



Published in final edited form as:

Biochem Biophys Res Commun. 2012 August 3; 424(3): 549–553. doi:10.1016/j.bbrc.2012.06.153.

Intestinal Pdx1 mediates nutrient metabolism gene networks and maternal expression is essential for perinatal growth in mice

Chin Chen, Tripp Leavitt, and Eric Sibley

Division of Pediatric Gastroenterology, Stanford University School of Medicine, Stanford, CA

Abstract

The homeodomain transcription factor Pdx1 is essential for pancreas formation and functions in pancreatic islets cells to regulate genes involved in maintenance of glucose homeostasis. In order to investigate a role for Pdx1 in intestinal cells, we analyzed the functions and networks associated with genes differentially expressed by *Pdx1* overexpression in human Caco-2 cells. In agreement with previous results for intestine isolated from mice with *Pdx1* inactivation, functional analysis of genes differentially expressed with Pdx1 overexpression revealed functions significantly associated with nutrient metabolism. Similarly, network analysis examining the interactions among the differentially expressed genes revealed gene networks involved in lipid metabolism. Consistent with defects in maternal nutrient metabolism, mouse pups born to dams with intestine-specific *Pdx1* inactivation are underweight and fail to thrive in the neonatal period compared to pups born to control dams. We conclude that Pdx1 mediates lipid metabolism gene networks in intestinal cells and that maternal expression is essential for perinatal growth in mice.

Keywords

transcription factors; gene regulation; expression profile

INTRODUCTION

The homeodomain-containing transcription factor Pdx1 is essential for pancreas formation during embryogenesis and subsequent maintenance of islet function for normal glucose homeostasis. Pdx1 regulates expression of genes in the pancreas necessary for maintaining pancreatic identity and function including insulin, glucose transporter 2, glucokinase, islet amyloid polypeptide and somatostatin.^{1–8} Mutations in the human *Pdx1* gene are linked to maturity-onset diabetes of the young, type 4 (MODY4) and type 2 diabetes mellitus.^{9–12} With respect to nutritional and hormonal regulation controlling pancreatic *Pdx1* expression, glucose, GLP-1, insulin, T³, HB-EGF, and TNF- α all positively regulate the *Pdx1* gene promoter in pancreatic β - cells.¹³ The role of Pdx1 expressed in the intestine, however, is not well defined.

© 2012 Elsevier Inc. All rights reserved.

Correspondence: Eric Sibley, M.D., Ph.D., Department of Pediatrics (Gastroenterology), Stanford University School of Medicine, 300 Pasteur Drive, Grant Building G310, Stanford, CA 94305-5208, Telephone: (650) 723-5070, Telefax: (650) 724-3106, eric.sibley@stanford.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

DISCLOSURES

No conflicts of interest are declared by the author(s).

Pdx1 is expressed in the anterior duodenal region of the small intestine and decreases in expression distally.¹⁴ Mice homozygous for a *Pdx1* null mutation (*Pdx1*^{-/-}) fail to form a pancreas and die in the neonatal period within a week of birth.^{15, 16} Therefore, in order to investigate roles for *Pdx1* expressed in the intestine, mice with *Pdx1* inactivation restricted to the intestinal epithelium (*Pdx1*^{flx/flx}; *VilCre*) have been generated.¹⁷ *Pdx1*^{flx/flx}; *VilCre* mice survive through adulthood and have pancreata and small intestines with gross morphologies that are indistinguishable from those of controls. Expression profiling identified genes differentially expressed in duodenal segments isolated from mature *Pdx1*^{flx/flx}; *VilCre* and control mice.¹⁸ Pathway analysis of the differentially expressed genes revealed functions that are significantly associated with metabolism of nutrients such as lipids, carbohydrates, amino acids, vitamins and minerals. In addition, network analysis examining the interactions among the differentially expressed genes revealed gene networks involved in metabolism of lipids and minerals.¹⁸

In order to identify additional novel *Pdx1*-regulated genes, human intestinal epithelial Caco-2 cells have been engineered to overexpress *Pdx1* and gene expression profiles relative to control cells were assessed.¹⁹ Fatty acid binding protein 1, liver, *FABP1*, a gene with known intestinal cell expression, was identified as a candidate *Pdx1* target through such analysis. In the present study, we sought to analyze the functions and networks associated with genes differentially expressed by *Pdx1* overexpression in human intestinal Caco-2 cells. Upon identifying associations between *Pdx1* overexpression and gene networks associated with nutrient and lipid metabolism, we proceeded to investigate the role of intestinal *Pdx1* expression in supporting maternal-fetal nutrition in mice.

