

## Review Article

# The role of the TGF- $\beta$ family in wound healing, burns and scarring: a review

Jack W Penn, Adriaan O Grobbelaar, Kerstin J Rolfe

*The Institute of Plastic Surgery, Research and Education. Dept Plastic Surgery, The Royal Free Hospital, Pond St Hampstead, London UK, NW3 2QG*

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**Abstract:** It is estimated worldwide that over 6 million people per annum experience a burn injury. Despite advances in management and improved survival rates, the incidence of hypertrophic scarring remains high. These scars are particularly common after burns and are often raised, red, hard and may cause abnormal sensations. Such pathological scarring can lead to severe functional impairment, psychological morbidity, and costly long term healthcare. Wound healing is an inherent process which restores the integrity of the skin after injury and although scarring is a frequent by-product, the scarless wound healing observed in early human gestational fetuses suggests that it is not an essential component of the response. This has led to a large body of research attempting to understand the mechanisms behind scarring and in turn prevent it. One of the main focuses of recent research has been the role played by the growth factor TGF- $\beta$  in the process of both wound healing and scar formation. The three isoforms (TGF- $\beta$ 1, TGF- $\beta$ 2 and TGF- $\beta$ 3) appear to have overlapping functions and predominantly mediate their effects through the intracellular SMAD pathway. Initial research suggested that TGF- $\beta$ 1 was responsible for the fibrotic scarring response whereas the scarless wound healing seen in fetal wounds was due to increased levels of TGF- $\beta$ 3. However, the reality appears to be far more complex and it is unlikely that simply altering the ratio of TGF- $\beta$  isoforms will lead to scarless wound healing. Other aspects of the TGF- $\beta$  system that appear promising include the downstream mediator CTGF, the proteoglycan decorin and the binding protein p311. Other putative mechanisms which may underlie the pathogenesis of hypertrophic scars include excessive inflammation, excessive angiogenesis, altered levels of matrix metalloproteinases, growth factors, and delayed apoptosis of fibrotic myofibroblasts either due to p53 genetic alterations or tensile forces across the wound. If an effective treatment for hypertrophic scars following burns injury is to be developed then further work must be carried out to understand the basic mechanisms of pathological scarring.

**Keywords:** Burn scarring, hypertrophic scarring, pathological scarring, regeneration, TGF- $\beta$ , wound healing

## Introduction

It has been estimated that over 6.6 million people world-wide suffer a burns injury annually [1]. Though many burn injuries require no medical intervention, a number require medical treatment and the American Burn Association estimate that 500,000 patients seek medical attention a year [2]. Survival following extensive burns has improved over recent years with advancement in fluid resuscitation, new antibiotics, skin replacements and specialised burns centres. However, the incidence, treatment and prevention of scarring, particularly hypertrophic scarring has not improved with survival [3]. Patients following a burn injury may require long term medical intervention for their scars and

their related issues. Patients suffering pathological scarring such as hypertrophic scars can suffer from disfigurement, disability, stigmatization, disruption of daily activities, as well as psychological issues [4-7].

Hypertrophic scars are often raised, red, hard, and usually have abnormal sensations, which can include pain and tenderness [8, 9]. The incidence of hypertrophic scarring has varied in studies between 32-67% but rises to 75% in children, young adults and those with pigmented skin [10-14]. Some hypertrophic scars particularly those associated with thermal injuries are associated with contractures [15, 16], which are not only disfiguring but when occurring over a joint can result in loss of functional

ity and disability [17].

