Oblique views; all solvates, disorder, and hydrogen atoms attached to sp²carbons removed for clarity. B. Side view of the porpholactones **5aH**₂ and **15-II-Zn** in comparison to an comparable view to the chromophore of [*meso*-tetrakis(4pyridyl)porphyrinato]Zn(pyridine) **1pyZn**; *meso*-aryl substituents, axially coordinated pyridine (if present), and hydrogen atoms attached to sp²-carbons removed for clarity. See ESI for details on the crystal structure analyses.

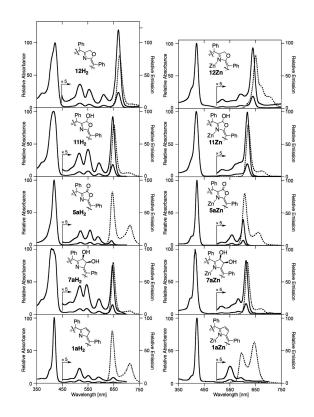


Figure 2.

Normalized UV-visible (solid traces) and fluorescence spectra (broken traces) of the compounds indicated (all in CH_2Cl_2 at ambient temperature). Their extinction coefficients are presented in the experimental section.

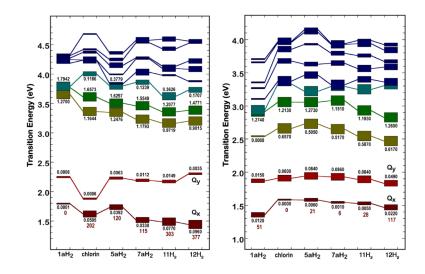
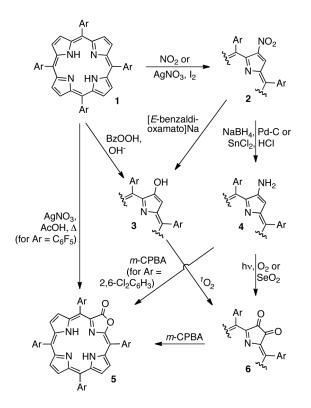


Figure 3.

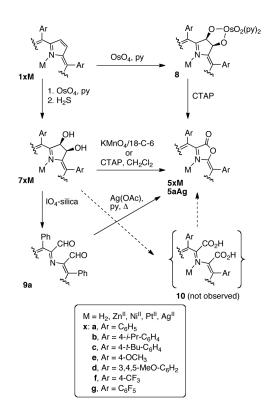
Level ordering of the low-lying electronic transitions of the chromophores investigated. Each electronic transition is represented by a rectangle with vertical position corresponding to the transition energy and height proportional to the oscillator strength. The oscillator strengths for selected transitions are shown in black directly above or below selected transition markers. The red numbers at bottom represent the ratio of the Q_x oscillator strength divided by the total oscillator strength of the Soret bands multiplied by 10^4 ; cf. to Table 1. Chlorin refers to the parent 2,3-dihydroporphin structure ($C_{20}N_4H_{16}$). Left panel: Results based on level two SAC-CI molecular orbital theory. These calculations were completed on modified chromophores in which the phenyl groups were replaced by hydrogens. Right panel: Results based on MNDO-PSDCI molecular orbital theory. The MNDO-PSDCI calculations were carried out on chromophores that included the phenyl groups.

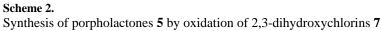


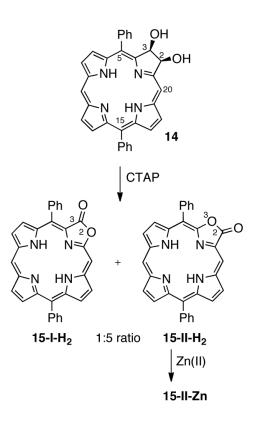


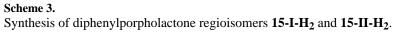
Scheme 1.

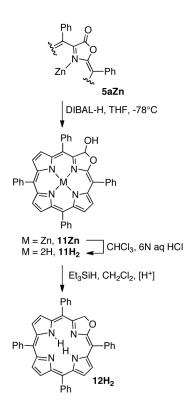
Literature-known syntheses of porpholactone **5**.^a ^a Only free bases are shown but some transformations may require metal complexation, or metal insertion takes place during the transformation.



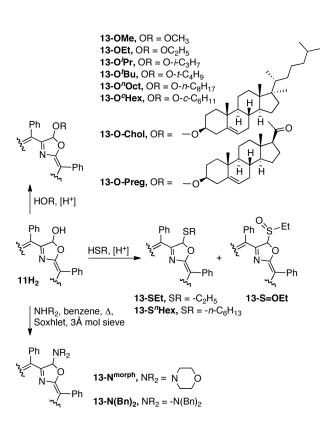






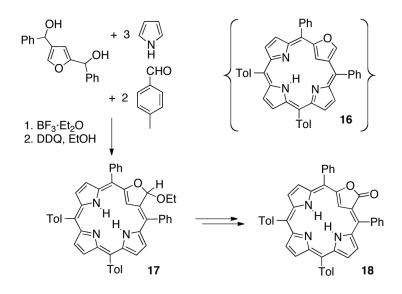


Scheme 4. Step-wise reduction of porpholactones 5.



Scheme 5.

Derivatization of porpholactol $11 H_2$ by nucleophilic substitution of the lactol hydroxy group.



Scheme 6.

Synthesis of 2-oxa-21-carbaporphyrin **16**, 2-oxa-21-carbachlorin **17**, and 2-oxa-3-oxo-21-carbaporphyrin **18**.

Table 1

Ratios of the oscillator strength of the Q_x band divided by the oscillator strength of the Soret band as a function of solvent and comparison to computed ratios.

Solvent	$1aH_2$	$5aH_2$	$7aH_2$	$11H_2$	12H ₂
Exp. Average ^a	82	69	230	348	499
SAC-CI ^b	0	120	115	303	377
MNDO-PSDCIC	51	21	9	28	117
Dipole Moment (D) ^d	0.0000	5.0056	3.5992	1.3018	1.1781
Dipole Moment* $(D)^{e}$	0.0000	4.7023	0.8262	1.2315	1.3615

exane, acetone, EtOAc, MeCN, MeOH, THF, pyridine, toluene). For a detailed listing of the solvatochromic properties of ŝ 4 ñ 12H2, see the ESI.

 $b_{
m From \, Figure \, 3}$ (left panel).

 $c_{\rm From \ Figure \ 3}$ (right panel).

 $d^d_{
m B3LYP/6-31G(d)}$ dipole moments computed without the phenyl rings.

 e B3LYP/6-31G(d) dipole moments computed with the phenyl rings.