MATERIALS AND METHODS

Animals

Mice with intestinal epithelium-specific *Pdx1* inactivation (*Pdx1*^{flx/flx}; *VilCre*) were generated by intercross mating between *VilCre* and *Pdx1*^{flx/flx} mouse strains as previously described.¹⁷ To investigate the extent of defects in nutrient metabolism in mothers with *Pdx1* inactivation, a *Pdx1*^{flx/flx}; *VilCre* virgin female was mated with a *Pdx1*^{flx/flx}; *VilCre* stud male. A littermate control *Pdx1*^{flx/flx} virgin female was mated with the same *Pdx1*^{flx/flx}; *VilCre* stud in the same cage. Identical normal diet feed was accessible to all mice. The protocol for animal use was reviewed and approved by the Stanford University Institutional Animal Care and Use Committee (IACUC).

Functional analysis of genes differentially expressed in Caco-2 cells with *Pdx1* overexpression

Microarray data, previously generated for human intestinal epithelial Caco-2 cells stably transfected with a vector driving mouse *Pdx1* cDNA expression or with empty vector alone¹⁹, was analyzed with the web-based software and database, Ingenuity Pathways Analysis (IPA version 8.8, Ingenuity Systems, www.ingenuity.com). Specifically, functions and interactions of genes exhibiting significant differential expression >4-fold were analyzed.

IPA Functional analysis was performed to find significant associations of the differentially expressed genes to molecular and cellular functions. Under the primary categories, subcategories were classified, consisting of specific, basic level functions populated with a group of genes or chemicals, based on the findings stored in the Ingenuity Knowledge Base. Statistically significant, non-random associations of the differentially expressed genes with the specific functions and subcategories were indicated by a *p* value less than 0.05 following right-tailed Fisher's exact test.

Network analysis of differentially expressed genes

IPA Network analysis was performed to examine and visualize interactions among genes exhibiting significant changes in expression with *Pdx1* overexpression in Caco-2 cells by generating statistically significant, non random networks. The differentially expressed genes served as “seeds” and connected to other genes or chemicals in the Ingenuity Knowledge Base via direct or indirect interactions. Networks were limited to 35 genes or chemicals to maximize specificity of the connections. Network analysis complements functional analysis, because functional analysis considers the differentially expressed genes alone. The statistical significance, or scores, of generated networks were calculated with right-tailed Fisher’s Exact Test. The higher the score, the lower the probability of finding the observed number of differentially expressed genes in a given network by random chance

RESULTS

Pdx1 overexpression differentially regulates genes with functions associated with nutrient metabolism

In order to identify functions for genes differentially expressed in intestinal cells with Pdx1 overexpression, microarray data was analyzed for human intestinal epithelial Caco-2 cells engineered to overexpress *Pdx1* relative to control cells. The microarray data was previously generated from experiments using RNA isolated from Caco-2 cells stably transfected with a Pdx1 cDNA expression or with empty vector alone at 9 days post confluency.¹⁹

Microarray data was analyzed by a computer software, Ingenuity Pathway Analysis (IPA), for significant association ($p < 0.05$) with biological functions represented by the genes differentially expressed > 4 -fold by *Pdx1* overexpression in post-confluent Caco-2 cells. The association was examined by analysis for molecular and cellular functional annotations (Table 1). In post-confluent Caco-2 cells, metabolism of drugs and nutrients such as lipids, carbohydrates, amino acids, nucleic acids, vitamins and minerals was likely affected by *Pdx1* overexpression (Table 1).

The findings are in agreement with the previous gene profiling study using mice with intestinal epithelium-specific *Pdx1* inactivation, showing that functions of the genes differentially expressed in mature duodenum are also significantly associated with nutrient and drug metabolism.¹⁸ In post-confluent Caco-2 cells overexpressing *Pdx1*, products of the differentially expressed genes involved in lipid metabolism include biosynthetic enzymes, transport proteins, kinases, ligand-dependent nuclear receptors and cytokines (Table 2).