### Scarring

Wound healing is an inherent process, which restores the integrity of skin as quickly as possible. Restoration of the skin is essential, due to the skin's importance in survival through the prevention of infection, fluid loss and other vital functions. Some have suggested that wound healing evolved for speed, to allow the wound to heal quickly reducing the risk of infection [18]. Wound healing is a dynamic process with at one end of the spectrum an over-exuberance resulting in pathological scarring while at the other end a non-healing or chronic wound. However, it has been known since the 1970's that scarring is not required for wound healing with early human gestational fetuses healing cutaneous wounds perfectly without the formation of scar tissue [19]. Variation in the outcome of wound healing is not just seen between racial groups, individuals, gender and age but can also vary within the same individual. Hypertrophic scarring has been shown to be more common following certain injuries such as burns, delayed epithelisation or wounds occurring in areas of high tension for example the deltoid and sternal regions or areas of movement [20-22]. Others, have suggested that the depth of the wound may be associated with hypertrophic scarring, with fibroblasts derived from the deep dermis resembling fibroblasts from hypertrophic scarring [23]. Singer and Clark suggest that hypertrophic scarring is caused by an aberrant form of wound healing demonstrated by a constitutively active proliferative phase [24]. Though other theories have been proposed and will be discussed later.

Hypertrophic scars are raised, abnormally pigmented and can cause itching or abnormal sensations. Unlike keloids, hypertrophic scars remain within the boundary of the original injury and hypertrophic scars can regress with time. Hypertrophic scars have been shown to have a preponderance of collagen type III fibres orientated parallel to the epidermal surface. They are often composed of nodules containing myofibroblasts, differentiated fibroblasts expressing  $\alpha$ -smooth muscle actin, collagen filaments and other extracellular matrices [25].

### TGF- $\beta$ and hypertrophic scars

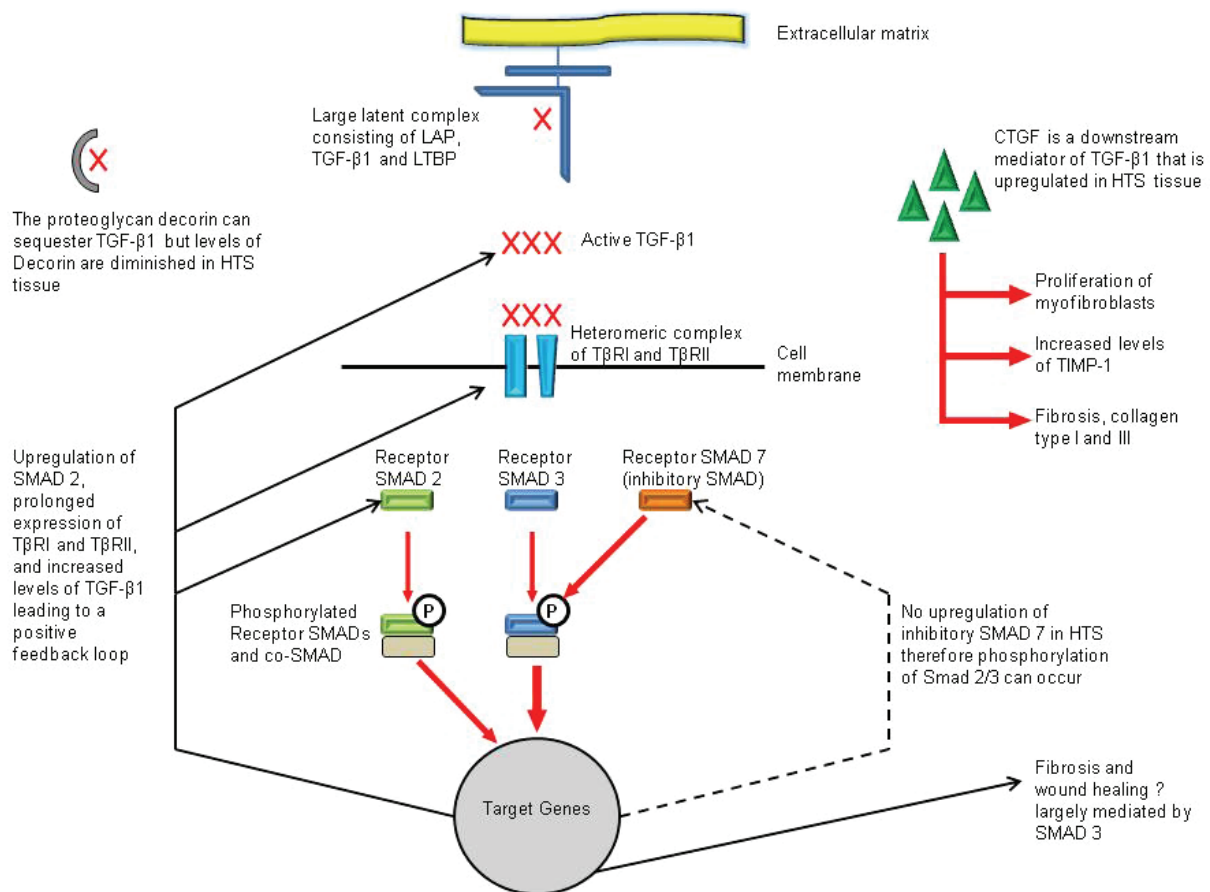
TGF- $\beta$  is a family of growth factors involved in a

