

Complete Genome Sequences of *Mycoplasma leachii* Strain $PG50^{T}$ and the Pathogenic *Mycoplasma mycoides* subsp. *mycoides* Small Colony Biotype Strain Gladysdale

Kim S. Wise,^a Michael J. Calcutt,^b Mark F. Foecking,^a Ramana Madupu,^c Robert T. DeBoy,^c Kerstin Röske,^d Miranda L. Hvinden,^b Tara R. Martin,^b A. Scott Durkin,^c John I. Glass,^c and Barbara A. Methé^c

Department of Molecular Microbiology and Immunology, University of Missouri, Columbia, Missouri, USA^a; Department of Veterinary Pathobiology, University of Missouri, Columbia, Missouri, USA^b; J. Craig Venter Institute, Rockville, Maryland, USA^c; and Saxony Academy of Sciences Leipzig, Leipzig, Germany^d

Mycoplasma mycoides subsp. *mycoides* small colony biotype (SC) is the high-consequence animal pathogen causing contagious bovine pleuropneumonia. We report the complete genome sequences of the pathogenic strain *M. mycoides* subsp. *mycoides* SC Gladysdale and a close phylogenetic relative, *Mycoplasma leachii* PG50^T, another bovine pathogen of the *M. mycoides* phylogenetic clade.

Contagious bovine pleuropneumonia (CBPP) is among the most important blights of cattle in countries in which CBPP is endemic (8). Although the study of the etiologic agent, *Mycoplasma mycoides* subsp. *mycoides* small colony biotype (SC), has benefited greatly from the availability of the genome sequence of PG1^T (9), the avirulence of this reference isolate (2) limits understanding of pathogenic strains. Reported herein is the complete genome sequence of *M. mycoides* subsp. *mycoides* SC strain Gladysdale, a pathogenic isolate (6), which is also employed at the Foreign Animal Disease Center (PIADC) to experimentally demonstrate acute-disease pathology.

Genomic DNA from a clonal isolate (MU clone SC5) was prepared at PIADC and safety tested before transfer to the J. Craig Venter Institute (JCVI; Rockville, MD). The genome was sequenced to $8 \times$ coverage by the whole-genome shotgun (WGS) approach using paired-end Sanger sequencing. Scaffold-directed gap closure yielded a single complete sequence. This resulting assembly was autoannotated via the JCVI pipeline, with manual curation applied to specific genes encoding phase-variable surface lipoproteins or associated with genomic islands. The circular chromosome (1,193,808 bp) is slightly smaller than that of M. mycoides subsp. mycoides SCPG1^T (1,211,703 bp) due primarily to copy number variation of discrete gene blocks. No prophages or prototypic mycoplasmal integrative conjugative element (ICE) units occur, although a region with similarity to Mycoplasma capricolum Tra Island I (5) is present in both strains. Gene sets that endow cytotoxic H₂O₂ production, the principal virulence factor identified for M. mycoides subsp. mycoides SC (4), also occur in both strains. Accordingly, we surmise that differences in virulence may be enciphered not by macroscale insertion or deletion of genomic regions but rather within the multiple single nucleotide polymorphisms (SNPs) and indel differences dispersed throughout the chromosomes.

To enable comparative genomic analyses of bovine pathogens belonging to the *M. mycoides* phylogenetic clade (2), WGS Sanger sequencing (paired-end approach) and assembly were similarly completed for *Mycoplasma leachii* strain $PG50^{T}$ (MU clone A8). This species, formerly known as *Mycoplasma* sp. bovine group 7 (2), causes pneumonia, mastitis, polyarthritis, and abortion (7). Notable features of the 1,008,951-bp chromosome are the paucity

of insertion sequence (IS) incursions in comparison to the ISladen genomes of *M. mycoides* subsp. *mycoides* SC strains and the presence of an integrative element that encodes multiple palindromic amphipathic repeat coding element (PARCEL) domain proteins (5). In addition, a novel portfolio of phase-variable lipoprotein genes, predicting combinatorial expression governed by indel mutations in poly(TA) tract-containing promoters, verifies the widespread presence of this stochastic mechanism of surface diversification among pathogens of the *M. mycoides* clade (3, 10) and further supports its likely role in host niche adaptation, immune avoidance, or disease transmission.