Network analysis of differentially expressed genes indicates that Pdx1 overexpression may impact lipid metabolism

To complement the functional analysis described above, networks were generated to analyze the relational interactions among differentially expressed genes. Gene networks visualize the relationships among genes differentially expressed > 4 -fold in response to *Pdx1* overexpression in post-confluent Caco-2 cells. The relationships examined include direct and indirect interactions between the genes of interest. Direct interactions refer to physical binding relationships such as protein-DNA binding, while examples for indirect interactions include activation, transcription, phosphorylation, or localization. The differentially expressed genes were used as “seeds” and connected as many of them into a network. Other molecules (genes or chemicals) in the Ingenuity Knowledge Base were also included to connect multiple smaller gene networks into a larger network, thus providing insights into possible functional roles for Pdx1 in the intestinal cell culture.

The representative network shown in Figure 1 has a high significance score of 30 and contains a high number (16) of genes differentially expressed > 4-fold in response to *Pdx1* overexpression in post-confluent Caco-2 cells. The score of 30 indicates that the chance is 1 in 10^{30} to form a network of 35 molecules by randomly selecting from the Ingenuity database and including at least 16 differentially expressed genes by *Pdx1* overexpression. Network analysis examining the interactions among the differentially expressed genes revealed genes involved in lipid metabolism, including acyl-CoA synthetase long-chain family member 1 (*ACSL1*), fatty acid binding protein 1, liver (*FABP1*) and UDP glucuronosyltransferase 2 family, polypeptide B15 (*UGT2B15*). This network suggests a functional role of *Pdx1* in modulating lipid metabolism.

Offspring of *Pdx1*^{flox/flox}; *VilCre* dam were underweight and failed to thrive postnatally

To investigate the extent of defects in nutrient metabolism in mice with *Pdx1* inactivation, a *Pdx1*^{flox/flox}; *VilCre* virgin female was mated with a *Pdx1*^{flox/flox}; *VilCre* stud male, along with a littermate control *Pdx1*^{flox/flox} virgin female. Identical normal diet feed was accessible to all mice. The weight and condition of the offspring from both dams were tracked upon birth through weaning and compared (Figure 2). The offspring from both dams were born on the same day and the size of the litters was similar (7 pups to *Pdx1*^{flox/flox}; *VilCre* dam and 10 to the littermate control dam). However, the size of the litter to *Pdx1*^{flox/flox}; *VilCre* dam continued to decrease; with 5 pups remaining 3 days after birth, 3 pups 10 days after birth, and 1 surviving until weaning at 23 days that died shortly thereafter, compared with 10 pups born to the control dam that thrived throughout the postnatal period.

At postnatal day 3, the offspring of *Pdx1*^{flox/flox}; *VilCre* dam appeared smaller in size, pale and lethargic compared to those of the control dam (Figure 2A). Although milk was present in the stomach of pups from *Pdx1*^{flox/flox}; *VilCre* dam, indicating that they were fed, the average weight of the pups (1.1g) was less than that of control pups (1.6g) (Figure 2D). Weight gain was delayed at postnatal day 10 for pups from the *Pdx1*^{flox/flox}; *VilCre* dam, with an average weight of 3.4g in contrast to that of 5g for control pups (Fig. 2D). In comparison to control pups at postnatal day 10, the pups born to the *Pdx1*^{flox/flox}; *VilCre* dam remained smaller overall in size, appeared pale in extremities, and lacked fur coat coverage from the ear down (Figure 2B). At postnatal day 23, pups were weaned. The remaining pup from the *Pdx1*^{flox/flox}; *VilCre* dam showed further growth retardation, weighing less than half (4g) of the average weight (9.2g) of the control pups (Figure 2D). The appearance of the remaining pup was also consistent with growth retardation showing small physique, fur coat abnormality, pale limbs, ears and tail (Figure 2C). Pups born to the littermate control dam had no apparent developmental abnormalities.