number of essential cellular functions. The three isoforms of TGF- $\beta$  (TGF- $\beta$ 1, - $\beta$ 2, - $\beta$ 3) are secreted as inactive latent precursors that require activation prior to binding to the TGF- $\beta$  receptors [26]. The three isoforms, share 60-80% homology and are encoded by different genes. However, the isoforms are believed to activate the same intracellular signalling pathways and appear *in vitro* to have overlapping biological functions, though with differing *in vivo* expression [27]. However, knockout mice have shown that the three isoforms appear to play different roles in both development and homeostasis [28-31]. All three isoforms appear to be present in wound healing, and even wounds from early human foetuses, which repair cutaneous wounds perfectly, have been shown to contain all three TGF- $\beta$  isoforms [32]. However, these isoforms in fetal wounds not only differ in levels of expression, duration in the wound, but also in their biological activity [32-37]. TGF- $\beta$  is involved in a number of processes in wound healing: inflammation, stimulating angiogenesis, fibroblast proliferation, collagen synthesis and deposition and remodelling of the new extracellular matrix [38, 39]. Interestingly chronic, non-healing wounds often show a loss of TGF- $\beta$ 1 signalling [40, 41].

All three isoforms are believed to bind and signal through the two TGF- $\beta$  receptors (T $\beta$ RI and T $\beta$ RII) [26]. T $\beta$ RII is constitutively phosphorylated and on binding of the ligand, phosphorylates T $\beta$ RI. Phosphorylation of the receptor complex activates the SMAD intracellular signalling pathway through the receptor Smads (Smad-2 and Smad-3) and co-Smad 4. The receptor SMADs and Smad-4 cross over the nuclear membrane where they regulate a number of genes [26, 41]. TGF- $\beta$  can also activate a number of non-Smad signalling pathways whose function in TGF- $\beta$  remains to be elucidated [26].

Fibroblasts derived from hypertrophic scars have been shown, by a number of groups, to have an altered phenotype compared to fibroblasts derived from normal scars or uninjured dermis [43-45]. Wang and colleagues showed that hypertrophic derived fibroblasts and hypertrophic scar tissue produced more mRNA and protein for TGF- $\beta$ 1 than normal skin or fibroblasts derived from normal skin, suggesting a possible role for TGF- $\beta$ 1 in hypertrophic scar formation [45]. Not only have hypertrophic derived fibroblasts shown more TGF- $\beta$ 1 expression, but they have also been shown to have a

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**Figure 1.** Summary of TGF- $\beta$  signalling in hypertrophic scarring.

prolonged expression of the TGF- $\beta$  receptors compared to normal skin [46]. The group suggest that this expression of TGF- $\beta$  is persistent compared to normal wound healing where receptor expression decreases during the remodelling phase [46]. The group suggest that this persistence of receptor expression may result in a feedback loop resulting in a fibrotic phenotype [46]. Others have suggested that the three isoforms of TGF- $\beta$  have different temporal effects on wound healing and scarring, and any disruption in this expression pattern may result in hypertrophic scar formation [47].