With genome sequences determined for all taxa within the *M. mycoides* clade (2), a framework is established to expound the pangenome of this group and to identify possible gene patterns that correlate with ruminant host specificity or disease severity. Furthermore, as members of this group are model organisms for genome synthesis and transplantation (1), these data should be a valuable adjunct in designing novel genomes through synthetic biology.

Nucleotide sequence accession numbers. The complete genome sequences are available in GenBank under accession numbers CP002107.1 (*M. mycoides* subsp. *mycoides* SC Gladysdale) and CP002108.1 (*M. leachii* PG50^T).

ACKNOWLEDGMENTS

This research was supported by the USDA-ARS Program for Prevention of Animal Infectious Diseases (1940-32000-039-08S) at the University of Missouri and the Office of Science (BER), U.S. Department of Energy, grant no. DE-FC02-02ER63453 to the J. Craig Venter Institute.

We thank Timothy S. Gorton and William R. White for facilitating the preparation and handling of genomic DNA at the PIADC.

REFERENCES

1. Gibson DG, et al. 2010. Creation of a bacterial cell controlled by a chemically synthesized genome. Science 329:52–56.

Received 3 May 2012 Accepted 4 June 2012 Address correspondence to Kim S. Wise, wisek@missouri.edu. Copyright © 2012, American Society for Microbiology. All Rights Reserved. doi:10.1128/JB.00761-12

- Manso-Silván L, et al. 2009. Mycoplasma leachii sp. nov. as a new species designation for Mycoplasma sp. bovine group 7 of Leach, and reclassification of Mycoplasma mycoides subsp. mycoides LC as a serovar of Mycoplasma mycoides subsp. capri. Int. J. Syst. Evol. Microbiol. 59:1353–1358.
- Persson A, Jacobsson K, Frykberg L, Johansson K-E, Poumarat F. 2002. Variable surface protein Vmm of *Mycoplasma mycoides* subsp. *mycoides* small colony type. J. Bacteriol. 184:3712–3722.
- Pilo P, et al. 2005. A metabolic enzyme as a primary virulence factor of Mycoplasma mycoides subsp. mycoides small colony. J. Bacteriol. 187:6824–6831.
- Röske K, et al. 2010. A versatile palindromic amphipathic repeat coding sequence horizontally distributed among diverse bacterial and eucaryotic microbes. BMC Genomics 11:430.
- Rurangirwa FR, Masiga WN, Mtui B. 1976. An anamnestic response to challenge with virulent *Mycoplasma mycoides* subsp. *mycoides* of cattle immune to contagious bovine pleuropneumonia. J. Comp. Pathol. 86: 381–386.
- Tardy F, Maigre L, Poumarat F, Citti C. 2009. Identification and distribution of genetic markers in three closely related taxa of the *Mycoplasma mycoides* cluster: refining the relative position and frontiers of the *Mycoplasma* sp. bovine group 7 taxon (*Mycoplasma leachii*). Microbiology 155: 3775–3787.
- 8. Thiaucourt F, et al. 2011. Mycoplasma mycoides, from "mycoides Small Colony" to "capri." A microevolutionary perspective. BMC Genomics 12:114.
- Westberg J, et al. 2004. The genome sequence of *Mycoplasma mycoides* subsp. *mycoides* SC type strain PG1^T, the causative agent of contagious bovine pleuropneumonia (CBPP). Genome Res. 14:221–227.
- 10. Wise KS, et al. 2006. Distinctive repertoire of contingency genes conferring mutation-based phase variation and combinatorial expression of surface lipoproteins in *Mycoplasma capricolum* subsp. *capricolum* of the *Mycoplasma mycoides* phylogenetic cluster. J. Bacteriol. 183:4926-4941.