DISCUSSION

In order to investigate a role for *Pdx1* in intestinal cells, the functions and networks associated with genes differentially expressed by *Pdx1* overexpression in human intestinal Caco-2 cells were analyzed. The functions of genes differentially expressed by *Pdx1* overexpression are significantly associated with nutrient and lipid metabolism. Analysis of the relationships among the genes also supports a role for *Pdx1* in mediating networks associated with lipid metabolism. These findings are in agreement with our previous gene profiling study of mice with intestinal epithelium-specific *Pdx1* inactivation, which showed that functions of the genes differentially expressed in mature duodenum are also significantly associated with nutrient and lipid metabolism.¹⁸

Having identified associations between both *Pdx1* overexpression and *Pdx1* inactivation and gene networks associated with nutrient and lipid metabolism, we investigated the role of

intestinal *Pdx1* expression in supporting maternal-fetal nutrition in mice. Consistent with defects in maternal nutrient metabolism, *Pdx1^{flox/flox}; VilCre* mice born to dams with intestine-specific *Pdx1* inactivation are underweight and fail to thrive in the neonatal period compared to pups born to control dams. These findings are also consistent with our previous report demonstrating that mature *Pdx1^{flox/flox}; VilCre* mice with intestinal epithelium-specific *Pdx1* inactivation have altered metabolism for nutrients such as lipid and iron, even when a normal diet was fed.¹⁸ Failure to gain weight and the growth abnormalities observed in the offspring from *Pdx1^{flox/flox}; VilCre* dam are most likely due to malnutrition resulting from the defects in maternal nutrient metabolism following *Pdx1* inactivation in the dam's intestinal epithelium, regardless of the offspring's own genotypes, because the pups born to the littermate control dam show no developmental abnormalities. We conclude that *Pdx1* mediates nutrient metabolism gene networks in intestinal cells and that maternal expression is essential for perinatal growth in mice.

Acknowledgments

This research was supported by National Institute of Diabetes and Digestive and Kidney Diseases Grants DK72416 and DK60715 (to E. Sibley) and DK56339 (to the Stanford Digestive Diseases Center).

REFERENCES

- Bretherton-Watt D, Gore N, Boam DS. Insulin upstream factor 1 and a novel ubiquitous factor bind to the human islet amyloid polypeptide/amylin gene promoter. *Biochem J.* 1996; 313(Pt 2):495–502. [PubMed: 8573083]
- Carty MD, Lillquist JS, Peshavaria M, Stein R, Soeller WC. Identification of cis- and trans-active factors regulating human islet amyloid polypeptide gene expression in pancreatic beta-cells. *J Biol Chem.* 1997; 272:11986–11993. [PubMed: 9115263]
- Leonard J, Peers B, Johnson T, Ferreri K, Lee S, Montminy MR. Characterization of somatostatin transactivating factor-1, a novel homeobox factor that stimulates somatostatin expression in pancreatic islet cells. *Mol Endocrinol.* 1993; 7:1275–1283. [PubMed: 7505393]
- Miller CP, McGehee RE Jr, Habener JF. IDX-1: a new homeodomain transcription factor expressed in rat pancreatic islets and duodenum that transactivates the somatostatin gene. *Embo J.* 1994; 13:1145–1156. [PubMed: 7907546]
- Ohlsson H, Karlsson K, Edlund T. IPF1, a homeodomain-containing transactivator of the insulin gene. *Embo J.* 1993; 12:4251–4259. [PubMed: 7901001]
- Serup P, Jensen J, Andersen FG, et al. Induction of insulin and islet amyloid polypeptide production in pancreatic islet glucagonoma cells by insulin promoter factor 1. *Proc Natl Acad Sci U S A.* 1996; 93:9015–9020. [PubMed: 8799146]
- Waeber G, Thompson N, Nicod P, Bonny C. Transcriptional activation of the GLUT2 gene by the IPF-1/STF-1/IDX-1 homeobox factor. *Mol Endocrinol.* 1996; 10:1327–1334. [PubMed: 8923459]
- Watada H, Kajimoto Y, Umayahara Y, et al. The human glucokinase gene beta-celltype promoter: an essential role of insulin promoter factor 1/PDX-1 in its activation in HIT15 cells. *Diabetes.* 1996; 45:1478–1488. [PubMed: 8866550]
- Cockburn BN, Bermanno G, Boodram LL, et al. Insulin promoter factor-1 mutations and diabetes in Trinidad: identification of a novel diabetes-associated mutation (E224K) in an Indo-Trinidadian family. *J Clin Endocrinol Metab.* 2004; 89:971–978. [PubMed: 14764823]
- Stoffers DA, Stanojevic V, Habener JF. Insulin promoter factor-1 gene mutation linked to early-onset type 2 diabetes mellitus directs expression of a dominant negative isoprotein. *J Clin Invest.* 1998; 102:232–241. [PubMed: 9649577]
- Hani EH, Stoffers DA, Chevre JC, et al. Defective mutations in the insulin promoter factor-1 (IPF-1) gene in late-onset type 2 diabetes mellitus. *J Clin Invest.* 1999; 104:R41–R48. [PubMed: 10545531]
- Macfarlane WM, Frayling TM, Ellard S, et al. Missense mutations in the insulin promoter factor-1 gene predispose to type 2 diabetes. *J Clin Invest.* 1999; 104:R33–R39. [PubMed: 10545530]