Fibroblasts derived from hypertrophic scars have been shown to have an alteration in TGF- $\beta$  signalling. Studies have indicated increased expression and phosphorylation of the receptor Smads-2 and/or 3 in hypertrophic scarring [48, 49]. Xie and colleagues showed that the Smad inhibitor, Smad 7, showed no up regulation in

response to TGF- $\beta$ 1 compared to fibroblasts derived from normal skin [48]. Kopp et al showed that over expressing Smad 7 prevented collagen contraction in both normal and hypertrophic scar derived fibroblasts [49]. Interestingly, animals lacking Smad-3 show improved wound-healing (increased rate of re-epithelization, reduced infiltration of monocytes). While in a bleomycin-lung fibrosis model, mice lacking Smad-3 showed suppression of type I procollagen mRNA expression and an attenuation of the fibrotic process [50, 51; **Figure 1**].

Rorison et al [52] showed that plasma levels of TGF- $\beta$  maybe a predictive indicator in children who develop hypertrophic scars after a burn. They showed that children whose burns healed well and without hypertrophic scarring showed elevation of TGF- $\beta$  in their plasma for two weeks post burn. However, those who developed hy-

hypertrophic scarring did not demonstrate this early increase in TGF- $\beta$  levels [52]. Others have shown an increased frequency of CD4+/TGF- $\beta$  producing T cells in blood from burn patients, and this was further identified in hypertrophic scar tissue [53]. Wang and colleagues suggested that these cells may regulate the functions of dermal fibroblasts resulting in hypertrophic scar formation. They showed that dermal fibroblasts showed increased proliferation, alpha smooth muscle expression and collagen gel contraction and collagen synthesis when treated with medium derived from CD4+ T lymphocytes from a burn patient compared to normal patients [53].

CTGF (CCN2) a downstream mediator of TGF- $\beta$ 1, though co-ordinately expressed, CTGF has been shown to be involved in the formation of fibrosis and TGF- $\beta$ 1's pro-contractile activity is believed to be mediated by CTGF [54, 55; **Figure 1**]. CTGF has been shown to be elevated in fibroblasts derived from hypertrophic scars in both unstimulated hypertrophic scar fibroblasts and following stimulation with any of the three isoforms of TGF- $\beta$  [56]. Sisco and colleagues showed in an animal model that inhibiting the action of CTGF through antisense oligonucleotides limited hypertrophic scar formation through the reduction of myofibroblasts, decreased TIMP-1 and collagens types I and III, but did not alter wound closure, therefore CTGF may be a potential future therapeutic target in the prevention of hypertrophic scarring [57].

Proteoglycans are specialised glycoproteins which also contain linear polysaccharides, glycosaminoglycans. Proteoglycans are known to play a role in cell signalling and can interact and modulate proteins found in the extracellular matrix. The proteoglycan decorin is known to bind to the three TGF- $\beta$  isoforms and inhibits their activity by sequestering the isoforms to the extracellular matrix. Decorin also plays a role in regulating collagen fibrillogenesis, and has been shown to interact with other growth factors regulating their action, including CTGF [58]. Both fibromodulin and decorin have been shown to have lower levels or delayed expression in post-burn hypertrophic scars [59, 60]. This low or reduced expression may explain the irregular collagen organisation and increased extracellular matrix production in pathological scarring [59]. The use of recombinant human decorin in *in vitro* studies has shown that

decorin plays a role in reducing hypertrophic fibroblast proliferation, collagen synthesis and collagen contraction with decorin inhibiting both basal and TGF- $\beta$ 1 enhanced contraction in both normal and hypertrophic scar fibroblasts [61, 62].

P311 a binding protein of the TGF- $\beta$ 1 latency associated protein, has been suggested to be involved in myofibroblast differentiation and fibrosis [63], though its biological function remains largely unknown. P311 has been found to be over expressed in hypertrophic scar tissue and other fibrotic lesions [64, 65], and appears involved in wound healing as it is expressed by myofibroblasts, though is absent once the wound is healed [63]. The role that P311 has on TGF- $\beta$  expression remains unclear, in human derived skin cells forced expression of p311 increases both TGF- $\beta$ 1 and collagen type 1 (COL1A1) mRNA expression [66], other human fibrotic tissues show elevation of both P311 and TGF- $\beta$ 1 suggesting a role for both in fibrosis [65]. However, in the mouse fibroblast cell lines, NIH 3T3 and C3H10, P311 inhibited TGF- $\beta$ 1 and T $\beta$ RII expression with a subsequent decrease in collagen expression [63].