13. Campbell SC, Macfarlane WM. Regulation of the *pdx1* gene promoter in pancreatic beta-cells. *Biochem Biophys Res Commun.* 2002; 299:277–284. [PubMed: 12437983]
14. Guz Y, Montminy MR, Stein R, et al. Expression of murine STF-1, a putative insulin gene transcription factor, in beta cells of pancreas, duodenal epithelium and pancreatic exocrine and endocrine progenitors during ontogeny. *Development.* 1995; 121:11–18. [PubMed: 7867492]
15. Jonsson J, Carlsson L, Edlund T, Edlund H. Insulin-promoter-factor 1 is required for pancreas development in mice. *Nature.* 1994; 371:606–609. [PubMed: 7935793]
16. Offield MF, Jetton TL, Labosky PA, et al. PDX-1 is required for pancreatic outgrowth and differentiation of the rostral duodenum. *Development.* 1996; 122:983–995. [PubMed: 8631275]
17. Chen C, Fang R, Davis C, Maravelias C, Sibley E. Pdx1 inactivation restricted to the intestinal epithelium in mice alters duodenal gene expression in enterocytes and enteroendocrine cells. *Am J Physiol Gastrointest Liver Physiol.* 2009; 297:G1126–G1137. [PubMed: 19808654]
18. Chen C, Sibley E. Expression profiling identifies novel gene targets and functions for Pdx1 in the duodenum of mature mice. *Am J Physiol Gastrointest Liver Physiol.* 2012; 302:G407–G419. [PubMed: 22135308]
19. Chen C, Fang R, Chou LC, Lowe AW, Sibley E. PDX1 regulation of FABP1 and novel target genes in human intestinal epithelial Caco-2 cells. *Biochem Biophys Res Commun.* 2012; 423:183–187. [PubMed: 22640736]

Highlights

- Pdx1 overexpression regulates genes with functions associated with nutrient metabolism.
- Network analysis indicates that Pdx1 overexpression may impact lipid metabolism.
- Offspring of mothers with intestine-specific Pdx1 inactivation fail to thrive.