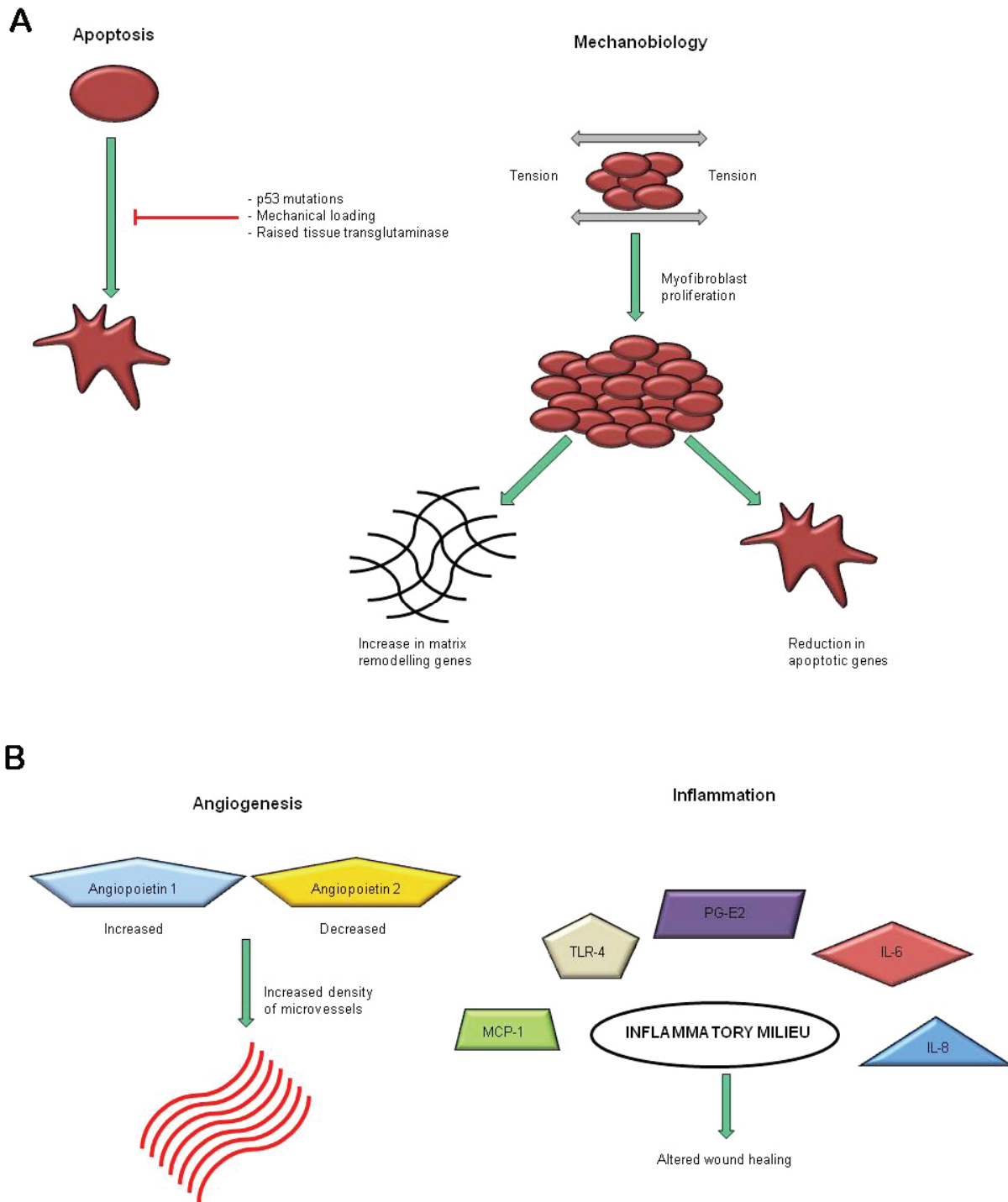
### Other mechanisms in the development of hypertrophic scarring