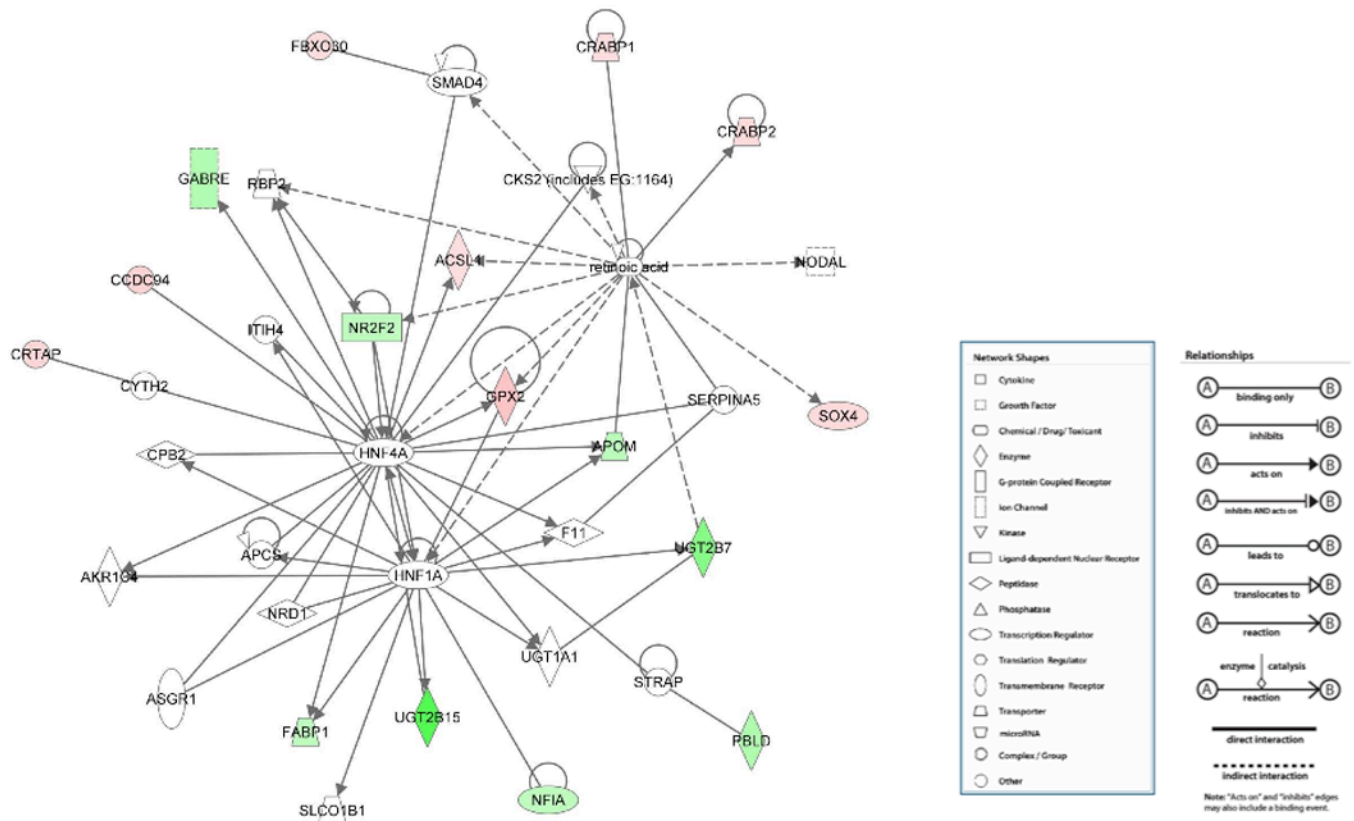


Figure 1. Network analysis suggesting a role for Pdx1 in modulating lipid metabolism in Caco-2 cells

Genes significantly upregulated in expression by >4-fold are in red and genes significantly downregulated are in green. This network scored high statistical significance with right-tailed Fisher's Exact Test and contained a high fraction of differentially expressed genes. Genes or chemicals in white were not included in the 129 genes with >4-fold changes in expression¹⁹ or on the microarray chips screened.