Though the exact pathophysiology of hypertrophic scars is unknown a number of theories besides the role of TGF- $\beta$  have been proposed (**Figure 2**). **Table 1** shows examples of other mechanisms shown to be aberrant in hypertrophic scars. Understanding the mechanisms behind hypertrophic scars will help with therapeutic treatments to help prevent or reduce hypertrophic scarring.

### Treatment

To date, there remains no definitive treatment to either prevent or reduce any form of scarring. Peer reviewed data on the effectiveness of scar prevention treatments are few and far between. Clinical studies on current treatments are often inadequate due to small numbers of patients, lack of well designed controls and a lack of standardisation in scar outcome measurements [86]. A number of mechanisms associated with the development of hypertrophic scarring have been used to manipulate the scarring potential. **Table 2** reviews a number of the current and

## TGF- $\beta$ family in wound healing



**Figure 2.** Summary of some of the mechanisms believed to be involved in the formation of hypertrophic scars.

potential therapies involved in different mechanisms associated with developing hypertrophic scarring.

A number of groups are studying the manipulation of TGF- $\beta$  in the prevention of both normal scarring and hypertrophic scarring. TGF- $\beta$  neu-

## TGF- $\beta$ family in wound healing

**Table 1.** Potential mechanisms involved in the development of hypertrophic scarring. Summarised in Figure 2

Mechanism	Results	Reference
Apoptosis	<ul style="list-style-type: none"> <li>-p53 alterations in hypertrophic scar derived cells</li> <li>-Mechanical loading inhibits cellular apoptosis</li> <li>-Bcl-2 increased in peripheral blood mononuclear cell fractions from burns patients with hypertrophic scars compared to burns patients who had normal healing</li> <li>-failure to undergo apoptosis in hypertrophic scar fibroblasts due to over expression of tissue transglutaminase</li> <li>- myofibroblasts from hypertrophic scars fail to undergo apoptosis in response to apoptotic inducers</li> <li>-Decreased Fas expression in vivo and in vitro</li> </ul>	[44, 67-71]
Mechanobiology, tension	<ul style="list-style-type: none"> <li>-Cyclical stretching influences expression of genes, growth factors etc</li> <li>-Stretching causes number of myofibroblasts to increase</li> <li>-mechanical strain upregulates matrix remodelling genes but down regulates cellular apoptosis genes</li> </ul>	[72-74]
Angiogenesis and angiogenic factors	<ul style="list-style-type: none"> <li>-Decrease in angiopoietin1/angiopoietin 2 ratio in hypertrophic scars (following surgery), microvessel density higher in hypertrophic scars</li> </ul>	[75]
Inflammation	<ul style="list-style-type: none"> <li>- Increased toll-like receptor-4, prostaglandin E2, IL-6, IL-8 and MCP-1 in hypertrophic derived fibroblasts (following burn) compared to normal matched dermal fibroblasts</li> <li>-Increase in IL-6 in hypertrophic burn scar fibroblasts compared to normal control fibroblasts</li> <li>-Increased IL-15 in hypertrophic scars compared to normal scars and skin</li> <li>-HTS after a burn showed a polarized Th2 systemic response- increased T cells and Th2 fibrogenic cytokines</li> </ul>	[76-79]
MMP	<ul style="list-style-type: none"> <li>-Higher TIMP-1 and 2 mRNA in hypertrophic scars</li> <li>Higher MMP2 mRNA</li> <li>Higher TIMP-1 in sera than normal scar</li> <li>-Higher MMP2 in hypertrophic compared to normal</li> </ul>	[80,81]
Extra cellular matrix	<ul style="list-style-type: none"> <li>-Distribution and organization of fibrillin-1 and elastin different between normal skin and pathological scars (hypertrophic and keloids)</li> </ul>	[82]
Growth factors	<ul style="list-style-type: none"> <li>-Increased expression TGF-<math>\beta</math>1, <math>\beta</math>2, <math>\beta</math>3, bFGF and VEGF in keratinocytes from burn scars at 1 month compared to matched normal. TGF<math>\beta</math>3 elevated in hypertrophic scars than normal.</li> <li>-IGF-1 – increased number of cells producing IGF1 in hypertrophic scar tissue compared to normal skin samples</li> </ul>	[83-85]