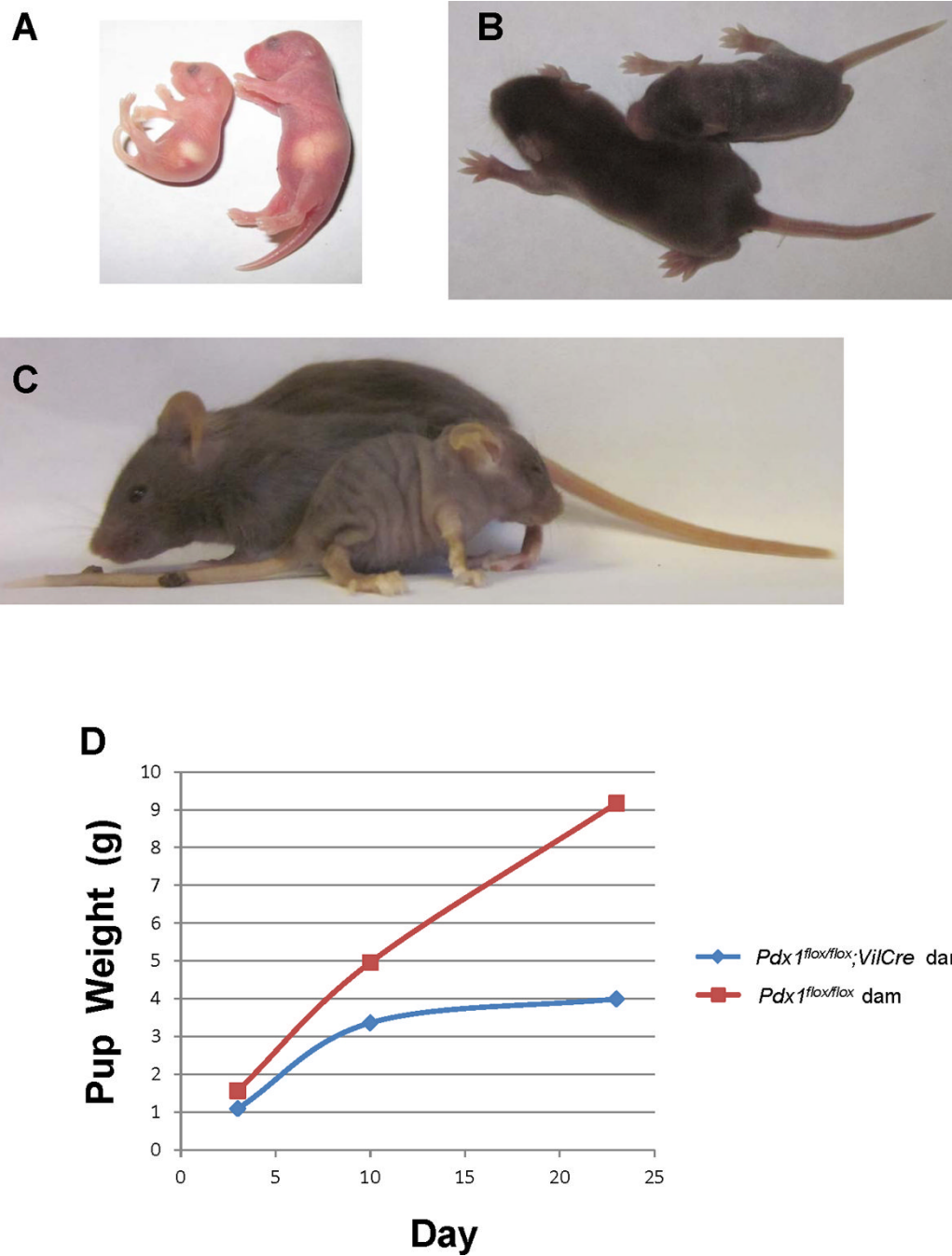


Figure 2. Maternal *Pdx1* inactivation effects perinatal growth

Pups born to *Pdx1^{flox/flox}; VilCre* dams are underweight (A) and fail to thrive (B,C) compared to those born to littermate control *Pdx1^{flox/flox}* dams. D). Weights of of pups born to a *Pdx1^{flox/flox}; VilCre* dam (blue line) compared to pups born to a littermate control *Pdx1^{flox/flox}* dam (red line).

Table 1

Pdx1 overexpression in Caco-2 cells alters expression of genes with functions significantly associated with metabolism of nutrients and drugs ($p < 0.05$).

Category	<i>P</i> Value
Lipid Metabolism	1.30E-04 – 1.31E-02
Carbohydrate Metabolism	3.98E-04 – 1.10E-02
Nucleic Acid Metabolism	1.59E-03 – 5.22E-03
Drug Metabolism	1.90E-03 – 6.65E-03
Vitamin and Mineral Metabolism	6.65E-03 – 6.65E-03
Amino Acid Metabolism	1.90E-03 – 1.28E-02

Table 2

Genes with functions in lipid metabolism increased or decreased in expression >4-fold by Pdx1 overexpression in Caco-2 cells ($p < 0.05$)

Genes with Functions in Lipid Metabolism	Gene Symbol	Fold Change
acyl-CoA synthetase long-chain family member 1	<i>ACSL1</i>	4.04
angiotensinogen (serpin peptidase inhibitor, clade A, member 8)	<i>AGT</i>	-6.24
apolipoprotein M	<i>APOM</i>	-4.32
cytochrome P450, family 27, subfamily A, polypeptide 1	<i>CYP27A1</i>	-4.95
estrogen receptor 1	<i>ESR1</i>	-4.18
fatty acid binding protein 1, liver	<i>FABP1</i>	-4.07
fibronectin 1	<i>FN1</i>	11.17
kininogen 1	<i>KNG1 (includes EG:3827)</i>	-14.58
mitogen-activated protein kinase 8	<i>MAPK8</i>	-5.62
sema domain, immunoglobulin domain (Ig), short basic domain, secreted, (semaphorin) 3A	<i>SEMA3A</i>	4.20
tumor necrosis factor (ligand) superfamily, member 10	<i>TNFSF10</i>	-5.17
UDP glucuronosyltransferase 2 family, polypeptide B4	<i>UGT2B4</i>	-10.54
UDP glucuronosyltransferase 2 family, polypeptide B7	<i>UGT2B7</i>	-7.69
UDP glucuronosyltransferase 2 family, polypeptide B15	<i>UGT2B15</i>	-11.06