tralisising antibodies have been shown to inhibit fibrosis in a number of animal models [97, 98], however, reduction in TGF- $\beta$  signalling has been linked with chronic or non-healing wounds [40, 41]. Blocking TGF- $\beta$  through a number of natural TGF- $\beta$  inhibitors, such as decorin, biglycan, LAP, may block the fibrotic TGF- $\beta$  response, but not affect the TGF- $\beta$  immune response [99]. Other methods of manipulating TGF- $\beta$  include blocking the TGF- $\beta$  receptors with kinase inhibitors (for example SD-208), a dominant negative TGF $\beta$ RII (TbetaRIIDeltacyt), which have been shown to prevent fibrosis in animal models by blocking profibrotic gene expression [100, 101]. Ahn and colleagues in 2010 showed that betaglycan (the TGF- $\beta$  III receptor) inhibited Smad

signalling and Akt and ERK phosphorylation [102], and *in vivo* models suggest that soluble betaglycan may prevent fibrosis in animal models [103]. Few clinical studies have been performed on the manipulation of TGF- $\beta$ 1 and its isoforms to prevent dermal scarring. Ferguson and colleagues published three double-blind placebo controlled studies (phase I/II) using the administration of TGF- $\beta$ 3 prophylactically [104], however the Phase 3 trial appears to have been unsuccessful [105].

### Problems

Though there has been a huge array of research conducted on hypertrophic scarring, however,

**Table 2.** Therapies and potential therapies for hypertrophic scarring

Mechanism	Drug	Reference
Apoptosis/ reducing proliferation	Intralesional corticosteroids, IFN- $\alpha$ 2b Combinations of treatment Compression Fibrostat	[87-92]
Extracellular matrix	Silicone gel (various mechanisms including remodelling of extra cellular matrix) AZX-100 (Capstone Therapeutics; peptide analogue of HSP 20) Minocycline	[93-95]
Inflammation	IFN- $\alpha$ 2b	[88]
Growth factors	Receptor tyrosine kinase inhibitors (SU9518, SU11657, Imatinib/Gleevec)	[96]

our understanding of its pathophysiology, and potential therapeutic agents remain unclear. There are problems with current research with as yet no definitive animal model available, though studies have described a number of animal models including the rabbit and the red Duroc pig [106, 107]. However it remains to be seen if these models are an adequate comparison compared to human scarring. Further, comparisons between research studies has been hampered due to a number of variations between studies for example the aetiology of the hypertrophic scarring (thermal injury v injury v surgery), maturity of the scar, any previous treatments, and the location of the hypertrophic scar which may all impact research findings. Research is almost always conducted after the fact; there have been few if any research conducted on the tissue prior or during the injury, and then following on after the injury. In human patients this may be difficult due to ethical permission, but this may indicate not only potentially susceptible individuals but also where wound healing goes awry.

### Conclusion

Hypertrophic scarring is a common abnormality of wound healing often associated with thermal injuries. To date, the mechanism behind this form of scarring remains unclear with much research focussing on the pro-fibrotic growth factor TGF- $\beta$ 1. Further, as yet there is no definitive treatment to reduce or prevent any scarring though the market place is large with many patients finding their scarring unacceptable.

**Address correspondence to:** Dr. K Rolfe, The Institute of Plastic Surgery Research and Education, Dept

Plastic Surgery, The Royal Free Hospital, Pond St Hampstead, London, UK, NW3 2QG E-mail: work@kerstinrolfe.com

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